

1 **An Optical Scattering Based Cost-Effective Approach Towards**
2 **Quantitative Assessment Of Turbidity And Particle Size Estimation In Drinking Water**
3 **Using Image Analysis**
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ABSTRACT

Contaminated water consumption primarily for drinking purposes is the cause of approximately 502,000 global deaths every year mostly in economically challenging countries indicating the need for a cheap, easy to use a yet robust and scientifically proven method for determination of water quality. In this work, we have characterized the water quality utilizing the principles of optical scattering by the suspended particulate matter using a low-cost wireless-enabled camera. The images grabbed by the camera on an optically lit cast screen on a red and a blue dot were allowed to arrive through a “model scattering medium”. An estimate of the amount of light reaching the detector camera essentially provide Optical Density of the medium. Edge blurring of the captured images reveals information of the suspended particulates (sizes) in the medium. The individual pixel information was analyzed and the 'edge blurring' phenomenon was shown on an RGB intensity curve. The average diameter of the dominant suspended particles presents in the model scattering medium is also estimated from the fitting parameters and compared with that from commercially available Dynamic Light Scattering (DLS) instrument. The system is effective in measuring bacterial growth and the acquired data have been compared with that of the growth curve obtained from the gold standard method. Limit of Detection (LOD) of the set-up was found to be 48 ppm. The extremely cost-effective nature of the set-up, the innovative method of analysis, and easy availability of components would expectedly make water quality assessment very easy and user friendly.

1. INTRODUCTION

Increasing environmental pollution is a matter of grave concern in modern society(Wang, Webber, Finlayson, & Barnett, 2008). Pollution extends from air, sound, and water (Burningham & Thrush, 2004; Gorman, 2001). Among these, water pollution has shown a significant increase with the growing population index particularly in Low and Middle-Income Countries (LMIC) (Suk et al., 2016; Thomas, Wickramasinghe, Mendis, Roberts, & Foster, 2015). A worldwide minimum of 2 billion people consumes water for drinking, contaminated with fecal matter (Kimani-Murage & Ngindu, 2007). Contaminated water is the root cause of deadly diseases such as diarrhea, cholera, dysentery as well as typhoid, and its consumption results in 502,000 diarrheal deaths annually (Dwight, Fernandez, Baker, Semenza, & Olson,

76 2005; Kimani-Murage & Ngindu, 2007). These data indicate the urgent need for quantitative
77 assessment of water quality including lakes(Li et al., 2007) and bigger water bodies with online
78 determination of results indicating the readiness of consumption of available drinking water.
79 Water quality is determined by its chemical, physical, and biological content (Lawson, 2011;
80 Ramalho, Cunha, Teixeira, & Gibbs, 2001; Sadeghi, Mohammadian, Nourani, Peyda, &
81 Eslami, 2007). The assessment is mainly a manual process and conventionally it is done by the
82 collection of water samples and using chemical and other methods of analysis (Farré & Barceló,
83 2003). The processes are complex, suitable only for a trained person (Tebbutt, 1997).
84 Moreover, they are time-consuming and offline (Lenat, 1988). With the advent and
85 development of sensor-based systems, substantial research has been carried out to automate
86 and real-time monitor the water quality and Internet of Things (IoT) enable devices are in
87 demand for immediate intimation of human action needed anywhere (O'Flynn et al., 2007).
88 Such sensor-based systems mainly focus on the total dissolved solvents (TDS) and pH
89 properties of water and few such sensors have been made commercially available also. While
90 online sensors ensure immediate data availability and trigger the need for urgent action, their
91 calibration, reliability, and water-induced stains become an important concern (Lambrou,
92 Anastasiou, Panayiotou, & Polycarpou, 2014). Some alternate experimental methods were also
93 tried by researchers like using the bioscreen microbiological growth analyser (Johnston, 1998)
94 and underwater imaging systems(Ouyang et al., 2013; Selmo et al., 2017).

95 Various methods of probing water quality have been tried and researched by various
96 scientists (Association & Association, 1989). The contemporary research in this direction
97 includes the measurement of ocean watercolor and estimation of its effect on marine biology
98 (Barale, 1991; Shujing & Zhihua, 2001). RGB analysis has been used to determine the salinity
99 index of water by using the ration of R to B and B to G was used to determine the chlorophyll
100 content of water (Goddijn & White, 2006). Airborne digital image photography has also been

101 used to map water pollution and overcome the problem of cloud cover scenes (Kallio et al.,
102 2001). Recently, computer vision and artificial intelligence have witnessed their application in
103 the measurement of water turbidity and related parameters (Zion, 2012). Various methods of
104 estimation of coliforms in drinking water have been tried as a measure to estimate water
105 quality (Rompré, Servais, Baudart, De-Roubin, & Laurent, 2002). In our present study, we
106 have used image analysis of a Red and a Blue dots on an optically lit cast screen across a model
107 turbid medium to estimate the optical density (turbidity) and computational analysis of the
108 captured image-edge blurring phenomena to conclude on the diameter of dominant suspended
109 particulate matter in the turbid colloidal solution. We have also explored the possibility of using
110 a submersible camera to acquire data for long term data acquisition of a natural water body.
111 Data acquired remotely has been analyzed in our indigenously developed software for online
112 monitoring. The proposed set-up finally produces real-time data of particle size estimation and
113 fitness of consumption of contaminated water samples with sub-micron suspended particulate
114 matter, which are difficult to assess via visual inspection. The developed set-up efficiently
115 estimates the presence of suspended particulate matter including micro-organism to a level of
116 48 ppm (and hence defines the LOD of the system) which is well below the WHO level of 300
117 ppm in drinking water (Organization, 2003). Water samples with coarser particles will be
118 easier to identify and screen for consumption visually have not been included in this study.

119 **2. MATERIALS AND METHODS**

120 **2.1. Cast Screen**

121 An optically backlit cast screen was used to draw the Red and Blue dots. The 220 VAC LED
122 lamp was purchased from Philips and was fitted with an optical diffuser to block the direct
123 beam saturating the receiver (camera). The dots of 1 cm diameter were printed from a calibrated
124 true color printer (HP 2280). The wavelengths corresponding to individual colors were
125 determined from the reverse calculation of RGB value with wavelength correlation. The

126 submersible camera (QAWACHH) was purchased online. “Y-camera” app was used to acquire
127 live images on a laptop or smartphone. Quartz cuvette was held in between as a sample holder
128 ensuring a clear field of view of the cast screen through the sample.

129 **2.2. Preparation of Samples**

130 The same quality milk samples were prepared at various concentrations starting from 1 μ L to
131 40 μ L in 1 mL of whole raw milk. The purchased sample was maintained at the highest purity
132 level to the best of knowledge. The standard pipetting apparatus (Accupipet) was used to
133 extract the exact amount of solution under test.

134 **2.3. Camera Characterization**

135 Computer vision mainly suffers from the problem of auto-brightness and auto saturation of
136 pixels (Hu, Gallo, Pulli, & Sun, 2013). This leads to unequal referencing of data under various
137 ambient light conditions. However, the choice of camera was made to be able to manually
138 adjust the focus and exposure. Moreover, important considerations were taken to ensure
139 submerged condition water protection for electronics and camera optics. To tackle this
140 problem, a mobile endoscope camera enabled with in-built wireless LAN was used to capture
141 images and transfer to a distant computer or mobile in real-time.

142 **2.4. Development of low-cost instrument and Its Working Principle**

143 The working principle of this device is primarily based on the scattering and absorption of light
144 by suspended particulate matters in a colloidal solution. Fig. 1 shows the schematic diagram of
145 the experimental arrangement. A backlit screen (diffusor) has been used as a cast screen. Two
146 colors Red and Blue have been utilized as a marker of distinctly apart wavelength with no
147 overlap of the spectrum. Light from the screen travels to the wireless camera after interacting
148 with the sample in the cuvette. The camera has been strategically placed keeping in mind the
149 view angle to cover both dots which are kept ensuring equal illumination behind both. The
150 camera can be kept submerged under real-life situations and is enabled with wireless LAN to

151 ensure remote monitoring of the sample. Light traveling from the screen will suffer absorption
152 by the sample guided by Beer-Lambert's law. However, light traveling at the edge of the sample
153 will suffer multiple reflections and will result in blurring of the edges as depicted conceptually
154 in Fig. 2(a). The indigenously developed machine-computer interface will acquire live images
155 and will do a pixel analysis of the entire image frame to quickly calculate the amount of light
156 absorbed as well as particle size estimation of the dominant component. Fig. 2(b) shows the
157 pattern of fade experienced by the edge of the image as one moves away from the center of the
158 circle.

159 **2.5. Interfacing Software Design**

160 A LabVIEW based program is designed to acquire and process data from the instrument via a
161 USB port. A Microsoft-Windows based on-board computer is used to run the developed
162 software in real-time to acquire data. The interface is made to be simple and intuitive, thus, no
163 requirement of any additional training to operate the software. The software identifies the
164 attached camera and grabs video frames. It then does frame by frame pixel RGB analysis and
165 extracts the Red and Blue information from the similar color dots respectively. The software
166 also plots the value in real-time and tries to identify the edge and performs an online fitting of
167 a sigmoidal function. The derived values are a marker of various parameters of the suspended
168 particulate matter.

169 **2.6. Bacterial Growth curve experiment**

170 The bactericidal activity is performed using MRSA (*methicillin-resistant staphylococcus*
171 *aureus*) bacteria cells. The cells are cultured in a Luria Broth (LB) medium under an incubator
172 shaker at 37 °C for 24 h. The optical density of freshly grown overnight culture is fixed to 0.1
173 in LB medium initially. The culture is then put in a cuvette and incubated at 37°C with shaking
174 for 9 h. The absorbance is taken at every hour interval and plotted against time with baseline
175 correction. The minimum detectable concentration of MRSA was determined using the onset

176 of the growth curve. To estimate the limit of detection (LOD) of the suspended micro-organism
177 (MRSA), we have converted the concentration of the micro-organism in the media from
178 CFU/ml to ppm unit.

179 **2.7. Crystal Violet (CV) Staining Assay**

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181 The freshly diluted culture of MRSA is spread over a biofilm and kept in an incubator at 37°C
182 for 24 h. Then, 1% of CV solution is spread over biofilm and incubated for 3 h. After washing
183 with water, the biofilm is exposed under a microscope (Leica digital inverted microscopes
184 DMI8).

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3. RESULT AND DISCUSSION

186 The acquired video is analyzed frame by frame. Individual frames were performed a raster scan
187 for pixel RGB information. Fig. 3(a) shows the intensity plot of Red value (from RGB analysis)
188 obtained from Red-colored dot. Similar results were obtained from the blue dot after extraction
189 of blue value (from the RGB analysis) as evident from Fig. 3(b). The curves were fitted with a
190 sigmoidal function. The fitting parameters obtained were found to be markers of absorption
191 and scattering parameters of the sample under test. The fitted equation was,

$$y = A_2 + \frac{(A_1 - A_2)}{\left(1 + e^{((x-x_0)/dx)}\right)} \quad (i)$$

192 The value of X_0 obtained from the fitting function from individual curves is plotted against
193 concentration for light red n blue dots as shown in Fig. 4(a & b) respectively. The parameter is
194 found to be indicative of the broadening of the edges due to Rayleigh scattering of optical
195 signals by the suspended particulate matter. The pixel profile plot is expected to show a shift
196 of intensities towards the negative X-axis to represent the broadening to the colored dot on the
197 left edge. This was confirmed by plotting X_0 profile with concentration as shown in Fig. 3 (a &
198 b). The curve clearly shows the linearly decreasing profile with increasing concentration
199 suggesting a significant broadening of edges which is a signature of number and size

200 distribution of colloidal substance present in an optically turbid solution. Hence, the broadening
 201 of edges becomes the signature of the number of scattering materials present in the colloidal
 202 suspension.

203 The diameter of particles in colloidal suspension can be estimated from the following well
 204 known Rayleigh scattering equation (ii). The intensity I of light scattered by anyone of the
 205 small spheres of diameter d and refractive index n from a beam of unpolarized light of
 206 wavelength λ and intensity I_0 is given by

$$I = I_0 \frac{1 + \cos^2 \theta}{2R^2} \left(\frac{2\pi}{\lambda}\right)^4 \left(\frac{n^2 - 1}{n^2 + 1}\right)^2 \left(\frac{d}{2}\right)^6 \quad (\text{ii})$$

207 where R is the distance to the particle and θ is the scattering angle.

208 Our experimental set-up dictates the use of two distinct wavelengths which was derived by
 209 conversion of RGB parameters to respective colors and further to specific wavelengths. The
 210 derived wavelengths were found to be $\lambda = 700$ nm for red color and $\lambda=450$ nm for blue color.

211 The above equation (ii) can be re-written as the following:

$$d^6 = \frac{1}{(1 + \cos^2 \theta) \left(\frac{m^2 - 1}{m^2 + 2}\right)^2} \frac{8 R^2 \lambda^4 I}{\pi^4 I_0} \quad (\text{iii})$$

212 Therefore,

$$d^6 = K \frac{I}{I_0} \quad (\text{iv})$$

213 Where K is the constant and is governed by the equation

$$K = \frac{1}{(1 + \cos^2 \theta) \left(\frac{m^2 - 1}{m^2 + 2}\right)^2} \frac{8 R^2 \lambda^4}{\pi^4} \quad (\text{v})$$

214 After calculation using the above-mentioned values of parameters we get,

215 $K=2.589 \times 10^{-30}$ for blue (considering $\lambda_{\text{Blue}}= 450\text{nm}$, $R=4\text{mm}$ and $(1+\cos^2\theta)=1.99952$) and

216 $K = 1.516 \times 10^{-29}$ for red (considering $\lambda_{\text{Red}}= 700\text{nm}$, $R=4\text{mm}$ and $(1+\cos^2\theta)=1.99956$)

$$6 \log d = \log k + \log \frac{I}{I_0} \quad (\text{vi})$$

$$6 \log d = \log k - \log \frac{I_0}{I} \quad (\text{vii})$$

217 The term $\log \frac{I_0}{I}$ is the signature of Beer Lambert's law which is synonymous to the parameter
218 A_2-A_1 of our fitting function. From equation (vii) we have calculated out for 450 nm
219 wavelength as 252 nm and for 700 nm wavelength to be 730 nm which is in very close
220 approximation with the standard DLS data as shown in Fig. 5. indicating the variation of the
221 diameter size of the dominant scatterer present in the sample with increasing concentration. It
222 was found that the diameter estimated using our set-up was in close agreement with the results
223 from the DLS using the gold standard instrument.

224 The fitting parameter (A_2-A_1) represents the extinction coefficient of the light. The amount of
225 light traveling from the screen to the camera suffers absorption and scattering from the sample
226 media. The difference of pixel information from normal spots to colored dotted spots represents
227 the amount of light lost during forwarding travel towards the camera. The two dots represent
228 two dominant wavelengths and carry spectroscopic information relating to the colloidal
229 sample. (A_2-A_1) is an indicative parameter towards the Optical Density of the sample governed
230 by Beer Lambert's law as shown in Fig. 6 (a & b). The choice of milk as a simulation of turbid
231 water samples was found to be appropriate as we found a substantial similarity between particle
232 size estimation from DLS instrument using the refractive index of various particles found in
233 real-world water samples e.g. silica with the same. Fig. 7 shows the comparison of number
234 concentration data obtained via DLS instrument using the refractive index of silica and milk
235 respectively. Using our set-up, the effective diameters were found to be 6.8 μm using blue dot
236 whereas using the red dot we arrived at the diameter of 10.9. Once again better approximation
237 was observed using blue dot depicting the effectiveness of the set-up in a lower range of visible
238 wavelengths.

239 **Fig. 8** attributes the standardized growth curve of MRSA. Herein, after the short (of 1h) lag
240 phase, optical density is exponentially increased up to 8 h (log phase). The measurement of
241 population growth is also manifested by image processing of red and blue dots. It is found that
242 the growth curve exhibited by the blue dot showed much higher sensitivity and quick response
243 to the growth of MRSA. However, it also exhibits quick saturation commensurate with a
244 standardized growth curve. In contrast, the response obtained from the Red curve is found to
245 be less responsive compared to the standard growth curve but showed no signs of saturation
246 with increasing time. It can be concluded that from this strategy, we can get two sensing curves,
247 one will better sensitivity as well as a response but the lower dynamic range and the other with
248 a lesser response but with a wider dynamic range. The bacterial growth curve was acquired
249 using blue and red dot images. The minimum detectable concentration from the blue curve was
250 found to be 48 ppm and the corresponding red curve was found to be 448 ppm. The LOD for
251 the gold standard method was found to be 52 ppm.

252 The average hydrodynamic diameter of MRSA is found to be 1 μ m from DLS as evident from
253 Fig 9 (a). As shown in Fig 9 (b) the microscopic image of MRSA designates the cell diameter
254 is approximately 0.8 μ m which is in good agreement to our findings 5.6 μ m from our developed
255 technique. It is to be noted the values are sometimes over-estimated due to the accumulation
256 of multiple molecules as shown in Fig 9 (b).

257 **4. CONCLUSIONS**

258 In this work, we have presented a simple, innovative, and cost-effective technique involving
259 the analysis of a video captured from a camera and estimate the amount of turbidity and also
260 suspended particle size. The data have been analyzed using an indigenously developed software
261 for online analysis of the model turbid medium. The set-up is found to be effective in
262 calculating the above-mentioned parameters quickly and with a fair amount of accuracy. We
263 also have investigated the possibility of assessment of bacterial presence in water with a fair

264 amount of accuracy. We hope the developed strategy with quick, easy, and precise
265 determination of water quality with reasonably low LOD of suspended particulate matter (48
266 ppm) would offer an affordable alternative in a low resource setting for developing countries.
267 The technology can be further applied to assess air quality and visibility assessment in a foggy
268 atmosphere. However, more experimentation is required before the same can be established.

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273 **6. CONFLICT OF INTEREST**

274 The authors declare no conflict of interest.

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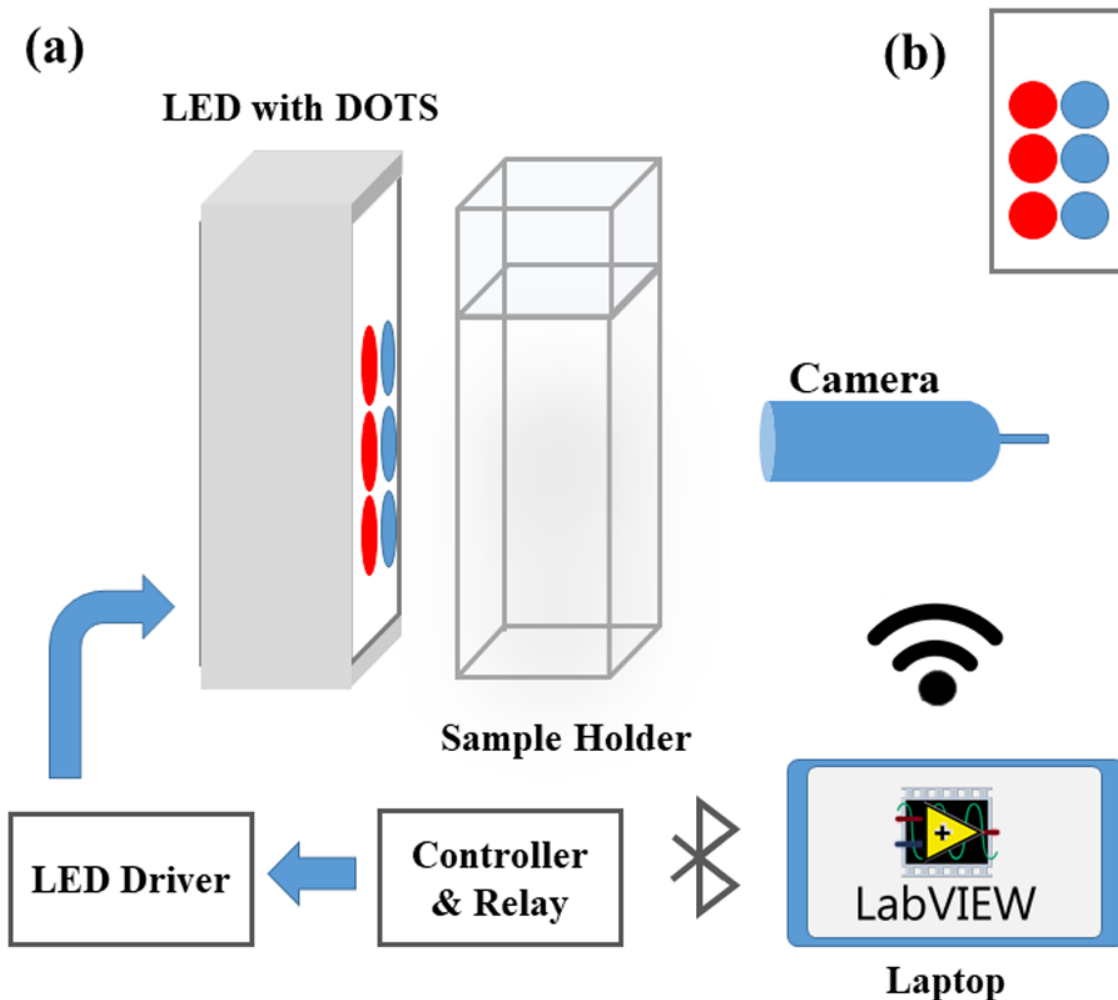
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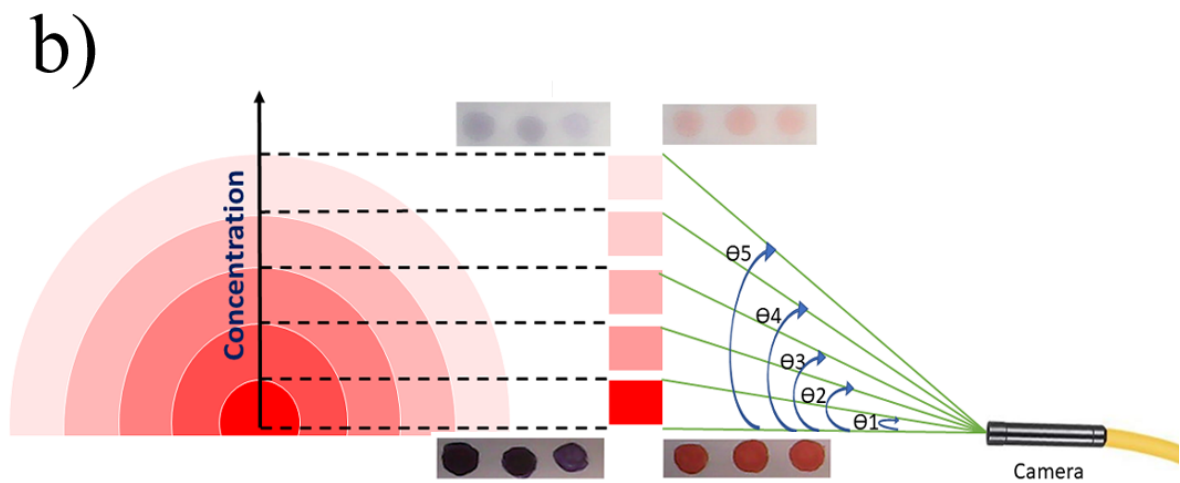
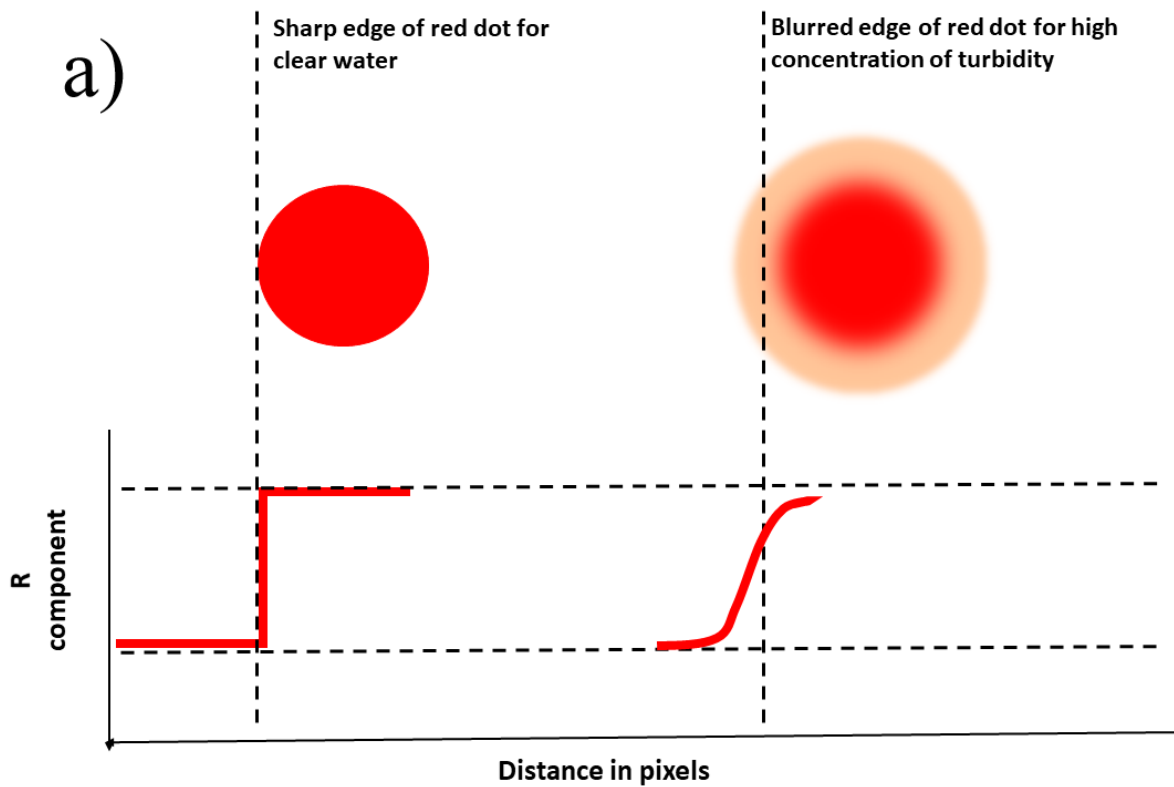
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FIGURES AND TABLES

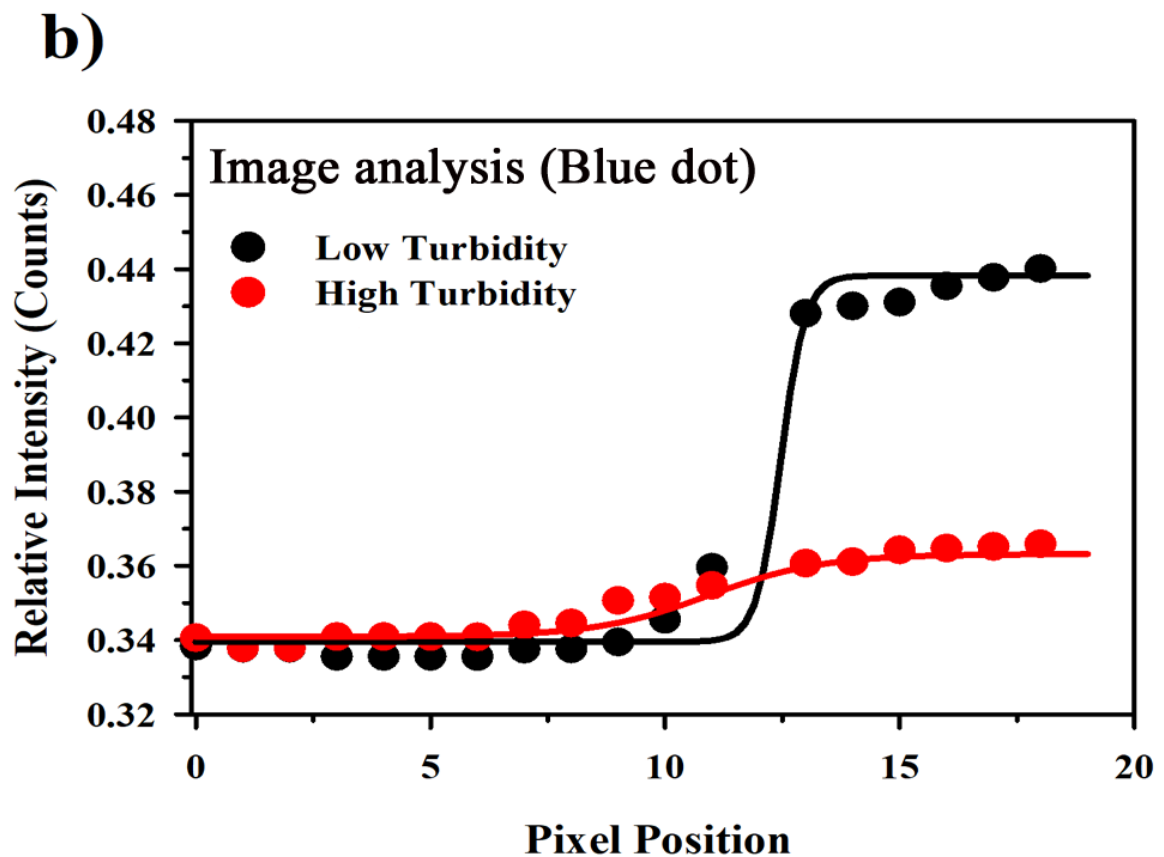
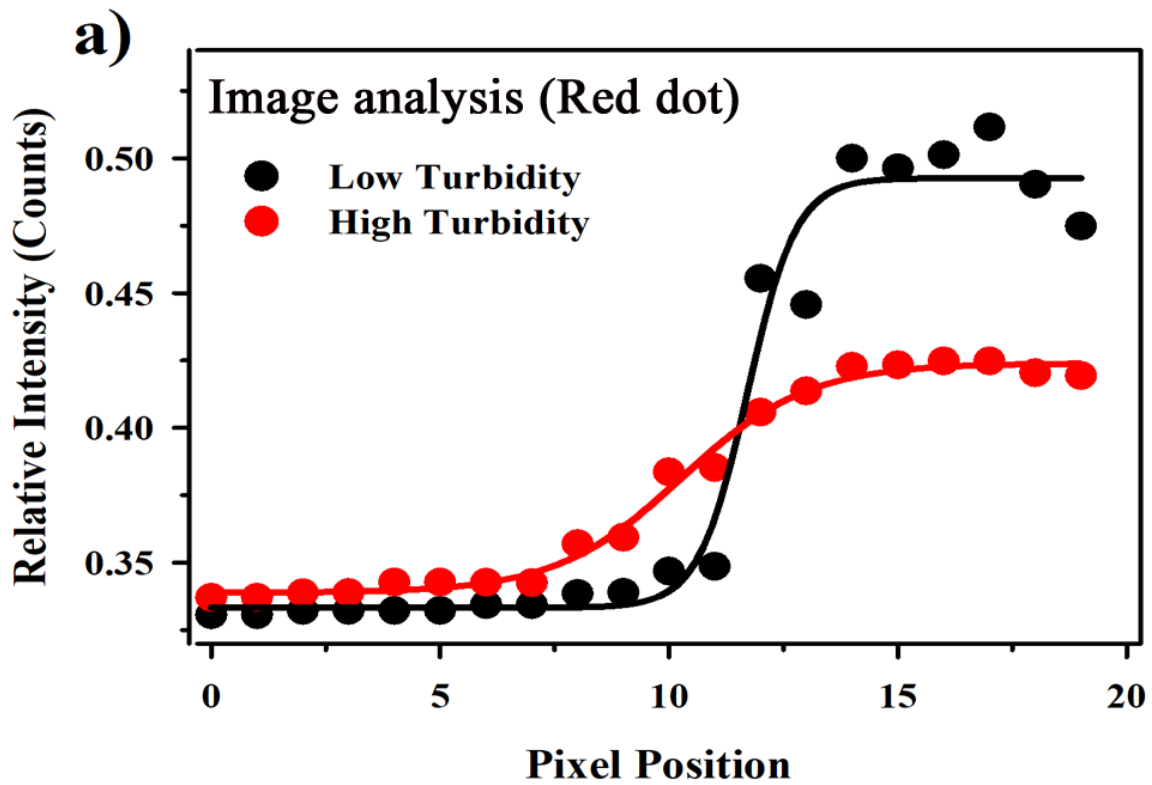


358
359 *Figure. 1:* (a) Schematic of the experimental arrangement for measuring optical signals from
360 backlit cast screen (b) The dots of particular color representing the various segments of visible
361 spectra
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Figure. 2: a) Schematic of the image analysis method. Clearwater image of a dot as seen by a camera. The pixel intensity plot relative to the pixel position gives a sharp rising edge of a pulse function. Pixel intensity plot of turbid water and schematic of turbid water image b) Schematic of the relationship of blurring the edge of acquired images in turbid water



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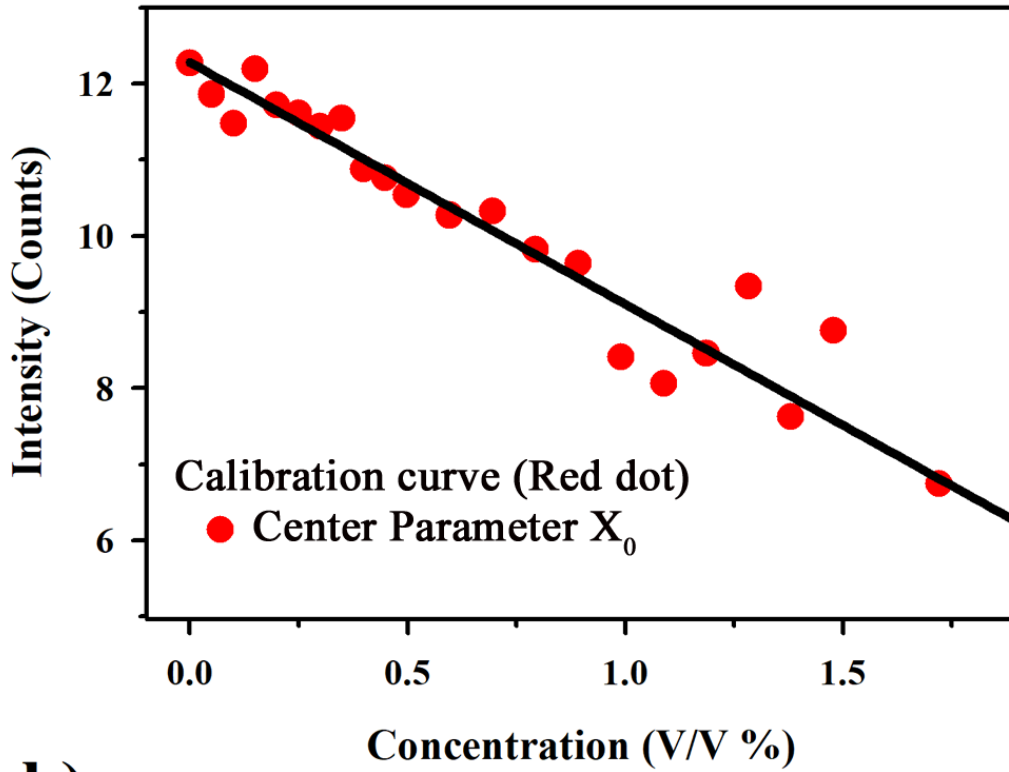
369 Figure. 3: a) Pixel intensity plot of acquired images of the Red dot in clear and turbid water and their respective Boltzman fitting (b) Pixel intensity plot of acquired images of the Blue dot

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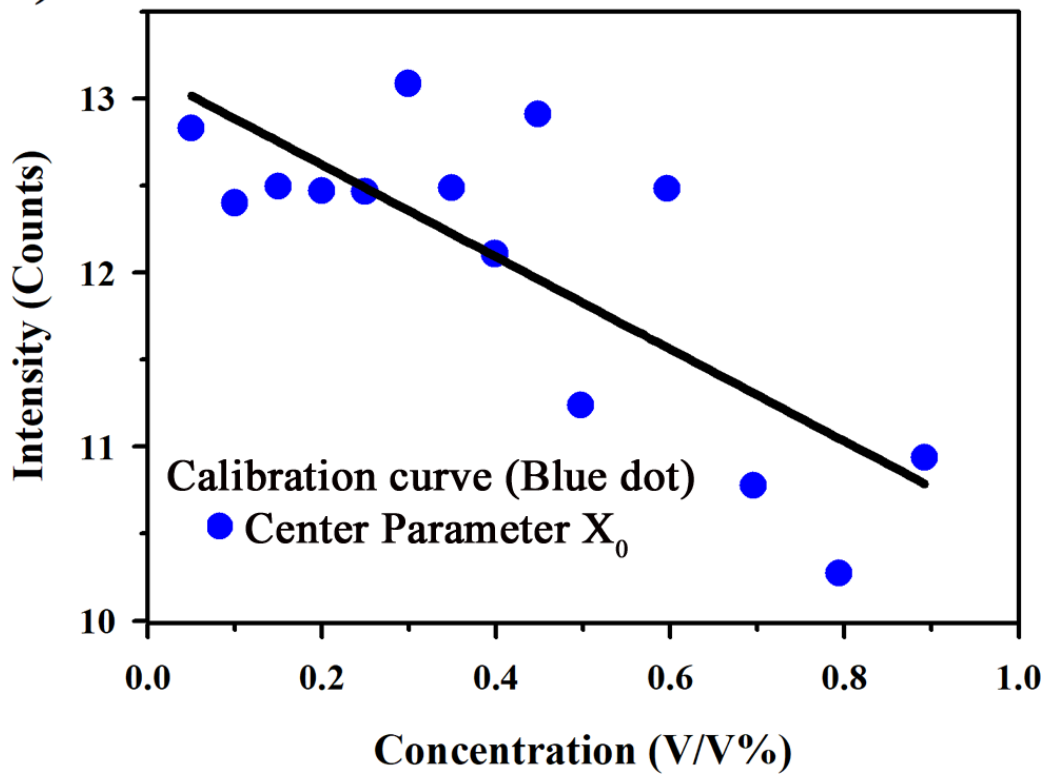
in clear and turbid water and their respective Boltzman fittings (solid lines)

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a)



b)

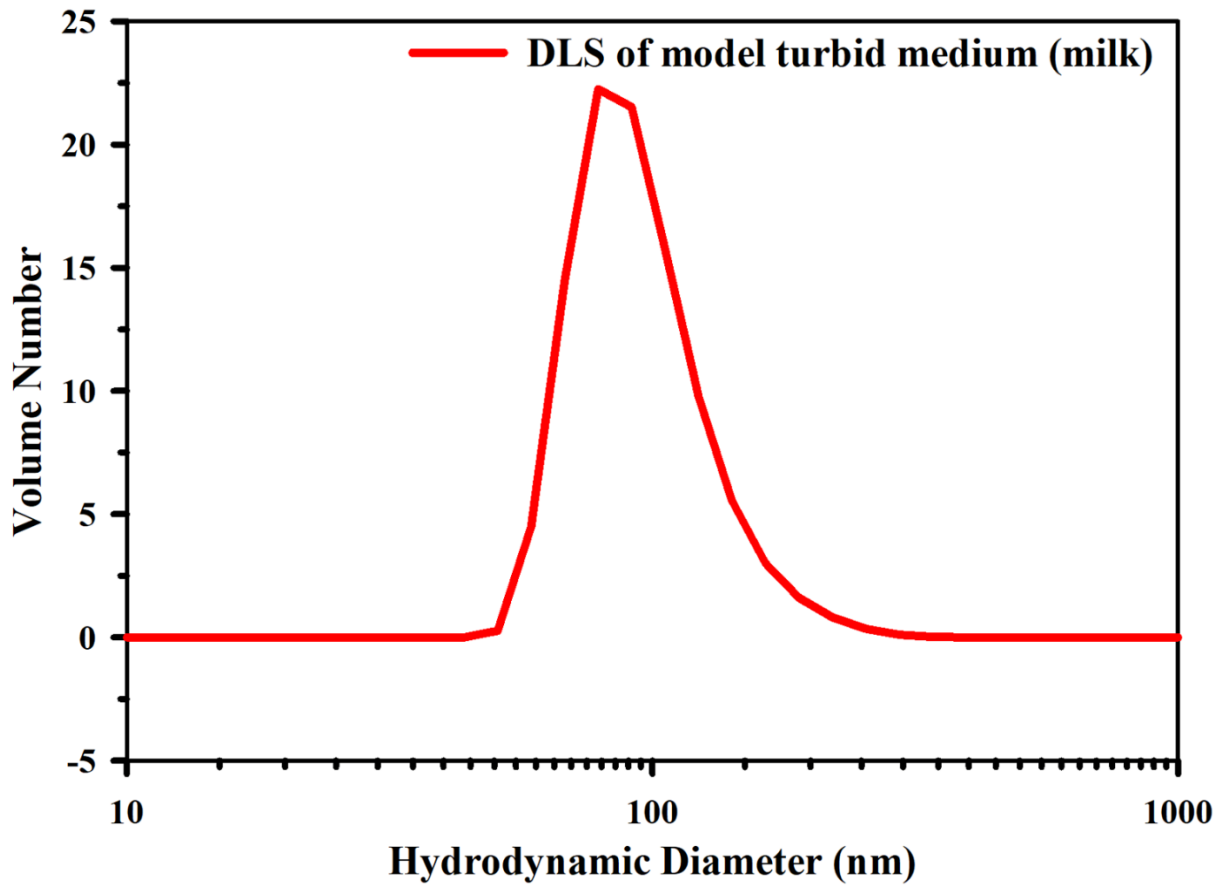


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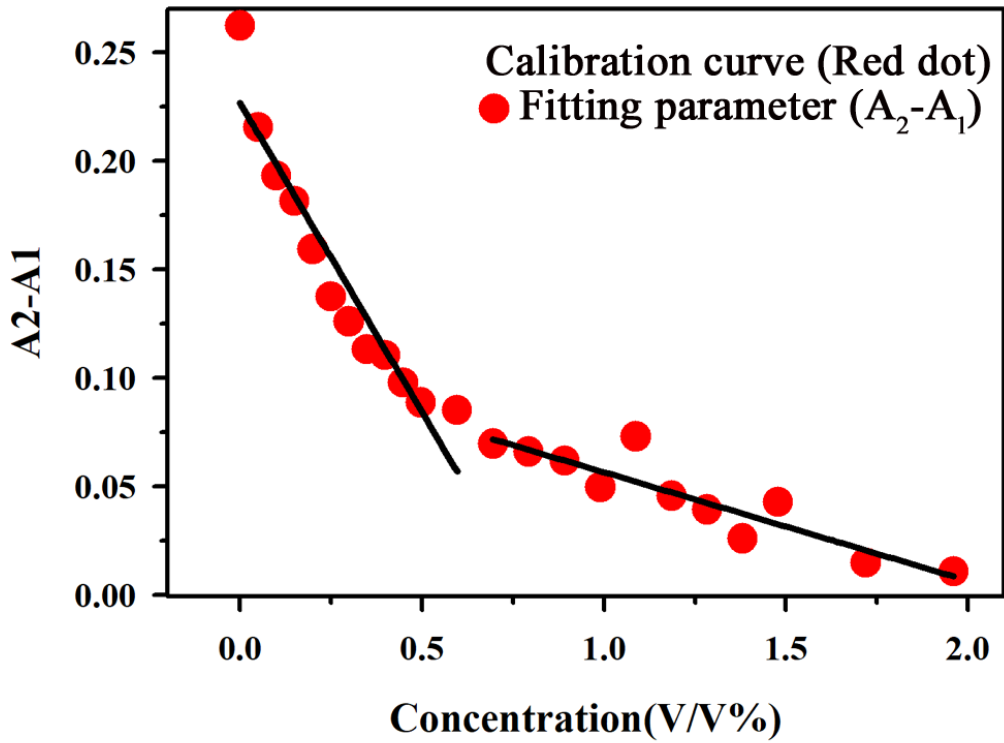
Figure 4: a) Plot of fitted center parameter X_0 with concentration for red dot b) Plot of fitted center parameter X_0 with concentration for the blue dot



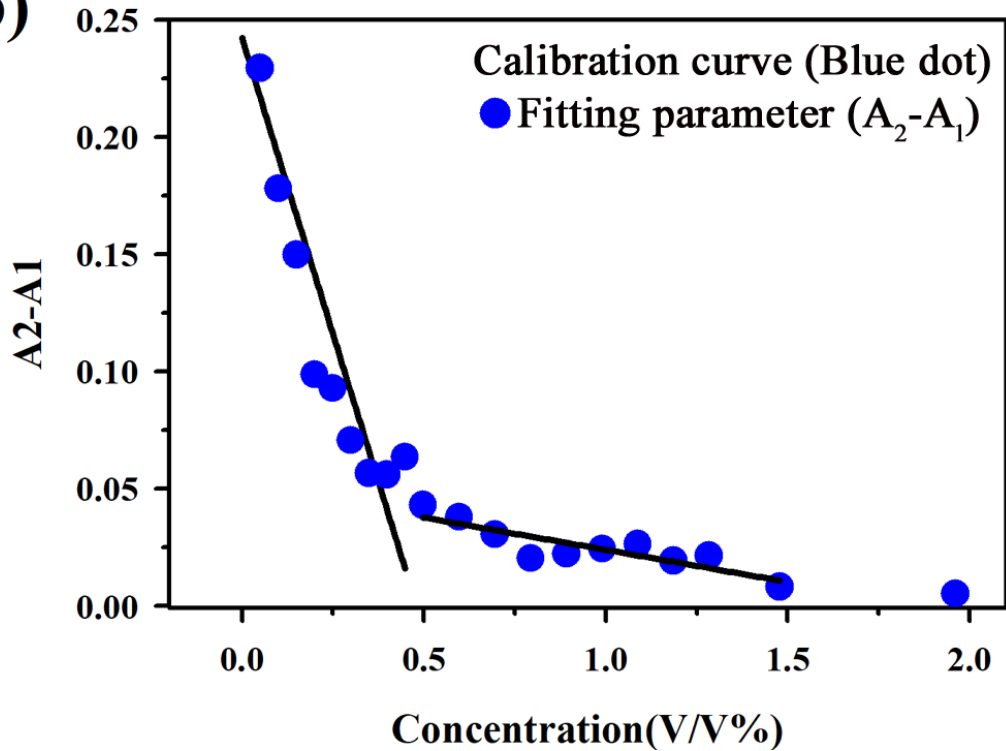
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Figure. 5: DLS data of the fitting function of a sample colloidal solution (Milk)

a)

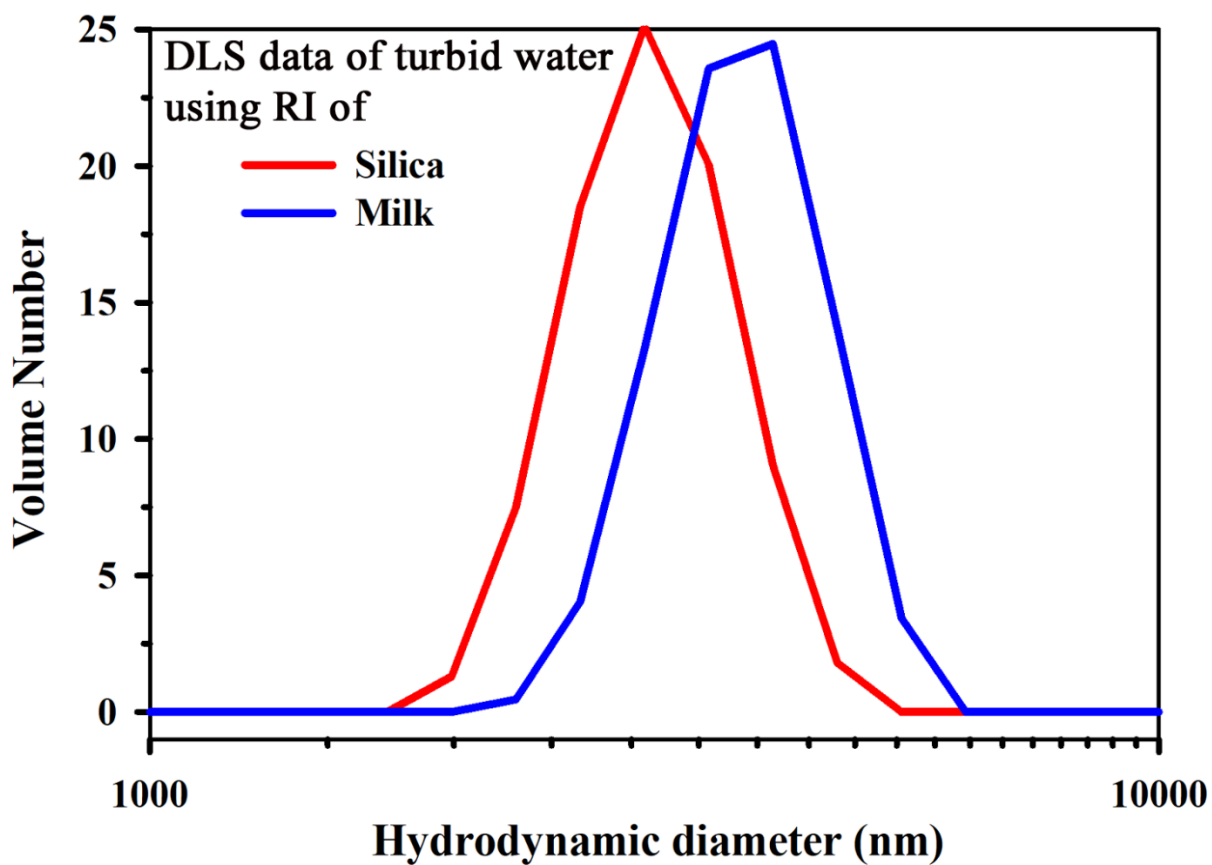


b)



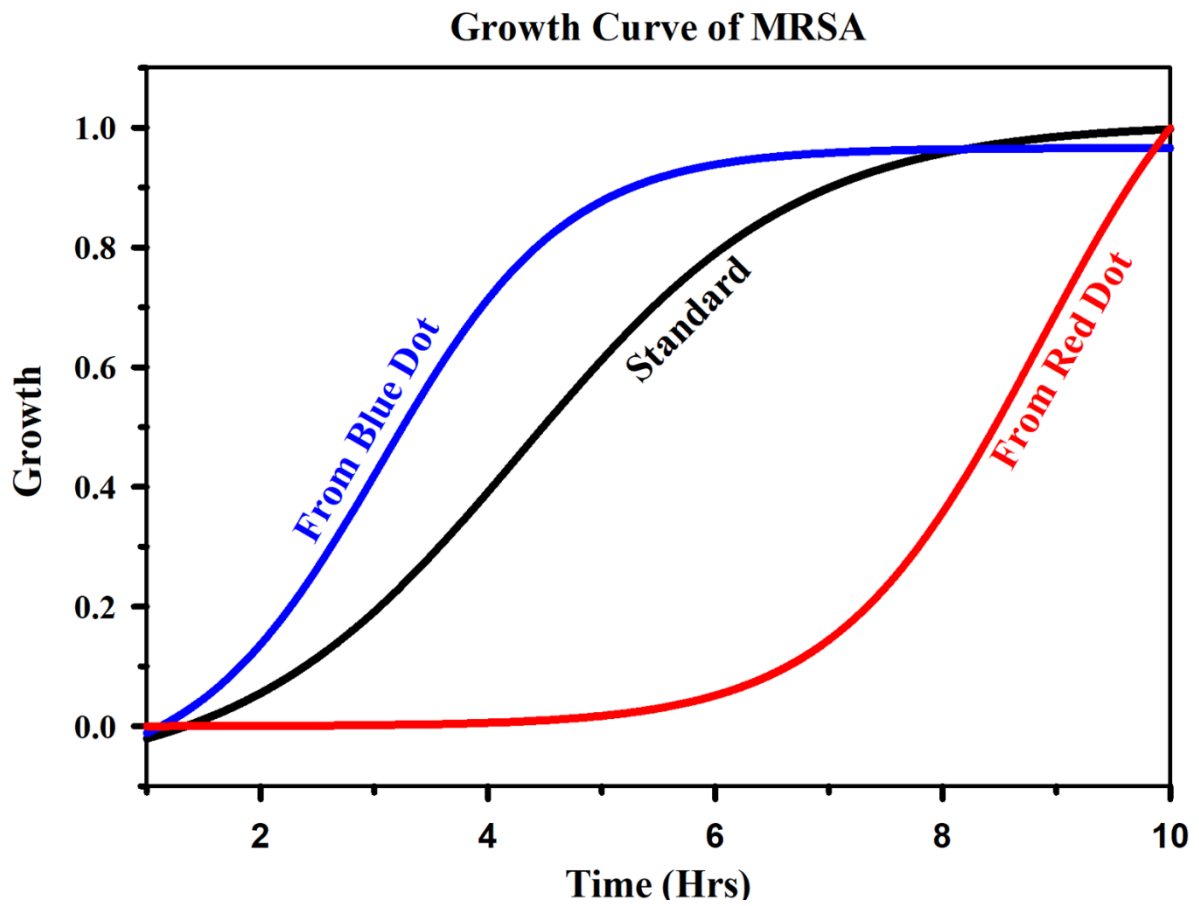
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378 Figure. 6.: a): Plot of the fitted parameter indicating a difference of initial and final amplitudes
379 (A_2-A_1) and its dual linear fitting function for red dot (b) Plot of the fitted parameter indicating
380 a difference of initial and final amplitudes (A_2-A_1) and its dual linear fitting function for the
381 blue dot.



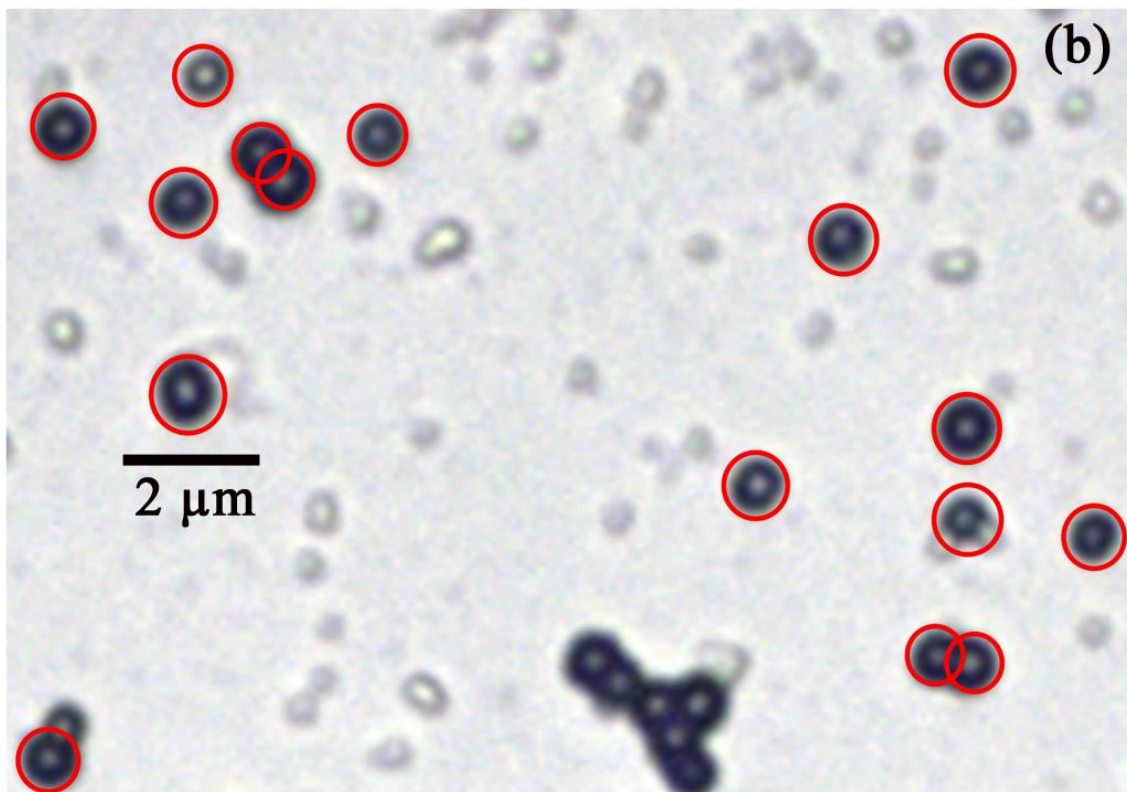
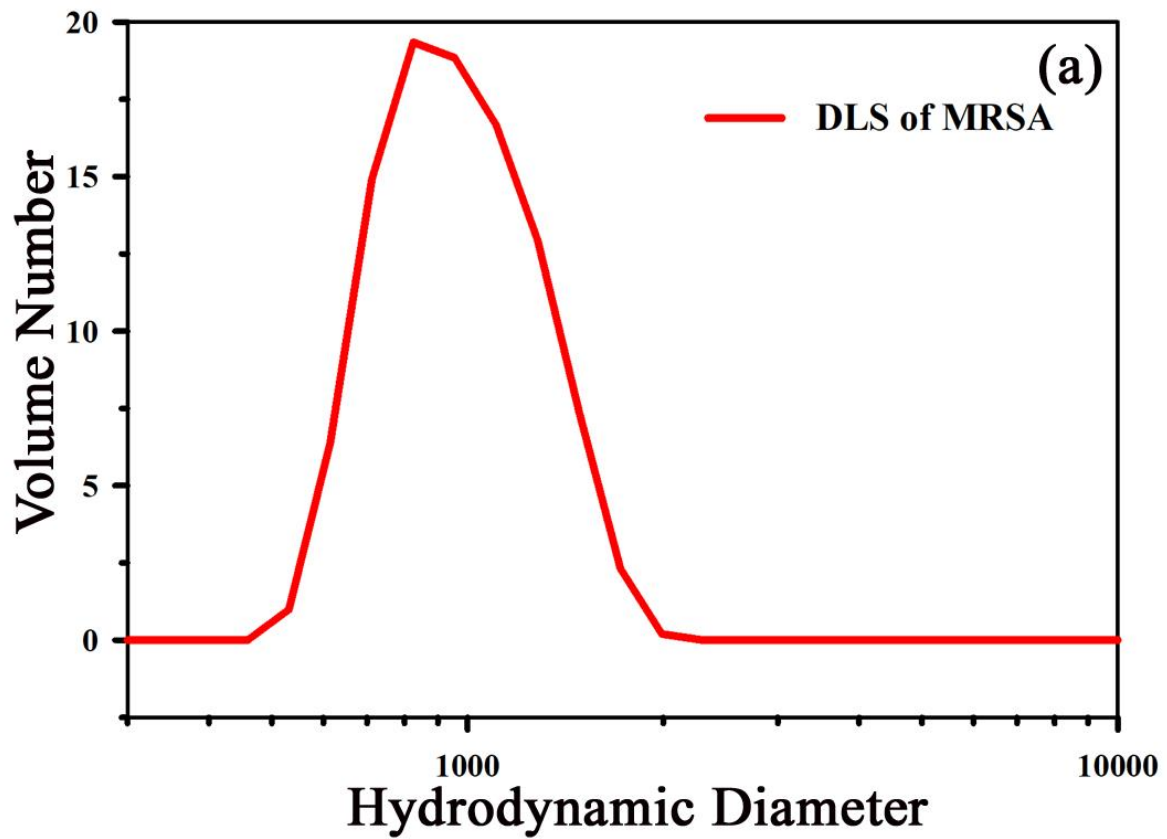
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Figure. 5: Particle size distribution of turbid water samples collected locally and measured with DLS using refractive index (RI) of silica and milk respectively.



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Figure. 6: Growth curve as obtained from the standard process in comparison to the developed strategy.



429 Figure. 7: (a) DLS data of MRSA exhibiting hydrodynamic diameter around 1 micron. (b)
430 Microscopic image of MRSA confirming the diameter of around 1 micron..
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