

1 **Non-additive genetic effects induce novel phenotypic distributions in male mating traits of F1**
2 **hybrids**

3
4

5 **Abstract**

6 Hybridization is a source of phenotypic novelty and variation because of increased additive genetic
7 variation. Yet, the roles of non-additive allelic interactions in shaping phenotypic mean and variance
8 of hybrids have been underappreciated. Here we examine the distributions of male-mating traits in
9 F1 hybrids via a meta-analysis of 3,208 effect sizes from 39 animal species pairs. Although additivity
10 sets phenotypic distributions of F1s to be intermediate, F1s also showed recessivity and resemblance
11 to maternal species. F1s expressed novel phenotypes (beyond the range of both parents) in 65% of
12 species pairs, often associated with increased phenotypic variability. Overall, however, F1s expressed
13 smaller variation than parents in 51% of traits. While genetic divergence between parents did not
14 impact phenotypic novelty, it increased phenotypic variability of F1s. By creating novel phenotypes
15 with increased variability, non-additivity of heterozygotic genome may play key roles in determining
16 mating success of F1s, and their subsequent extinction or speciation.

17

18 **Key words**

19 Meta-analysis, Transgressive segregation, Developmental stability,
20 Heterosis, Outbreeding depression, Phenotypic uniformity

21

22 **Introduction**

23 The long history of animal and plant breeding has taught us that interspecific hybridization is a
24 powerful source of phenotypic novelty. While hybrid populations usually distribute their phenotypes
25 throughout the range of parental species, they often exhibit novel phenotypes, as well as novel
26 variability (i.e. more extreme phenotypic mean or variability than observed in either parent species)
27 (Edmands 1999; Janick 1999; Stelkens and Seehausen 2009). Novel phenotypes enable hybrids to
28 exploit novel niches and ultimately become new species (Rieseberg et al. 1999; Kagawa and
29 Takimoto 2018). Naturally, the distribution of hybrids' phenotypic traits has attracted much attention
30 of evolutionary biologists. Despite long interests of breeding and evolutionary biologists, however,
31 hybrid phenotype has never been subjected to the formal meta-analysis that accounts for different
32 types of statistical non-independence and mean-variance relationship (Rieseberg et al. 1999; Stelkens
33 and Seehausen 2009; Nakagawa et al. 2015).

34
35 Novel phenotypes are produced by various mechanisms, such as mutation, chromosomal
36 recombination, dominance, epistasis, and cross direction effects including epigenetics (DeVicente
37 and Tanksley 1993; Cockett et al. 1996; Lamkey and Edwards 1999; Rieseberg et al. 1999).
38 Chromosomal recombination and rearrangements increase additive (heritable) genetic variation in
39 hybrid populations at second and later generations. In F1 hybrids characterized by heterozygous
40 genome, additive effects are expected to result in the averaged trait size of both parental species.
41 However, F1 hybrids often resemble more one of the two parents (dominance or recessivity), and
42 have different phenotypes between reciprocal hybrids (cross direction effects) (Thompson et al.
43 2021). Moreover, F1 hybrids sometimes exhibit novel phenotypes (Lamkey and Edwards 1999;
44 Rieseberg et al. 1999; Stelkens and Seehausen 2009). Although novel variability — especially
45 smaller phenotypic variation in F1 hybrids — has been utilized in agriculture to enhance yield
46 stability (Riggs 1988; Janick 1999), taxon-wide prevalence of novel variability has not been
47 examined (Vetukhiv 1953; Wallace 1955; Edmands 1999). Fragmented evidence above suggests that
48 non-additive genetic effects are important sources of novel phenotypic distributions of hybrid
49 populations (Alibert and Auffray 2003; Chen 2013; Wei and Zhang 2018). Because phenotypic
50 distribution determines survival and reproductive success of hybrids, it is particularly relevant to the
51 gene flow and backcrossing of hybrid populations, and, ultimately, speciation.

52
53 Genetic divergence between parental species has been hypothesized as a factor shaping phenotypic
54 distribution of F1 hybrids. Heterozygotic loci in F1 hybrids can increase phenotypic novelty by

55 inducing novel allelic interactions (e.g., dominance and epistasis) (Lamkey and Edwards 1999;
56 Rieseberg et al. 1999). Genetic divergence between parents, which enhances heterozygosity of F1
57 hybrids, is thus expected to increase phenotypic novelty of F1 hybrids (Stelkens and Seehausen
58 2009). Further, accrued genetic difference between parents can harm hybrid fitness (i.e. outbreeding
59 depression) due to increasing deleterious allelic interactions (i.e. genetic incompatibility) (Moyle and
60 Nakazato 2010) or collapse of beneficial gene interactions that evolved within the parental species
61 (Dagilis et al. 2019). Outbreeding depression of F1 hybrids could result in smaller trait size than both
62 parents (Arnold and Hodges 1995; Wei and Zhang 2018), and increase phenotypic variability by
63 inducing developmental instability (Alibert and Auffray 2003). Also, F1 hybrids between genetically
64 close parents may perform better than both parents by being released from deleterious allelic
65 interactions within species (i.e. heterosis or hybrid vigor) (Dagilis et al. 2019). Under heterosis, F1
66 hybrids can exhibit greater trait size and / or reduced trait variability than parents because of
67 enhanced developmental stability (Arnold and Hodges 1995; Edmands 1999).

68

69 Here we investigate phenotypic mean and variability of F1 hybrids of male mating trait size (i.e.
70 traits used during mating that females do not possess) across animal taxa (Fig. 2a). By focusing on
71 male traits, we exclude the possible biases resulting from any sex differences in phenotypic
72 distributions. Given a wide prevalence of sexual dimorphism (Poissant et al. 2010), sexes should be
73 distinguished when comparing phenotypic distributions between F1 hybrids and parents. Phenotypic
74 distribution can be sexually-biased in F1 hybrids, but not in parents, due to sex-biased mortality in
75 F1 hybrids (Haldane 1922; Schilthuizen et al. 2011). Male mating traits also directly relate to the
76 reproductive success and have been reported relatively well in the literature. In contrast, little
77 information is available for female-specific sexual traits and mate preference of female hybrids.

78

79 By employing formal phylogenetically controlled meta-analytic techniques, we test if phenotypic
80 mean and variability of F1 hybrids are larger or smaller than mid-species value (Fig. 2b), and are
81 affected by cross direction (father from species A with mother from species B, or vice versa; Fig. 2c).
82 Further, we provide the first quantification of how often F1 phenotype is more / less variable than
83 parents. By using a phylogenetic comparative method, we investigate potential factors including
84 genetic divergence between parents that influence novelty and variability of male mating trait size of
85 F1 hybrids (Fig. 4 and 5: hypotheses for each factor are summarized in Supplementary Table S1).

86

87

88 **Material and methods**

89

90 ***Data collection***

91 **Literature search.** We conducted systematic review of literature and followed Preferred Reporting
92 Items for Meta-Analyses (PRISMA, Fig. 1) when reporting our meta-analytic workflow. We
93 searched journal articles that contain ‘hybrid*’ AND ‘male*’ AND (‘grow*’ OR ‘size.’ OR ‘length.’
94 OR ‘mass’ OR ‘weight.’ OR ‘behav*’ OR ‘trait.’ OR ‘phenotyp*’) in the title, abstract or keywords,
95 by using Web of Science and Scopus on October 20, 2017. We also searched journal articles through
96 backward / forward citations of reviews of hybrid fitness and phenotype (Haldane 1922; Arnold and
97 Hodges 1995; Burke and Arnold 2001; Reinhold 2002; Turelli and Moyle 2007; Stelkens and
98 Seehausen 2009; Schilthuizen et al. 2011; Reinhold and Engqvist 2013).

99

100 **Inclusion criteria.** To be included in our analysis, the primary study had to report means and
101 variance of male sexual phenotypes in hybrids and both non-hybrid crosses (i.e. parental species;
102 note that we use the offspring from non-hybrid crosses as the proxy of the phenotypes of the parental
103 species, the “parents”); separately report the phenotype of reciprocal hybrid crosses; use different
104 species or subspecies for the crosses; experimentally obtain all crosses and raise them in same
105 environment; provide all necessary statistics (means, standard deviations/errors and sample sizes) for
106 effect size calculations. Relevant studies deemed to meet the above criteria, based on titles and
107 abstracts, were screened as full-texts by two of the authors (KA and ML).

108

109 **Data extraction.** From primary studies, we extracted the phenotypic measures and the number of
110 individuals in each group of animals. We extracted data from figures using R package *metaDigitse*
111 1.0 (Pick et al. 2019). We estimated standard deviation from range data, if necessary (Wan et al.
112 2014). The observations that had any negative trait sizes were removed from the dataset, but this
113 procedure did not reduce included studies and species.

114

115 ***Dominance and cross direction effects***

116 **Effect sizes.** For phenotypic means, we calculated the log ratio ($\ln RR$ (Hedges et al. 1999)) from the
117 non-hybrid parent with the smaller trait size (small non-hybrid) to each reciprocal hybrid cross
118 (arrows 2–3 in Fig. 2b), mid-species (arrow 4 in Fig. 2b) and the other parent (non-hybrids with
119 larger trait size — large non-hybrid: arrow 1 in Fig. 2b). We evaluated phenotypic variability in the
120 analogous way with phenotypic mean, but we used the log ratio between coefficient of variation

121 (lnCVR, Nakagawa et al. 2015) and aligned parents by phenotypic variability (small and large
122 non-hybrids for parents with small and large coefficient of variation of trait size, respectively).

123

124 **Meta-analyses.** To estimate the overall mean of each effect size, we first estimated meta-analytic
125 means of each effect size (effect sizes 1–3 in Fig. 2b). We then asked whether phenotypic mean or
126 variability is greater or smaller than mid-species (comparing effect sizes 2–3 with 4 in Fig. 2b), by
127 using meta-analytic model that included mid-species value as an intercept. To investigate cross
128 direction effects, we tested whether the reciprocal hybrid crosses differ in their mean phenotype
129 (compared effect size 2 with 3 in Fig. 2b).

130

131 All meta-analytic models were phylogenetically controlled and included following random effects:
132 study ID, trait observation ID and strain ID that discriminated intraspecific genetic strains or
133 populations. Phylogenetic effects were included as a random effect as a covariance matrix for large
134 non-hybrid parental species. We did not include species as random effect because this effect should
135 be accommodated by the strain random effect. A phylogenetic tree was created based on Open Tree
136 of Life database (Hinchliff et al. 2015) (topology of the tree: Fig. S1–2). We identified species in the
137 Open Tree Taxonomy using the R package *rotl* 3.0.10 (Michonneau et al. 2016) and computed
138 branch lengths using the default settings of the *compute.brLen* function in the R package *ape* 5.3
139 (Paradis and Schliep 2019). All meta-analytic models were implemented using the R package
140 *metafor* 2.1-0 (Viechtbauer 2010). All analyses were conducted in R 3.6.3 (R Core Team 2020).

141

142 ***Novelty in phenotypic means and variabilities***

143 **Effect sizes.** For phenotypic mean, we calculated lnRR of large non-hybrid to hybrid crosses, and
144 lnRR of hybrid crosses to small non-hybrid at each trait observation (Fig. 2c). As the sign of these
145 effect sizes should be positive when hybrid phenotypic mean lies within the ranges of parents,
146 negative effect sizes indicate novel phenotypic means. These effect sizes also indicate the relative
147 trait size compared with parents – whether mean trait size is greater or smaller than both parents and
148 whether hybrids exceed maternal or paternal species' trait size (Fig. 2c). Again, we evaluated
149 phenotypic variability in the analogous way with phenotypic mean by using lnCVR and aligning
150 parents by phenotypic variability size.

151

152 **Predictor variables.** We additionally collected genetic distance between parental species and
153 heterogametic sex because we *a priori* expected them to influence phenotypic distribution of hybrids.

154 We took the mitochondria *cytochrome c oxidase I (COI)* DNA sequences (up to 20 sequences per
155 parental species depending on their availability) from GenBank
156 (<http://www.ncbi.nlm.nih.gov/Genbank/>) and aligned them using ClustalW (Thompson et al. 1994).
157 Alignments with 450 bp as the minimum length were manually optimized for calculating the genetic
158 distance. We calculated maximum composite likelihood distance with uniform substitution rate and
159 homogenous pattern (Tamura et al. 2004) in the software MEGA 7.0.26 (Kumar et al. 2016). If
160 multiple sequences were available for a pair of species, we calculated the average of all possible
161 pairs of sequences. We assigned heterogametic sex at family or genus level, if deemed evolutionary
162 constant within the taxon (Ashman et al. 2014).

163

164 **Bayesian models.** To find the factors affecting novelty in phenotypic means and variances, we
165 conducted logistic regression with the novelty status of phenotypic mean or variance (whether
166 phenotypic mean or variance of hybrids is more extreme than of either parents) as a binary response
167 variable (0 = not novel; 1 = novel). We ran the analogous logistic models for phenotypic means and
168 variances. As predictors, the models included: (i) natural logarithm of genetic divergence between
169 parents; (ii) phenotypic divergence between parents in the focal trait – absolute value of $\ln RR$; (iii)
170 range overlaps between parental species – allopatry or not; (iv) viability of reciprocal hybrids –
171 completely inviable or not; (v) trait type – morphology or sound traits; and the relative trait size or
172 variability compared to parents – (vi) greater or smaller and (vii) exceeding maternal or paternal
173 species. The model also included interaction terms of species or trait level factors ($i-v$) with parental
174 species compared (vi and vii). Range overlaps and viability of hybrids were assessed based on the
175 primary studies.

176

177 The main effects indicate how species- and trait-level characteristics influence the probability of F1
178 hybrids to exhibit novel phenotypic means or variability in any directions ($i-v$), and whether
179 phenotypic means or variability is biased toward certain direction (vi and vii). The interaction terms
180 indicate how species and trait level characteristics affected the relative trait size or variability
181 compared to parents (interactions with vi, greater trait size than both parents; those with vii, trait size
182 exceed maternal species value). Two (all) fish and three orthoptera species-pairs were not used in
183 regression because of the lack of data for genetic distance or sex-determination system. To visualize
184 significant factors' influences on the relative trait size or variability (Fig. 4c–e and 5c–e), we ran
185 simpler models for each significant factor. As predictors, we included the focal factors (either one of
186 $i-v$), the compared parental species – large or small (vi), and the interaction between them. As

187 random effects, these models included trait study ID, strain ID (discriminated genetic strains or
188 populations within a species), and standard error of the response variable. Phylogenetic effects were
189 included as a random effect as a covariance matrix for large parental species (Fig. S1–2). Standard
190 error of the binary response variable was estimated as $\frac{\pi}{\sqrt{3n}}$, where n is the summation of sample sizes
191 of the hybrid and non-hybrid crosses compared.

192
193 Lastly, we conducted an ordinal regression mixed-effects model to examine the link between
194 phenotypic novelty and variability. The model included relative trait variability of hybrids compared
195 to parents (ordered: greater than both parents > within the range of parents > smaller than both
196 parents) as a response, and novelty in phenotypic means (novel or non-novel) as a predictor variable.
197 This model also included trait study ID, strain ID, trait observation ID, cross direction and phylogeny
198 (tree topology, Fig. S3) as random effects.

199
200 All Bayesian analyses were conducted in R package *MCMCglmm* v.2.29 (Hadfield 2010), using
201 phylogenetic comparative hierarchical models described in Hadfield and Nakagawa (2010). We used
202 a weakly informative Gelman prior for the fixed effects, and inverse-Wishart priors for the variances
203 of the random effects. The residual variance was fixed to 1, as this cannot be estimated with binary
204 or trinary data. Parameter estimates were subsequently scaled under the assumption that the true
205 residual variance is 0. We ran the analysis for 60,000 iterations with a burn-in of 5,000 and a thinning
206 interval of 10.

207 208 209 **Results**

210 211 ***Dataset***

212 Although trait inheritance pattern is a key question of evolutionary biology, a modest number of
213 publications reported results of crossing species reciprocally while separating offspring trait data by
214 sexes. We found 25 such published studies (Supplementary material, section *Data description*). We
215 extracted two sets of 1,604 effect sizes comparing phenotypic mean and variability, respectively,
216 between parents and hybrids, based on 401 male traits observations from 39 species pairs. More than
217 half species crossings involved *Drosophila* (Diptera, 59%, 23/39 species pairs, Fig. 2a) and only
218 13% of species pairs (5/39) were vertebrates, including bony fish, frog, bird and rodent species. Most

219 species pairs were male heterogametic (92%, 34/37 species pairs, excluding two fishes of which
220 sex-determination system is unknown). Both reciprocal hybrids were viable in 59% of species pairs
221 (23/39); and geographic ranges of parental lineages overlap in 36% of species pairs (14/39).

222

223 ***Dominance in phenotypic means***

224 F1 hybrids of both cross directions showed partial recessivity. Hybrids whose maternal species had
225 larger trait size (large non-hybrid mother \times small non-hybrid father) and the reciprocal hybrids (large
226 father \times small mother) on average showed 12.5% and 16.9% smaller phenotypic mean than
227 mid-species value, respectively (Fig. 3a; $P < 0.001$ each, Table S3). This trend was qualitatively the
228 same when we excluded traits showing novel phenotypic means (i.e. values outside the range of both
229 parental species; Fig. 3c). Nonetheless, the magnitude of phenotypic difference between hybrids and
230 parental species is highly heterogeneous, much of which are either due to phylogeny or unexplained
231 (total $I^2 = 99.7\%$, which are partitioned into phylogeny $I^2 = 38.8\%$, study $I^2 = 1.1\%$, crossed lineage
232 $I^2 = \sim 0\%$, residual $I^2 = 59.8\%$; Fig 3a–b). Such a high heterogeneity suggests that any inheritance
233 patterns are plausible in F1 hybrids (e.g., dominance, cross direction effect and exhibiting novel
234 phenotypic means). –

235

236 ***Crossing direction effects in phenotypic means***

237 Hybrids were more similar to the males of maternal species than of paternal species: F1 hybrids from
238 large mother species exhibited 4.9% larger trait size than the reciprocal hybrids (large father \times small
239 mother) (Fig. 3a; $P < 0.001$, Table S4). Same tendency was detected when we excluded traits
240 showing novel phenotype (hybrids having larger maternal species had 8.7% larger phenotypic
241 means; $P < 0.001$, Table S4).

242

243 ***Dominance and cross direction effects in phenotypic variabilities***

244 F1 hybrids exhibited similar phenotypic variability (coefficient of variation, CV) to the mid-species
245 value (Fig. 3b, Table S5). Phenotypic variability did not significantly differ between F1 hybrids
246 whose maternal species had larger trait variability and the reciprocal hybrids (Table S6). Yet, these
247 do not necessarily mean that phenotypic variability inherits additively. In most trait observations,
248 hybrid phenotypic variability is either larger or smaller than that of both parents (74.9%, see *Novelty*
249 *in phenotypic variabilities*). We also note that detection power of the differences in CVs is lower than
250 in comparisons of means or variances alone, because errors of CVs contain sampling errors of both
251 means and variances (Nakagawa et al. 2015; Senior et al. 2020). The magnitude of phenotypic

252 variability was highly heterogenous, and was partially dependent upon phylogeny and study
253 identities (total $I^2 = 75.6\%$, partitioned into: phylogeny $I^2 = 19.5\%$, study $I^2 = 11.4\%$, crossed lineage
254 $I^2 = \sim 0\%$, residual $I^2 = 44.7\%$; Fig. 3c, d).

255

256 *Novelty in phenotypic means*

257 F1 hybrids exhibit novel phenotypic means in 64.7% (22/34) of species pairs, and 42.9% (143/333)
258 of trait observations (Fig. 4a). This indicates that non-additive genetic interaction is a powerful
259 source of phenotypic novelty in male mating traits.

260

261 Genetic divergence between parents did not significantly associate with novelty in phenotypic means
262 (main effect: $\beta = 0.26$, 95% credible interval [CI] = -0.41 – 0.9, $P = 0.420$; Fig. 4b). Rather, genetic
263 divergence significantly reduced the probability of hybrids to have larger trait size than both parents
264 (interaction term of genetic divergence with the relative trait size: $\beta = -0.59$, CI = -1.19 – 0.02, $P =$
265 0.041; Fig. 4b–c). Increasing phenotypic divergence between parental species generally reduced the
266 novelty (main effect of phenotypic divergence: $\beta = -0.8$, CI = -1.48 – -0.23, $P < 0.001$). Compared to
267 F1 hybrids with viable reciprocal hybrids, those without viable reciprocal hybrids were more likely
268 to exhibit smaller trait mean than both parents (interaction term of reciprocal hybrids' viability with
269 the relative trait size compared to parents: $\beta = 2.51$, CI = 1.01 – 4.01, $P = 0.001$; Fig. 4b and e) and,
270 albeit non-significant, tended to show novel phenotypic means (main effect of reciprocal hybrids'
271 viability: $\beta = -1.36$, CI = -2.97 – 0.04, $P = 0.064$; Fig. 4b). Novel phenotypic means were not biased
272 toward any directions, as F1 hybrids exhibited either larger or smaller trait size than parents (main
273 effect of relative trait size, $P > 0.05$, Table S7) and significantly exceeded phenotypic means of either
274 maternal or paternal species (main effect of exceeding maternal or paternal species, $P > 0.05$, Table
275 S7). This indicates that recessivity and crossing direction effect had little impact on novelty in
276 phenotypic means.

277

278 *Novelty in phenotypic variabilities*

279 Phenotypic variability of F1 hybrids rarely lied within the range of parental species (6.1%, 31/33 of
280 species pairs; 25.1%, 248/331 of trait observations, Fig. 5a). Compared to the parents, hybrids
281 exhibited smaller phenotypic variability in 51.4% of trait observations and had greater variability less
282 frequently (24.8%, Fig. 5a).

283

284 Novel phenotypes were more variable than non-novel phenotypes of which mean lie within the range

285 of parents (ordinal phylogenetic random regression: $\beta = 1.01$, CI = 0.45 – 1.62, $P < 0.001$). That is,
286 novel phenotypes, regardless of relative trait size compared to parents, were more likely to exhibit
287 greater variability than parents (30.0 % of trait observations) in comparison to non-novel phenotypes
288 (18.5 %).

289
290 F1 hybrids between genetically distant parents were likely to be phenotypically more diverse than
291 parents, whereas those between genetically close parents were typically more homogenous than
292 parents (interaction term of genetic divergence with the relative trait variability compared to parents:
293 $\beta = 1.25$, CI = 0.47 – 2.00, $P = 0.002$: Fig. 5b–c and Table S8). F1 hybrids were more phenotypically
294 variable than parents in traits with large parental divergence (interaction term of phenotypic
295 divergence with the relative trait variability: $\beta = 0.83$, CI = 0.01 – 1.61, $P = 0.036$: Fig. 5b). Note that
296 novelty in variabilities did not associate with phenotypic divergence when species-level moderators
297 were not included in the model (Fig. 5d). F1 hybrids tended to vary less than parents in sound traits,
298 and to vary more than parents in morphology (interaction term of sound traits vs. morphological
299 traits with the relative trait variability: $\beta = -3.29$, CI = -5.21 – -1.38, $P = 0.001$: Fig. 5b and e).

300

301

302 **Discussion**

303

304 By synthesizing 401 male traits observation data from 39 species pairs across the animal kingdom,
305 we illuminated important roles of non-additivity in phenotypic distribution of F1 hybrids. We
306 detected recessivity and cross direction effect on phenotypic means of the hybrids (Fig. 3a and c).
307 Novel phenotypic means were found in 65% of species pairs (Fig. 4a), highlighting the importance
308 of non-additive allelic interactions in creating novel phenotypes. Novelty in phenotypic means was
309 not associated with parental genetic divergence but enhanced by genetic incompatibility (when
310 reciprocal hybrids are inviable, Fig. 4b). Similar to observations made in plant agricultural breeding
311 (Riggs 1988; Janick 1999), F1 hybrids exhibited smaller phenotypic variability than both parents in
312 majority of trait observations (Fig. 5a). We showed that genetic differentiation between parents
313 increases phenotypic variability of F1 hybrids (Fig. 5b).

314

315 Recent synthesis showed that resemblance to maternal species was the norm in F1 hybrids across
316 animals and plants (Thompson et al. 2021). Our analysis also revealed resemblance to maternal
317 species of F1 hybrids, which is counterintuitive because we focused on male traits. Given that

318 maternal effects are particularly pervasive for morphology, including body size (Moore et al. 2019),
319 mothers can directly influence male-mating morphological trait size, and even sound traits that are
320 partially determined by morphology and body size (Ryan and Brenowitz 1985; Bennet-Clark 1998).
321 Alternatively, maternally inherited mitochondrial genome may influence F1 male phenotypes.

322

323 *Phenotypic novelty*

324 We suggest that hybrid populations may express novel phenotypes much more frequently than
325 previously thought. A recent comparative analysis, using non-male-mating traits, showed that F1
326 hybrids express novel phenotype only in 20% of species pairs across any plants and animal taxa
327 (Thompson et al. 2021), which is one-third of the estimate from our study (65%). Discordance
328 between these two results suggests that male-mating traits of animals more frequently express novel
329 phenotype during hybridization, and/or using sex-aggregated data in the earlier study has led to
330 underestimation of novel phenotype expression frequency. Alternatively, a large portion of novel
331 phenotype observations in our dataset may arise from heterosis or outbreeding depression, though
332 Thompson et al. (2021) attempted to exclude heterosis and outbreeding depression by choosing traits
333 seemingly unrelated to fitness. Our dataset included any mating traits that are likely to reflect body
334 growth (e.g., genitalia size) or behavioral deficit (e.g., compromised vocal traits). Hence, F1 hybrids
335 may exhibit larger or smaller male trait sizes than both parents by growing faster or slower than
336 parents (heterosis and outbreeding depression, respectively). Outbreeding depression, at least, is
337 possibly reflected in our result — trait size declined in F1 hybrids without viable reciprocal hybrids
338 and from genetically distant parents (Fig. 4b–c and e).

339

340 While non-additive interactions can enhance phenotypic novelty in any generation of hybrids, studies
341 on quantitative trait loci (QTL) suggests that recombination after F1 hybrids is the major source of
342 phenotypic novelty (DeVicente and Tanksley 1993; Rieseberg et al. 2003; Koide et al. 2019). Novel
343 phenotype is thus expected to be more frequent in later generation hybrids than in F1 hybrids. Yet,
344 other comparative studies, included F1 and later generation hybrids across animals, found that 29–
345 31% of traits showed novel phenotypes (Rieseberg et al. 1999; Stelkens and Seehausen 2009), which
346 is about half of our estimate. We exclusively focused on experimentally-derived F1 hybrids while
347 previous studies included natural hybrid populations (Rieseberg et al. 1999; Stelkens and Seehausen
348 2009). In natural hybrid populations included in the previous syntheses, extrinsic natural selection
349 may have removed novel phenotypes and led to underestimation of phenotypic novelty (Rieseberg et
350 al. 1999; Stelkens and Seehausen 2009).

351

352 We anticipated that genetic divergence between parental species positively relates to phenotypic
353 novelty. This is because parent divergence should increase the number of heterozygotic loci in F1
354 hybrids, and heterozygosity is expected to enhance phenotypic novelty (Lamkey and Edwards 1999).
355 However, in our results, genetic divergence did not associate with the probability of F1 hybrids to
356 express novel phenotypic means (Fig. 4b). This is in line with the previous finding across animals
357 and plants that phenotypic means of F1 hybrids do not increase their degree of novelty along with
358 genetic divergence between parents (Thompson et al. 2021). Our prediction was not supported,
359 presumably because the effects of inter-allelic interactions can be diverse. We assumed that
360 inter-allelic interactions among QTLs act in the same direction (e.g., all interactions increase trait
361 size) (Lamkey and Edwards 1999). If the sign varies among the interactions, however, genetic
362 differentiation no longer necessarily increases phenotypic novelty because interactions will cancel
363 out effects of each other. Alternatively, F1 hybrid fitness can be maximized when genetic distance
364 between the parents is small or moderate because of strong heterosis combined with weak genetic
365 incompatibility (Moll et al. 1965; Wei and Zhang 2018; Dagilis et al. 2019). When heterosis
366 increases trait size through improving fitness, novel phenotype expression can become frequent in
367 hybrids between non-genetically distant parents. Genetic divergence between parents thus could be a
368 poor predictor of phenotypic novelty of F1 hybrids.

369

370 Based on the similar reasoning with genetic divergence, we expected that F1 hybrids are likely to
371 exhibit novel phenotypic means in traits with greater parental phenotypic divergence. However, our
372 analysis detected the opposite pattern (Fig. 4b and d). When parental species are phenotypically
373 similar, their QTL could be heterozygous. Here, some of F1 hybrids will be homozygous at QTL.
374 Additive effects of QTL enable homozygous hybrids to express novel phenotype (Stelkens and
375 Seehausen 2009). Hence, additivity and heterozygosity of parents may explain the novel phenotypic
376 means of F1 hybrids from phenotypically close parents. Furthermore, we found that novel
377 phenotypes (mean outside the range of parents) were more variable than non-novel phenotypes
378 (mean within the range of parents). If phenotypic variability reflects developmental instability
379 (Edmands 1999; Alibert and Auffray 2003), observed pattern suggests that novel phenotypes arise
380 from developmental instability of hybrids. That is, hybrids with stable development may exhibit less
381 extreme phenotype (within the range of parents), whereas hybrids suffering developmental instability
382 may tend to exhibit novel phenotype. Moreover, F1 hybrids tend to exhibit novel phenotype when
383 genetic incompatibility is strong (i.e. the reciprocal hybrids is inviable, Fig. 4b). We therefore

384 suggest that developmental instability arising from genetic incompatibility is an important source of
385 phenotypic novelty in F1 hybrids.

386

387 *Phenotypic variability*

388 We revealed F1 hybrids tend to be less phenotypically variable than parents (Fig. 5a). Yet, this does
389 not necessarily indicate reduced evolvability of hybrid populations — later-generation hybrids can
390 expand phenotypic variation through recombination even if F1 hybrids exhibit smaller phenotypic
391 variability than parents (Edmands 1999; Rieseberg et al. 1999). Rather, our results emphasize the
392 time lag between hybridizing event and expression of novel phenotypic variation in hybrid
393 population (Grant and Grant 2019).

394

395 As expected, genetic divergence between parental species enhanced phenotypic variability of F1
396 hybrids (Fig. 5b). Increasing phenotypic variability with parental genetic divergence was also
397 reported in later generation hybrids (Stelkens and Seehausen 2009; Stelkens et al. 2009). The
398 common pattern in F1 and later generation hybrids implies that non-additive interactions contribute
399 to phenotypic distribution of hybrid population. Genetically close parents yielded F1 hybrids with
400 smaller phenotypic variability, possibly due to heterosis enhancing developmental stability (Edmands
401 1999), rather than due to selective mortality caused by outbreeding depression (i.e. individuals with
402 anomalous phenotypes, resulting from epistatic interactions, are likely to die). This is because
403 heterosis is often observed in hybrids between genetically similar parental species (Dagilis et al. 2019),
404 but outbreeding depression become more likely as parents genetically diverge (Moyle and Nakazato
405 2010; Dagilis et al. 2019).

406

407 We found that F1 hybrids tended to vary less than parents in sound traits, and to vary more than
408 parents in morphology (Fig. 5b). Larger variability in mating-related traits may facilitate
409 backcrossing due to greater overlap in phenotypic range with parental species. Hence, taxa relying
410 predominantly on morphology-based mating traits (e.g., genitalia and coloration) might be more
411 prone to gene flow resulting from backcrossing.

412

413 *Knowledge gap and future directions*

414 In addition to each individual trait size, trait correlation (covariance) can influence the attractiveness
415 of F1 males, because multiple mating traits often interactively determine sexual attractiveness
416 (Rosenthal 2013). Our additional analysis revealed traits can vary in the strength and directions of

417 dominance within hybrids (i.e. several traits resemble one parent, but other traits resemble the other
418 parent: Supplementary material, section *Trait mosaicism*), which was previously shown in non-male
419 mating traits (Thompson et al. 2021). This indicates that trait correlation within parental species
420 could easily break in F1 hybrids, as reported in several studies (Rieseberg and Ellstrand 1993;
421 Matsubayashi et al. 2010; Selz et al. 2014; Thompson et al. 2021). Moreover, a variety of trait
422 correlation patterns can arise in F1 individuals because phenotypic variability varies among traits
423 (Fig. 3b and 5b). Despite the importance of determining fitness and mating pattern, correlation
424 among mating traits of F1 hybrids has received little attention (Parsons et al. 2011). Mating pattern
425 of F1 hybrids also depends on mate preference of parents and hybrids (Svedin et al. 2008; Chen and
426 Pfennig 2020), of which inheritance pattern during hybridization varies across species pairs and
427 preference components (Table S2 for a summary of F1 female mate preferences). Yet, we are still far
428 from drawing general patterns of hybrid mate preference because it has rarely been studied. By
429 filling knowledge gaps in trait integration and mate preference of F1 hybrids, we can better
430 understand mechanisms of reproductive isolation and gene flow.

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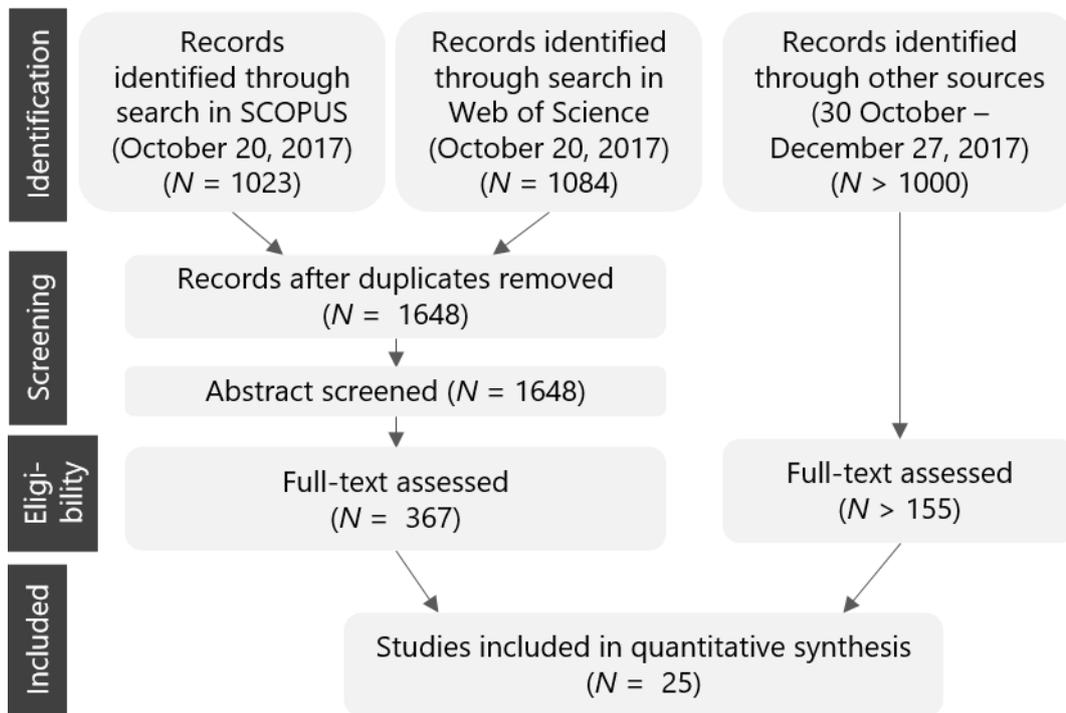
432 Non-additive interactions among species-specific alleles have received great attention, especially
433 regarding genetic incompatibilities damaging hybrid fitness (Turelli and Moyle 2007; Dagilis et al.
434 2019). In contrast, we still know very little about how allelic interactions influence the phenotypic
435 distribution of hybrid populations. Since allelic interactions are largely not heritable, increased
436 phenotypic novelty or variability in F1 hybrids do not directly indicate enhanced evolvability of
437 hybrid populations. Nevertheless, such non-additive interactions can appear in any hybrid
438 generations, including F1 hybrids. Indeed, QTL studies in rice have shown that epistasis underlies
439 novel phenotype expression by later generation hybrids (Mao et al. 2011; Koide et al. 2019).

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441 ***Conclusions***

442 By leveraging recent developments in meta-analysis, we have shown non-additive interactions are
443 powerful sources of phenotypic novelty and stability in male reproductive traits. By providing
444 phenotypic novelty and impacting phenotypic variation, non-additive allelic interactions may play
445 key roles in determining early succession and dynamics of hybrid populations, and thus, the course
446 of subsequent extinction or speciation. Finally, researchers can use the formal meta-analytic
447 techniques we have developed in this study to synthesize growing empirical articles and to generate
448 new insights into speciation.

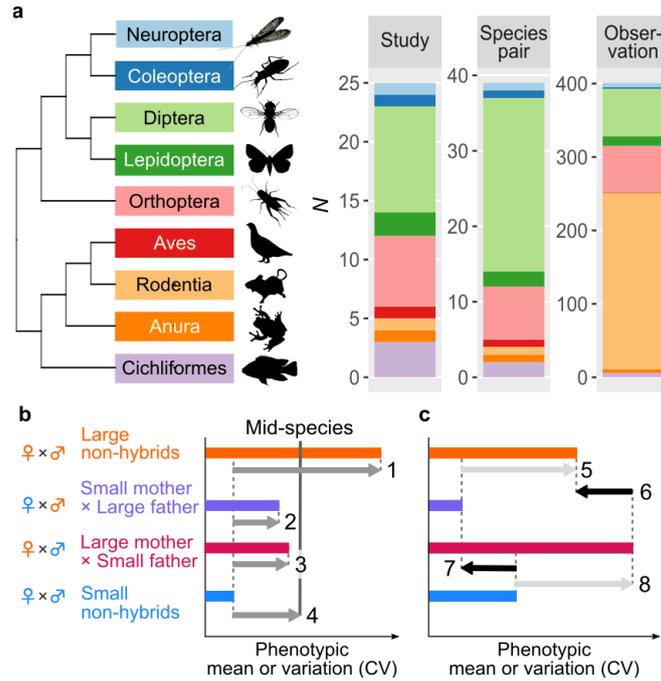
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453 **Figure 1 | PRISMA diagram.** The flow of inclusion and exclusion of studies identified during the
 454 literature search is shown. In brackets, we indicate the number of published literature (studies, *N*).
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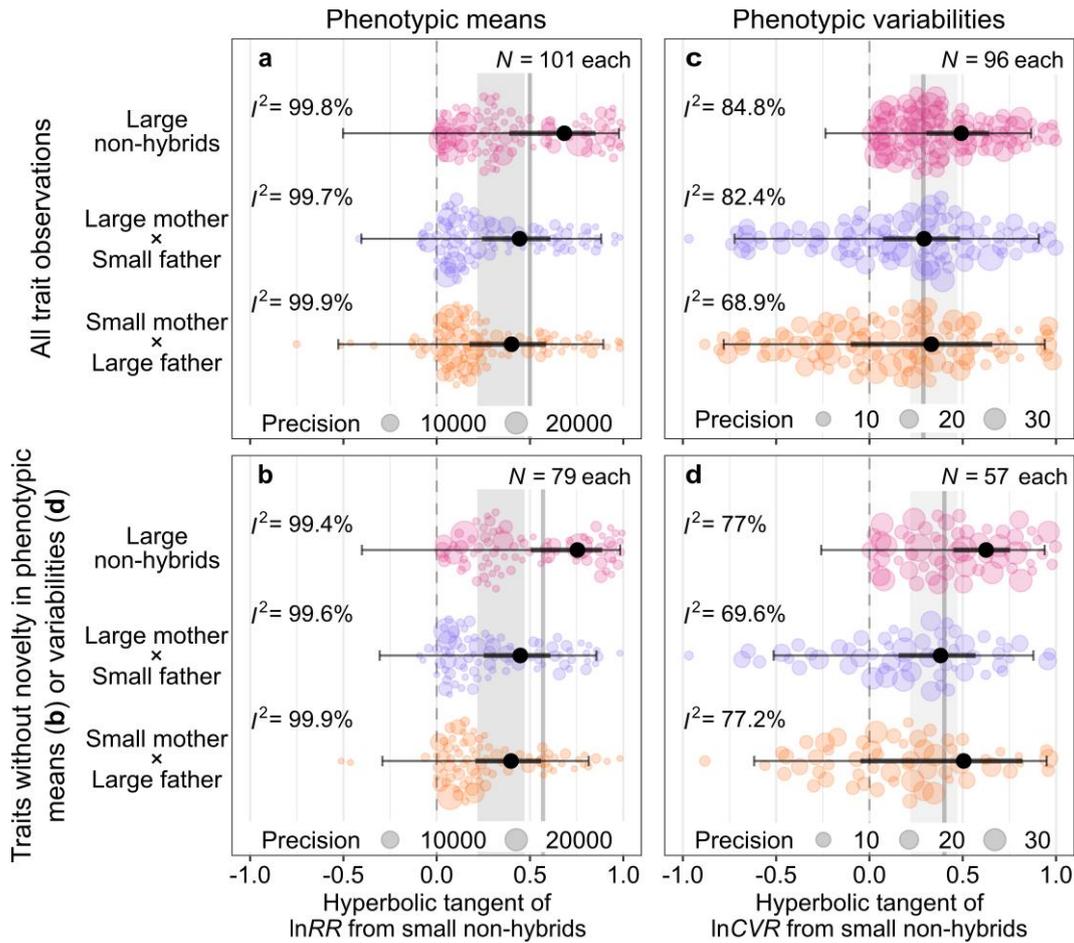
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Figure 2^[KA1] | **Dataset and effect size calculation.** **a** Taxonomic diversity of the data set, at the phylogeny, study, species and observation levels; shades of green refer to the main taxonomic groups, as on the phylogenetic tree. **b–c** Schematic representation on the ways used to calculate effect sizes that compared phenotypic mean and variation among males of hybrids and non-hybrid parental species (arrows). **b** Calculation of effect size used in the formal meta-analytic models (grey arrows). **c** Our approach to assess novel phenotype and variability expression. Black arrows indicate novel phenotypic means or variabilities, but light grey arrows do not. In this hypothetical example, hybrids from species with larger trait size or variability (large non-hybrids) females and species with smaller trait size or variability (small non-hybrids) males exceeded the large non-hybrids and maternal species (arrow 6), but the reciprocal hybrids (small female × large male) exceeded the small non-hybrids and father species (arrow 7).



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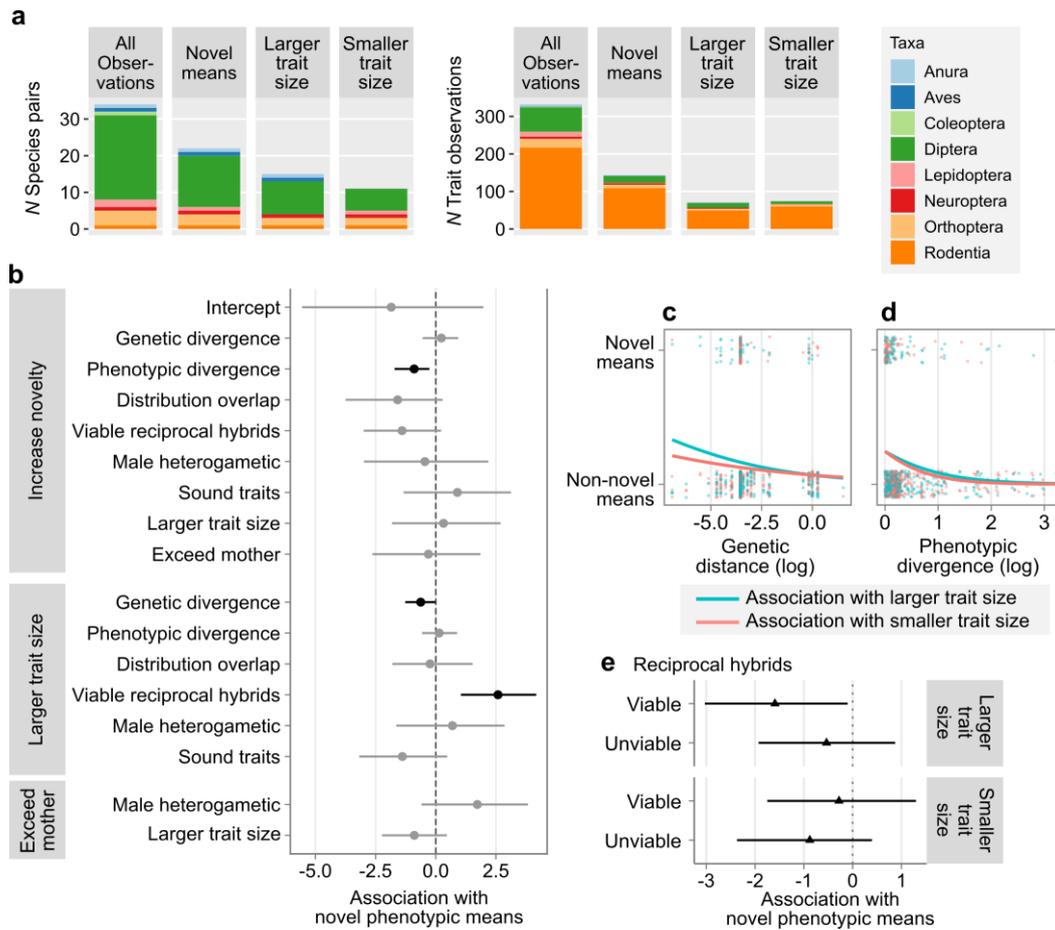
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Figure 3 | Relative phenotypic means and variabilities of hybrids and large non-hybrids

compared to small non-hybrids using orchard plots. The meta-analytic mean (mean effect size) is shown with its 95% confidence interval (thick line) and 95% prediction interval (thin line; see Nakagawa et al. 2021). Individual effect sizes are represented as dots proportional to sample sizes. Small and large non-hybrids indicate parental species with smaller and larger trait size for a given trait in **a** and **c**; species with smaller and larger trait size variability for a given trait in **b** and **d**. Dashed line indicates no difference from small non-hybrids, while grey vertical line indicates mid-species value (average of parental species in male trait size [**a** and **c**], and coefficient of variation of male trait size [**b** and **d**]).



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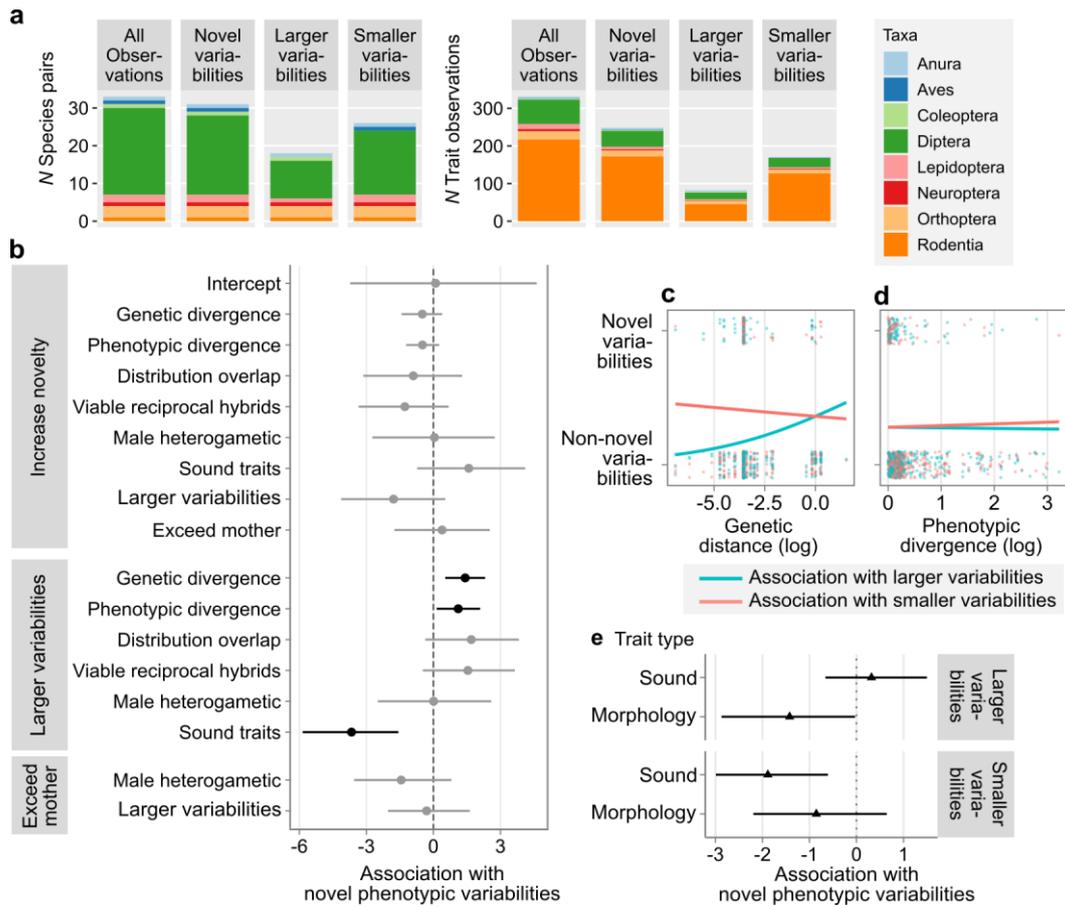
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Figure 4 | Novelty in phenotypic means. **a** Taxonomic distribution of novel phenotypic means. Novel phenotypic means was counted per species (count of species pairs whose hybrids exhibited novel phenotype in any traits; right) and per trait observation (when hybrids exhibited novel phenotype in any direction; left). Novel phenotypes with larger/smaller trait size compared to parents were also counted. **b** Result of the full model for the probability of hybrids to exhibit novel phenotypic means (point estimates with 95% Confidence Intervals). The main effects are categorized as the factors potentially “Increase novelty” in phenotypic means. The interaction terms with compared parental species (large vs. small, and maternal vs. paternal, see Fig. 2c) were labeled as “Larger trait size” and “Exceed mother”, respectively. Statistically significant and non-significant predictors are shown as black and grey, respectively. **c-e** Impacts of the three significant factors (identified in panel **b**) on novel phenotypic means with larger/smaller trait size.



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Figure 5 | Novelty in phenotypic variabilities. **a** Taxonomic distribution of novel phenotypic variability. Counts of novel phenotypic variability are analogous to Fig. 4a. **b** Result of the full model (point estimates with 95% CI). The categories of regression factors are analogous to Fig. 4b. Statistically significant and non-significant predictors are shown as black and grey, respectively. **c-e** Impacts of the three significant factors (identified in panel **b**) to the larger/smaller phenotypic variability.

506 **Competing interests**

507 The authors declare no competing interests.

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510 **References**

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