

**Does degeneration or genetic conflict shape gene content on UV sex
chromosomes?**

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1 **Abstract**

2 Studies of sex chromosomes have played a central role in understanding the
3 consequences of suppressed recombination and sex-specific inheritance among
4 several genomic phenomena. However, we argue that these efforts will benefit from a
5 more rigorous examination of haploid UV sex chromosome systems, in which both the
6 female-limited (U) and male-limited (V) experience suppressed recombination and sex-
7 limited inheritance, and both are transcriptionally active in the haploid and diploid states.
8 We review the life cycle differences that generate UV sex chromosomes and genomic
9 data showing that ancient UV systems have evolved independently in many eukaryotic
10 groups, but gene movement on and off the sex chromosomes, and potentially
11 degeneration continue to shape the current gene content of the U and V chromosomes.
12 Although both theory and empirical data show that the evolution of UV sex
13 chromosomes is shaped by many of the same processes that govern diploid sex
14 chromosome systems, we highlight how the symmetrical inheritance between the UV
15 chromosomes provide an important test of sex-limited inheritance in shaping genome
16 architecture. We conclude by examining how genetic conflict (over sexual dimorphism,
17 transmission ratio distortion, or parent-offspring conflict) may drive gene gain on UV sex
18 chromosomes, and highlight the role of breeding system in governing the action of
19 these processes. Collectively these observations demonstrate the potential for
20 evolutionary genomic analyses of varied UV sex chromosome systems, combined with
21 natural history studies, to understand how genetic conflict shapes sex chromosome
22 gene content.

23

24 **Introduction**

25 On this 50th anniversary of the International Association of Bryology, we look back
26 another five decades to the very beginnings of genetics and a remarkable period of
27 discovery in bryology. During this period, the first sex chromosomes documented in
28 plants were found in the liverwort *Sphaerocarpus donellii* (Allen 1917) and (Heitz 1928)
29 coined the now ubiquitous term "heterochromatin" for the dark-staining, heteromorphic
30 sex chromosomes in mosses. More recent work on sex chromosomes has led to key
31 discoveries regarding fundamental genetic processes such as recombination and gene
32 dosage (Muller 1932; Begun & Aquadro 1992). However, nearly all of this subsequent
33 foundational work has focused on species with XY and ZW sex chromosomes like in
34 Dipterans, Lepidopterans, Mammals, Birds, and few seed plants, organisms that do not
35 reflect the true diversity of sex chromosomes. Many bryophytes and algae possess UV
36 sex chromosomes, which are shaped by distinct evolutionary processes (reviewed in
37 (Bachtrog *et al.* 2011; McDaniel & Perroud 2012; Renner *et al.* 2017; Coelho *et al.*
38 2018). Only in the past few years have researchers brought new experimental tools to
39 observations first made more than a century ago to explore the untapped potential of
40 comparative genomic analyses among haploid-dioecious organisms.

41 The purpose of this review is to highlight the novel insights into genome evolution
42 that can be gained by studying UV sex chromosomes. We first present the two-locus
43 model for the evolution of sex chromosomes in the context of a UV system. Next we
44 describe how the symmetry of U and V sex chromosome transmission alters
45 established expectations for several central molecular population genetic parameters,
46 including recombination, effective population size, dosage, and the accumulation of

47 deleterious mutations, relative to XY or ZW chromosome systems. In the third section,
48 we focus on one of the key unanswered questions in sex chromosome evolution – what
49 are the roles of various forms of genetic conflict (sexual dimorphism, sex-ratio bias, and
50 parent-offspring conflict) in driving differences in the evolution of sex chromosome gene
51 content? We conclude by highlighting how this framework provides a clear motivation
52 for combining bryophyte genomics with systematic natural history observations and
53 classical genetic analysis.

54

55 **The evolution of UV sex chromosomes from a hermaphroditic ancestor**

56 All eukaryotes alternate between haploid and diploid stages in their life cycle. Meiosis
57 reduces the genome to a haploid state, while fertilization restores it to diploid.
58 Fertilization, of course, results from the fusion of two gametes, typically called sperm
59 and egg in anisogamous species. In species with separate males and females, the
60 segregation of a chromosome pair often determines sex. Where in the life-cycle sexual
61 differentiation occurs varies among lineages [BOX 1]. In organisms with dioecy in the
62 diploid stage, the sex chromosomes are referred to as either XY or ZW, depending on
63 which sex is heterogametic, while in haploid systems the sex chromosomes are always
64 UV (Fig.1; (Bachtrog *et al.* 2011)). The sex chromosome systems share much in
65 common with one another, but the differences among them can shed light on important
66 evolutionary processes.

67 Sex chromosomes are widely believed to evolve from ordinary autosomes, and
68 may first evolve in concert with dioecy. The evolution of genetically-determined dioecy
69 from a hermaphroditic ancestor requires linked male and female-sterility mutations

70 (Westergaard 1958; Charlesworth & Charlesworth 1978) (but see (Akagi *et al.* 2014;
71 Müller *et al.* 2020) for alternative routes). In the standard diploid formulation of this
72 model, a recessive, male-sterility mutation (i.e., one that renders a hermaphrodite
73 effectively a female) can increase in frequency in a hermaphroditic population when
74 inbred offspring are only half as fit as outbred offspring. That is, if inbreeding is
75 deleterious, male sterility may be favored because the increase in maternal fitness
76 compensates for the loss of fitness through paternity. This produces a gynodioecious
77 population containing females and hermaphrodites (Fig. 2A). Such populations are
78 widely known in angiosperms, but to our knowledge have not been experimentally
79 verified in bryophytes.

80 In a diploid gynodioecious population, a dominant, female suppressor (i.e., a
81 proto-Y chromosome) can increase in frequency, provided that a) it has compensatory
82 effects on male fitness and b) its inheritance is negatively correlated with the male-
83 sterility mutation (now the proto-X chromosome). The fixation of both mutations
84 produces a fully dioecious population with an XY sex chromosome system (Fig. 2A).
85 The conditions under which a female-sterility mutation evolves first, leading to an
86 androdioecious population, are more restrictive. In this case dioecy occurs following the
87 evolution of a dominant, male suppressor (i.e., a proto-W chromosome; Fig. 2A).
88 Recombination between the sterility factors produces some individuals that have both
89 male and female- sterility factors. Because these individuals cannot contribute to
90 subsequent generations, these circumstances favor the evolution of suppressed
91 recombination between the sterility loci, completing the transformation of an autosome
92 into a sex chromosome.

93 We should point out two important differences between the diploid and haploid
94 formulations of the two-locus model. First, both the androdioecy and gynodioecy
95 pathways lead to the evolution of UV sex chromosomes (Fig. 2B). Like diploid systems,
96 we expect that gynodioecious mating systems should be more frequent than
97 androdioecious systems, but we know of no rigorous evaluation of the frequencies of
98 such systems in any haploid mating species. Second, because the sexes are haploid,
99 dioecy in a UV system can evolve without a dominant male or female suppressor,
100 because dominance is irrelevant in a haploid system. Given that the evolution of a UV
101 system does not depend on the occurrence of rare dominant mutations, unlike both XY
102 and ZW systems, UV systems might evolve more readily than their diploid counterparts
103 (McDaniel *et al.* 2013; Villarreal & Renner 2013; Laenen *et al.* 2016). Interestingly, most
104 UV systems that have been studied to date are quite old (Ahmed *et al.* 2014; Bowman
105 *et al.* 2017; Carey *et al.* 2020).

106

107 **Symmetry in sex chromosome transmission alters evolutionary patterns**

108 The fundamental differences between XY/ZW and UV sex chromosomes stem from the
109 symmetrical transmission of U and V compared to the asymmetry of X and Y or Z and
110 W (Table 1). The most obvious implications of symmetry in transition are in the
111 expected amount of segregating variation on the sex chromosomes. In diploid systems,
112 each mated pair has three copies of an X or Z chromosome compared to four copies of
113 each autosome and only one Y or W. Thus, the baseline expected effective population
114 size (N_e) for a X or Z-linked locus is $\frac{3}{4}$ of an autosome and a Y or W-linked locus is $\frac{1}{4}$
115 (Bachtrog *et al.* 2011). In haploid-dioecious systems, each mated pair has one U and

116 one V for every two autosomes so both the U and the V chromosomes are expected to
117 have $\frac{1}{2} N_e$ of an autosome (McDaniel *et al.* 2013). Thus, in UV systems males and
118 females may have similar amounts of sex-linked genetic variation on which selection
119 can act, while in diploid systems the sexes are expected to have different amounts. We
120 should point out that demographic realities, including differential variance in
121 reproductive success between the sexes (Charlesworth 2009), for example due to
122 deviations from an equal ratio of males to females, or variance in female reproductive
123 output (Bengtsson & Cronberg 2009), may dramatically alter the patterns of N_e in sex
124 chromosomes relative to the autosomal expectation.

125 Regions of suppressed recombination, like sex chromosomes, also generally
126 have lower than expected levels of diversity due to selective sweeps and background
127 selection, which remove variation across these linked regions (reviewed in (Sayres
128 2018). In XY/ZW systems, suppressed recombination is confined to the male-specific Y
129 or the female-specific W chromosomes. In contrast, in UV systems, suppressed
130 recombination is present on both the female-specific U and male-specific V
131 chromosome (Table 1). Thus, both the male and female-specific chromosomes are
132 expected to experience an equivalent decrease in nucleotide diversity as consequence
133 of suppressed recombination, barring differences in mutation rate or the strength of
134 selection between the sexes. In addition to reduced genetic variation, suppressed
135 recombination can cause genes on the non-recombining chromosome to respond
136 slower to selection (positive or negative) (Comeron *et al.* 2008; Hough *et al.* 2017).
137 Consequently, such regions may show decreased codon bias (relative to autosomes)
138 and an increase in transposable element (TE) abundance, which in UV systems, is

139 expected to affect both equally, in contrast to the asymmetry in diploid systems (Table
140 1).

141 The decreased response to selection in non-recombining regions can promote
142 the accumulation of deleterious mutations and loss of functional genes, a process
143 referred to as degeneration (Charlesworth & Charlesworth 2000). In addition, because Y
144 and W chromosomes are always heterozygous with the X and Z, respectively, recessive
145 deleterious mutations are sheltered from purifying selection. In UV sex chromosomes,
146 because the diploids are always UV (not UU or VV; Fig. 1&2) the exposure to selection
147 is symmetrical between the male and female chromosome (Immler & Otto 2015).
148 Indeed, since the sex chromosomes are both expressed in haploids, UV sex
149 chromosomes overall should degenerate slower than either Y or W chromosomes
150 (Table 1; (Immler & Otto 2015)). Of course, a haploid-expressed gene could degenerate
151 on one of the sex chromosomes – for example, a gene specifically involved in sperm
152 production which is incorporated into the UV non-recombining region is likely to
153 experience strong purifying selection in males where its function is critical, but weak
154 purifying selection on the female U chromosome if its function is superfluous (Table 1).
155 Nevertheless, we have no a priori reason to expect that the U or V sex chromosome
156 should degenerate more than the other. The sparse data for UV systems largely support
157 these predictions. [BOX 2]

158 The lack of degeneration on UV sex chromosomes influences the potential for
159 dosage compensation, a common feature of XY/ZW systems. In diploid systems, the
160 homogametic sex has two copies of either an X or Z, and therefore two copies of sex-
161 linked genes to express. The heterogametic sex, in contrast, may have only one gene

162 copy if the Y or W-linked copies have been lost. The gene expression imbalance can
163 cause developmental problems or a variety of genetic disorders. Such problems are
164 averted by various forms of dosage compensation, in which one copy of many X-linked
165 genes is silenced in females, or the expression of X-linked genes is doubled in males. In
166 contrast, the sex chromosome composition of all life stages is balanced in the UV
167 system - the haploids are either U or V, and the diploids are homogeneous (Fig. 1).
168 While dosage compensation in diploid sex chromosomes is highly variable, and even
169 may be absent or incomplete (reviewed in (Mank 2013)), there is no expectation for any
170 such compensation in haploid systems.

171

172 **The role of genetic conflict in the expansion of UV chromosome systems**

173 The population genetic processes that we have discussed so far suggest that sex
174 chromosome evolution follows a well-circumscribed path. This view masks the diversity
175 in sex chromosome size and gene content that is found among, and even within many
176 lineages of eukaryotes (Bull 1983; Of Sex Consortium & Others 2014; Furman *et al.*
177 2020). Clearly degeneration may not play a major role in the evolution of gene content
178 on UV chromosomes, suggesting that other factors, potentially related to life history,
179 may explain variation in sex chromosome size. Surveying the UV species that have
180 been studied so far (Box 2), the proportion of the genome residing on the sex
181 chromosomes ranges from >1% in the brown algae *Ectocarpus*, ~1% in the green algae
182 *Volvox*, ~2% in the liverwort *Marchantia*, and as high as 30% of the genome size in the
183 moss *Ceratodon* (Ferris *et al.* 2010; Ahmed *et al.* 2014; Bowman *et al.* 2017; Carey *et*
184 *al.* 2020). It is possible that the variation in UV sex chromosome size is simply a

185 consequence of different patterns of degeneration, like the accumulation of TEs.
186 However, based on the data currently available each of these UV pairs shows a similar
187 proportion of shared versus sex-specific genes, suggesting that the chromosomes that
188 currently possess more genes have, in fact, also lost more genes from either the U or V.
189 Instead, the size of the sex-limited portion of the U and V chromosomes within a
190 species shows a remarkable correspondence with potential for new mutations to have
191 different fitness effects in males and females, based on the life history of each species
192 (Fig. 3).

193 If we consider the fate of a new mutation that increases the fitness of females
194 more than males (i.e., a sexually antagonistic allele), that mutation has a greater chance
195 of increasing in frequency if it is tightly linked to a U or W chromosome compared to an
196 autosome (Rice 1987). Conversely, a female-beneficial allele is more likely to fix in a
197 population if it is linked to a V or Y chromosome. Mutations that tighten the linkage
198 between sexually antagonistic alleles and the sex-determining region, such as
199 chromosomal inversions, translocations, or expansions of heterochromatin, also have a
200 greater chance of fixing. In spite of the intuitive appeal of this mechanism to explain the
201 evolution of sex linkage, the forces that drive the expansion of regions of suppressed
202 recombination on sex chromosomes remain poorly understood (Ironsides 2010). If sexual
203 antagonism is a major force driving the evolution of non-recombining sex chromosomes,
204 then species that experience more sexual antagonism should have more gene-rich sex
205 chromosomes.

206 Three major forms of genetic conflict can potentially shape the evolution of genes
207 on UV sex chromosomes, including sexual dimorphism, parent-offspring conflict, and

208 meiotic sex-ratio distortion. Some evidence supports the action of each of these forms
209 of conflict in species with UV systems. The prevalence of conflict is predicted to covary
210 with the breeding system and genetic diversity, both of which vary considerably among
211 bryophyte species. Although work on genetic conflict in UV systems is in its infancy, it is
212 clear that the forms of conflict may act synergistically, through both increasing
213 outbreeding levels and altering linkage patterns on the sex chromosomes. Here we
214 specifically explore how the prevalence of genetic conflict may drive variation in sex
215 chromosome content in bryophytes and other UV systems.

216

217 *Sexual dimorphism:* Perhaps the most obvious reason for sex chromosomes to grow
218 relates to their role in sexual dimorphism. Males and females achieve fitness through
219 different strategies (Bateman 1948; Robert 1972; Lessells & Parker 1999; Chapman
220 2006). Certainly genes related to sex-limited functions like sperm or egg production are
221 likely candidates for genes that could evolve sex linkage, although the number of genes
222 that are directly involved in gamete production may be relatively modest and is probably
223 similar among organisms. Nevertheless anisogamy can generate selection for sexual
224 dimorphism for a variety of additional traits between the sexes (Parker & Others 1979;
225 Bonduriansky *et al.* 2008).

226 The ubiquity of sexual dimorphism suggests selection frequently favors different
227 trait optima in males and females. For example, females of the moss *Ceratodon*
228 *purpureus* produce abundant volatile organic compounds (VOCs) while males produce
229 much less (Rosenstiel *et al.* 2012). Remarkably, the female VOCs attract moss sperm-
230 dispersing microarthropods (mites and springtails) in laboratory choice experiments,

231 suggesting that VOC production is part of a scent-based fertilization syndrome
232 analogous to flowering plant-pollinator mutualisms (Rosenstiel *et al.* 2012) Shortlidge et
233 al. submitted). Co-cultivating mosses and arthropods can increase both the number of
234 genotypes that reproduce and the overall number of sporophytes produced (Cronberg
235 *et al.* 2006); Shortlidge et al., submitted). The fact that males have lower VOC
236 production, however, suggests that scent-production may be costly. Thus, a mutation
237 that increases VOC production may be beneficial for females, but deleterious for males.
238 This sexual antagonism can be resolved by evolving either sex-linkage (i.e., moving the
239 genes VOC production to the sex chromosome, where they can evolve to their sex-
240 specific optima) or sex-biased gene expression, where the genes that control VOC
241 production may be expressed in different ways in males and females (Vicoso &
242 Charlesworth 2006; Ellegren & Parsch 2007). Sex-biased gene expression is very
243 common in animals, but the relative importance of sex-biased gene expression
244 compared to sex linkage in UV systems is unknown.

245 Sexual dimorphism is found in a wide variety of other traits in bryophytes,
246 suggesting that sexual antagonistic selection may drive the evolution of sex linkage. For
247 example, leaf size and juvenile growth in *C. purpureus* are sexually dimorphic
248 (McDaniel 2009) as are traits in the liverwort *Marchantia introflexa* (McLetchie &
249 Puterbaugh 2000; Fuselier 2008) and the brown algae *Ectocarpus* (Lipinska *et al.*
250 2015). The links between such trait variation and either male or female fitness are less
251 clear than for VOC production, but it remains a reasonable hypothesis that more
252 dimorphic species, or species with stronger male – male competition or female choice,
253 may possess more sex linked genes. We should point out that the relationship between

254 dimorphism and sex chromosome gene content may not be entirely straight-forward –
255 for example, several species of mosses, like the genera *Dicranum* and *Homalothecium*,
256 possess an extreme form of sexual dimorphism in which dwarf males grow epiphytically
257 on more typical sized females (Hedenäs & Bisang 2011; Rosengren & Cronberg 2014),
258 a form of sexual dimorphism that may actually decrease male-male competition.
259 Detailed studies relating the breeding system of a species to levels of sexual conflict are
260 fertile areas for research in bryophyte biology.

261

262 *Parent-offspring conflict over nutrient allocation:* The fact that males and females differ
263 in the amount of resources they allocate to reproduction also sets the stage for genomic
264 conflict (Arnqvist & Rowe 2005). Theory suggests conflict is particularly likely for
265 species that provision offspring through vivipary (Zeh & Zeh 2000). In embryophytes
266 (i.e., land plants), the embryo is retained on the haploid, female gametophyte and water
267 and nutrients are provided for a time through specialized transfer cells analogous to a
268 placenta. The males, in contrast, have no role in providing for offspring after fertilization.
269 A better provisioned embryo (sporophyte) is likely to make more and healthier spores,
270 increasing the fitness of both the male and female. However, in species where the
271 female can reproduce more than once, the allocation to one sporophyte may come at
272 the expense of other sibling sporophytes (Stark *et al.* 2009). Therefore, female fitness is
273 maximized by equal allocation to several offspring by modulating the transfer of
274 nutrients across the placenta (Haig 2013). In contrast, a male may not mate with the
275 same female again (nor any other female), so male fitness increases with every
276 additional viable spore that an offspring sporophyte produces. Thus, paternal genes in

277 the embryo are expected to evolve to extract as much of the female's resources as
278 possible to promote spore production (Haig 2013)). Recent data in *C. purpureus*
279 supports the parent-offspring hypothesis. In test crosses, the same female crossed to
280 two different males results in differences in sporophyte height, spore production, and
281 spore germination, suggesting the paternal genotype influences the transfer of nutrients
282 across the placenta (Shortlidge et al., submitted). Interestingly, the difference between
283 the interests of the maternal and paternal contributions becomes more intense as the
284 levels of outbreeding increase (Haig 2010), again highlighting the importance of the
285 breeding system for shaping genetic conflict.

286 We expect that sex chromosomes may be enriched in genes that influence the
287 flow of nutrients across the placenta from mother to offspring. In XY systems, only the
288 degenerate Y chromosome shows sex-limited inheritance, potentially minimizing its role
289 in parent-offspring conflict. However, autosomal genes that influence maternal transfer
290 may have parent-of-origin effects depending upon whether they are transferred from the
291 mother or father, an observation termed genomic imprinting (Reik & Walter 2001). What
292 is striking about UV systems is the unusual pattern of inheritance of the sex
293 chromosomes accomplishes what other systems do by genomic imprinting. Because
294 the U and V chromosomes do not recombine, their interests diverge considerably in the
295 developing UV-heterozygous embryo, providing a highly favorable environment for the
296 fixation of diploid-acting, sexually antagonistic alleles that influence the flow of nutrients
297 from the maternal (U) gametophyte to the offspring (UV) sporophyte. The U-linked
298 maternal genes within the embryo are genetically identical to its nurturing mother, and
299 therefore share a common interest, but the V-linked paternal genes in the embryo may

300 have different interests regarding the amount of offspring provisioning (Haig & Wilczek
301 2006). Formal tests linking parent-offspring conflict to the expansion of UV sex
302 chromosomes will likely involve a combination of placental transcriptomics, classical
303 genetics, and transgenic approaches.

304

305 *Sex-biased transmission ratio distortion*: A final form of sexual antagonism that is
306 common to all dioecious species with chromosomal sex determination is meiotic sex
307 ratio distortion. The meiotic sex ratio in a wide variety of eukaryotes deviates from the
308 expected 1:1 segregation of males and females (Lyttle 1993). At meiosis, any allele that
309 preferentially transmits itself to the next generation has an immediate benefit, thus
310 selection on these loci can be very strong because such alleles consistently have higher
311 frequency in offspring. In principle, any heterozygous locus could experience biased
312 transmission (Haig 2010). Distorter alleles may bias transmission by a wide variety of
313 mechanisms, but in UV systems distorters generally disable meiotic products carrying
314 the alternate allele. Mutations that result in more female offspring are likely to arise on,
315 or be inherited with, the U, while mutations that distorter the sex ratio in favor of males
316 will be associated with the V.

317 Most meiotic sex-ratio distorter systems typically consist of multiple distinct
318 genes. Distorter systems require a killer locus that halts development or otherwise
319 disables the products of meiosis, and a second locus which protects the distorting
320 chromosome from being disabled. Thus, a distorting U chromosome would have a killer
321 allele and an allele at the second locus that disarms the killer, often termed an
322 insensitive-responder allele. The V chromosome, in contrast, would lack the killer and

323 would carry a linked, sensitive-responder allele. Because the killer and insensitive-
324 responder must be inherited together, genomic regions that lack meiotic recombination,
325 like sex chromosomes, may be more likely to evolve distorter systems simply because
326 they contain more genes that can potentially evolve the killing and resistance functions.
327 As a meiotic killer system spreads, one sex could become rare, conditions which favor
328 the evolution of alleles at a third locus that suppress the killer allele, restoring fertility.
329 Although restorer alleles can arise on autosomes, the sex chromosomes are predicted
330 to be hot-spots for the evolution of restorer alleles, too. Thus, because they contain
331 large regions of suppressed recombination that have sex-limited inheritance, sex
332 chromosomes are likely to accumulate multiple genes that influence the outcome of
333 meiosis through either spore killing or fertility restoration.

334 A key difference between diploid and haploid mating systems is the action of a
335 meiotic distorter most often will result in spore death. Because each spore directly
336 contributes to forming the next generation of gametophytes, unlike sperm which
337 contribute indirectly through fertilization of an egg, spore death causes a loss of fitness.
338 Thus, the evolution of meiotic sex ratio distorters is likely to be held in check by the
339 fitness cost caused by spore-killing, in addition to the reduction in mating opportunity
340 cause by decreasing the frequency of one sex. Nevertheless, several spore killer
341 systems segregate in fungi (e.g., (Hammond *et al.* 2012)) and variation in spore death in
342 *C. purpureus* has a genetic basis (Shaw & Gaughan 1993; McDaniel *et al.* 2007; Norrell
343 *et al.* 2014). Numerous questions remain regarding the true fitness costs of meiotic sex-
344 ratio distortion, life history traits that promote the evolution of meiotic distorters, and the

345 potential interactions among different forms of genetic conflict in shaping the gene
346 content of the male and female sex chromosomes.

347

348 **The central role of natural history in evolutionary genomics**

349 In this review, we have argued that the wide variation in sex chromosome size and gene
350 content may be explained by variation in levels of genetic conflict, and further that
351 bryophytes provide the necessary variation to test this hypothesis. In UV systems,
352 suppressed recombination is not confounded with sex-specific mutational biases, sex-
353 specific inheritance, and sexual selection, making bryophytes well-suited for studying
354 how conflicts affect both male and female sex chromosomes. Clearly this effort will be
355 bolstered by sequencing additional bryophyte genomes, and we anticipate that soon we
356 will have a clearer picture of the gene content and evolutionary history of an increasing
357 number of sex chromosome systems. In addition, generating sex-linked markers will
358 open numerous research avenues in many previously genetically intractable systems,
359 enabling us to understand many sex-specific demographic phenomena (dispersal,
360 growth, mating success, male-male competition, female choice, biased mortality, or
361 effective population size), in addition to more effectively designing studies of sexual
362 dimorphism, parent-offspring conflict, and meiotic sex-ratio variation.

363 Equally important, however, is the role that natural history and phylogenetics can
364 play in contextualizing the results of genomic analyses. Life history features relating to
365 patterns of mating in nature, in particular, may be central to modulating the role of
366 genetic conflict in genome evolution. Bryophytes are one of the few terrestrial groups
367 that contain large proportions of both hermaphrodites and species with separate sexes.

368 This circumstance makes bryophytes unusually well suited for linking sex chromosome
369 history and molecular evolution to patterns of diversification. How often is speciation
370 associated with a change in mating system? Are sex chromosomes more likely than
371 autosomes to be exchanged among distant populations, or more likely to host loci that
372 promote reproductive isolation? Does genetic conflict promote elevated rates of
373 speciation (Crespi & Nosil 2013), or extinction (Price *et al.* 2010; Werren 2011), and
374 under what conditions? Critically, bryophytes possess underexplored life history and
375 taxonomic variation that may be used to draw general solutions to key questions
376 regarding the long-term evolution of sex chromosomes across eukaryotes.

377

378 **BOX 1: UV sex chromosomes are found in haploid-dioecious life cycles**

379 In animals and vascular plants with genetically-determined separate sexes, sexual
380 differentiation occurs in the diploid stage of the life cycle. Therefore, the sex of an
381 individual is determined by the genotype at fertilization. In the XY system, the fusion of
382 Y-bearing sperm with an X-bearing egg produces a male zygote (the heterogametic
383 sex), while the fusion of an X-bearing sperm with an X-bearing egg produces a
384 homogametic, female zygote. In the ZW system, in contrast, the female is the
385 heterogametic sex, meaning it produces Z or W-bearing eggs, which fuse with Z-
386 bearing sperm. In these diploid systems, the non-recombining Y and W chromosomes
387 are sex-specific (i.e., transmitted only through the male or female line, respectively),
388 while the X and Z are shared between both sexes and freely recombine, much like an
389 autosome, in the homogametic sex. In both the XY or ZW systems, sexual
390 differentiation is a diploid phenomenon.

391 In other multicellular lineages, including the Phaeophytes (brown algae),
392 Rhodophytes (red algae), Chlorophytes (Ulvophyceae and Chlorophyceae green algae),
393 and Bryophytes (mosses, liverworts, and hornworts), sex determination occurs in the
394 haploid stage of the life cycle. As such, only one copy of a sex chromosome is present
395 during sex expression, with individuals inheriting the U correlating with females and V
396 with males. Indeed, since the sexes are already haploid, gametes are produced by
397 mitosis not meiosis (Fig. 1). Fertilization of a U-linked egg and a V-linked sperm
398 produces a zygote. Importantly, the zygote is monomorphic (i.e., non-sexed) and
399 heterozygous at the sex determining factor - homogametic UU or VV individuals are
400 never formed. At maturity the diploid individual, the U and V pair at meiosis and
401 segregate independently into females and males (Bachtrog *et al.* 2011). Therefore, in
402 UV systems both the U and V are sex-specific.

403 The U and V chromosomes are in some ways similar to mating-type (MT) loci
404 found in fungi, algae, and other protists (e.g., (Ferris *et al.* 2010; Bazzicalupo *et al.*
405 2019). In these organisms, however, the gametes bearing alternate MT are not
406 phenotypically distinct from one another (i.e., isogamous) and therefore do not have
407 defined sexes (Fig. 3). Thus, while MT loci provide valuable systems for studying the
408 evolution of suppressed recombination in the absence of anisogamy (Branco *et al.*
409 2017; Bazzicalupo *et al.* 2019), we restrict our focus here to true UV sex chromosomes.

410 **BOX 2: Genomic analyses of UV systems**

411 A growing number of genomic analyses of UV sex chromosomes systems (reviewed in
412 (Coelho *et al.* 2018)) provide a foundation for comparative analyses between haploid

413 and diploid sex chromosome systems (Table 1). One of the best genomically studied
414 UV systems to date is the brown algae *Ectocarpus sp.* An ancestral-state reconstruction
415 of sexual condition in brown algae found dioecy evolved near the origins of this lineage
416 (Luthringer *et al.* 2014) and molecular evidence indicate *Ectocarpus* and the kelps
417 (Laminariales) may share a common UV system that evolved at least ~80-110 million
418 years ago (MYA) (Lipinska *et al.* 2017) The U and V chromosomes in *Ectocarpus sp.*
419 are relatively small (0.93 and 0.92 megabases (Mb) of the 205 Mb genome,
420 respectively); (Ahmed *et al.* 2014). The non-recombining portions of the U and V have
421 similar numbers of genes at 22 and 20, respectively, and share 11 homologous genes
422 (Ahmed *et al.* 2014). Avia *et al.* (2018) found the *Ectocarpus* U and V-linked loci had an
423 N_e about $\frac{1}{2}$ the autosomal average, suggesting these chromosomes experience levels
424 of selection similar to autosomal loci (Avia *et al.* 2018). As a result of suppressed
425 recombination, the sex chromosomes show decreased codon bias (Lipinska *et al.* 2017)
426 and increased TEs relative to autosomes in both sex-determining and pseudoautosomal
427 regions (Ahmed *et al.* 2014; Luthringer *et al.* 2015). Consistent with the effects of
428 degeneration, Lipinska *et al.* (2017) found evidence of gene loss from the ancestral
429 brown algae sex chromosome, in addition to gene gain and gene movement to the
430 autosomes (Lipinska *et al.* 2017).

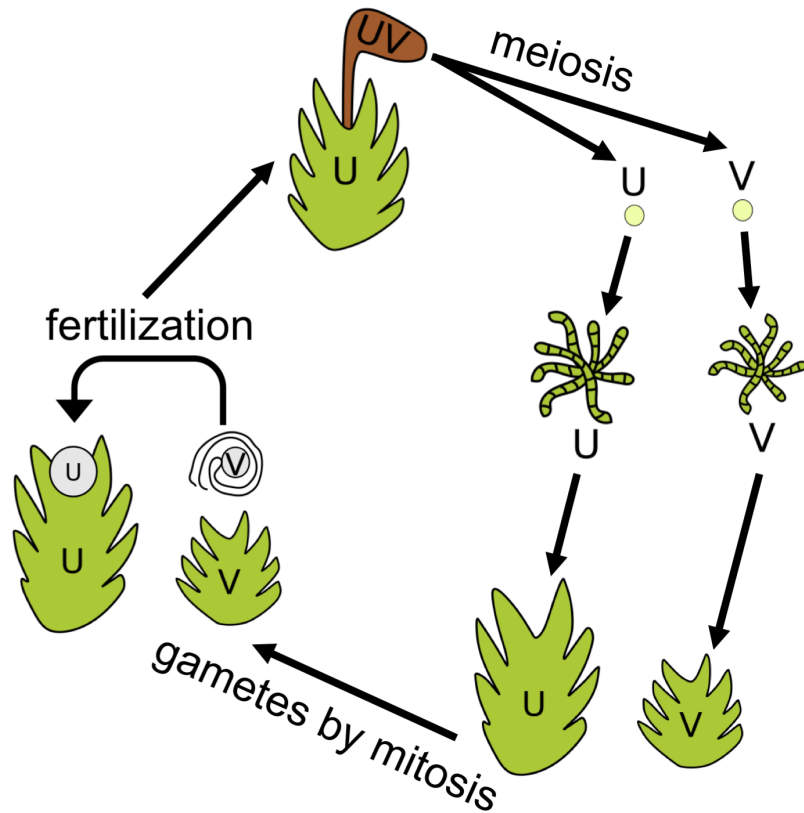
431 Within the chlorophyte green algae, at least two UV sex chromosome systems
432 evolved independently. Dioecy and MT loci evolved in the ancestor of the volvocine
433 algae, including *Gonium pectorale*, *Volvox carteri*, and *Chlamydomonas reinhardtii* at
434 least 200 MYA (Herron *et al.* 2009; Hamaji *et al.* 2016). Although, strictly speaking, only
435 *Volvox* possess anisogamy and therefore true UV sex chromosomes, we mention the

436 other species because the MT chromosomes and UV sex chromosomes are clearly
437 homologous. The *Volvox* U and V are 1.51 and 1.13 of the 131 Mb genome and also
438 have similar numbers of genes at 55 and 60, respectively, 50 of which are homologs
439 between the two (Ferris *et al.* 2010). Consistent with the effects of suppressed
440 recombination, the *Volvox* sex chromosomes have accumulated TEs (Ferris *et al.* 2010)
441 and comparisons between *Gonium*, *Chlamydomonas*, and *Volvox* found evidence for
442 gene loss (Hamaji *et al.* 2016). A second UV sex chromosome evolved in the lineage
443 including *Ulva partita* (Yamazaki *et al.* 2017). The 1.5 Mb *Ulva* U (referred to as MT+)
444 and 1.0 Mb V (referred to as MT-) have some asymmetry in gene numbers with 67 and
445 46 genes, respectively, and share 23 homologs. While the size of the *Ulva partita*
446 genome is currently unknown, the recent *Ulva mutabilis* genome is 98.5 Mb and *U.*
447 *partia* may be similar (De Clerck *et al.* 2018). The discovery of sex-linked PCR markers
448 in red algae (Guillemin *et al.* 2012), suggests that UV sex chromosomes independently
449 evolved in this group, too, but genomic analyses are currently lacking.

450 Within the bryophytes, more than half of the approximately 20,000 described
451 species are dioecious. Heteromorphic sex chromosomes have been observed using
452 cytological methods in 16 unrelated genera (Allen 1945; Renner *et al.* 2017) and sex-
453 linked molecular markers have been tested in several additional species (McLetchie &
454 Puterbaugh 2000; McDaniel *et al.* 2007; Korpelainen *et al.* 2008; Baughman *et al.*
455 2017). Ancestral-state reconstructions of sexual condition first showed that bryophyte
456 sex chromosomes may have ancient origins (McDaniel *et al.* 2013; Laenen *et al.* 2016)
457 and emerging phylogenomic data support in liverworts they evolved at least 400 MYA
458 and in mosses 300 MYA (Carey *et al.* 2020).

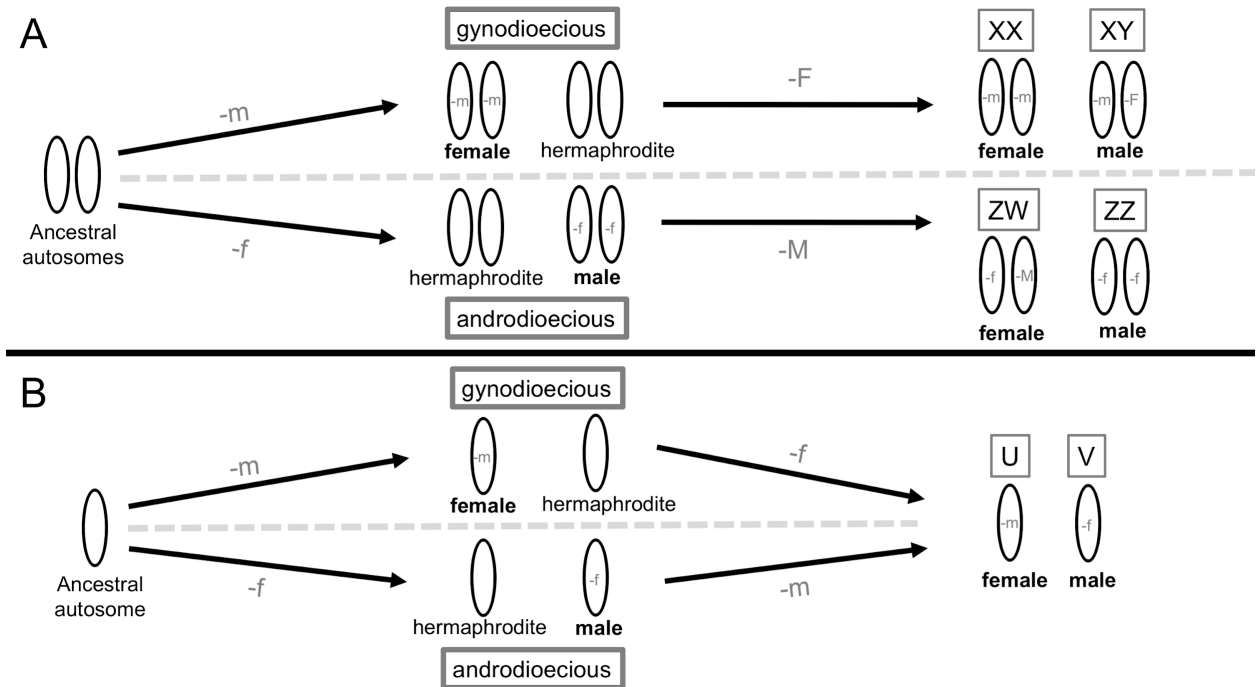
459 To date, the best studied bryophyte UV sex chromosomes are in the liverwort
460 *Marchantia polymorpha*. The 4.37 Mb U chromosome contains 74 genes and the 6 Mb
461 V contains 105 genes, with 20 shared homologs between the U and V, suggesting
462 some asymmetry in size and gene number between the sexes (total genome size 225.8
463 Mb; (Okada *et al.* 2001; Yamato *et al.* 2007; Bowman *et al.* 2017). The gene density in
464 the U and V-linked regions is ~5x lower than the autosomes, consistent with the
465 accumulation of TEs due to suppressed recombination (Bowman *et al.* 2017). Early data
466 in the moss *Ceratodon purpureus*, suggested the U and V sex chromosomes were
467 large, and the recent chromosome-scale genomes found they are each ~110 Mb of the
468 360 Mb genome and harbor hundreds over 3400 genes. (Voglmayr 2000; McDaniel *et*
469 *al.* 2007; Szövényi *et al.* 2015; Carey *et al.* 2020). Although, in contrast to *Ectocarpus*
470 (*Avia et al.* 2018), sex-linked genes in *C. purpureus* exhibited lower nucleotide diversity
471 than an panel of autosomal loci (McDaniel *et al.* 2013), suggesting selection or
472 demographic processes may shape these chromosomes.

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Figure 1. Life cycle of a dioecious moss. In mosses, the products of meiosis are haploid spores. For dioecious species, meiosis also segregates the U and V sex chromosomes in females and males, respectively. The spores germinate and produce protonemal filaments, upon which gametophores will develop. Because they are haploid, the male and female gametophores will produce gametes by mitosis. A V-containing sperm fuses with a U-containing egg, undergoing fertilization, and a diploid embryo (sporophyte) will develop on the maternal plant. The sporophyte embryo has both U and V sex chromosomes, but is considered a non-sexed stage, and within which the spores are produced.



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492 **Figure 2. Two-locus model for the evolution of XY, ZW, and UV sex**
 493 **chromosomes.** For diploid sex chromosomes (A) that originate from an ancestral pair
 494 of autosomes, a recessive male-sterility mutation (-m) first evolves, making a
 495 gynodioecious population (females and hermaphrodites). From the gynodioecious
 496 population a dominant female-sterility mutation (-F) arises making males. Once
 497 recombination is suppressed between these sex-determining loci, sex chromosomes
 498 are formed. This pathway leads to an XY sex chromosome system. If from an ancestral
 499 autosomal pair, a recessive female-sterility mutation (-f) evolves first, making an
 500 androdioecious population, followed by a dominant male-sterility mutation (-M), the
 501 result is a ZW sex chromosome system. In contrast in haploid sex chromosome
 502 evolution (B), if a male or female-sterility mutation evolves first, followed by sterility
 503 mutation of the opposite sex, the result is always a UV sex chromosome.

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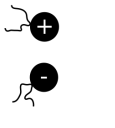


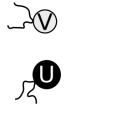


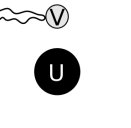

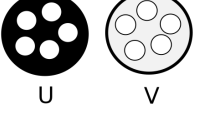
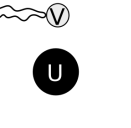
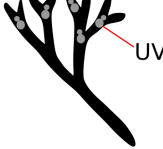
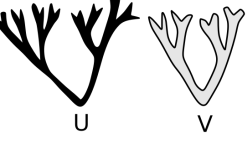

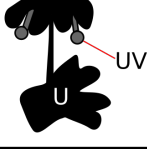




505 **Table 1. Comparison of XY, ZW, and UV characteristics.** Males (M); females (F);506 Effective population size (N_e); Transposable elements (TE)

Characteristic	X	Y	Z	W	U	V
Inheritance pattern	M and F	M	M and F	F	F	M
Hemizyosity	M	M	F	F	diploid phase	diploid phase
Expected N_e , relative to autosomes	3/4	1/4	3/4	1/4	1/2	1/2
Suppressed recombination	NA	Yes	NA	Yes	Yes	Yes
TE content, relative to autosomes	similar	higher	similar	higher	higher	higher
Gene density, relative to autosomes	similar	lower	similar	lower	lower	lower
Gene-content bias	NA, F, or M-biased	M-biased	NA, F, or M-biased	F-biased	F-biased	M-biased

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	Species	Gametes	Diploid	Haploid
Level of genomic conflict	<i>Chlamydomonas reinhardtii</i>			
	<i>Ectocarpus sp.</i>			
	<i>Volvox carteri</i>			
	<i>Gracilaria chilensis</i>			
	<i>Marchantia polymorpha</i>			
	<i>Ceratodon purpureus</i>			

536 **Figure 3. Predicted intensity of genomic conflict among UV systems.** The level of
537 genomic conflict is lower in mating-type loci because they do not have separate sexes,
538 so cannot be sexually dimorphic, have parent-offspring conflict, or sex-ratio distortion.
539 Across UV systems, the level of genomic conflict increases as the number or intensity of
540 conflicts increases. Sexual dimorphism tends to be low in algae and is generally
541 greater in magnitude in bryophytes. Parent-offspring conflict depends on the amount of
542 time the embryo remains nutritionally dependent on the maternal plant, which tends to
543 be longer in bryophytes. Sex-ratio distortion is possible in all systems, except perhaps in

544 mating-type loci. Although we note there are limited studies on these metrics across UV
545 systems and other closely-related species may have higher or lower levels of these
546 genomic conflicts.

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