

The Three Ts of Pathogen Evolution During Zoonotic Emergence

Elisa Visher^{1*}, Claire Evensen², Sarah Guth¹, Edith Lai³, Marina Norfolk⁴, Carly Rozins⁵, Nina A. Sokolov¹, Melissa Sui⁴, Michael Boots^{1,6}

1. Department of Integrative Biology, University of California, Berkeley, CA, 94720, USA
2. Mathematical Institute, University of Oxford, OX2 6GG, UK
3. College of Natural Resources, University of California, Berkeley, CA, 94720, USA
4. College of Letters & Sciences, University of California, Berkeley, CA, 94720, USA
5. Department of Science and Technology Studies, Division of Natural Science, York University, Toronto, Ontario, M3J 1P3, CA
6. Centre for Ecology and Conservation, College of Life and Environmental Sciences, University of Exeter, Penryn Campus, Penryn, TR10 9FE, UK

*Corresponding Author: elisa_visher@berkeley.edu

Author ORCID Information

EV: 0000-0003-3984-4748
CE: 0000-0002-2060-2362
SG: 0000-0001-5533-9456
EL: 0000-0002-0968-4547
MN: 0000-0001-6034-3417
CR: 0000-0003-1503-4871
NAS: 0000-0002-2920-3106
MS: 0000-0002-3422-2723
MB: 0000-0003-3763-6136

Abstract

When novel zoonotic diseases like Sars-CoV-2 emerge, they are likely to be poorly adapted to humans. Effective control measures will suppress transmission before significant evolution can occur, but extended transmission in human populations allows time for selection pressures to act. In this review, we discuss the factors shaping zoonotic pathogens' transmissibility and virulence at spillover and the selection pressures acting on these traits during emergence into human populations. We discuss how selection pressures during epidemics of emerging zoonotic disease are determined by the three Ts: trade-offs, transmission, and time scales. In short, virulence and transmission may trade-off, but transmission is likely to be favored by selection early in emergence. However, the relative selection pressures on transmission and virulence shift depending on the time scale of the epidemic. Predicting pathogen evolution in zoonoses therefore depends critically on understanding both the trade-offs of transmission-improving mutations and the time scales of selection.

42 **Introduction**

43 The current pandemic has emphasized that zoonotic emerging infectious diseases are
44 undeniably a grave public health concern (Woolhouse et al., 2012). Clearly, these diseases
45 necessitate rapid research upon emergence to uncover the pathogen's biology and modes of
46 transmission in order to develop diagnostics, public health recommendations, treatments, and
47 vaccines (Holmes et al., 2018). If successful, these interventions can stop transmission chains
48 and end the epidemic. If these interventions are unsuccessful, however, extended circulation in
49 humans can create selective pressures on these zoonotic pathogens (Plowright et al., 2017).
50 Therefore, in extended outbreaks, some attention should turn towards monitoring and
51 understanding potential pathogen evolution. Robust public health surveillance systems that
52 include viral sequencing can identify potential adaptive variants (Korber et al., 2020; Rambaut et
53 al., 2020) and evolutionary theory can help us understand how host, pathogen, and ecological
54 traits shape selective pressures to determine possible evolutionary outcomes (Bonneaud &
55 Longdon, 2020; Day et al., 2020).

56 Many recently emerged zoonotic pathogens have been viruses, particularly RNA viruses,
57 whose high mutation rates mean that multiple variants reach high frequencies early in epidemics
58 (Geoghegan et al., 2016). Most mutations in viruses have deleterious or neutral fitness effects
59 (Sanjuán, 2010), but the small proportion of mutations with beneficial fitness effects might be
60 particularly important for emerging zoonotic pathogens adapting to human hosts (Parrish et al.,
61 2008; Plowright et al., 2017). However, even when these beneficial mutations occur early in the
62 epidemic, they are slow to spread because selection pressures are weak relative to stochastic
63 factors like drift in small populations (MacLean et al., 2020). Potential adaptive variants can also
64 be difficult to identify because phylogenetic patterns are often complicated by human
65 demographic factors and founder effects (Villabona-Arenas et al., 2020). For example, a variant
66 of Ebola virus in the 2016 epidemic seemed to be associated with increased human

67 transmissibility in phylodynamic and *in vitro* assays, but did not correlate with higher viral titers or
68 shedding in macaques (Diehl et al., 2016; Marzi et al., 2018; Urbanowicz et al., 2016).

69 Despite these challenges, stories about mutations often spark public concern about
70 pathogens evolving to be more deadly, more transmissible, or to evade vaccines and treatments
71 (Grubaugh, Petrone, et al., 2020). Alternatively, historical theories of evolution towards avirulence
72 still pervade the public consciousness and sometimes lead to the prediction that the virus will
73 quickly evolve to become less dangerous (Smith, 1904). During the current Sars-CoV-2 epidemic,
74 reports of a mutational variant (D614G) increasing in frequency set off these debates in May
75 (Korber et al., 2020). Early responses cautioned against the overinterpretation of these reports
76 (Grubaugh, Hanage, et al., 2020; Grubaugh, Petrone, et al., 2020; MacLean et al., 2020;
77 Villabona-Arenas et al., 2020), but recent experiments in human cell culture and *in vivo* rodent
78 models have confirmed that this D614G variant may improve human transmission through higher
79 infectivity and replication in upper respiratory tissues (Hou et al., 2020; Plante et al., 2020). More
80 recently, the B.1.1.7 lineage with multiple spike mutations emerged in the UK and seems likely to
81 increase transmission rate (Rambaut et al., 2020). Despite these increases in transmission, the
82 D614G variant does not seem to be associated with changes in clinical severity (Volz et al., 2020)
83 and primary reports suggest that the B.1.1.7 lineage may not cause increased mortality either
84 (Davies et al., 2020). What is clear is that these examples show the potential for evolutionary
85 change during disease emergence.

86 Given the plausibility of Sars-CoV-2 adapting to improve human transmission and public
87 fascination with the topic, it is important that the broad scientific community have a clear
88 understanding of virulence evolution theory to quickly combat any false narratives. The study of
89 virulence and transmission evolution in epidemics of emerging infectious disease has been an
90 active but often separate area of research in both evolutionary virology and eco-evolutionary
91 theory (Cressler et al., 2016; Geoghegan & Holmes, 2018). In this review, we will integrate insights

92 from disease ecology, virology, computational genomics, and population genetics and eco-
93 evolutionary theory to form a more complete understanding of the factors shaping pathogen
94 evolution (Visher & Boots, 2020). We will discuss: how a pathogen's evolutionarily stable (long
95 term 'optimal') strategy depends on trade-off shape; what predicts pathogen virulence at the
96 spillover barrier; why selection pressures favor transmission improvements in maladapted
97 zoonotic pathogens; and how these selection pressures change over time during epidemics.
98 Through this, we describe predictions for pathogen evolution during epidemics of emerging
99 zoonotic disease and how they change depending on pathogen factors and host population
100 structure.

101

102 **Introduction to the Three Ts: Trade-offs, Transmission, and Time Scales**

103 The adaptive evolution of any trait depends on the presence of variation and the ability of
104 selection pressures to act on that variation. It is clear that pathogens, particularly RNA viruses,
105 can quickly generate and maintain large amounts of variation (Geoghegan & Holmes, 2017).
106 Selection pressures on these variants are weak compared to stochastic and demographic
107 pressures at the start of an epidemic, but gain strength as the number of infections increase
108 (MacLean et al., 2020). An extensive body of literature suggests that selection pressures on
109 virulence during epidemics of emerging zoonotic disease are determined by the three Ts: trade-
110 offs, transmission, and time scales (Anderson & May, 1982; Bull & Ebert, 2008; Day et al., 2020;
111 Lenski & May, 1994). See Figure 1 for graphical summary.

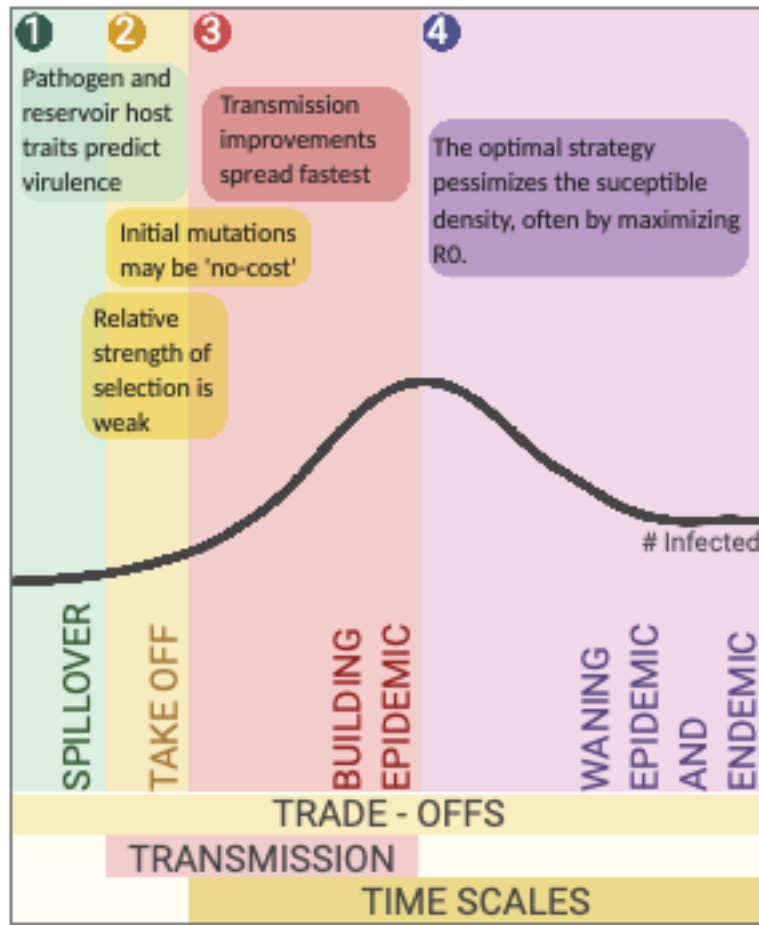
112 Theory has often assumed, and empirical data has increasingly shown us, that many
113 pathogen traits, like transmission and virulence, **trade-off** with each other (Acevedo et al., 2019;
114 Anderson & May, 1982; Cressler et al., 2016; Frank, 1996; Table 1). The **trade-off** theory is
115 important because it explains how different intermediate virulence, transmission, and recovery

116 rates can be optimal for a pathogen due to constraints between these key traits (Anderson & May,
117 1982; Cressler et al., 2016; Frank, 1996). It is often assumed that these trade-offs arise because
118 these traits all correlate with within-host pathogen replication rates, although this is not necessary
119 if symptoms directly correlate with transmission (Bonneaud et al., 2020). A large body of eco-
120 evolutionary theory has shown that the shapes of these trade-offs determine the pathogen's
121 optimum strategy and thus the direction of selection pressures (Anderson & May, 1982; Frank,
122 1996).

123 Emerging zoonotic pathogens typically do not have histories of selection in human
124 populations and thus are likely to be maladapted for human-to-human transmission (Warren &
125 Sawyer, 2019; Woolhouse et al., 2005). Meta-analyses of field data on recently emerged
126 pathogens can tell us patterns associated with a novel zoonotic pathogen's ability to transmit in
127 humans (Geoghegan et al., 2016; Guth et al., 2019; Olival et al., 2017). This can tell us about the
128 extent of maladaptation to humans and establishes a starting point for selection. In theory, this
129 maladaptation means that emerging zoonotic pathogens may initially have 'no-cost' mutations
130 available that improve transmission without impacting traits like virulence (Bull & Ebert, 2008). In
131 these cases, selection pressures for **transmission** improvements are likely to be most important
132 (Bull & Ebert, 2008).

133 Finally, transmission increases continue to be the most important selection pressure on
134 pathogens at the start of an epidemic, even when they trade-off with virulence (Lenski & May,
135 1994). Theory combining population genetics and eco-evolutionary approaches has shown how
136 the relative selection pressures on different pathogen traits shift as the density of infected and
137 susceptible hosts changes during an epidemic (Day & Proulx, 2004; Lenski & May, 1994).
138 Therefore, the pathogen's optimum strategy changes over **time** during an epidemic. Preliminary
139 evolutionary epidemiology modelling of Sars-CoV-2 has shown that evolution can vary depending

140 on these three Ts: trade-offs, transmission, and time scales (Day et al., 2020). We will describe
141 each of these in detail below.



142 Figure 1: The Three Ts of Pathogen Evolution During
143 Zoonotic Emergence

143

144 **The Virulence-Transmission Trade-Off Hypothesis**

145 Evolutionary biologists have long been entranced by the question of why pathogens harm
146 their hosts, or cause virulence (See Box 1) (Fenner & Ratcliffe, 1965). Based on the assumption
147 that host damage was always detrimental to parasite fitness, early ideas predicted that all
148 parasites should evolve towards avirulence (Alizon et al., 2009; Smith, 1904). This was
149 considered the 'conventional wisdom' until the 1980s, when foundational papers began to
150 appreciate that virulence might be linked to other parasite traits like transmission or recovery rate

151 and therefore could have an evolutionary optimum (Anderson & May, 1982; Ewald, 1983). Any
152 trade-offs between these traits would mean that low virulence could come at a cost of low
153 transmission or fast recovery and that avirulence would therefore hinder parasite fitness. This
154 virulence and transmission trade-off is now fundamental to our theories on pathogen evolution.

155 Theory on the virulence and transmission trade-off typically suggests that virulence and
156 transmission are both functions of the within-host exploitation or replication rate (Alizon et al.,
157 2009; Cressler et al., 2016). Because faster replicating pathogens generate larger population
158 sizes, they increase their transmission rate while causing more host damage (Cressler et al.,
159 2016; Frank, 1996). This damage increases host mortality, thereby decreasing the host's
160 infectious period and providing a shorter window for the infected host to contact susceptible hosts
161 (Anderson & May, 1982). In short, faster within-host replication increases the likelihood of
162 infection upon contact while decreasing the overall duration of infection (Anderson & May, 1982;
163 Frank, 1996). Under the trade-off hypothesis, parasites are therefore selected for exploitation
164 rates that balance virulence and transmission (Anderson & May, 1982; Cressler et al., 2016;
165 Frank, 1996).

166 Several other trade-offs have been proposed that don't depend on virulence and
167 transmission trading off through the within-host exploitation rate. A virulence-recovery trade-off
168 can occur if low replication rates make pathogens easier to clear such that lower virulence trades
169 off with faster recovery rates (Anderson & May, 1982). Alternatively, a transmission-recovery
170 trade-off can happen if the immune response is activated in a density dependent manner so that
171 high replication rates have high transmission, but fast recovery (Alizon, 2008). A sickness
172 behavior-transmission trade-off may happen if faster replication rates make the host feel sick and
173 isolate themselves so that high replication leads to higher transmission rates, but fewer contacts
174 (Ewald, 1994). Finally, the virulence and transmission trade-off does not necessarily depend on
175 changes to the within-host replication rate if symptoms themselves are needed for transmission

176 (Bonneaud et al., 2020). These alternative trade-offs can all still lead to selection for parasites to
177 balance their virulence or transmission metrics with other traits.

178 In simple host-parasite models, pathogens are selected to maximize the epidemiological
179 R_0 (i.e. the number of secondary infections that a parasite produces during its infectious period in
180 an entirely susceptible population) (See Box 2) (Anderson & May, 1982; but see Lion & Metz,
181 2018). The virulence-transmission trade-off predicts that these two traits are positively correlated,
182 but the shape of this relationship is critical to the predictions of evolutionary theory (Anderson &
183 May, 1982; Frank, 1996). When the trade-off is linear, pathogens evolve maximum virulence; but
184 when the trade-off is saturating (such that virulence is acceleratingly costly in terms of
185 transmission), pathogens will evolve towards an intermediate virulence (Alizon et al., 2009;
186 Anderson & May, 1982). Given the centrality of the trade-off hypothesis to our understanding of
187 virulence, it is noticeable that there are an increasing number of empirical studies that have found
188 support for the core idea (See Table 1) (Acevedo et al., 2019).

189 While virulence evolution has traditionally been discussed in terms of R_0 maximization, R_0
190 does not directly correlate with pathogen fitness. A more universal rule is that parasites are
191 selected following a pessimisation principle where the evolutionarily stable strategy is that which
192 can be sustained in the lowest quality environment (Lion & Metz, 2018; Metz et al., 2008; Mylius
193 & Diekmann, 1995). In virulence and transmission trade-off models, this is the strategy with the
194 lowest susceptible population at ecological equilibrium. Under the pessimisation principle, the key
195 insights of the trade-off hypothesis still hold in more varied, complex ecological circumstances
196 (Lion & Metz, 2018). If virulence trades off with other parasite fitness components, selection will
197 balance the negative fitness contributions of virulence with the positive fitness contributions of
198 traits like transmission (Lion & Metz, 2018).

199

Box 1. Defining virulence

A textbook definition of virulence is “Whereas ‘pathogenicity’ refers to the capacity of micro-organisms to cause disease, the essentially synonymous term virulence is generally used to note variations in degree. Virulence encompasses two features of an organism’s disease-producing capacity: infectivity (i.e., the ability to colonize and invade a host) and

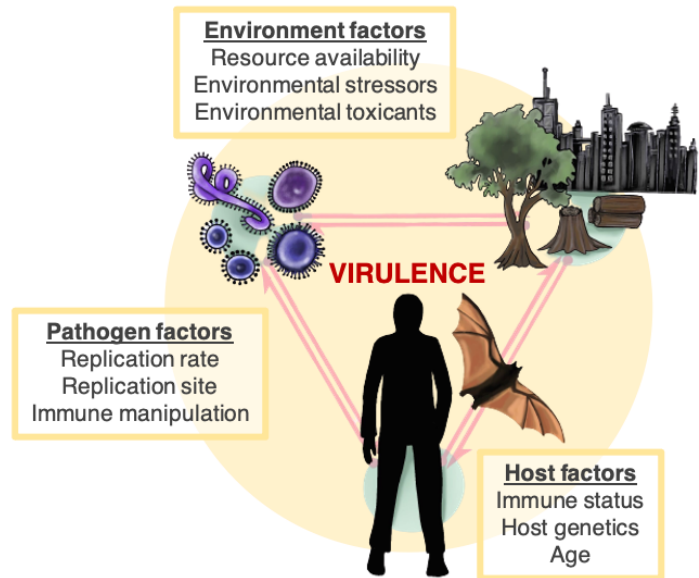


Figure 2: Disease Triangle of Virulence

severity of the disease that is produced” (Davis et al., 1990; Read, 1994). Different subfields, however, emphasize different parts of this definition with plant pathologists focusing more on infectivity and animal disease focusing more on severity. In the context of the virulence and transmission trade-off theory, virulence is defined more narrowly as the additional rate of mortality due to infection (Read, 1994). In these models, virulence is therefore a host outcome that is mediated by host, pathogen, and environmental traits. Host and pathogen traits involved in virulence are similar in human and other animal systems and include traits like host age and genetics and pathogen replication rate and immune manipulation. Environmental factors involved in causing virulence for humans include resource availability (including access to healthcare), exposure to environmental toxicants, and environmental stressors (including chronic stress from social inequities and racism) (Barber, 2020).

Box 2. Deriving R_0 maximization

We can look at a simplistic SI model to understand the math behind R_0 maximization.

First, we set up our system of equations for the host-parasite system before mutation.

$$\frac{dS}{dt} = b - \beta SI - dS$$

$$\frac{dI}{dt} = \beta SI - dI - \alpha I$$

In this system, we have natural birth (b) and death (d), density dependent transmission (β), and virulence (α), but no recovery.

We can then solve for the ecological equilibrium of this system.

$$(1) \quad S = \frac{b}{d}, \quad I = 0$$

$$(2) \quad S = \frac{d + \alpha}{\beta}, \quad I = \frac{b}{d + \alpha} - \frac{d}{\beta}$$

The first equilibrium is simply when there is no infection in the system, so we focus on the second. This second equation is the ecological equilibrium of the system infected only by the resident strategy.

Now, we want to conduct an invasion analysis asking what mutant values (m) can invade the ecological equilibrium set by the resident strategy (r).

$$\frac{dS}{dt} = b - \beta_r SI_r - \beta_m SI_m - dS$$

$$\frac{dI_r}{dt} = \beta_r SI_r - dI_r - \alpha_r I_r$$

$$\frac{dI_m}{dt} = \beta_m SI_m - dI_m - \alpha_m I_m$$

To see when the mutant can invade, we determine the stability of the mutant-free equilibrium,

$$S = \frac{d + \alpha_r}{\beta_r}, \quad I_r = \frac{b}{d + \alpha_r} - \frac{d}{\beta_r}, \quad I_m = 0$$

The equilibrium is not stable when an emerging rare mutant can increase in number. This gives the invasion criteria.

$$\frac{\beta_m}{d + \alpha_m} > \frac{\beta_r}{d + \alpha_r}$$

In this simplistic example, the invasion criteria may be familiar as a form of R_0 , the basic reproductive number. This means that the mutant with the highest R_0 can invade any population at equilibrium.

Table 1. Empirical tests of virulence evolution theory			
Paper	System	Virulence	Results
The virulence and transmission trade-off			
(Anderson & May, 1982)	<i>Oryctolagus cuniculus</i> / Myxoma virus	Mortality rate	R_0 was maximized at an intermediate virulence that had slower recovery and mortality rates
(Mackinnon & Read, 1999)	<i>Mus musculus</i> / <i>Plasmodium chabaudi</i>	Body mass loss and anemia	Virulence and transmission stage density are both positively correlated with replication rate
(Mackinnon & Read, 2004)	<i>Homo sapiens</i> / <i>Plasmodium falciparum</i>	Mortality rate	Parasite fitness peaks at intermediate virulence values with higher parasite replication and lower mortality
(Jensen et al., 2006)	<i>Daphnia magna</i> / <i>Pasteuria ramosa</i>	Time to host death in an obligately killing, castrating parasite	Transmission stage production peaked at intermediate virulence
(Fraser et al., 2007)	<i>Homo sapiens</i> / HIV-1	Duration of asymptomatic infection	R_0 peaks at intermediate viral set point load and virulence
(Roode et al., 2008)	<i>Danaus plexippus</i> / <i>Ophryocystis elektroscirrha</i>	Emergence and mating probability, adult lifespan and fecundity	Parasite lifetime fitness peaks at intermediate replication rates
(Atkins et al., 2013)	<i>Gallus gallus domesticus</i> / Marek's disease virus	Host lifespan	R_0 peaks at intermediate virulence
(Doumayrou et al., 2013)	<i>Brassica rapa</i> / Cauliflower mosaic virus	Symptom severity	Virulence and transmission show a positive, saturating relationship, but the relationship with replication rate is not clear
(Tardy et al., 2019)	<i>Haemorrhous mexicanu</i> / <i>Mycoplasma gallisepticum</i>	Body mass loss, symptom severity, and putative mortality rate	Virulence increases with parasite replication rate in isolates before, but not after host resistance evolution

(Bonneaud et al., 2020)	<i>Haemorrhous mexicanu</i> / <i>Mycoplasma gallisepticum</i>	Host mortality and symptom severity	R_0 peaks at intermediate virulence, even when the relationship between transmission and virulence is not dependent on replication rate
(Acevedo et al., 2019)	Meta-analysis of multiple systems		Strong evidence of increasing relationships between virulence and replication and transmission and replication
Virulence evolution during epidemics			
(Berngruber et al., 2013)	<i>Escherichia coli</i> / bacteriophage lambda	Horizontal transmission through lysis (rather than vertical)	Virulent, lytic phage is strongly favored during competition at the start of an epidemic, but latent virus outcompetes it as the epidemic progresses
Virulence evolution in spatially structured populations			
(Kerr et al., 2006)	<i>Escherichia coli</i> / T4 coliphage	Competitive ability and productivity	Prudent strategies dominate with spatially restricted migration, while virulent phages dominate with global migration
(Boots & Meador, 2007)	<i>Plodia interpunctella</i> / granulosis virus	Proportion of hosts infected in an obligate killer	Spatially structure selects for less infective, more prudent virus
(Berngruber et al., 2015)	<i>Escherichia coli</i> / bacteriophage lambda	Horizontal transmission through lysis (rather than vertical)	Latent, more prudent virus outcompetes lytic virus in spatially structured populations
Virulence evolution with environmental transmission			
(Ogbunugafor et al., 2013)	HeLa cells / vesicular stomatitis virus	Host cell death	There is a trade-off between transmission and the formation of environmentally persistent particles
(Wasik et al., 2015)	BHK cells / vesicular stomatitis virus	Plaque size	There is a trade-off between viral fecundity and the formation of environmentally persistent particles
(Walther & Ewald, 2004)	<i>Homo sapiens</i> / respiratory tract pathogens	Case fatality rate	Respiratory pathogens that survive longer in the environment are more virulent

203 **Virulence and transmission trade-offs acting at spillover**

204 As we have outlined, theory on the virulence and transmission trade-off is based upon the
205 idea that pathogens will be selected towards an optimal level of virulence within the host
206 populations to which they are adapted (Cressler et al., 2016). Recently emerged zoonotic
207 diseases do not have this evolutionary history with human populations and are therefore unlikely
208 to be at their evolutionary optimum when they first emerge (Guth et al., 2019; Mollentze et al.,
209 2020; Woolhouse et al., 2005). However, emerging pathogens may still be regulated by an
210 underlying virulence and transmission trade-off. In meta-analyses of recently emerged viral
211 zoonoses, excessively high virulence is associated with a lower R_0 (Brierley et al., 2016;
212 Geoghegan et al., 2016; Guth et al., 2019) and this negative association supports the theoretical
213 prediction that high virulence impedes pathogen fitness. Theory also predicts a cost to
214 excessively low virulence, an effect that is not supported in these analyses (Anderson & May,
215 1982; Guth et al., 2019). However, this could easily result from discovery bias because we are
216 unlikely to notice low-transmission zoonoses that cause only a few infections and have low
217 virulence (Bonneaud & Longdon, 2020). As such, there is little evidence to not expect emerging
218 diseases to be governed by trade-offs once they emerge into human populations.

219

220 **What predicts the virulence of disease when it first gets to humans?**

221 Emerging zoonoses vary widely in their virulence and transmission rates, but there are
222 some pathogen and reservoir host characteristics that are associated with the pathogen's
223 phenotype in humans (Geoghegan et al., 2016; Guth et al., 2019; Olival et al., 2017). In particular,
224 meta-analyses of recently emerged viral zoonoses have supported phylogenetic trends in
225 zoonotic potential (Guth et al., 2019). The phylogenetic distance between a pathogen's reservoir
226 host and humans predicts the pathogen's probability of being zoonotic (Olival et al., 2017),
227 virulence (Guth et al., 2019; Longdon et al., 2015), and R_0 in human populations (Geoghegan et

228 al., 2016; Guth et al., 2019). Mammalian hosts closely related to humans (e.g. primates) harbor
229 zoonoses associated with lower human mortality and higher capacity for transmission, while more
230 distantly related hosts (most notably, bats) harbor highly virulent zoonoses that appear to be
231 relatively maladapted for human-to-human transmission (Guth et al., 2019).

232 These phylogenetic trends can be understood if pathogens from distantly related reservoir
233 hosts have evolved replication strategies adapted to their reservoir host's more dissimilar
234 immunology, physiology, and ecology (Guth et al., 2019; Mollentze et al., 2020). There may also
235 be host orders with unique features in their biology beyond host dissimilarity that influence
236 pathogen traits in humans (Brook & Dobson, 2015). Specifically, bats seem to harbor unusually
237 virulent viruses (Brook & Dobson, 2015; Guth et al., 2019) which may, in part, result from their
238 high viral tolerance selecting for high replication rates (Brook et al., 2020).

239

240 **Transmission mode changes may cause shifts in virulence**

241 Zoonotic pathogens often have different virulence in human hosts than in their reservoir
242 hosts (Guth et al., 2019). However, pathogens that alter their transmission modes upon
243 emergence may be expected to have especially large shifts in virulence (Ewald, 1991). Most
244 pathogens have multiple possible modes of transmission, where their primary mode is determined
245 by factors like host social behavior and the environment (Antonovics et al., 2017). Pathogens can
246 undergo immediate shifts in transmission route upon emergence when human behavior promotes
247 the primary use of transmission routes not preferred in their reservoir hosts or when the receptors
248 that they bind to are located in different tissues (Antonovics et al., 2017). This can lead to
249 immediate shifts in virulence due to changes in pathogen inoculum size and anatomical site of
250 infection (Leggett et al., 2012; McMahon et al., 2018). Over longer evolutionary time scales,
251 different transmission pathways may create novel selection pressures on virulence (Ewald, 1991).

252 Thus, changes in a parasite's dominant mode of transmission during emergence can lead to both
253 immediate changes in and selection pressures for future changes in virulence.

254

255 **Virulence and transmission relationships are likely maladapted in emerging pathogens**

256 Pathogen virulence and capacity for transmission in humans loosely trade-off in a meta-
257 analysis of zoonotic viruses (Guth et al., 2019). Despite this trend, there is a substantial amount
258 of noise in the relationships between virulence and transmission. Some of this noise is likely due
259 to the fundamental complication of predictive evolution that each pathogen will have a unique
260 trade-off curve dependent on the nuances of its biology (Ebert & Bull, 2003). However, entirely
261 maladapted phenotypes also exist below the trade-off curve (Bull & Ebert, 2008; Shoval et al.,
262 2012). Simply, novel zoonotic pathogens can be bad at both transmission and virulence. Overall,
263 then, pathogens will vary in virulence and transmission because they have unique trade-off
264 shapes that predict different optimum values and because they can be maladapted below the
265 trade-off.

266 The concept of Pareto fronts describes such scenarios where phenotypes can be in the
267 region of sub-optimal phenotype space below the trade-off front (Shoval et al., 2012). The trade-
268 off front (or Pareto front) separates these accessible, maladapted phenotype combinations from
269 impossible, ideal phenotypes (Li et al., 2019; Shoval et al., 2012). In the sub-optimal region below
270 the Pareto front, improvements in one trait may not affect the other trait as simple adaptations
271 can be made before costs are incurred. Applied to virulence evolution, this concept means that
272 recently emerged diseases, even if broadly regulated by trade-offs, may select for no-cost
273 improvements in transmission that do not affect or can actually decrease their virulence (See
274 Figure 3a) (Bull & Ebert, 2008). The relationship between virulence and R_0 in recently emerged
275 zoonotic viruses seems to display such a Pareto front where phenotypes exist below, but not
276 above, a trade-off front (See Figure 3b).

277 For an illustrative example of how virulence and transmission can break trade-offs in
278 maladapted zoonosis, we can use H5N1 as a case study (Wasik et al., 2019). Concern over the
279 pandemic potential of highly pathogenic avian H5N1 influenza A led to two experimental evolution
280 studies examining the virus's ability to evolve respiratory droplet transmission in a ferret model
281 system (Herfst et al., 2012; Imai et al., 2012). Both studies found that the virus could evolve
282 respiratory droplet transmission, which would increase its transmission rate. However, this higher
283 transmission rate actually correlated with substantial decreases in virulence (Herfst et al., 2012;
284 Imai et al., 2012). This was because avian influenzas recognize a sialic acid (Sia α 2,3Gal) that is
285 found in ferrets' (and humans') lower respiratory tracts while human influenzas recognize a sialic
286 acid (Sia α 2,6Gal) found in the upper respiratory tract (Herfst et al., 2012). In the lab, H5N1 was
287 able to evolve Sia α 2,6Gal recognition and localize to the upper respiratory tract tissues that
288 allowed for droplet transmission. This change in replication site led to more efficient transmission,
289 but also lower host mortality despite no selection against virulence in the experiment (Herfst et
290 al., 2012; Imai et al., 2012). Notably for our understanding of the virulence and transmission
291 trade-off, these changes were dependent on changes in replication site, not replication rate. In
292 some sense, these no-cost transmission improvements could only happen because the virus was
293 so maladapted to mammalian hosts that it was using suboptimal binding sites. After these no-cost
294 adaptations brought the virus to the Pareto front, further adaptation would have to involve changes
295 to transmission and virulence that trade-off with each other through processes like replication
296 rates.
297

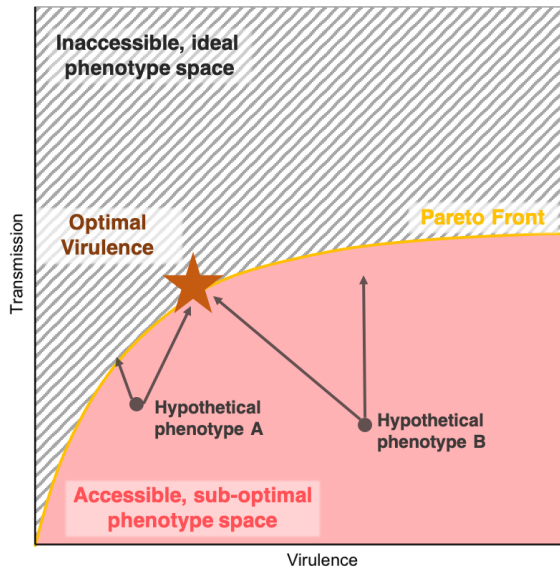


Figure 3a. Conceptual Diagram of the Pareto front between virulence and transmission. Possible phenotypes can be selected to improve transmission along any pathway within the accessible phenotype space. Since each pathogen's function determining their virulence and transmission trade-off varies, we cannot know where a hypothetical phenotype sits below the Pareto front. Selection for improved transmission can therefore involve decreases, no changes, or increases in virulence depending on the pathogen's starting point and mutational availability.

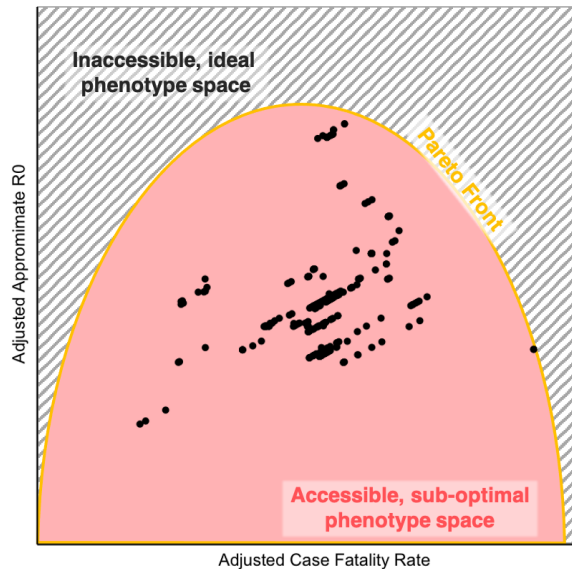


Figure 3b. Recently emerged viral zoonoses follow a Pareto front of virulence and reproductive capacity. Data is from a published dataset of recently emerged viral zoonoses from mammalian hosts (Guth et al. 2019). Approximate R0 is classified from 1 (no recorded human to human transmission) to 4 (endemic transmission) and adjusted by virus family and number of citations. Case fatality rate is adjusted by virus family and number of citations. Dots represent adjusted CFR and R0 values for individual epidemics of different viral zoonoses.

298

299

300 **Do we expect to see adaptive evolution of transmission and virulence in recently**
 301 **emerged diseases?**

302 While there are certainly selection pressures on recently emerged zoonotic pathogens,
 303 this does not necessarily mean that there will be adaptive evolution (Grubaugh, Petrone, et al.,
 304 2020; MacLean et al., 2020). A key tenant of evolutionary theory is that selection pressures must
 305 act through a background of stochasticity and drift to result in adaptive evolution (Crow & Kimura,
 306 2009). As small population sizes mean that both stochasticity and drift are relatively strong, the
 307 inevitably small population of infected individuals at the start of an epidemic means that these
 308 factors are likely to overwhelm selection and determine the spread of mutants (Hartl & Clark,
 309 1997).

310 Stochastic effects are additionally exasperated by the existence of founder effects during
311 epidemic range expansions resulting in spatial stochasticity analogous to genetic drift (Slatkin &
312 Excoffier, 2012). Thus, founder effects and variation in transmission due to host behavior and
313 stochasticity likely determine the fate of mutants at the start of epidemics (MacLean et al., 2020).
314 However, as the population size of infected individuals increases or if there are mutations of large
315 enough effect size, the balance between selection and stochasticity may shift towards selection
316 and result in adaptive evolution.

317 Finally, the adaptive evolution of acute, respiratory pathogens may additionally be
318 constrained by the small bottleneck sizes of transmission events, which also increase
319 stochasticity (McCrone et al., 2018; McCrone & Lauring, 2018). The normally short infectious
320 periods of acute diseases mean that only limited amounts of mutation and selection can occur
321 before transmission. Small bottleneck sizes mean that only a few genetic variants are transmitted.
322 Together, these factors mean that it is less likely for an acute, respiratory virus to have enough
323 time within a host to generate adaptive mutations and select on those variants strongly enough
324 for them to reach high enough frequencies to transmit through tight bottlenecks to other
325 individuals (McCrone et al., 2018). This can impede adaptive evolution at the population level
326 (Morris et al., 2020). This may mean that individuals with chronic infections are especially
327 important for adaptive evolution in acute, respiratory pathogens as they have longer infectious
328 periods that allow for the fixation of beneficial mutations (Rambaut et al., 2020; Xue et al., 2017).

329

330 **What are the selective pressures on transmission and virulence in recently emerged** 331 **diseases?**

332 Standard eco-evolutionary theory assumes that ecological and evolutionary time scales
333 are decoupled such that ecological equilibrium is reached before new mutants invade (Metz et
334 al., 1995). Epidemics are definitionally not at ecological equilibrium and high mutation rates mean

335 that new variants are likely to arise early in epidemics (Bull & Ebert, 2008). Therefore, the
336 assumptions of decoupled timescales must be relaxed to examine how selection pressures on
337 virulence and transmission change over the course of an epidemic (Lenski & May, 1994).

338

339 **Selection on virulence and transmission during epidemics**

340 Selection pressures on virulence and transmission during epidemics can be explored by
341 using models that do not assume separation of time scales, often using population genetic
342 approaches (Bolker et al., 2010; Bull & Ebert, 2008; Day & Gandon, 2007; Day & Proulx, 2004;
343 Lenski & May, 1994). These models allow for the existence of multiple simultaneous mutants so
344 that the competitive fitness of each can be assessed over shifting ecological conditions in time.
345 They show that strategies with higher transmission rates (betas) and virulence can be selected
346 during epidemic growth stages, despite R_0 optimized (intermediate virulence) strategies
347 dominating at endemic equilibrium (Day & Proulx, 2004; Lenski & May, 1994). This is because
348 strategies with higher transmission rates spread fastest at the start of the epidemic when the
349 density of susceptible hosts is high (Day & Proulx, 2004; Lenski & May, 1994).

350 Intuitively, these results can be explained as: an infected host during the early stages of
351 an epidemic encounters mostly susceptible hosts, so strains with higher transmission rates will
352 have faster population growth rates since they have shorter generation times than strains with
353 higher R_0 s (but lower transmission rates) that produce more secondary infections more slowly
354 over a longer infectious period. Therefore, improvements in transmission rate are the most
355 important at the start of an epidemic and can be selected for even if they increase virulence. This
356 also demonstrates that the high density of susceptible hosts early in epidemics crucially influences
357 selection pressures (Bull & Ebert, 2008; Cressler et al., 2016; Day & Proulx, 2004; Lenski & May,
358 1994).

359

360 **Selection on virulence and transmission in structured populations**

361 Simple virulence evolution trade-off theory assumes that transmission happens randomly
362 in a homogeneously mixing population (Cressler et al., 2016). However, natural populations
363 almost always have heterogeneous mixing patterns due to spatial structure and social networks
364 (Boots & Sasaki, 1999; van Baalen, 2002). In these structured populations, transmission mostly
365 happens between neighboring individuals. This can lead to ‘self-shading’ where highly infectious
366 strains rapidly deplete their local susceptible populations and compete for available hosts with
367 related strains (Boots & Sasaki, 1999; Boots & Sasaki, 2000; Lion & Boots, 2010). Both these
368 components of ‘self-shading’, the ecological clustering of infected individuals and the genetic
369 clustering of related strains, slow the rate of spread of highly infectious strains. On the other hand,
370 less infectious strains maintain higher local densities of susceptible individuals and have higher
371 onward transmission (Boots & Sasaki, 1999; Boots & Sasaki, 2000; Lion & Boots, 2010). Thus,
372 structured host populations select for lower pathogen infectivity and virulence at endemic
373 equilibrium.

374 However, the high availability of susceptible hosts at the start of an epidemic is likely to
375 reduce the impact of self-shading. Instead, we see that pathogens need to have higher
376 transmission rates to seed an epidemic in a spatially structured population than in a well-mixed
377 one (Keeling, 1999). Before ecological equilibrium, the invasion front of a spatially structured
378 epidemic also has a high local supply of susceptible hosts. This leads to a dynamic where virulent,
379 high beta strains are selected at the invasion front and then are succeeded by more prudent
380 strategies as the local dynamics approach equilibrium (Griette et al., 2015; Lion & Gandon, 2016).
381 Overall then, it is possible that structure in host populations temporarily selects for higher
382 virulence while the epidemic is spreading through mostly susceptible populations. However, if
383 there are also trade-offs where high virulence impedes host movement, then the spatial front of

384 the epidemic might instead have lower virulence (Hawley et al., 2013; Osnas et al., 2015). As
385 such, it is unclear how population structure and movement overall will select emerging pathogens.

386

387 **Selection on virulence with environmental transmission**

388 Simple virulence evolution trade-off theory assumes that pathogens only transmit by direct
389 contact between hosts. However, many pathogens also transmit through the environment
390 (Bonhoeffer et al., 1996; Ewald, 1983; Gandon, 1998; Kamo & Boots, 2004). The ‘curse of the
391 pharaoh’ hypothesis suggested that parasites can have higher virulence when they transmit
392 through the environment because transmission is not linked to the host’s infectious period (Ewald,
393 1983). However, at ecological equilibrium, environmental transmission can select for higher
394 virulence only if hosts can be multiply infected or if transmission can happen from environmental
395 pools after host death (Day, 2002; Day & Gandon, 2006; Gandon, 1998). Propagule survival in
396 spatially structured populations may actually increase self-shading and select for even lower
397 virulence (Kamo & Boots, 2004).

398 Importantly, environmental transmission also selects for higher virulence during the
399 epidemic stage if propagule dynamics are faster than host dynamics (Bonhoeffer et al., 1996).
400 This result holds even if hosts are singly infected and do not transmit after death because it
401 instead relates to the relative speed of pathogen population growth rates (Bonhoeffer et al., 1996;
402 Lenski & May, 1994). Under ‘curse of the pharaoh’, more virulent strategies with shorter infectious
403 periods will be more represented in the environmental reservoir and will therefore have higher
404 population growth rates when the susceptible density is high. Overall then, it is likely that
405 environmental transmission will select for higher virulence in epidemics (Bonhoeffer et al., 1996;
406 Day, 2002; Day & Gandon, 2006; Ewald, 1983; Gandon, 1998). However, it can be costly to make
407 environmentally persistent particles if they require more host resources or impede attachment to
408 host cells (Ogbunugafor et al., 2013). This can alter the dynamics of ‘curse of the pharaoh’ models

409 and potentially lead to bistability or branching resulting in sudden shifts and diversity in virulence
410 (Boldin & Kisdi, 2012; Caraco & Wang, 2008; Roche et al., 2011).

411

412 **Selection on virulence with antigenic escape**

413 Finally, simple virulence evolution trade-off theory assumes that recovered hosts are fully
414 immune such that host immunity does not wane and pathogens do not evolve to escape such
415 immunity. However, some, but not all, viral pathogens exhibit antigenic evolution to escape
416 neutralizing antibodies conferred by previous infections or vaccines (Drexler et al., 2014; Kennedy
417 & Read, 2017; Mclean, 1998; Rambaut et al., 2008; Wong et al., 2017). Notably though, selection
418 for antigenic or vaccine escape evolution is significantly slower and less efficient than for drug
419 resistance—likely due to differences in the timing and breadth of selective pressures (Debbink et
420 al., 2017; Kennedy & Read, 2017; Morris et al., 2020). When antigenic escape occurs, however,
421 it means that recovered individuals are newly susceptible to evolved strains and essentially
422 ‘resets’ the timescale of an epidemic by replenishing the density of susceptible hosts. This effect
423 had been postulated to transiently select for transmission-maximizing strategies with higher
424 virulence (Bull & Ebert, 2008), but has recently been shown to select for the long term persistence
425 of more acute, highly transmissible and virulent pathogens (Sasaki et al., 2021). However, the
426 mutations conferring antigenic escape likely trade-off with other pathogen traits like receptor
427 binding avidity, folding, and expression and therefore may constrain the possible virulence and
428 transmission phenotypes for such mutants (Greaney et al., 2020; Hensley et al., 2009).

429

430 **Other factors shaping selection on virulence**

431 Many other factors influence the evolution of virulence and have been reviewed elsewhere
432 (Cressler et al., 2016). In brief, multiple infection or co-infection may select for more virulent
433 pathogens due to within-host competition for resources (Alizon & van Baalen, 2008). Host

434 demographic features like immigration and density dependent mortality or fecundity may also alter
435 selection on virulence (Cressler et al., 2016). Additionally, host heterogeneities like age (Iritani et
436 al., 2019), genetic diversity (Osnas & Dobson, 2012; Regoes et al., 2000), and resistance
437 (Gandon, 2004) may select for virulence optimized on certain types of host.

438

439 **How might public health measures shape selection on virulence?**

440 The question of whether public health measures can purposely or inadvertently drive
441 pathogen evolution naturally arises when discussing virulence evolution. It is likely to be very
442 difficult to purposefully manage virulence evolution because it is so difficult to fully predict (Ebert
443 & Bull, 2003). For one, zoonotic pathogens can evolve in unpredictable ways if they start below
444 the Pareto front of the virulence and transmission trade-off. Additionally, selection pressures on
445 virulence are dependent on trade-offs that vary for each disease and moreover host population
446 characteristics that change rapidly. Finally, as we have discussed, selection pressures on
447 virulence are likely to be weak compared to stochastic effects at the start of epidemics.

448 Public health measures intentionally driving the evolution of virulence may therefore be
449 quixotic fantasies for emerging diseases. However, we can gain insight into how public health
450 measures can inadvertently select on virulence. Non-pharmaceutical public health interventions
451 for epidemics primarily aim to decrease transmission and therefore either stop the epidemic or
452 slow it until vaccines and treatments can be developed (Lai et al., 2020). This decreases the total
453 number of infected individuals, which will have the greatest impact on the total mortality burden
454 of any epidemic (Day et al., 2020).

455 However, some of these interventions may also contribute to the selection pressures
456 acting on the pathogen (Day et al., 2020; Ebert & Bull, 2003). First, increased environmental
457 sanitation raises the propagule death rate in the environment, thus potentially selecting for lower
458 pathogen virulence under the 'curse of the pharaoh' hypothesis (Bonhoeffer et al., 1996). Second,

459 decreased travel and extra-household contacts should alter the spatial and social structure of the
460 population to make a more structured transmission network (Boots & Sasaki, 1999). Third,
461 quarantine of symptomatic individuals may select for decreased or altered symptoms (Knell, 2004;
462 Saad-Roy et al., 2020). Finally, vaccines can sometimes create selection pressures on pathogens
463 with potential evolutionary impacts to consider (Kennedy & Read, 2020). However, recent models
464 have explored potential vaccine induced selection for proposed Sars-CoV-2 vaccines and suggest
465 that they are unlikely to select for higher virulence (Miller & Metcalf, 2020).

466 While the most human mortality will be prevented by simply preventing transmission,
467 considering the effects of control measures on virulence evolution can, in principle, lead to better
468 epidemic management (Day et al., 2020). Understanding host population characteristics creating
469 strong selection pressures for high transmission strategies could help distribute public health
470 effort if there are limited resources (Day et al., 2020). Importantly, weak epidemic control
471 measures that allow for extended transmission in humans increase the evolutionary potential of
472 zoonotic pathogens because they allow for stronger selection pressures and more mutations (Day
473 et al., 2020). Thus, the best evolutionary management practice for an epidemic of a zoonotic
474 infectious disease would be to suppress transmission using strong, rapid public health
475 interventions.

476

Box 3. Future Research Questions

There are several gaps in our understanding of the patterns and predictors of virulence evolution that require interdisciplinary, integrative approaches across often siloed subfields.

- What are the costs to transmission in human populations? Human pathogens are fairly rarely limited by host mortality, so models that purely define virulence as ‘the additional mortality rate due to infection’ are often inappropriate for virulence evolution in human

populations. This gap is currently hindering the uptake of trade-off theory amongst applied virology and medical fields (Bull & Luring, 2014) and several other trade-offs like time to recovery and sickness behavior may be more applicable for most human pathogens (Alizon, 2008; Anderson & May, 1982; Ewald, 1994). Effort should be put towards determining the empirical evidence for such trade-offs and exploring their consequences for evolutionary theory.

- What is the distribution of mutational fitness effects for pathogen traits like transmission and virulence in emerging zoonotic diseases and what are their trade-offs (or lack thereof)? Many studies of mutational fitness effects use simple metrics of fitness that do not capture how different components of fitness may trade-off (Sanjuán, 2010; Visher et al., 2016), but see (Greaney et al., 2020). Unmeasured evolutionary constraints and trade-offs may alter the distribution of mutational fitness effects outside of simple laboratory conditions (Visher & Boots, 2020). As selection may act on different fitness components differently at various time scales, being able to disentangle their relative contributions is important to build stronger predictive theory.
- How do selection pressures and their strengths relative to stochasticity vary across within host, between host, and population level scales? Many models do not include the heterogeneous selection pressures that arise from changes in pathogen population sizes at transmission, infection progression across tissues, temporally varying immune pressures, and heterogeneous hosts (Visher & Boots, 2020). Models that include these nuanced empirical circumstances can sometimes better describe population level evolutionary dynamics (Mideo et al., 2008; Morris et al., 2020; Ogbunugafor et al., 2010).

478 **Conclusion**

479 Novel zoonotic pathogens emerge into the human population maladapted to human hosts
480 and, although it is difficult to predict their emergence, pathogen and reservoir host traits can
481 loosely predict their virulence and transmission phenotypes (Guth et al., 2019; Olival et al., 2017).
482 Broadly, virulence is thought to trade-off with transmission leading to an optimal, intermediate
483 level of both. However, maladapted virulence and transmission phenotypes may start below the
484 Pareto front, so selection for transmission can have decoupled effects on virulence (Bull & Ebert,
485 2008). Selection pressures on pathogens are weak compared to drift during the highly stochastic
486 early epidemic period, but can become relevant if epidemic control fails and extended
487 transmission occurs (Hartl & Clark, 1997). A nuanced body of theory describes the selection
488 pressures acting on pathogen transmission and virulence (Cressler et al., 2016). These selection
489 pressures follow the three Ts—trade-offs, transmission, and time scales. Trade-offs between
490 traits mean that pathogens are selected to balance the benefits of transmission with the costs of
491 virulence, but the relative balance depends on the time scale of the epidemic (Anderson & May,
492 1982; Bull & Ebert, 2008). When the density of susceptible hosts is high early in the epidemic,
493 pathogens are selected for higher transmission rates even if they trade-off with higher virulence
494 (Bull & Ebert, 2008; Lenski & May, 1994). To predict how a pathogen’s virulence will evolve then,
495 we must understand the fitness impacts and trade-offs of transmission-improving mutations and
496 the population structure of the host (Day et al., 2020). This makes virulence evolution an
497 academically interesting topic with a rich body of theory surrounding it, but no universal
498 predictions (Ebert & Bull, 2003). However, this will not be a problem if strong, rapid public health
499 measures suppress transmission early on since this will both decrease the evolutionary potential
500 of such pathogens and decrease the total mortality burden by limiting the number of people
501 infected.

502

503 **Acknowledgements**

504 We would like to thank members of the Boots lab for helpful discussions. We would also like to
505 thank the many virologists and evolutionary biologists on twitter whose threads on viral evolution
506 helped shaped this paper.

507

508 **Author Contributions**

509 All authors researched and edited the paper. EV and MB conceptualized and wrote the paper.

510

511 **Data accessibility**

512 No novel data is used in this manuscript; data used is publicly available from (Guth et al., 2019).

513 The annotated R script used for analysis is in the supplementary materials.

514

515 **Funding**

516 EV, SG, and NAS acknowledge funding from NSF GRFP DGE 1752814 grants. EV and MN

517 acknowledge funding from the UC Berkeley SURF-SMART program. EV acknowledges funding

518 from the Philomathia Foundation Graduate Student Fellowship in the Environmental Sciences.

519 EV and MB acknowledge funding from NSF DEB 2011109. CR and MB acknowledges funding

520 from NIH/R01-GM122061-03.

521

522 **Competing Interests**

523 We declare no competing interests.

524

525

526

527

528

529

530

531 **Works Cited**

532

- 533 Acevedo, M. A., Dilleuth, F. P., Flick, A. J., Faldyn, M. J., & Elderd, B. D. (2019). Virulence-
534 driven trade-offs in disease transmission: A meta-analysis*. *Evolution*, *73*(4), 636–647.
535 <https://doi.org/10.1111/evo.13692>
- 536 Alizon, S., Hurford, A., Mideo, N., & Baalen, M. V. (2009). Virulence evolution and the trade-off
537 hypothesis: History, current state of affairs and the future. *Journal of Evolutionary*
538 *Biology*, *22*(2), 245–259. <https://doi.org/10.1111/j.1420-9101.2008.01658.x>
- 539 Alizon, Samuel. (2008). Transmission-Recovery Trade-Offs to Study Parasite Evolution. *The*
540 *American Naturalist*, *172*(3), E113–E121. <https://doi.org/10.1086/589892>
- 541 Alizon, Samuel, & van Baalen, M. (2008). Multiple Infections, Immune Dynamics, and the
542 Evolution of Virulence. *The American Naturalist*, *172*(4), E150–E168.
543 <https://doi.org/10.1086/590958>
- 544 Anderson, R. M., & May, R. M. (1982). Coevolution of hosts and parasites. *Parasitology*, *85*(2),
545 411–426. <https://doi.org/10.1017/S0031182000055360>
- 546 Antonovics, J., Wilson, A. J., Forbes, M. R., Hauffe, H. C., Kallio, E. R., Leggett, H. C., Longdon, B.,
547 Okamura, B., Sait, S. M., & Webster, J. P. (2017). The evolution of transmission mode.
548 *Philosophical Transactions of the Royal Society B: Biological Sciences*, *372*(1719),
549 20160083. <https://doi.org/10.1098/rstb.2016.0083>
- 550 Atkins, K. E., Read, A. F., Savill, N. J., Renz, K. G., Islam, A. F., Walkden-Brown, S. W., &
551 Woolhouse, M. E. J. (2013). Vaccination and Reduced Cohort Duration Can Drive
552 Virulence Evolution: Marek's Disease Virus and Industrialized Agriculture. *Evolution*,
553 *67*(3), 851–860. <https://doi.org/10.1111/j.1558-5646.2012.01803.x>
- 554 Barber, S. (2020). Death by racism. *The Lancet Infectious Diseases*, *20*(8), 903.
555 [https://doi.org/10.1016/S1473-3099\(20\)30567-3](https://doi.org/10.1016/S1473-3099(20)30567-3)
- 556 Berngruber, T. W., Froissart, R., Choisy, M., & Gandon, S. (2013). Evolution of Virulence in
557 Emerging Epidemics. *PLOS Pathogens*, *9*(3), e1003209.
558 <https://doi.org/10.1371/journal.ppat.1003209>
- 559 Berngruber, T. W., Lion, S., & Gandon, S. (2015). Spatial Structure, Transmission Modes and the
560 Evolution of Viral Exploitation Strategies. *PLOS Pathogens*, *11*(4), e1004810.
561 <https://doi.org/10.1371/journal.ppat.1004810>
- 562 Boldin, B., & Kisdi, É. (2012). On the evolutionary dynamics of pathogens with direct and
563 environmental transmission. *Evolution*, *66*(8), 2514–2527.
564 <https://doi.org/10.1111/j.1558-5646.2012.01613.x>
- 565 Bolker, B. M., Nanda, A., & Shah, D. (2010). Transient virulence of emerging pathogens. *Journal*
566 *of The Royal Society Interface*, *7*(46), 811–822. <https://doi.org/10.1098/rsif.2009.0384>
- 567 Bonhoeffer, S., Lenski, R. E., & Ebert, D. (1996). The curse of the pharaoh: The evolution of
568 virulence in pathogens with long living propagules. *Proceedings of the Royal Society of*
569 *London. Series B: Biological Sciences*, *263*(1371), 715–721.
570 <https://doi.org/10.1098/rspb.1996.0107>
- 571 Bonneaud, C., & Longdon, B. (2020). Emerging pathogen evolution. *EMBO Reports*, *21*(9),
572 e51374. <https://doi.org/10.15252/embr.202051374>

573 Bonneaud, C., Tardy, L., Hill, G. E., McGraw, K. J., Wilson, A. J., & Giraudeau, M. (2020).
574 Experimental evidence for stabilizing selection on virulence in a bacterial pathogen.
575 *Evolution Letters*, n/a(n/a). <https://doi.org/10.1002/evl3.203>

576 Boots, M., & Sasaki, A. (1999). "Small worlds" and the evolution of virulence: Infection occurs
577 locally and at a distance. *Proceedings of the Royal Society B: Biological Sciences*,
578 266(1432), 1933–1938.

579 Boots, M., & Sasaki, A. (2000). The evolutionary dynamics of local infection and global
580 reproduction in host-parasite interactions. *Ecology Letters*, 3(3), 181–185.

581 Boots, Michael, & Meador, M. (2007). Local Interactions Select for Lower Pathogen Infectivity.
582 *Science*, 315(5816), 1284–1286. <https://doi.org/10.1126/science.1137126>

583 Brierley, L., Vonhof, M. J., Olival, K. J., Daszak, P., & Jones, K. E. (2016). Quantifying Global
584 Drivers of Zoonotic Bat Viruses: A Process-Based Perspective. *The American Naturalist*,
585 187(2), E53-64. <https://doi.org/10.1086/684391>

586 Brook, C. E., Boots, M., Chandran, K., Dobson, A. P., Drosten, C., Graham, A. L., Grenfell, B. T.,
587 Müller, M. A., Ng, M., Wang, L.-F., & van Leeuwen, A. (2020). Accelerated viral dynamics
588 in bat cell lines, with implications for zoonotic emergence. *ELife*, 9, e48401.
589 <https://doi.org/10.7554/eLife.48401>

590 Brook, C. E., & Dobson, A. P. (2015). Bats as 'special' reservoirs for emerging zoonotic
591 pathogens. *Trends in Microbiology*, 23(3), 172–180.
592 <https://doi.org/10.1016/j.tim.2014.12.004>

593 Bull, J. J., & Ebert, D. (2008). Invasion thresholds and the evolution of nonequilibrium virulence.
594 *Evolutionary Applications*, 1(1), 172–182. [https://doi.org/10.1111/j.1752-](https://doi.org/10.1111/j.1752-4571.2007.00003.x)
595 4571.2007.00003.x

596 Bull, J. J., & Luring, A. S. (2014). Theory and Empiricism in Virulence Evolution. *PLOS*
597 *Pathogens*, 10(10), e1004387. <https://doi.org/10.1371/journal.ppat.1004387>

598 Caraco, T., & Wang, I.-N. (2008). Free-living pathogens: Life-history constraints and strain
599 competition. *Journal of Theoretical Biology*, 250(3), 569–579.
600 <https://doi.org/10.1016/j.jtbi.2007.10.029>

601 Cressler, C. E., McLeod, D. V., Rozins, C., Van Den Hoogen, J., & Day, T. (2016). The adaptive
602 evolution of virulence: A review of theoretical predictions and empirical tests.
603 *Parasitology*, 143(7), 915–930. <https://doi.org/10.1017/S003118201500092X>

604 Crow, J. F., & Kimura, M. (2009). *An Introduction to Population Genetics Theory*. Blackburn
605 Press.

606 Davies, N. G., Barnard, R. C., Jarvis, C. I., Kucharski, A. J., Munday, J., Pearson, C. A. B., Russell, T.
607 W., Tully, D. C., Abbott, S., Gimma, A., Waites, W., Wong, K. L., Zandvoort, K. van, Group,
608 C. C.-19 W., Eggo, R. M., Funk, S., Jit, M., Atkins, K. E., & Edmunds, W. J. (2020).
609 Estimated transmissibility and severity of novel SARS-CoV-2 Variant of Concern
610 202012/01 in England. *MedRxiv*, 2020.12.24.20248822.
611 <https://doi.org/10.1101/2020.12.24.20248822>

612 Davis, B. D., Dulbecco, R., & Eisen, H. N. (1990). *Microbiology*. Lippincott.

613 Day, T. (2002). Virulence evolution via host exploitation and toxin production in spore-
614 producing pathogens. *Ecology Letters*, 5(4), 471–476. [https://doi.org/10.1046/j.1461-](https://doi.org/10.1046/j.1461-0248.2002.00342.x)
615 0248.2002.00342.x

616 Day, T., & Gandon, S. (2006). Insights From Price's Equation into Evolutionary Epidemiology. In
617 Z. Feng, U. Dieckmann, & S. A. Levin (Eds.), *Disease Evolution: Models, Concepts, and*
618 *Data Analyses* (Vol. 13). American Mathematical Soc.

619 Day, T., & Gandon, S. (2007). Applying population-genetic models in theoretical evolutionary
620 epidemiology. *Ecology Letters*, *10*(10), 876–888. [https://doi.org/10.1111/j.1461-](https://doi.org/10.1111/j.1461-0248.2007.01091.x)
621 [0248.2007.01091.x](https://doi.org/10.1111/j.1461-0248.2007.01091.x)

622 Day, T., Gandon, S., Lion, S., & Otto, S. P. (2020). On the evolutionary epidemiology of SARS-
623 CoV-2. *Current Biology*, *30*(15), R849–R857. <https://doi.org/10.1016/j.cub.2020.06.031>

624 Day, T., & Proulx, S. R. (2004). A General Theory for the Evolutionary Dynamics of Virulence. *The*
625 *American Naturalist*, *163*(4), E40–E63. <https://doi.org/10.1086/382548>

626 Debbink, K., McCrone, J. T., Petrie, J. G., Truscon, R., Johnson, E., Mantlo, E. K., Monto, A. S., &
627 Lauring, A. S. (2017). Vaccination has minimal impact on the intrahost diversity of H3N2
628 influenza viruses. *PLOS Pathogens*, *13*(1), e1006194.
629 <https://doi.org/10.1371/journal.ppat.1006194>

630 Diehl, W. E., Lin, A. E., Grubaugh, N. D., Carvalho, L. M., Kim, K., Kyawe, P. P., McCauley, S. M.,
631 Donnard, E., Kucukural, A., McDonel, P., Schaffner, S. F., Garber, M., Rambaut, A.,
632 Andersen, K. G., Sabeti, P. C., & Luban, J. (2016). Ebola Virus Glycoprotein with Increased
633 Infectivity Dominated the 2013–2016 Epidemic. *Cell*, *167*(4), 1088–1098.e6.
634 <https://doi.org/10.1016/j.cell.2016.10.014>

635 Doumayrou, J., Avellan, A., Froissart, R., & Michalakis, Y. (2013). An Experimental Test of the
636 Transmission-Virulence Trade-Off Hypothesis in a Plant Virus. *Evolution*, *67*(2), 477–486.
637 <https://doi.org/10.1111/j.1558-5646.2012.01780.x>

638 Drexler, J. F., Grard, G., Lukashev, A. N., Kozlovskaya, L. I., Böttcher, S., Uslu, G., Reimerink, J.,
639 Gmyl, A. P., Taty-Taty, R., Lekana-Douki, S. E., Nkoghe, D., Eis-Hübinger, A. M., Diedrich,
640 S., Koopmans, M., Leroy, E. M., & Drosten, C. (2014). Robustness against serum
641 neutralization of a poliovirus type 1 from a lethal epidemic of poliomyelitis in the
642 Republic of Congo in 2010. *Proceedings of the National Academy of Sciences*, *111*(35),
643 12889–12894. <https://doi.org/10.1073/pnas.1323502111>

644 Ebert, D., & Bull, J. J. (2003). Challenging the trade-off model for the evolution of virulence: Is
645 virulence management feasible? *Trends in Microbiology*, *11*(1), 15–20.
646 [https://doi.org/10.1016/S0966-842X\(02\)00003-3](https://doi.org/10.1016/S0966-842X(02)00003-3)

647 Ewald, P. W. (1983). Host-Parasite Relations, Vectors, and the Evolution of Disease Severity.
648 *Annual Review of Ecology and Systematics*, *14*, 465–485. JSTOR.

649 Ewald, P. W. (1991). Transmission modes and the evolution of virulence. *Human Nature*, *2*(1),
650 1–30. <https://doi.org/10.1007/BF02692179>

651 Ewald, P. W. (1994). *Evolution of Infectious Disease*. Oxford University Press.

652 Fenner, F., & Ratcliffe, F. N. (1965). Myxomatosis. *Myxomatosis*.
653 <https://www.cabdirect.org/cabdirect/abstract/19662202443>

654 Frank, S. A. (1996). Models of Parasite Virulence. *The Quarterly Review of Biology*, *71*(1), 37–78.
655 <https://doi.org/10.1086/419267>

656 Fraser, C., Hollingsworth, T. D., Chapman, R., de Wolf, F., & Hanage, W. P. (2007). Variation in
657 HIV-1 set-point viral load: Epidemiological analysis and an evolutionary hypothesis.
658 *Proceedings of the National Academy of Sciences*, *104*(44), 17441–17446.
659 <https://doi.org/10.1073/pnas.0708559104>

660 Gandon, S. (1998). The curse of the pharaoh hypothesis. *Proceedings of the Royal Society B:*
661 *Biological Sciences*, 265(1405), 1545–1552.

662 Gandon, Sylvain. (2004). EVOLUTION OF MULTIHOST PARASITES. *Evolution*, 58(3), 455–469.
663 <https://doi.org/10.1111/j.0014-3820.2004.tb01669.x>

664 Geoghegan, J. L., & Holmes, E. C. (2017). Predicting virus emergence amid evolutionary noise.
665 *Open Biology*, 7(10), 170189. <https://doi.org/10.1098/rsob.170189>

666 Geoghegan, J. L., & Holmes, E. C. (2018). The phylogenomics of evolving virus virulence. *Nature*
667 *Reviews Genetics*, 19(12), 756–769. <https://doi.org/10.1038/s41576-018-0055-5>

668 Geoghegan, J. L., Senior, A. M., Giallonardo, F. D., & Holmes, E. C. (2016). Virological factors that
669 increase the transmissibility of emerging human viruses. *Proceedings of the National*
670 *Academy of Sciences*, 113(15), 4170–4175. <https://doi.org/10.1073/pnas.1521582113>

671 Greaney, A. J., Starr, T. N., Gilchuk, P., Zost, S. J., Binshtein, E., Loes, A. N., Hilton, S. K.,
672 Huddleston, J., Eguia, R., Crawford, K. H. D., Dingens, A. S., Nargi, R. S., Sutton, R. E.,
673 Suryadevara, N., Rothlauf, P. W., Liu, Z., Whelan, S. P. J., Carnahan, R. H., Crowe, J. E., &
674 Bloom, J. D. (2020). Complete Mapping of Mutations to the SARS-CoV-2 Spike Receptor-
675 Binding Domain that Escape Antibody Recognition. *Cell Host & Microbe*.
676 <https://doi.org/10.1016/j.chom.2020.11.007>

677 Griette, Q., Raoul, G., & Gandon, S. (2015). Virulence evolution at the front line of spreading
678 epidemics. *Evolution*, 69(11), 2810–2819. <https://doi.org/10.1111/evo.12781>

679 Grubaugh, N. D., Hanage, W. P., & Rasmussen, A. L. (2020). Making Sense of Mutation: What
680 D614G Means for the COVID-19 Pandemic Remains Unclear. *Cell*, 182(4), 794–795.
681 <https://doi.org/10.1016/j.cell.2020.06.040>

682 Grubaugh, N. D., Petrone, M. E., & Holmes, E. C. (2020). We shouldn't worry when a virus
683 mutates during disease outbreaks. *Nature Microbiology*, 5(4), 529–530.
684 <https://doi.org/10.1038/s41564-020-0690-4>

685 Guth, S., Visher, E., Boots, M., & Brook, C. E. (2019). Host phylogenetic distance drives trends in
686 virus virulence and transmissibility across the animal–human interface. *Philosophical*
687 *Transactions of the Royal Society B: Biological Sciences*, 374(1782), 20190296.
688 <https://doi.org/10.1098/rstb.2019.0296>

689 Hartl, D. L., & Clark, A. G. (1997). *Principles of population genetics* (Vol. 116). Sinauer associates.

690 Hawley, D. M., Osnas, E. E., Dobson, A. P., Hochachka, W. M., Ley, D. H., & Dhondt, A. A. (2013).
691 Parallel Patterns of Increased Virulence in a Recently Emerged Wildlife Pathogen. *PLOS*
692 *Biology*, 11(5), e1001570. <https://doi.org/10.1371/journal.pbio.1001570>

693 Hensley, S. E., Das, S. R., Bailey, A. L., Schmidt, L. M., Hickman, H. D., Jayaraman, A.,
694 Viswanathan, K., Raman, R., Sasisekharan, R., Bennink, J. R., & Yewdell, J. W. (2009).
695 Hemagglutinin Receptor Binding Avidity Drives Influenza A Virus Antigenic Drift. *Science*,
696 326(5953), 734–736. <https://doi.org/10.1126/science.1178258>

697 Herfst, S., Schrauwen, E. J. A., Linster, M., Chutinimitkul, S., de Wit, E., Munster, V. J., Sorrell, E.
698 M., Bestebroer, T. M., Burke, D. F., Smith, D. J., Rimmelzwaan, G. F., Osterhaus, A. D. M.
699 E., & Fouchier, R. A. M. (2012). Airborne transmission of influenza A/H5N1 virus
700 between ferrets. *Science (New York, N.Y.)*, 336(6088), 1534–1541.
701 <https://doi.org/10.1126/science.1213362>

702 Holmes, E. C., Rambaut, A., & Andersen, K. G. (2018). Pandemics: Spend on surveillance, not
703 prediction. *Nature*, 558(7709), 180–182. <https://doi.org/10.1038/d41586-018-05373-w>

704 Hou, Y. J., Chiba, S., Halfmann, P., Ehre, C., Kuroda, M., Dinnon, K. H., Leist, S. R., Schäfer, A.,
705 Nakajima, N., Takahashi, K., Lee, R. E., Mascenik, T. M., Graham, R., Edwards, C. E., Tse,
706 L. V., Okuda, K., Markmann, A. J., Bartelt, L., Silva, A. de, ... Baric, R. S. (2020). SARS-CoV-
707 2 D614G variant exhibits efficient replication ex vivo and transmission in vivo. *Science*.
708 <https://doi.org/10.1126/science.abe8499>

709 Imai, M., Watanabe, T., Hatta, M., Das, S. C., Ozawa, M., Shinya, K., Zhong, G., Hanson, A.,
710 Katsura, H., Watanabe, S., Li, C., Kawakami, E., Yamada, S., Kiso, M., Suzuki, Y., Maher, E.
711 A., Neumann, G., & Kawaoka, Y. (2012). Experimental adaptation of an influenza H5 HA
712 confers respiratory droplet transmission to a reassortant H5 HA/H1N1 virus in ferrets.
713 *Nature*, 486(7403), 420–428. <https://doi.org/10.1038/nature10831>

714 Iritani, R., Visher, E., & Boots, M. (2019). The evolution of stage-specific virulence: Differential
715 selection of parasites in juveniles. *Evolution Letters*, 3(2), 162–172.
716 <https://doi.org/10.1002/evl3.105>

717 Jensen, K. H., Little, T., Skorping, A., & Ebert, D. (2006). Empirical Support for Optimal Virulence
718 in a Castrating Parasite. *PLOS Biology*, 4(7), e197.
719 <https://doi.org/10.1371/journal.pbio.0040197>

720 Kamo, M., & Boots, M. (2004). The curse of the pharaoh in space: Free-living infectious stages
721 and the evolution of virulence in spatially explicit populations. *Journal of Theoretical*
722 *Biology*, 231(3), 435–441. <https://doi.org/10.1016/j.jtbi.2004.07.005>

723 Keeling, M. J. (1999). The effects of local spatial structure on epidemiological invasions.
724 *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 266(1421),
725 859–867. <https://doi.org/10.1098/rspb.1999.0716>

726 Kennedy, D. A., & Read, A. F. (2017). Why does drug resistance readily evolve but vaccine
727 resistance does not? *Proceedings of the Royal Society B: Biological Sciences*, 284(1851),
728 20162562. <https://doi.org/10.1098/rspb.2016.2562>

729 Kennedy, D. A., & Read, A. F. (2020). Monitor for COVID-19 vaccine resistance evolution during
730 clinical trials. *PLOS Biology*, 18(11), e3001000.
731 <https://doi.org/10.1371/journal.pbio.3001000>

732 Kerr, B., Neuhauser, C., Bohannan, B. J. M., & Dean, A. M. (2006). Local migration promotes
733 competitive restraint in a host–pathogen “tragedy of the commons.” *Nature*, 442(7098),
734 75–78. <https://doi.org/10.1038/nature04864>

735 Knell, R. J. (2004). Syphilis in renaissance Europe: Rapid evolution of an introduced sexually
736 transmitted disease? *Proceedings of the Royal Society B: Biological Sciences*, 271(Suppl
737 4), S174–S176.

738 Korber, B., Fischer, W. M., Gnanakaran, S., Yoon, H., Theiler, J., Abfalterer, W., Hengartner, N.,
739 Giorgi, E. E., Bhattacharya, T., Foley, B., Hastie, K. M., Parker, M. D., Partridge, D. G.,
740 Evans, C. M., Freeman, T. M., de Silva, T. I., Angyal, A., Brown, R. L., Carrilero, L., ...
741 Montefiori, D. C. (2020). Tracking Changes in SARS-CoV-2 Spike: Evidence that D614G
742 Increases Infectivity of the COVID-19 Virus. *Cell*, 182(4), 812–827.e19.
743 <https://doi.org/10.1016/j.cell.2020.06.043>

744 Lai, S., Ruktanonchai, N. W., Zhou, L., Prosper, O., Luo, W., Floyd, J. R., Wesolowski, A.,
745 Santillana, M., Zhang, C., Du, X., Yu, H., & Tatem, A. J. (2020). Effect of non-
746 pharmaceutical interventions to contain COVID-19 in China. *Nature*, 585(7825), 410–
747 413. <https://doi.org/10.1038/s41586-020-2293-x>

748 Leggett, H. C., Cornwallis, C. K., & West, S. A. (2012). Mechanisms of Pathogenesis, Infective
749 Dose and Virulence in Human Parasites. *PLOS Pathogens*, *8*(2), e1002512.
750 <https://doi.org/10.1371/journal.ppat.1002512>

751 Lenski, R. E., & May, R. M. (1994). The Evolution of Virulence in Parasites and Pathogens:
752 Reconciliation Between Two Competing Hypotheses. *Journal of Theoretical Biology*,
753 *169*(3), 253–265. <https://doi.org/10.1006/jtbi.1994.1146>

754 Li, Y., Petrov, D. A., & Sherlock, G. (2019). Single nucleotide mapping of trait space reveals
755 Pareto fronts that constrain adaptation. *Nature Ecology & Evolution*, 1–13.
756 <https://doi.org/10.1038/s41559-019-0993-0>

757 Lion, S., & Gandon, S. (2016). Spatial evolutionary epidemiology of spreading epidemics.
758 *Proceedings of the Royal Society B: Biological Sciences*, *283*(1841), 20161170.
759 <https://doi.org/10.1098/rspb.2016.1170>

760 Lion, Sébastien, & Boots, M. (2010). Are parasites “prudent” in space? *Ecology Letters*, *13*(10),
761 1245–1255. <https://doi.org/10.1111/j.1461-0248.2010.01516.x>

762 Lion, Sébastien, & Metz, J. A. J. (2018). Beyond R0 Maximisation: On Pathogen Evolution and
763 Environmental Dimensions. *Trends in Ecology & Evolution*, *33*(6), 458–473.
764 <https://doi.org/10.1016/j.tree.2018.02.004>

765 Longdon, B., Hadfield, J. D., Day, J. P., Smith, S. C. L., McGonigle, J. E., Cogni, R., Cao, C., &
766 Jiggins, F. M. (2015). The Causes and Consequences of Changes in Virulence following
767 Pathogen Host Shifts. *PLOS Pathogens*, *11*(3), e1004728.
768 <https://doi.org/10.1371/journal.ppat.1004728>

769 Mackinnon, M. J., & Read, A. F. (1999). Genetic Relationships Between Parasite Virulence and
770 Transmission in the Rodent Malaria Plasmodium Chabaudi. *Evolution*, *53*(3), 689–703.
771 <https://doi.org/10.1111/j.1558-5646.1999.tb05364.x>

772 Mackinnon, M. J., & Read, A. F. (2004). Virulence in malaria: An evolutionary viewpoint.
773 *Philosophical Transactions of the Royal Society B: Biological Sciences*, *359*(1446), 965–
774 986. <https://doi.org/10.1098/rstb.2003.1414>

775 MacLean, O. A., Orton, R. J., Singer, J. B., & Robertson, D. L. (2020). No evidence for distinct
776 types in the evolution of SARS-CoV-2. *Virus Evolution*, *6*(1).
777 <https://doi.org/10.1093/ve/veaa034>

778 Marzi, A., Chadinah, S., Haddock, E., Feldmann, F., Arndt, N., Martellaro, C., Scott, D. P., Hanley,
779 P. W., Nyenswah, T. G., Sow, S., Massaquoi, M., & Feldmann, H. (2018). Recently
780 Identified Mutations in the Ebola Virus-Makona Genome Do Not Alter Pathogenicity in
781 Animal Models. *Cell Reports*, *23*(6), 1806–1816.
782 <https://doi.org/10.1016/j.celrep.2018.04.027>

783 McCrone, J. T., & Luring, A. S. (2018). Genetic bottlenecks in intraspecies virus transmission.
784 *Current Opinion in Virology*, *28*, 20–25. <https://doi.org/10.1016/j.coviro.2017.10.008>

785 McCrone, J. T., Woods, R. J., Martin, E. T., Malosh, R. E., Monto, A. S., & Luring, A. S. (2018).
786 Stochastic processes constrain the within and between host evolution of influenza virus.
787 *ELife*, *7*, e35962. <https://doi.org/10.7554/eLife.35962>

788 Mclean, A. R. (1998). Vaccines and their impact on the control of disease. *British Medical*
789 *Bulletin*, *54*(3), 545–556. <https://doi.org/10.1093/oxfordjournals.bmb.a011709>

790 McMahan, D. P., Wilfert, L., Paxton, R. J., & Brown, M. J. F. (2018). Chapter Eight - Emerging
791 Viruses in Bees: From Molecules to Ecology. In C. M. Malmstrom (Ed.), *Advances in Virus*

792 *Research* (Vol. 101, pp. 251–291). Academic Press.
793 <https://doi.org/10.1016/bs.aivir.2018.02.008>
794 Metz, J. A. J., Geritz, S. A. H., Meszina, G., Jacobs, F. J. A., & Heerwaarden, J. S. van. (1995,
795 September). *Adaptive Dynamics: A Geometrical Study of the Consequences of Nearly*
796 *Faithful Reproduction* [Monograph]. <http://pure.iiasa.ac.at/id/eprint/4497/>
797 Metz, J. A. J., Mylius, S. D., & Diekmann, O. (2008, June). *When Does Evolution Optimise?*
798 [Monograph]. IR-08-013. <http://pure.iiasa.ac.at/id/eprint/8769/>
799 Mideo, N., Alizon, S., & Day, T. (2008). Linking within- and between-host dynamics in the
800 evolutionary epidemiology of infectious diseases. *Trends in Ecology & Evolution*, 23(9),
801 511–517. <https://doi.org/10.1016/j.tree.2008.05.009>
802 Miller, I. F., & Metcalf, C. J. E. (2020). No current evidence for risk of vaccine-driven virulence
803 evolution in SARS-CoV-2. *MedRxiv*, 2020.12.01.20241836.
804 <https://doi.org/10.1101/2020.12.01.20241836>
805 Mollentze, N., Streicker, D. G., Murcia, P. R., Hampson, K., & Biek, R. (2020). Virulence
806 mismatches in index hosts shape the outcomes of cross-species transmission.
807 *Proceedings of the National Academy of Sciences*, 202006778.
808 <https://doi.org/10.1073/pnas.2006778117>
809 Morris, D. H., Petrova, V. N., Rossine, F. W., Parker, E., Grenfell, B. T., Neher, R. A., Levin, S. A., &
810 Russell, C. A. (2020). Asynchrony between virus diversity and antibody selection limits
811 influenza virus evolution. *ELife*, 9, e62105. <https://doi.org/10.7554/eLife.62105>
812 Mylius, S. D., & Diekmann, O. (1995). On Evolutionarily Stable Life Histories, Optimization and
813 the Need to Be Specific about Density Dependence. *Oikos*, 74(2), 218–224.
814 <https://doi.org/10.2307/3545651>
815 Ogbunugafor, C. B., Alto, B. W., Overton, T. M., Bhushan, A., Morales, N. M., & Turner, P. E.
816 (2013). Evolution of Increased Survival in RNA Viruses Specialized on Cancer-Derived
817 Cells. *The American Naturalist*, 181(5), 585–595. <https://doi.org/10.1086/670052>
818 Ogbunugafor, C. B., Basu, S., Morales, N. M., & Turner, P. E. (2010). Combining mathematics
819 and empirical data to predict emergence of RNA viruses that differ in reservoir use.
820 *Philosophical Transactions: Biological Sciences*, 365(1548), 1919–1930. JSTOR.
821 Olival, K. J., Hosseini, P. R., Zambrana-Torrel, C., Ross, N., Bogich, T. L., & Daszak, P. (2017).
822 Host and viral traits predict zoonotic spillover from mammals. *Nature*, 546(7660), 646–
823 650. <https://doi.org/10.1038/nature22975>
824 Osnas, E. E., & Dobson, A. P. (2012). Evolution of Virulence in Heterogeneous Host Communities
825 Under Multiple Trade-Offs. *Evolution*, 66(2), 391–401. <https://doi.org/10.1111/j.1558-5646.2011.01461.x>
826 Osnas, E. E., Hurtado, P. J., & Dobson, A. P. (2015). Evolution of Pathogen Virulence across
827 Space during an Epidemic. *The American Naturalist*, 185(3), 332–342.
828 <https://doi.org/10.1086/679734>
829 Parrish, C. R., Holmes, E. C., Morens, D. M., Park, E.-C., Burke, D. S., Calisher, C. H., Laughlin, C.
830 A., Saif, L. J., & Daszak, P. (2008). Cross-Species Virus Transmission and the Emergence
831 of New Epidemic Diseases. *Microbiology and Molecular Biology Reviews : MMBR*, 72(3),
832 457–470. <https://doi.org/10.1128/MMBR.00004-08>
833 Plante, J. A., Liu, Y., Liu, J., Xia, H., Johnson, B. A., Lokugamage, K. G., Zhang, X., Muruato, A. E.,
834 Zou, J., Fontes-Garfias, C. R., Mirchandani, D., Scharton, D., Bilello, J. P., Ku, Z., An, Z.,

836 Kalveram, B., Freiberg, A. N., Menachery, V. D., Xie, X., ... Shi, P.-Y. (2020). Spike
837 mutation D614G alters SARS-CoV-2 fitness. *Nature*, 1–9.
838 <https://doi.org/10.1038/s41586-020-2895-3>

839 Plowright, R. K., Parrish, C. R., McCallum, H., Hudson, P. J., Ko, A. I., Graham, A. L., & Lloyd-
840 Smith, J. O. (2017). Pathways to zoonotic spillover. *Nature Reviews Microbiology*, 15(8),
841 502–510. <https://doi.org/10.1038/nrmicro.2017.45>

842 Rambaut, A., Loman, N. J., Pybus, O. G., Barclay, W., Barrett, J., Carabelli, A., Connor, T.,
843 Peacock, T., Robertson, D. L., Volz, E., & COVID-10 Genomics Consortium UK. (2020,
844 December 18). *Preliminary genomic characterisation of an emergent SARS-CoV-2 lineage*
845 *in the UK defined by a novel set of spike mutations*. Virological.
846 [https://virological.org/t/preliminary-genomic-characterisation-of-an-emergent-sars-cov-](https://virological.org/t/preliminary-genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-the-uk-defined-by-a-novel-set-of-spike-mutations/563)
847 [2-lineage-in-the-uk-defined-by-a-novel-set-of-spike-mutations/563](https://virological.org/t/preliminary-genomic-characterisation-of-an-emergent-sars-cov-2-lineage-in-the-uk-defined-by-a-novel-set-of-spike-mutations/563)

848 Rambaut, A., Pybus, O. G., Nelson, M. I., Viboud, C., Taubenberger, J. K., & Holmes, E. C. (2008).
849 The genomic and epidemiological dynamics of human influenza A virus. *Nature*,
850 453(7195), 615–619. <https://doi.org/10.1038/nature06945>

851 Read, A. F. (1994). The evolution of virulence. *Trends in Microbiology*, 2(3), 73–76.
852 [https://doi.org/10.1016/0966-842X\(94\)90537-1](https://doi.org/10.1016/0966-842X(94)90537-1)

853 Regoes, R. R., Nowak, M. A., & Bonhoeffer, S. (2000). Evolution of Virulence in a Heterogeneous
854 Host Population. *Evolution*, 54(1), 64–71. [https://doi.org/10.1111/j.0014-](https://doi.org/10.1111/j.0014-3820.2000.tb00008.x)
855 [3820.2000.tb00008.x](https://doi.org/10.1111/j.0014-3820.2000.tb00008.x)

856 Roche, B., Drake, J. M., & Rohani, P. (2011). The curse of the Pharaoh revisited: Evolutionary bi-
857 stability in environmentally transmitted pathogens. *Ecology Letters*, 14(6), 569–575.
858 <https://doi.org/10.1111/j.1461-0248.2011.01619.x>

859 Roode, J. C. de, Yates, A. J., & Altizer, S. (2008). Virulence-transmission trade-offs and
860 population divergence in virulence in a naturally occurring butterfly parasite.
861 *Proceedings of the National Academy of Sciences*, 105(21), 7489–7494.
862 <https://doi.org/10.1073/pnas.0710909105>

863 Saad-Roy, C. M., Wingreen, N. S., Levin, S. A., & Grenfell, B. T. (2020). Dynamics in a simple
864 evolutionary-epidemiological model for the evolution of an initial asymptomatic
865 infection stage. *Proceedings of the National Academy of Sciences*.
866 <https://doi.org/10.1073/pnas.1920761117>

867 Sanjuán, R. (2010). Mutational fitness effects in RNA and single-stranded DNA viruses: Common
868 patterns revealed by site-directed mutagenesis studies. *Philosophical Transactions of*
869 *the Royal Society B: Biological Sciences*, 365(1548), 1975–1982.
870 <https://doi.org/10.1098/rstb.2010.0063>

871 Sasaki, A., Lion, S., & Boots, M. (2021). The impact of antigenic escape on the evolution of
872 virulence. *BioRxiv*, 2021.01.19.427227. <https://doi.org/10.1101/2021.01.19.427227>

873 Shoval, O., Sheftel, H., Shinar, G., Hart, Y., Ramote, O., Mayo, A., Dekel, E., Kavanagh, K., & Alon,
874 U. (2012). Evolutionary trade-offs, Pareto optimality, and the geometry of phenotype
875 space. *Science*, 1217405.

876 Slatkin, M., & Excoffier, L. (2012). Serial Founder Effects During Range Expansion: A Spatial
877 Analog of Genetic Drift. *Genetics*, 191(1), 171–181.
878 <https://doi.org/10.1534/genetics.112.139022>

879 Smith, T. (1904). Some Problems in the Life History of Pathogenic Microorganisms. *Science*,
880 20(520), 817–832. <https://doi.org/10.1126/science.20.520.817>

881 Tardy, L., Giraudeau, M., Hill, G. E., McGraw, K. J., & Bonneaud, C. (2019). Contrasting evolution
882 of virulence and replication rate in an emerging bacterial pathogen. *Proceedings of the*
883 *National Academy of Sciences*, 116(34), 16927–16932.
884 <https://doi.org/10.1073/pnas.1901556116>

885 Urbanowicz, R. A., McClure, C. P., Sakuntabhai, A., Sall, A. A., Kobinger, G., Müller, M. A.,
886 Holmes, E. C., Rey, F. A., Simon-Loriere, E., & Ball, J. K. (2016). Human Adaptation of
887 Ebola Virus during the West African Outbreak. *Cell*, 167(4), 1079-1087.e5.
888 <https://doi.org/10.1016/j.cell.2016.10.013>

889 van Baalen, M. (2002). Contact Networks and the Evolution of Virulence. In *Adaptive Dynamics*
890 *of Infectious Diseases: In Pursuit of Virulence Management*. Cambridge University Press.

891 Villabona-Arenas, C. J., Hanage, W. P., & Tully, D. C. (2020). Phylogenetic interpretation during
892 outbreaks requires caution. *Nature Microbiology*, 5(7), 876–877.
893 <https://doi.org/10.1038/s41564-020-0738-5>

894 Visher, E., & Boots, M. (2020). The problem of mediocre generalists: Population genetics and
895 eco-evolutionary perspectives on host breadth evolution in pathogens. *Proceedings of*
896 *the Royal Society B: Biological Sciences*, 287(1933), 20201230.
897 <https://doi.org/10.1098/rspb.2020.1230>

898 Visher, E., Whitefield, S. E., McCrone, J. T., Fitzsimmons, W., & Luring, A. S. (2016). The
899 Mutational Robustness of Influenza A Virus. *PLOS Pathogens*, 12(8), e1005856.
900 <https://doi.org/10.1371/journal.ppat.1005856>

901 Volz, E., Hill, V., McCrone, J. T., Price, A., Jorgensen, D., O’Toole, Á., Southgate, J., Johnson, R.,
902 Jackson, B., Nascimento, F. F., Rey, S. M., Nicholls, S. M., Colquhoun, R. M., Filipe, A. da
903 S., Shepherd, J., Pascall, D. J., Shah, R., Jesudason, N., Li, K., ... Connor, T. R. (2020).
904 Evaluating the effects of SARS-CoV-2 Spike mutation D614G on transmissibility and
905 pathogenicity. *Cell*, 0(0). <https://doi.org/10.1016/j.cell.2020.11.020>

906 Walther, B. A., & Ewald, P. W. (2004). Pathogen survival in the external environment and the
907 evolution of virulence. *Biological Reviews*, 79(4), 849–869.
908 <https://doi.org/10.1017/S1464793104006475>

909 Warren, C. J., & Sawyer, S. L. (2019). How host genetics dictates successful viral zoonosis. *PLOS*
910 *Biology*, 17(4), e3000217. <https://doi.org/10.1371/journal.pbio.3000217>

911 Wasik, B. R., Bhushan, A., Ogbunugafor, C. B., & Turner, P. E. (2015). Delayed transmission
912 selects for increased survival of vesicular stomatitis virus. *Evolution*, 69(1), 117–125.
913 <https://doi.org/10.1111/evo.12544>

914 Wasik, B. R., de Wit, E., Munster, V., Lloyd-Smith, J. O., Martinez-Sobrido, L., & Parrish, C. R.
915 (2019). Onward transmission of viruses: How do viruses emerge to cause epidemics
916 after spillover? *Philosophical Transactions of the Royal Society B: Biological Sciences*,
917 374(1782), 20190017. <https://doi.org/10.1098/rstb.2019.0017>

918 Wong, A. H. M., Tomlinson, A. C. A., Zhou, D., Satkunarajah, M., Chen, K., Sharon, C., Desforges,
919 M., Talbot, P. J., & Rini, J. M. (2017). Receptor-binding loops in alphacoronavirus
920 adaptation and evolution. *Nature Communications*, 8(1), 1735.
921 <https://doi.org/10.1038/s41467-017-01706-x>

922 Woolhouse, M. E. J., Haydon, D. T., & Antia, R. (2005). Emerging pathogens: The epidemiology
923 and evolution of species jumps. *Trends in Ecology & Evolution*, 20(5), 238–244.
924 <https://doi.org/10.1016/j.tree.2005.02.009>

925 Woolhouse, M., Scott, F., Hudson, Z., Howey, R., & Chase-Topping, M. (2012). Human viruses:
926 Discovery and emergence. *Philosophical Transactions of the Royal Society B: Biological*
927 *Sciences*, 367(1604), 2864–2871. <https://doi.org/10.1098/rstb.2011.0354>

928 Xue, K. S., Stevens-Ayers, T., Campbell, A. P., Englund, J. A., Pergam, S. A., Boeckh, M., & Bloom,
929 J. D. (2017). Parallel evolution of influenza across multiple spatiotemporal scales. *ELife*,
930 6, e26875. <https://doi.org/10.7554/eLife.26875>
931