

1 A new frontier in molecular applications of ecological stoichiometry to understand global soil  
2 organic matter decomposition

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12 **Abstract.**

13 Coupled biogeochemical cycles drive ecosystem ecology by influencing individual-to-  
14 community scale behaviors; yet the development of integrative process-based models remains  
15 elusive. Soil organic matter (SOM) decomposition in particular is regulated by resource  
16 stoichiometry that dictates microbial nutrient acquisition ('ecological stoichiometry'). Ecological  
17 stoichiometry has revealed promising patterns of global ecosystem functions and informed  
18 process-based biogeochemical models. Despite its basis in biogeochemical modeling, ecological  
19 stoichiometry is largely absent from implementations of high-resolution microbial  
20 measurements, and the metabolic models they inform. Such fine-scale studies are critical  
21 components of larger scale models by developing transferrable relationships. One of the  
22 challenges to integrating models across molecular resolutions is that models of each scale use  
23 different underlying frameworks with few common threads to connect them. To address this  
24 challenge, we contend that ecological stoichiometry provides a framework for merging state-of-  
25 science biogeochemical models with microbial metabolic models to predict SOM decomposition.  
26 This article discusses new approaches to genome-enabled experiments and to models leveraging  
27 stoichiometric theory. We highlight two gaps that limit our understanding of SOM  
28 decomposition: (1) understanding how individual microorganisms alter metabolic strategies in  
29 response to substrate stoichiometry and (2) translating this knowledge to the scale of  
30 biogeochemical models. We suggest iterative information exchange to refine the objectives of  
31 high-resolution investigations and to specify limited dynamics for representation in large-scale  
32 models through integrated genome-enabled reaction networks. We propose that advancing  
33 technologies in the context of stoichiometric theory provides an untapped framework for

- 34 interpreting molecular data and further distilling this information into reduced complexity SOM  
35 decomposition models.  
36  
37 **Key words:** carbon use efficiency, carbon cycling, nitrogen cycling, ecosystem models, soil  
38 nutrients, microbiome

## 39 **Introduction.**

40           The world's soil ecosystems contain a pool of carbon (C, ~1500 Gt C) that is larger than  
41 vegetation and atmospheric stocks combined, and its destabilization has the potential to  
42 dramatically alter future climates (Crowther et al. 2019). Much of this belowground C is  
43 contained by soil organic matter (SOM). Soil organic matter decomposition is regulated by  
44 resource availability that determines the metabolic pathways used for microbial nutrient  
45 acquisition (Hungate et al. 2003; Wang & Houlton 2009; Zhang et al. 2014). It is rate-limited by  
46 microbial processes that are notoriously difficult to measure and predict beyond the scale of  
47 localized experimental plots (Bond & Lamberty et al. 2016; Naylor et al. 2020). While microbial  
48 processes can now be measured using state-of-science molecular approaches, and theoretically  
49 used to establish scalable principles, interpreting vast molecular data types using standard  
50 statistical approaches is challenging. Therefore, SOM decomposition at scales most relevant to  
51 climate change largely continues to be predicted through generalized environmental proxies such  
52 as moisture, temperature, mineralogy, and total soil C or nutrient pool sizes (Bailey et al. 2018).  
53 These parameters leave a substantial amount of uncertainty surrounding model predictions  
54 (Todd-Brown et al. 2014; Todd-Brown et al. 2013). The shortcomings of predicting SOM  
55 decomposition beyond localized scales point to a need to incorporate more understanding of  
56 scalable spatial and biochemical processes into SOM decomposition models (Todd-Brown et al.  
57 2013). Recent work has improved conceptual, empirical, and numerical representations of  
58 mineral protection and hydrologic mechanisms that regulate microbial access to SOM  
59 (Blankinship et al. 2018; Brangarí et al. 2020; Waring et al. 2020; Woolf & Lehmann 2019). In  
60 contrast, detailed representations of the microbial metabolisms that directly convert SOM into  
61 carbon dioxide are still missing from predictive models at the ecosystem-scale.

62 Progress in understanding the processes that drive SOM decomposition historically has  
63 advanced in parallel between biogeochemical and microbial research domains (Fig. 1). Recent  
64 attempts have represented fine- (individual microbial metabolisms and metabolites),  
65 intermediate- (groups of microorganisms and/or elemental pool composition), and coarse-  
66 resolution (multiple interacting microbial groups and/or elemental pool sizes) connectivity of  
67 microbial drivers and biogeochemical outcomes in predictive models at a particular level of  
68 resolution (Fig. 1 and 2). Yet, we lack integrative process-based models that link across  
69 molecular to ecosystem scales and can enable more robust predictions of SOM decomposition in  
70 next-generation models (Chowdhury et al. 2019). Translating the depth of information that now  
71 can be obtained by molecular approaches to the ecosystem-level is pressing line of scientific  
72 inquiry (Hall et al. 2018). A major challenge to this effort is that that models of each scale use  
73 different underlying frameworks such that there are few common threads with which to connect  
74 models.

75 While many biogeochemical models are built on the principles of ecological  
76 stoichiometry (i.e., the role of elemental composition in regulating ecological interactions,  
77 Sterner & Elser 2002; Van de Waal et al. 2018) through nutrient-regulated decomposition rates,  
78 ecological stoichiometry is largely absent from research using high-resolution molecular  
79 measurements that allow unprecedented insight into soil microbiomes. Ecological stoichiometry  
80 uses the elemental (e.g., C:H:N:O:P:S) imbalance between organisms and their substrates to  
81 predict that nutrient limitations universally control biological activity (Elser et al. 2000; Elser et  
82 al. 1996; Reiners 1986; Sterner 1995). It has been leveraged to decipher overarching patterns in  
83 the processes governing SOM decomposition (Billings & Ballantyne IV 2013) and is the guiding

84 framework for decades of biogeochemical investigations that provide the foundation for many  
85 process-based models.

86         Here, we argue that by using an ecological stoichiometry framework to combine new  
87 microbiological and computational tools with biogeochemistry, we have a currently unexploited  
88 opportunity to integrate predictive models across measurement scales and to better understand  
89 global patterns in microbial SOM decomposition. Our intent in this paper is not to  
90 comprehensively review all existing types of microbiological and biogeochemical models, rather  
91 to highlight an opportunity for future research while providing necessary background  
92 information. We propose the use of top-down and bottom-up information exchange guided by  
93 ecological stoichiometry to iteratively (1) refine the objectives of high molecular resolution  
94 investigations and (2) specify a limited set of dynamics for representation in large-scale models  
95 through integrated genome-enabled reaction networks (Fig. 3). Top-down approaches leverage  
96 bulk biogeochemical pool sizes to identify nutrient limitation (s) that then guide parameters  
97 selection and calibration of reduced complexity models. Bottom-up approaches use  
98 stoichiometric principles to guide the analysis and interpretation of large molecular datasets by  
99 establishing predictive rules that are translatable across scales. By using ecological stoichiometry  
100 as a conceptual framework to transfer knowledge across scales and couple to existing models, we  
101 have the potential to develop a new generation of reduced complexity genome-enabled models.

102

### 103 **Ecological Stoichiometry as a Guiding Framework in Soil Organic Matter Decomposition.**

104

105         Microbial (i.e., fungal, bacterial, archaeal, and viral) degradation of chemically diverse  
106 SOM is central to global C cycles and is reliant on broader organismal and soil stoichiometry

107 (Buchkowski et al. 2015; Buchkowski et al. 2019; van Groenigen et al. 2006). Nutrient  
108 availabilities in soil determine the metabolic pathways (and their efficiencies) and substrates that  
109 are used during SOM decomposition. Much belowground N and S is stored within organic  
110 matter that is mineralized by soil microbial communities to provide inorganic nutrients  
111 accessible to plants (Freney 1986; Jenkinson 1990; Kirkby et al. 2011). While P availability is  
112 often associated with mineral weathering; the decomposition of organic matter is a key source of  
113 P in many ecosystems (Heuck et al. 2015; Margalef et al. 2017). Microbial decomposition also  
114 increases the bioavailability of SOM by cleaving polymeric compounds into simple C substrates  
115 that can be directly assimilated by microorganisms. This process is often catalyzed by  
116 extracellular enzymes (EE) that release nutrients from organic matter, connecting C, N, S, and P  
117 cycling (Allison et al. 2014). Viral predation can also influence these dynamics by lysing specific  
118 clades of microorganisms and by influencing organic matter accessibility to microorganisms  
119 (Kuzyakov & Mason-Jones 2018; Weinbauer 2004; Wilhelm & Suttle 1999). Global patterns in  
120 the importance of these mechanisms in governing SOM decomposition, and the specific  
121 environmental conditions under which certain metabolic pathways are favored by microbial  
122 communities, are active areas of research.

123         Understanding SOM decomposition mechanisms across different ecosystems is  
124 challenging because heterogeneity in nutrient statuses, substrate qualities and energetics, and  
125 microbial communities can lead to different apparent controls in different soils (Milcu et al.  
126 2011; Sullivan et al. 2014). Rates of decomposition have been linked to microbial adjustments in  
127 the acquisition of specific elements in response to differences between microbial biomass and  
128 substrate stoichiometries (i.e., nutrient limitations, Billings & Ballantyne IV 2013; Billings &  
129 Ziegler 2008; Frost et al. 2005; Manzoni 2017; Manzoni et al. 2008; Spohn 2016; Sterner &

130 Elser 2002). This work is rooted in the paradigm that organisms exhibit stoichiometric  
131 homeostasis in which they maintain stable biomass elemental ratios regardless of substrate  
132 stoichiometry. It leads to the prediction that nutrient limitations universally regulate biological  
133 activity (Spohn 2016). Specifically, the relative strength of the relationship between microbial  
134 metabolisms involved in N, S, or P acquisition and SOM decomposition should be strongest in  
135 soils with the lowest underlying proportion of that element. Major enzymatic pathways  
136 associated with organic N-, S-, and P- acquisition (e.g., proteases, sulfatases, and phosphatases)  
137 should therefore vary across soils based on nutrient status. Indeed, the stoichiometry of microbial  
138 biomass features little global variation (60:7:1 C:N:P) (Cleveland & Liptzin 2007) and substrate  
139 stoichiometries are highly variable (Frost et al. 2005; Manzoni 2017; Manzoni et al. 2008; Spohn  
140 2016; Sterner & Elser 2002), supporting a dynamic in which microorganisms must adapt to  
141 account for soil nutrient concentrations.

142         The dependency of SOM decomposition on soil nutrients has been investigated most  
143 frequently using a suite of biogeochemical techniques in observational or fertilization studies  
144 such as CO<sub>2</sub> flux, mass loss, bulk chemistry (e.g., C:N), microbial biomass measurements, and/or  
145 potential EE activity measurements (Sullivan et al. 2014). In contrast to current ‘omics  
146 approaches that can directly measure the metabolic pathways involved in SOM decomposition,  
147 each of these approaches provides indirect evidence for nutrient-regulated microbial  
148 decomposition using comparatively low molecular resolution. Additionally, the majority of  
149 researchers quantify total or coarsely-defined lumped C pools with minimal characterization of  
150 the abundance of specific SOM compound classes or their stoichiometry. For instance, the  
151 classical pool and flux model focuses on three or four operationally defined C pools that are  
152 connected by biological drivers of decomposition, namely extracellular enzymes (Abramoff et al.



153 2018). As such, current estimates of nutrient limitations on SOM decomposition vary widely  
154 (Cleveland et al. 2006; Grandy et al. 2008; Kirkby et al. 2013; Kirkby et al. 2014; Sullivan et al.  
155 2014; Takriti et al. 2018), and the majority of research has focused on a few elements, generally  
156 on linkages between C and N cycling (Cleveland et al. 2006; Grandy et al. 2008; Khan et al.  
157 2016; Kirkby et al. 2013; Kirkby et al. 2014).

158         Potential EE activity remains the primary tool in biogeochemical research for measuring  
159 the decomposition of specific compound classes in SOM pools. EEs are ubiquitous in soils and  
160 catalyze the depolymerization of complex SOM compounds into simpler molecules for direct  
161 uptake (Allison et al. 2010). The standard suite of enzyme assays target organic substrates  
162 including including proteins, carbohydrates, amino sugars, organic phosphates, and lignins  
163 (Allison et al. 2007; Burns 1982; Sinsabaugh & Shah 2011). However, certain extracellular  
164 enzymes (e.g., NAG, BG, phosphatases) are more widely assayed than others (e.g., phenol  
165 oxidase, sulfatases), and methodological limitations have constrained the development of  
166 accurate assays for some extracellular enzymes (Allison et al. 2010; Graham et al. 2019).  
167 Additionally, while the ratios of EE activities have been used to assess nutrient limitations  
168 (Grandy et al. 2008; Hill et al. 2014; Jing et al. 2020; Moorhead et al. 2013; Sinsabaugh et al.  
169 2008), these assays can be influenced by pH and only provide potential rates. They are low  
170 molecular resolution compared to many ‘omics-based approaches because they target chemical  
171 bonds contained by classes of SOM rather than specific molecules, and they can have long  
172 turnover times that decouple them from SOM cycling at a specific point in time (Schimel et al.  
173 2017).

174         Microbial investigations into nutrient-regulated SOM decomposition have traditionally  
175 used a complementary set of tools, revealing differences in metabolic strategies to maintain

176 stoichiometric homeostasis using lumped traits(e.g. community-level variation between bacteria  
177 and fungi or copiotrophic and oligotrophic metabolisms, Elser et al. 2003; Fierer et al. 2007;  
178 Strickland & Rousk 2010). Changes in microbial heterotrophy has also been suggested to vary  
179 microbial preferences for C vs. N globally, an inference drawn from primarily biogeochemical  
180 data (Taylor & Townsend 2010). Understanding how changes in metabolic strategies of  
181 individual microorganisms respond to changes in substrate stoichiometry scale to a microbial  
182 community's elemental use efficiency and capacity for decomposition is a key unknown in  
183 process-based modelling.

184         To address this challenge, we propose using ecological stoichiometry to inform the  
185 collection and analysis of high-resolution microbial data types. This approach guides the  
186 development of more precise model representations that in turn can be valuable tools for  
187 interpreting new high-resolution datasets.

188         ‘Omics-based characterization of microbial communities (i.e., metagenomic,  
189 metatranscriptomic, metaproteomic, and metabolomic) and other high molecular resolution  
190 approaches (e.g., chemical probes) provide direct means to evaluate how stoichiometry regulates  
191 the ecology of decomposition. ‘Omics approaches have been highlighted as emerging tools in  
192 ecological stoichiometry but are not yet widely implemented (Van de Waal et al. 2018).  
193 Microbial genes and their expression patterns can reveal the genetic potential (genes), expression  
194 (transcripts), and translation (proteins) of specific enzymes involved in SOM decomposition and  
195 more directly evaluate if N, S, and P acquisition vary predictably with SOM stoichiometry.  
196 While these modern microbial approaches have their own limitations (e.g., data annotation and  
197 discrepancies in turnover time when compared to rates), they provide a more complete  
198 representation of biogeochemical cycles than EE potential assays.

199           Several potential analysis targets for stoichiometric regulation of SOM decomposition  
200 have been identified in ‘omics pipelines. For example, Finn et al. (2020) and Wilhelm et al.  
201 (2019) proposed lists of relevant KO and CaZY annotations. These contain enzymes involved in  
202 the decomposition of organic C as well as N, S, P, and iron cycles, including EEs commonly  
203 measured in EE assays. They span the following classes of enzymes: (1) cellulose (e.g.,  
204 cellobiosidase, endoglucanase), hemi-celluloses (e.g., xylanase), lignin (ligninase), and  
205 cellobiose (e.g., beta-glucosidase) degradation; (2) mineral and organic N cycling (e.g.,  
206 chitinase; leucine-aminopeptidase, N-acetyl glucosaminidase; and nitrate, nitrite and nitrous  
207 oxide reductases); (3) N-fixation (nitrogenase); (4) P cycling (monophosphatases); (5) methane  
208 cycling (e.g., methyl coenzyme M reductase, particulate methane monooxygenase); and (6)  
209 assimilatory and dissimilatory sulfate reductases. New annotations for molecular markers  
210 involved in SOM decomposition continue to be discovered, and our ability for molecular  
211 resolution into stoichiometry-based processes can help illuminate patterns where traditional  
212 biogeochemical methods have been insufficient.

213

#### 214 **Ecological Stoichiometry in SOM Decomposition Modelling.**

215

216           While ongoing research is steadily improving the accuracy of SOM decomposition  
217 models, the enormous amount of soil chemical and biological diversity and the depth of new  
218 molecular data types complicates efforts to formulate predictions based on stoichiometric  
219 relationships. These relationships underlie the kinetics of SOM decomposition and therefore are  
220 essential to genome-enabled model structures and calibrations. Still, data interpretation remains  
221 challenging for empiricists (Jansson & Hofmockel 2018), and most current SOM decomposition

222 models do not yet incorporate cutting-edge molecular approaches to represent detailed microbial  
223 processes. Additionally, there is a need to move beyond pool-based models, which rely on  
224 separating SOM by chemical, physical, and functional properties that can be difficult to measure  
225 (Abramoff et al. 2018), to more fluid and continuous relationships between belowground SOM,  
226 microorganisms, and decomposition (Jinyun & Riley 2020; Waring et al. 2020). Microbial data  
227 mining approaches have made incremental advances to this effort, but remaining obstacles  
228 highlight an opportunity for broader use of ecological stoichiometry to guide targeted  
229 investigations. Below, we discuss the current state of microbial models across different scales of  
230 community hierarchies as well as their counterparts in biogeochemical modelling (Fig. 1). We  
231 also discuss existing modelling approaches that integrate across fields and their use of ecological  
232 stoichiometry as a guiding framework (Fig. 2). Finally, we highlight opportunities for ecological  
233 stoichiometry to guide the development of high-resolution models such as genome-enabled  
234 reaction networks and their assimilation at larger scales via reduced complexity models (Fig. 3).

235         Many models emerging from microbiology are rooted in ecological representations at  
236 individual-to-community scales. At the individual level, common modeling approaches depict  
237 individual's growth rates and/or specific metabolic processes without consideration of  
238 stoichiometric balance. Single cell metabolic models typify process-based microbial models at  
239 the individual scale. This suite of models considers inputs in the context of a model organism's  
240 genome to make predictions of growth dynamics or specific metabolomic products (e.g., GEMS,  
241 Duarte et al. 2004; Loira et al. 2012; Lu et al. 2019). With respect to ecological stoichiometry at  
242 the individual scale, microorganisms maintain stoichiometric balance by adjusting growth rate or  
243 biomass stoichiometry and/or mining or excreting nutrients. This understanding could be  
244 integrated into individual-scale models by adjusting active metabolic processes based on

245 environmental nutrient context, for instance through metabolic flux analysis. At an intermediate  
246 scale (i.e., small groups of microorganisms), ecological stoichiometry operates by impacting the  
247 growth rates of organisms with particular traits – for example, microorganisms with specific  
248 nutrient acquisition pathways, r- and K- life strategies, stress tolerance, or other competitive  
249 advantages. However, stoichiometry is not captured in current process-based models at  
250 intermediate-scales. These models simulate interactions between individuals, limited sets of  
251 individuals, or their characteristic traits, to predict metabolic outputs or changes in the growth of  
252 specific organisms (Bouskill et al. 2012; Follows & Dutkiewicz 2011; Malik et al. 2020; Reed et  
253 al. 2014). Incorporation of stoichiometric principles such as substrate stoichiometry and/or  
254 environmental nutrient status at the intermediate scale can guide an individual's expressed  
255 metabolisms in the context of other microorganisms as well as the interactions of their  
256 metabolites and extracellular enzymes. Finally, at the community-scale (i.e., groups of microbial  
257 guilds), models tend to represent interactions among groups of several organisms through core  
258 ecological processes such as dispersal/transport, predator/prey, and mortality with limited  
259 consideration of stoichiometry (Abel et al. 2015; D'Acunto et al. 2019; Mattei et al. 2015); while  
260 ecological stoichiometry induces compositional shifts in metabolic or growth tendencies and/or  
261 subtle changes to community biomass C:N:P:S ratios. Changes in community stoichiometric  
262 balance should change along with compositional changes to inform community-level metabolism  
263 based on nutrient availability and substrate stoichiometries.

264 On the other hand, biogeochemical models, in particular those at intermediate to large  
265 scales, often employ stoichiometric principles. Fine-scale biogeochemical models include  
266 reaction networks and reactive transport models that represent molecular-level processes (e.g.,  
267 those involving specific SOM molecules, EEs, or inorganic nutrients, Porta et al. 2018; Riley et

268 al. 2014; Wilson et al. 2019). While stoichiometric principles underlying nutrient limitation(s)  
269 influence the elemental composition of organic matter and excretion of extracellular enzymes,  
270 these processes are mostly implicitly considered through the concentrations of specific molecules  
271 in fine-scale biogeochemical models. Thus, they represent an opportunity for model refinement  
272 through minimal adjustments. At intermediate scales, biogeochemistry considers the composition  
273 of pools of elements, often through relatively coarse parameters such as fast vs. slow cycling  
274 SOM pools, mineral SOM, chemical classes or C:N ratios of SOM, and/or organic vs. inorganic  
275 nutrient pools. Biogeochemical models at this scale are often built upon stoichiometric  
276 principles, as they constrain rates of SOM decomposition with parameters that represent nutrient  
277 availability, stoichiometries of organic substrates or microbial biomass, and carbon and nutrient  
278 use efficiencies. Process-based models at the intermediate scale have been a primary focus of  
279 many recent research efforts and include advances such as MEND (Wang et al. 2015), RESOM  
280 (Tang & Riley 2015), and MIMICS-CN (Kyker-Snowman et al. 2020). Finally, the largest scale  
281 considered here is comprised of seminal models that are generalized across environments,  
282 typified by the foundational CENTURY model (Parton 1996). These models focus on elemental  
283 stocks (e.g., total C, N, and/or P) and adjust SOM decomposition rates through nutrient  
284 limitations in line with ecological stoichiometry. Recent progress in large-scale pool models  
285 proposes increased accuracy from a tighter coupling of C and N dynamics (Berardi et al. 2020).

286 State-of-science SOM models use ecological stoichiometry to integrate between  
287 microbiology and biogeochemistry at intermediate scales. Due to computational limitations,  
288 these models generally are based on C pools with lumped characteristics and include two  
289 elements at most (Abramoff et al. 2018; Moorhead et al. 2012; Sulman et al. 2019; Wang et al.  
290 2015; Wieder et al. 2015). They use a small set of coarse parameters to denote microbial

291 processes, in particular C and nutrient use efficiencies (CUE, NUE), that describe C and nutrient  
292 assimilation into microbial biomass (Allison et al. 2010; Cleveland & Liptzin 2007; Geyer et al.  
293 2016; Min et al. 2016; Sihi et al. 2016; Sinsabaugh et al. 2013; Sinsabaugh et al. 2016; Wang et  
294 al. 2015; Wieder et al. 2014). CUE and NUE are typically estimated from stoichiometries of  
295 organic matter and microbial biomass or from the ratios of C- vs. nutrient-acquiring enzymes  
296 (Manzoni et al. 2012; Sinsabaugh et al. 2016). As such, CUE and NUE incorporate the principles  
297 of ecological stoichiometry into microbial-explicit models by serving as proxies for generalized  
298 microbial processes. Under nutrient limitation, microorganisms reduce their C uptake, acquire  
299 nutrients, and/or respire or excrete excess C such that C assimilation into biomass often declines  
300 (Anderson et al. 2005; Del Giorgio & Cole 1998; Manzoni 2017; Manzoni et al. 2017; Manzoni  
301 et al. 2008; Middelboe & Søndergaard 1993; Milcu et al. 2011; Mooshammer et al. 2014b; Smith  
302 & Prairie 2004; Sterner & Elser 2002). Thus, CUE typically decreases when nutrients are  
303 limiting. In contrast, when nutrients exist in excess, microorganisms adjust their uptake or rate of  
304 respiration/excretion to decrease NUE (Milcu et al. 2011; Mooshammer et al. 2014a).

305         The balance of these processes is difficult to predict due in part to microbial interactions  
306 (i.e., stoichiometry influences individual behavior rather than community-level behavior that  
307 ultimately regulates SOM decomposition, Manzoni 2017; Manzoni et al. 2012; Sterner & Elser  
308 2002). There is also large variation in microbial CUE and NUE, and their relationships to SOM  
309 decomposition, due to spatial and temporal changes in resource availability, microbial  
310 community structure, and soil physical properties (Frey et al. 2013; Herron et al. 2009; Malik et  
311 al. 2018; Manzoni et al. 2012; Qiao et al. 2019; Sinsabaugh et al. 2013; Sinsabaugh et al. 2016).  
312 Biogeochemical models have shown that small changes in CUE can have large impacts on SOM  
313 decomposition (Li et al. 2014; Six et al. 2006; Wieder et al. 2013), offering an intriguing

314 possibility for model improvement through detailed representations of nutrient-regulated  
315 microbial metabolic pathways to develop scalable and integrated genome-enabled reaction  
316 networks (Hagerty et al. 2018).

317

### 318 **Opportunities for Ecological Stoichiometry and High-Resolution Measurements in SOM** 319 **Decomposition Modelling.**

320

321         Recent work in biogeochemical modelling has improved predictions of SOM decomposition  
322 by more explicitly accounting for the stoichiometries of microbial biomass and SOM  
323 (Buchkowski et al. 2019) (Fig. 2). Fatichi et al. (2019) have proposed direct representation of  
324 microbial communities involved in coupled SOM and nutrient cycling, an approach that could  
325 not only aid in increasing the accuracy of SOM decomposition predictions but also become a  
326 useful tool for empiricists to interpret multidimensional microbial data. Below, we highlight  
327 opportunities to improve fine-scale predictive models as well as integration between  
328 biogeochemical and microbial models across scales of molecular resolution through  
329 stoichiometric principles that underlie the kinetics of SOM decomposition.

330         At the finest molecular resolution, there is a limited set of modelling approaches that  
331 currently predict rates of molecular processes involved in SOM decomposition, primarily arising  
332 from microbial modelling efforts. Genome-enabled metabolic models, in particular, infer  
333 function from molecular data types by placing annotations within the context of a cell's  
334 biochemical abilities to consume substrates, grow, and produce energy (Cuevas et al. 2016).  
335 These models are constrained by a set of biochemical reactions that define the possible functions  
336 of a microorganism or group or community of microorganisms and provide a structure in which



337 to root functional annotations (Seaver et al. 2020). Commonly used in systems biology to  
338 represent interconnected metabolic processes (Lieven et al. 2020; Seaver et al. 2020), they are  
339 capable of including thousands of genes, metabolites, fluxomes, transcriptomes, proteomes, and  
340 associated reactions. Flux balance analysis is widely used within genome-enabled model  
341 constructs to predict fluxes through the reactions in a metabolic network (Cuevas et al. 2016;  
342 Orth et al. 2010). Genome-enabled models can also be coupled to other methods of decoding  
343 high-resolution molecular data, such as thermodynamic theory (Garayburu-Caruso et al. 2020;  
344 Song et al. 2020), deep learning (Zampieri et al. 2019), or network modelling (Kessell et al.  
345 2020; McClure et al. 2020) to further extend their applicability. In order for fine-scale models to  
346 be used to develop transferrable understanding, continued expansion of these techniques under  
347 stoichiometrically constrained conditions is needed, with acknowledgement of the compositional  
348 nature of many stoichiometric metrics (Gloor et al. 2017; Tsilimigras & Fodor 2016).

349         In an idealized scenario, direct modelling of individual substrate uptake and release by all  
350 individuals in a soil ecosystem, as well as their interactions, should alleviate errors due to scaling  
351 across levels of community organization. To do so, microbial-explicit biogeochemical models  
352 would need to leverage genome-enabled or metabolic flux models that represent metabolic  
353 processes and species interactions at the individual-to-community scales. However, it is currently  
354 computationally unfeasible to model all microbial processes in a soil ecosystem as well as their  
355 interactions with each other, necessitating *a priori* choices of which metabolic pathways to  
356 include in a genome-enabled models. There are thousands of different metabolic pathways  
357 influencing SOM decomposition, and a large proportion of soil microbial communities consists of  
358 dark matter for which we can assign no function. Therefore, it is necessary to have a guiding

359 framework for determining which pathways to include in any metabolic model and further scale  
360 high molecular resolution predictions to the scale of a soil ecosystem (Zakem et al. 2020).

361 Ecological stoichiometry can address this need and link microbial and biogeochemical  
362 models across scales of molecular resolution through (1) top down approaches to inform model  
363 structures and (2) bottom up approaches for model calibration (Fig 3).

364 In top down approaches, ecological stoichiometry can be leveraged via bulk  
365 biogeochemical measurements to first identify nutrient limitation(s) that then guide model  
366 development at higher molecular resolutions. Patterns in nutrient concentrations and biochemical  
367 transformations can inform the selection of targeted parameters for inclusion in and calibration  
368 of reduced complexity models. For example, nutrient limitations identified from bulk  
369 measurements define a set of reactions that should most strongly impact rates of SOM  
370 decomposition. Examples include representing amino acid metabolism for N limitation,  
371 microbial biomass recycling for P limitation, and/or organosulfatases for S limitation (or  
372 combinations therein in the case of nutrient co-limitation) in integrated genome-enabled reaction  
373 networks (Fig. 3). Outcomes from this suite of models can then further refine parameters,  
374 pointing to specific microbial genes/transcripts/proteins in pathways associated with the rate  
375 limiting step in SOM decomposition. Using this approach, we can narrow down molecular data  
376 to specific microbial metabolisms or 'omics markers of interest to build genome-enabled models  
377 including only the most relevant parameters that are scalable to the level of soil ecosystems.

378 Conversely, stoichiometric theory can be used in bottom-up approaches to guide  
379 parameterization and calibration across scales of molecular resolution (Fig 3). Bottom-up  
380 approaches start with high-resolution measurements and use ecological stoichiometry as a  
381 framework for their interpretation. For example, data mining could be driven by a focus on

382 nutrient acquisition metabolic pathways (e.g., peptidase-, phosphatase-, and sulfatase-including  
383 metabolic pathways) in order of hypothesized nutrient limitations. Metabolic pathways with the  
384 most predictive power of SOM decomposition, inferred from transcripts, proteins, and  
385 metabolites mapping to metabolic pathways involved in nutrient regulation of SOM  
386 decomposition, then guide the portions of intermediate-scale models where greater resolution  
387 may lead to more accurate predictions. This could result in inclusion and/or calibration of  
388 specific portions of SOM pools that are most tightly coupled to decomposition rates [e.g., protein  
389 pools (low C:N) or necromass (low C:P)] or in the direct assimilation of specific metabolic  
390 pathways for representation instead of lower resolution measurements such as CUE and NUE.  
391 Dynamically activating genome-enabled reaction networks for a specific nutrient status could  
392 provide the maximum benefit of including high molecular resolution in SOM decomposition  
393 models while being computationally tractable. In turn, parameters and rate estimates made at the  
394 intermediate scale can then influence the relative importance of variables in low molecular  
395 resolution models such as CUE/NUE, bacterial:fungal biomass, and/or biomass C:N:S:P.

396         The result of these approaches, used in combination, is a new suite of models that  
397 represent a limited set of microbial metabolisms, guided by stoichiometric principles and high-  
398 resolution molecular measurements, that are computationally feasible and more accurately  
399 predict SOM decomposition (Fig. 3). This addresses a major challenge to the next generation of  
400 predictive models—deriving integrated genome-enabled reaction networks that can represent  
401 SOM decomposition mechanisms across molecular-to-ecosystem scales.

402

403 **Opportunities afforded by investments in new molecular and computational technologies**

404

405           The power of ecological stoichiometry to guide model integration across scales of  
406 biological complexity can increase even further when leveraged in combination with new  
407 advances in molecular technologies and machine learning algorithms. Research that  
408 simultaneously investigates microbial roles in C, N, S, and P cycling is rare, and EE assays are  
409 imprecise relative to ‘omics or probe-based assays. Additionally, in EE assays, targeted enzymes  
410 and substrates must be determined *a priori*, and several enzymes involved in SOM  
411 decomposition—notably phenol oxidases, peroxidases, and sulfatases—are underinvestigated  
412 due to methodological challenges (Sinsabaugh 2010). Investments in untargeted molecular  
413 approaches are opening new doors into soil microbial analyses by allowing detection of active  
414 metabolisms. These are nascent technologies that have the potential to improve our  
415 understanding of SOM decomposition but need greater usage and development to surmount  
416 remaining challenges. Stable isotope probing (SIP, Bernard et al. 2007; Dumont & Murrell 2005;  
417 Pepe-Ranney et al. 2016) paired with ‘omics-based approaches and new ultrahigh-resolution  
418 metabolomics (Tfaily et al. 2017) methods in particular are promising techniques that can  
419 overcome major limitations of EE assays. Below, we describe these technologies, interesting  
420 aspects of their potentials in the context of ecological stoichiometry, and current barriers to  
421 widespread usage. Our intent is to highlight emerging technologies that are beyond the suite of  
422 high-resolution measurements that are now common (e.g., metagenomics) and present a new  
423 class of technologies from which we can derive even more information.

424           In SIP, substrates enriched in heavy isotopes (e.g.,  $^{13}\text{C}$ ,  $^{18}\text{O}$ ,  $^{15}\text{N}$ ) are amended to  
425 microbial communities and then isotope-labeled biomarkers of active microbial populations are  
426 recovered and analyzed (Bernard et al. 2007; Dumont & Murrell 2005; Pepe-Ranney et al. 2016).  
427 While SIP has long been used in biogeochemistry, recent advances are increasing the tractability

428 of SIP within amplicon profiles, metagenomes, metatranscriptomes, and metaproteomes (Barnett  
429 & Buckley 2020; Jameson et al. 2017; Pepe-Ranney et al. 2016; Wilhelm et al. 2019; Youngblut  
430 et al. 2018a; Youngblut et al. 2018b). For instance, metaproteomic SIP can detect active  
431 metabolic pathways by assessing the rate of incorporation of heavy isotopes into proteins (Grob  
432 et al. 2015; Jehmlich et al. 2016; Von Bergen et al. 2013), and when SIP is used in metagenomic  
433 investigations, sequence assembly and binning into microbial genomes is vastly improved  
434 (Barnett & Buckley 2020; Wilhelm et al. 2019). These approaches have been used in  
435 environmental applications such detecting  $^{13}\text{C}$  transfer into microorganisms and metabolites (Li  
436 et al. 2019; Starr et al. 2018) and assessing taxon-specific growth rates (Hungate et al. 2015;  
437 Papp et al. 2020; Purcell et al. 2019) among other uses (Grob et al. 2015; Wilhelm et al. 2019).

438 SIP could be employed in the context of ecological stoichiometry for maximum benefit in  
439 either top down or bottom up approaches. For example, in a bottom up approach, broad  $^{13}\text{C}$  (e.g.,  
440  $^{13}\text{C}$ -labelled amino acids) and  $^{15}\text{N}$  (e.g.,  $^{15}\text{NO}_3$ ) tracers can evaluate linkages of C and N cycling  
441 with SOM decomposition by denoting microorganisms (via 16S rRNA or metagenomic  
442 sequencing) or active microbial pathways (via transcriptomics, proteomics, and metabolomics).  
443 In soil ecosystems that are constrained by N-availability, N cycles should notable impact SOM  
444 decomposition. An ecological stoichiometry framework therefore sets *a priori* expectations for  
445 pathways of interest in SOM decomposition (e.g., rate limiting pathways). In a top down  
446 approach, ecosystem nutrient status and/or the impact of specific portions of elemental cycles on  
447 SOM decomposition (e.g., amino acid cycling) can inform the choice of substrates used in SIP  
448 (e.g., a specific amino acid) to provide greater resolution into active microorganisms and  
449 metabolic pathways.

450           However, ‘omics-based SIP still faces a number of challenges to reach its full potential.  
451   The amount of label incorporation needed for downstream ‘omics measured can vary by  
452   technology, posing logistical issues because subsamples from a single isotopic enrichment  
453   cannot be easily analyzed for signatures in DNA, RNA, proteomic, and metabolomic data types.  
454   An unlabelled set of samples is needed in addition to enriched samples in many instances,  
455   doubling sample sizes in experimental designs. Further, extensive prototyping is often needed  
456   because label incorporation itself varies with time, across sample sets, and with microbial GC  
457   content. Additionally, microbial cross-feeding can present difficulties in tracing label  
458   incorporation (Coyotzi et al. 2016; Schwartz et al. 2016), and there is no standardized software  
459   available for data analysis. These drawbacks come at a large monetary cost that have constrained  
460   widespread application of SIP in ‘omics investigations. For SIP to reach its full potential,  
461   investment is needed to increase accessibility, determine the appropriate label amounts,  
462   incubation duration for specific soil types, and standardized software for data analysis as well as  
463   to streamline ‘omics integration with SIP.

464           In parallel, ultrahigh-resolution metabolomics, such as Fourier transform ion cyclotron  
465   resonance mass spectrometry (FTICR-MS, Tfaily et al. 2015; Tfaily et al. 2017), can illuminate  
466   new aspects of SOM decomposition by enabling the determination of elemental stoichiometry in  
467   thousands of SOM molecules simultaneously (Tfaily et al. 2017). In FTICR-MS, soils are  
468   extracted with a choice of solvent depending on the portion of the SOM pool of interest.  
469   Extractions can then be ionized via Electrospray ionization (ESI) to assist the transfer of ions  
470   from solution into the gaseous phase and injected into the FTICR-MS instrument to yield  
471   extremely high mass resolving power ( $>1M$ ) and mass measurement accuracy ( $<1$  ppm).  
472   Formulas assigned via ultrahigh-resolution metabolomics are a promising tool for evaluating

473 metabolisms influenced by ecological stoichiometry by providing information on the abundance  
474 of molecules with specific stoichiometries (e.g., C:N ratios of SOM molecules in a standing  
475 pool) or changes in these molecules during the course of experiments.

476         Yet, development of ultrahigh-resolution metabolomics and increased access to relevant  
477 technologies by the broader scientific community are needed to push its current boundaries. A  
478 primary constraint is that ionization efficiency and biases vary across soil types, and in some  
479 cases, even within soils in a single experiment. This makes quantification of metabolites  
480 unreliable such that data is most accurately used in a presence/absence format. Another weakness  
481 is that SOM must be extracted prior to analysis via most metabolomic technologies; however,  
482 there has been significant improvement in the development of new extraction techniques to  
483 minimize inefficiencies and target a suite of chemically distinct organic matter pools (Tfaily et  
484 al. 2015). Despite its limitations, ultrahigh-resolution metabolomics has been used to evaluate  
485 the relative balance of OM thermodynamics vs. N availability in regulating aerobic respiration,  
486 revealing its power when leveraged with ecological stoichiometry (Garayburu-Caruso et al.  
487 2020). Particularly when combined with other ‘omics technologies, it can provide molecular  
488 resolution into nutrient-regulated microbial metabolic pathways (Song et al. 2020). Investment in  
489 the use of tailored standards and advances in the calculation of ionization biases as well as  
490 improvements in extraction efficiencies can help these technologies reach their full potential.

491         In parallel with technological opportunities, the interpretation of untargeted molecular  
492 approaches is complicated by the inadequate description of metabolic pathways for soil  
493 microorganisms in existing databases and the unsuitability of standard statistical approaches in  
494 ecology for combining diverse multidimensional data types. Unsupervised machine learning is  
495 emerging as a tool both to decipher hidden patterns in complex data and to eliminate the needs

496 for *a priori* relationships between genes (e.g., via metabolic pathways or metagenome-assembled  
497 genomes) and for gap filling to infer absent data. There is an enormous diversity of machine  
498 learning algorithms with varying levels of complexity that are well-suited to discern patterns in  
499 data (Rana et al. 2020; Were et al. 2015). Techniques such as K-nearest-neighbor (Peterson  
500 2009), artificial neural network(ANN, Sarle 1994), support vector machines (SVM, Wang 2005),  
501 neuro-fuzzy (Nauck et al. 1997), decision tree classifiers (Safavian & Landgrebe 1991), and  
502 random forests (Liaw & Wiener 2002) are most commonly employed in microbiology and  
503 environmental disciplines, often in combination due to the various strengths and weaknesses of  
504 each (e.g., Cai et al. 2019; De Clercq et al. 2019; Dong & Chen 2019; Qdais et al. 2010;  
505 Rahimian Boogar et al. 2019; Thompson et al. 2019; Wang et al. 2020). Data-driven machine  
506 learning has been employed in a variety of gene annotation pipelines to reveal microbial dark  
507 matter (Arakawa et al. 2006; Aziz et al. 2008; Faria et al. 2018; Hyatt et al. 2010; Notebaart et al.  
508 2006), metabolite identification (Djoumbou-Feunang et al. 2019; Shen et al. 2013), gap-filling  
509 (Oyetunde et al. 2019), assimilation of multiple lines of data (Rana et al. 2020), pathway  
510 enrichment (Luo et al. 2009), and phenotype prediction (LaPierre et al. 2019; Pasolli et al. 2016).  
511 Still, we lack rigorous machine learning investigations in which sampling and experimental  
512 designs move beyond feature identification to translating selected features into meaningful  
513 ecological outcomes.

514 Machine learning approaches are particularly important for soils in which the vast  
515 majority of microorganisms have not been cultured. These methods have recently been used to  
516 describe complex environmental problems such as deriving microbial indicators of disturbances  
517 (Beall et al. 2016; Glasl et al. 2019), assessing microbial community dynamics (Cai et al. 2019;  
518 Guo et al. 2015; Lesnik & Liu 2017; Wang et al. 2020), and predicting soil C content (Chen et al.



519 2019; Hengl et al. 2017; Jardine & Siikamäki 2014; Keskin et al. 2019; Morellos et al. 2016)  
520 among other applications (Graham et al. 2018; Rahimian Boogar et al. 2019; Thompson et al.  
521 2019). Yet such applications are mostly descriptive and independent of any theoretical  
522 framework as a foundation for interpretation. Pairing machine learning with ecological theory for  
523 experimental hypothesis testing is needed to push fundamental biology forward (Rana et al.  
524 2020; Were et al. 2015). Using new high-resolution molecular data types in spatially- and  
525 temporally-distributed sampling, paired with machine learning, can offer unprecedented  
526 untargeted resolution into the nutrient-regulated pathways by which microorganisms decompose  
527 SOM (Manzoni et al. 2017; Mooshammer et al. 2014b).

528       Specifically, within the context of ecological stoichiometry, we propose that machine  
529 learning can parse nutrient-regulated mechanisms that control SOM decomposition rates across  
530 all scales of molecular resolution. For examine, at the finest molecular resolution, machine  
531 learning can uncover signals in gene, transcript, protein, or metabolite data (alone or in  
532 combination) that correspond to SOM decomposition rates. Signals can be interpreted within the  
533 context of stoichiometry informed *a priori* hypotheses to infer possible casual relationships for  
534 evaluation with experiments. Similar approaches can also be used in combination with metabolic  
535 pathway mapping or other grouping strategies (e.g., by N, S, or P cycle involvement) to further  
536 distill relationships into casual units that are representable in models.

537       Complementarily, investment in targeted approaches can improve the accuracy of  
538 existing enzyme-based approaches. Activity-based protein profiling (ABPP, Killinger et al.  
539 2019), for example, leverages chemical probes that react irreversibly with protein families to  
540 identify enzymes binding to specific molecules, and when paired with mass spectrometry-based  
541 approaches, can also quantify rates of enzyme activity (Killinger et al. 2019). In contrast to EE

542 assays that reveal potential enzyme rates, ABPP has the potential to dramatically improve  
543 investigations of SOM decomposition by revealing the true expression of decomposition  
544 enzymes; however, it is rarely used in soils (Sadler & Wright 2015; Whidbey & Wright 2018;  
545 Zegeye et al. 2020). Major challenges to soil-based ABPP include complexation with diverse  
546 chemical structures in soils (i.e., non-specificity) and probe extraction from the soil matrix  
547 (Killinger et al. 2019). A broad range of chemical probes have been developed to target enzymes  
548 such as serine hydrolases (Sanman & Bogyo 2014), cysteine proteases (Sanman & Bogyo 2014),  
549 metalloproteases (Saghatelian et al. 2004), kinases (Patricelli et al. 2007), glycosidases  
550 (Chauvigné-Hines et al. 2012), and ammonia monooxygenases (Bennett et al. 2016). These  
551 provide a solid foundation for further development and deployment in soils. ABPP could be  
552 employed in similar ways to the current usage of potential EE activities to evaluate  
553 stoichiometric relationships that underlie SOM decomposition with more precision. As ABPP  
554 use becomes more common in microorganisms (Sharifzadeh et al. 2018) and the diversity of  
555 available probes continues to expand (Killinger et al. 2019), there is potential for ABPP-based  
556 approaches to move beyond the coarse resolution of EE assays and to target specific enzymes  
557 involved in nutrient-regulated decomposition (Sadler & Wright 2015).

558

## 559 **Conclusions.**

560         Dynamic soil environments have led to the evolution of a large repository of  
561 interconnected microbial mechanisms for energy acquisition. Nutrient-driven metabolic  
562 regulation is the ultimate control over many biogeochemical processes including SOM  
563 decomposition (Manzoni et al. 2017; Mooshammer et al. 2014b). The stoichiometry of SOM  
564 decomposition in particular regulates biological activity across individual-to-community scales.

565 Ecological stoichiometry is largely absent from more modern implementations of high-resolution  
566 molecular measurements, despite promising relationships uncovered by decades of  
567 biogeochemical research that guide large scale predictive models. Using ecological  
568 stoichiometry as a framework to guide technological and model-based advances at the molecular  
569 scale can expediate progress towards overcoming a major challenge to understanding global  
570 SOM decomposition — interpreting the massive amount of data generated by high-resolution  
571 approaches to investigating diverse soil microbial communities. Ecological stoichiometry can  
572 provide a tractable set of metabolic reactions and parameters to depict detailed SOM cycling in  
573 genome-enabled reaction networks and scale across levels of molecular resolution within a  
574 common framework, yet it has not been leveraged in state-of-science experiments and models.  
575 We propose that investing in and deploying new technologies in the context of stoichiometric  
576 theory provides an untapped and promising framework for interpreting the vast amount of data  
577 we can now generate and further distilling this information into reduced complexity models that  
578 predict SOM decomposition.

579

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586

587 **Declarations.**

588

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594

595 *Conflicts of interest/Competing interests.* We declare no conflicts of interest/competing interests.

596

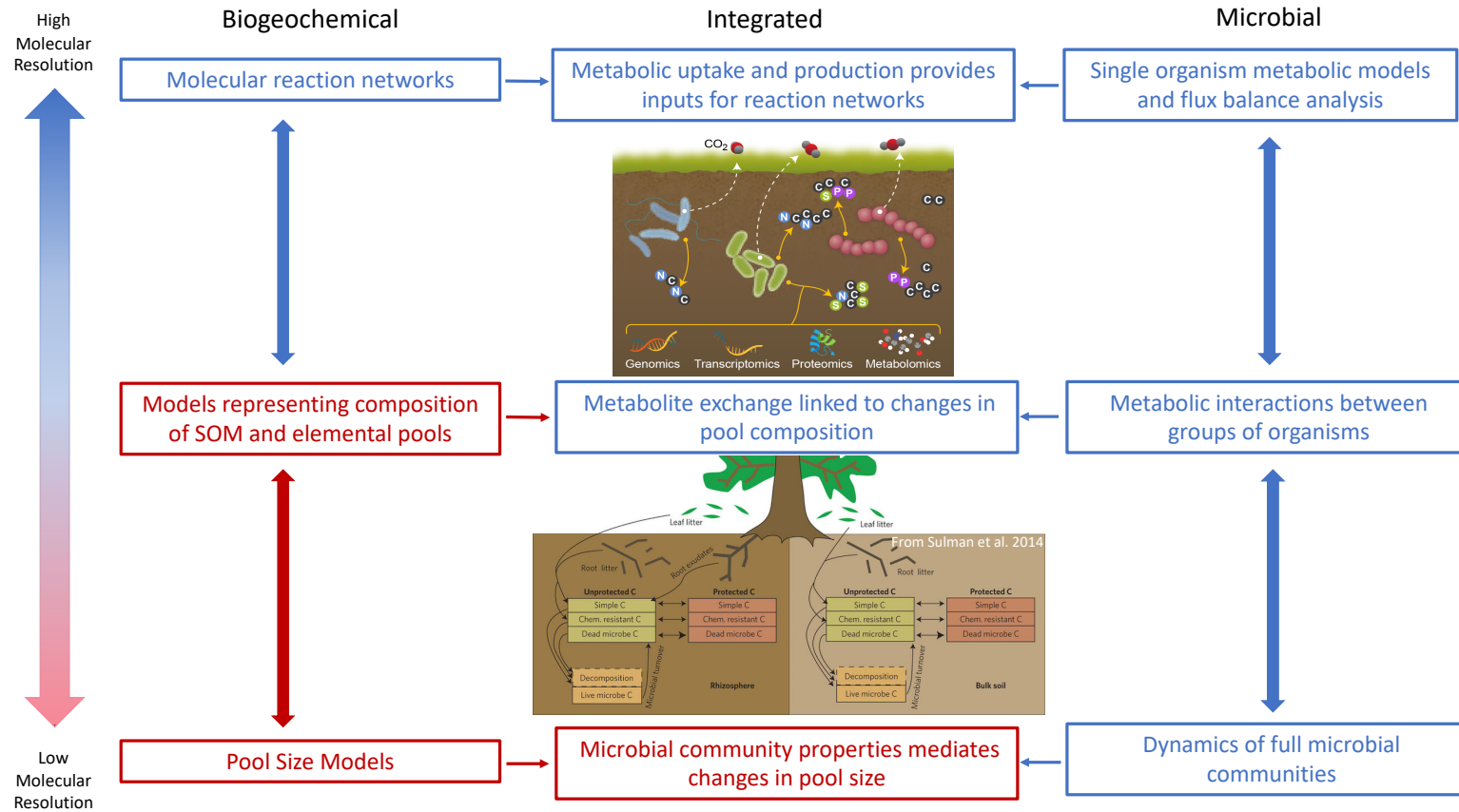
597 *Availability of data and material (data transparency).* Not applicable.

598

599 *Code availability (software application or custom code).* Not applicable.

600

601 *Authors' contributions.* EBG conceived and wrote the manuscript with significant contributions  
602 from KSH.

603 **Figures.**

604

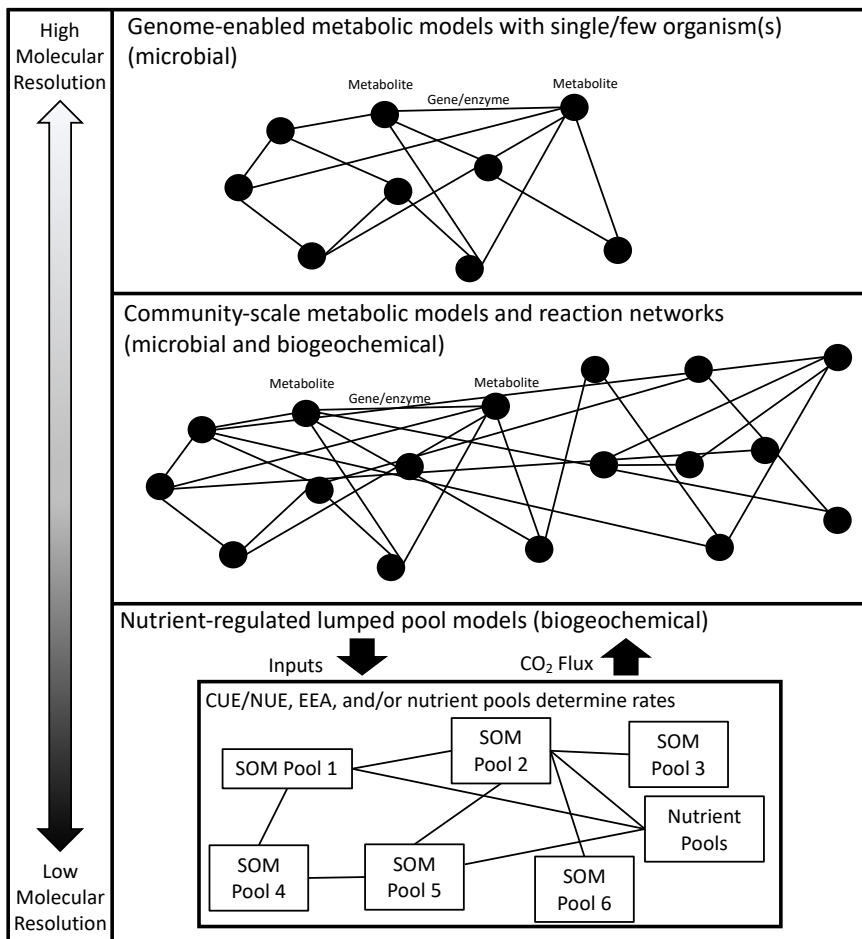
605 **Fig 1. Parallel development of processed-based biogeochemical and microbial models across scales of molecular resolution.**

606 Ecological stoichiometry has guided the development of many biogeochemical models at relatively coarse molecular resolution but

607 has not yet been assimilated into high-resolution modelling frameworks that are principally informed by microbiology. We present

608 opportunities for stoichiometry to serve as an underlying framework to integrate state-of-science SOM decomposition models across

609 ecological hierarchies – from individual microorganisms to bulk pools and fluxes – as well biogeochemical and microbial disciplines.  
610 Examples of common modelling frameworks are arranged vertically by level of molecular resolution and horizontally by scientific  
611 domain. Information exchange is denoted by arrows. Modelling frameworks that leverage ecological stoichiometry are presented in  
612 red, and opportunities to use these principles to guide multiscale model development are presented in blue. Insets denote the role of  
613 stoichiometry in SOM decomposition (top) and an example of state-of-science SOM decomposition models (bottom).

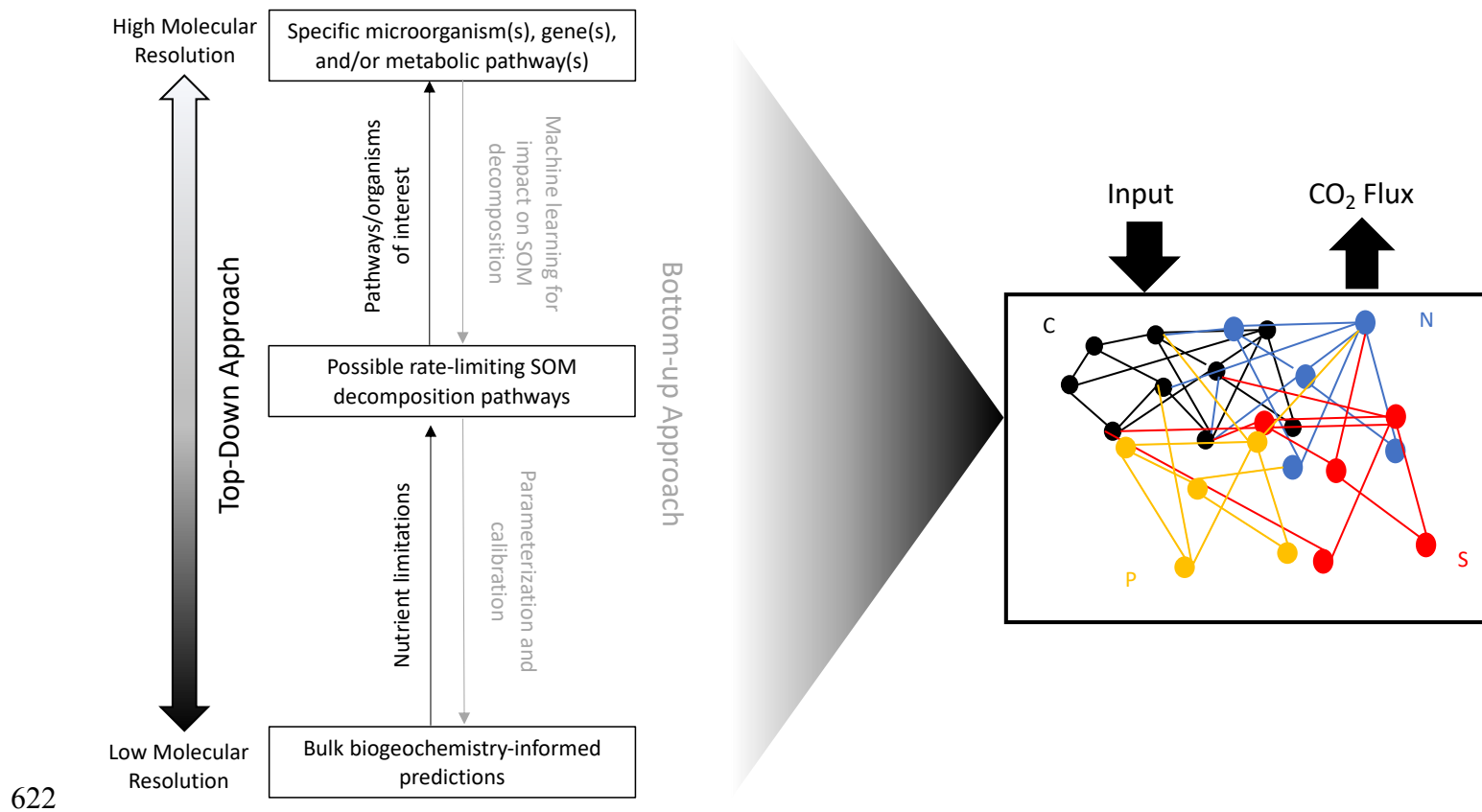


614

615 **Fig 2. State-of-science SOM decomposition models across scales of molecular resolution.** State-of-science models across scales of  
 616 ecological hierarchies arise from different scientific domains. At the highest level of molecular resolution (top), genome-enabled  
 617 models predict the production and consumption of specific metabolites by specific microorganisms or groups of microorganisms. This

618 class of models is rooted in fundamental microbiology. At intermediate scales, community-scale metabolic models and reaction  
619 networks have been largely separately developed in microbiology and biogeochemistry, respectively. Finally, at coarser levels of  
620 molecular resolution, biogeochemical models predict SOM decomposition using nutrient-regulated exchange between lumped SOM  
621 pools with specific chemical attributes.





**Fig 3. Schematic of opportunities for ecological stoichiometry to guide iterative approaches that generate genome-enabled reduced complexity models.** Ecological stoichiometry can guide a cycle of top-down and bottom-up approaches for more efficient generation of next generation models. Top-down approaches use nutrient limitations inferred from bulk biogeochemical measurements to guide the determination of rate-limiting pathways in SOM decomposition. In turn, rate-limiting pathways can reveal specific metabolic pathways and/or microorganism for detailed representation in predictive models. Bottom-up approaches use ecological

628 stoichiometry as a framework for guiding the interpretation of high molecular resolution data streams with the help of new machine  
629 learning algorithms, for example by focusing on metabolic pathways that tend to be involved in nutrient acquisition. Using these  
630 approaches to identify the rate-limiting steps of SOM decomposition can then aid in the parameterization and calibration of larger-  
631 scale models by informing the tuning of substrate use efficiencies and/or by revealing pathways for more detailed representation. By  
632 using both approaches iteratively, we can identify specific portions of genome-enabled reaction networks that add predictive power  
633 with detailed representation in large-scale SOM decomposition models.

634 **References.**

- 635
- 636 Abel S, Zur Wiesch PA, Chang H-H, Davis BM, Lipsitch M, Waldor MK (2015) Sequence tag–  
637 based analysis of microbial population dynamics. *Nature methods* 12(3): 223-226
- 638 Abramoff R, Xu X, Hartman M, O’Brien S, Feng W, Davidson E, Finzi A, Moorhead D,  
639 Schimel J, Torn M (2018) The Millennial model: in search of measurable pools and  
640 transformations for modeling soil carbon in the new century. *Biogeochemistry* 137(1-2): 51-71
- 641 Allison SD, Chacon SS, German DP (2014) Substrate concentration constraints on microbial  
642 decomposition. *Soil Biology and Biochemistry* 79: 43-49
- 643 Allison SD, Gartner TB, Holland K, Weintraub M, Sinsabaugh RL (2007) Soil enzymes: linking  
644 proteomics and ecological processes. In: *Manual of Environmental Microbiology, Third Edition.*  
645 *American Society of Microbiology.* p 704-711
- 646 Allison SD, Weintraub MN, Gartner TB, Waldrop MP (2010) Evolutionary-economic principles  
647 as regulators of soil enzyme production and ecosystem function. In: *Soil enzymology.* Springer.  
648 p 229-243
- 649 Anderson TR, Hessen DO, Elser JJ, Urabe J (2005) Metabolic stoichiometry and the fate of  
650 excess carbon and nutrients in consumers. *The American Naturalist* 165(1): 1-15
- 651 Arakawa K, Yamada Y, Shinoda K, Nakayama Y, Tomita M (2006) GEM System: automatic  
652 prototyping of cell-wide metabolic pathway models from genomes. *BMC bioinformatics* 7(1):  
653 168
- 654 Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass  
655 EM, Kubal M (2008) The RAST Server: rapid annotations using subsystems technology. *BMC*  
656 *genomics* 9(1): 75

- 657 Bailey VL, Bond-Lamberty B, DeAngelis K, Grandy AS, Hawkes CV, Heckman K, Lajtha K,  
658 Phillips RP, Sulman BN, Todd-Brown KE (2018) Soil carbon cycling proxies: understanding  
659 their critical role in predicting climate change feedbacks. *Global Change Biology* 24(3): 895-905
- 660 Barnett SE, Buckley DH (2020) Simulating metagenomic stable isotope probing datasets with  
661 MetaSIPSim. *BMC bioinformatics* 21(1): 37
- 662 Beall B, Twiss M, Smith D, Oyserman B, Rozmarynowycz M, Binding C, Bourbonniere R,  
663 Bullerjahn G, Palmer M, Reavie ED (2016) Ice cover extent drives phytoplankton and bacterial  
664 community structure in a large north-temperate lake: implications for a warming climate.  
665 *Environmental microbiology* 18(6): 1704-1719
- 666 Bennett K, Sadler NC, Wright AT, Yeager C, Hyman MR (2016) Activity-based protein  
667 profiling of ammonia monooxygenase in *Nitrosomonas europaea*. *Appl. Environ. Microbiol.*  
668 82(8): 2270-2279
- 669 Berardi D, Brzostek E, Blanc-Betes E, Davison B, DeLucia EH, Hartman MD, Kent J, Parton  
670 WJ, Saha D, Hudiburg TW (2020) 21st-century biogeochemical modeling: Challenges for  
671 Century-based models and where do we go from here? *GCB Bioenergy* 12(10): 774-788
- 672 Bernard L, Mougél C, Maron PA, Nowak V, Lévêque J, Henault C, Haichar FeZ, Berge O,  
673 Marol C, Balesdent J (2007) Dynamics and identification of soil microbial populations actively  
674 assimilating carbon from  $^{13}\text{C}$ -labelled wheat residue as estimated by DNA- and RNA-SIP  
675 techniques. *Environmental Microbiology* 9(3): 752-764
- 676 Billings SA, Ballantyne IV F (2013) How interactions between microbial resource demands, soil  
677 organic matter stoichiometry, and substrate reactivity determine the direction and magnitude of  
678 soil respiratory responses to warming. *Global Change Biology* 19(1): 90-102

- 679 Billings SA, Ziegler SE (2008) Altered patterns of soil carbon substrate usage and heterotrophic  
680 respiration in a pine forest with elevated CO<sub>2</sub> and N fertilization. *Global Change Biology* 14(5):  
681 1025-1036
- 682 Blankinship JC, Berhe AA, Crow SE, Druhan JL, Heckman KA, Keiluweit M, Lawrence CR,  
683 Marín-Spiotta E, Plante AF, Rasmussen C (2018) Improving understanding of soil organic  
684 matter dynamics by triangulating theories, measurements, and models. *Biogeochemistry* 140(1):  
685 1-13
- 686 Bond-Lamberty B, Epron D, Harden J, Harmon ME, Hoffman F, Kumar J, David McGuire A,  
687 Vargas R (2016) Estimating heterotrophic respiration at large scales: Challenges, approaches,  
688 and next steps. *Ecosphere* 7(6): e01380
- 689 Bouskill N, Tang J, Riley WJ, Brodie EL (2012) Trait-based representation of biological  
690 nitrification: model development, testing, and predicted community composition. *Frontiers in*  
691 *microbiology* 3: 364
- 692 Brangari AC, Manzoni S, Rousk J (2020) A soil microbial model to analyze decoupled microbial  
693 growth and respiration during soil drying and rewetting. *Soil Biology and Biochemistry*: 107871
- 694 Buchkowski RW, Schmitz OJ, Bradford MA (2015) Microbial stoichiometry overrides biomass  
695 as a regulator of soil carbon and nitrogen cycling. *Ecology* 96(4): 1139-1149
- 696 Buchkowski RW, Shaw AN, Sihi D, Smith GR, Keiser AD (2019) Constraining carbon and  
697 nutrient flows in soil with ecological stoichiometry. *Frontiers in Ecology and Evolution* 7: 382
- 698 Burns RG (1982) Enzyme activity in soil: location and a possible role in microbial ecology. *Soil*  
699 *biology and biochemistry* 14(5): 423-427

700 Cai W, Lesnik KL, Wade MJ, Heidrich ES, Wang Y, Liu H (2019) Incorporating microbial  
701 community data with machine learning techniques to predict feed substrates in microbial fuel  
702 cells. *Biosensors and Bioelectronics* 133: 64-71

703 Chauvigné-Hines LM, Anderson LN, Weaver HM, Brown JN, Koech PK, Nicora CD, Hofstad  
704 BA, Smith RD, Wilkins MJ, Callister SJ (2012) Suite of activity-based probes for cellulose-  
705 degrading enzymes. *Journal of the American Chemical Society* 134(50): 20521-20532

706 Chen D, Chang N, Xiao J, Zhou Q, Wu W (2019) Mapping dynamics of soil organic matter in  
707 croplands with MODIS data and machine learning algorithms. *Science of The Total*  
708 *Environment* 669: 844-855

709 Chowdhury TR, Lee J-Y, Bottos EM, Brislawn CJ, White RA, Bramer LM, Brown J, Zucker JD,  
710 Kim Y-M, Jumpponen A (2019) Metaphenomic responses of a native prairie soil microbiome to  
711 moisture perturbations. *Msystems* 4(4):

712 Cleveland CC, Liptzin D (2007) C: N: P stoichiometry in soil: is there a “Redfield ratio” for the  
713 microbial biomass? *Biogeochemistry* 85(3): 235-252

714 Cleveland CC, Reed SC, Townsend AR (2006) Nutrient regulation of organic matter  
715 decomposition in a tropical rain forest. *Ecology* 87(2): 492-503

716 Coyotzi S, Pratscher J, Murrell JC, Neufeld JD (2016) Targeted metagenomics of active  
717 microbial populations with stable-isotope probing. *Current opinion in biotechnology* 41: 1-8

718 Crowther TW, Van den Hoogen J, Wan J, Mayes MA, Keiser A, Mo L, Averill C, Maynard DS  
719 (2019) The global soil community and its influence on biogeochemistry. *Science* 365(6455):  
720 eaav0550

- 721 Cuevas DA, Edirisinghe J, Henry CS, Overbeek R, O'Connell TG, Edwards RA (2016) From  
722 DNA to FBA: how to build your own genome-scale metabolic model. *Frontiers in microbiology*  
723 7: 907
- 724 D'Acunto B, Frunzo L, Klapper I, Mattei M, Stoodley P (2019) Mathematical modeling of  
725 dispersal phenomenon in biofilms. *Mathematical biosciences* 307: 70-87
- 726 De Clercq D, Jalota D, Shang R, Ni K, Zhang Z, Khan A, Wen Z, Caicedo L, Yuan K (2019)  
727 Machine learning powered software for accurate prediction of biogas production: A case study  
728 on industrial-scale Chinese production data. *Journal of cleaner production* 218: 390-399
- 729 Del Giorgio PA, Cole JJ (1998) Bacterial growth efficiency in natural aquatic systems. *Annual*  
730 *Review of Ecology and Systematics* 29(1): 503-541
- 731 Djoumbou-Feunang Y, Fiamoncini J, Gil-de-la-Fuente A, Greiner R, Manach C, Wishart DS  
732 (2019) BioTransformer: a comprehensive computational tool for small molecule metabolism  
733 prediction and metabolite identification. *Journal of cheminformatics* 11(1): 1-25
- 734 Dong C, Chen J (2019) Optimization of process parameters for anaerobic fermentation of corn  
735 stalk based on least squares support vector machine. *Bioresource technology* 271: 174-181
- 736 Duarte NC, Herrgård MJ, Palsson BØ (2004) Reconstruction and validation of *Saccharomyces*  
737 *cerevisiae* iND750, a fully compartmentalized genome-scale metabolic model. *Genome research*  
738 14(7): 1298-1309
- 739 Dumont MG, Murrell JC (2005) Stable isotope probing—linking microbial identity to function.  
740 *Nature Reviews Microbiology* 3(6): 499-504
- 741 Elser J, Acharya K, Kyle M, Cotner J, Makino W, Markow T, Watts T, Hobbie S, Fagan W,  
742 Schade J (2003) Growth rate—stoichiometry couplings in diverse biota. *Ecology Letters* 6(10):  
743 936-943

- 744 Elser J, Sterner RW, Gorokhova Ea, Fagan W, Markow T, Cotner JB, Harrison J, Hobbie SE,  
745 Odell G, Weider L (2000) Biological stoichiometry from genes to ecosystems. *Ecology letters*  
746 3(6): 540-550
- 747 Elser JJ, Dobberfuhl DR, MacKay NA, Schampel JH (1996) Organism size, life history, and N:  
748 P stoichiometry: toward a unified view of cellular and ecosystem processes. *BioScience* 46(9):  
749 674-684
- 750 Faria JP, Rocha M, Rocha I, Henry CS (2018) Methods for automated genome-scale metabolic  
751 model reconstruction. *Biochemical Society Transactions* 46(4): 931-936
- 752 Fatichi S, Manzoni S, Or D, Paschalis A (2019) A Mechanistic model of microbially mediated  
753 soil biogeochemical processes: A reality check. *Global Biogeochemical Cycles* 33(6): 620-648
- 754 Fierer N, Bradford MA, Jackson RB (2007) Toward an ecological classification of soil bacteria.  
755 *Ecology* 88(6): 1354-1364
- 756 Finn D, Yu J, Penton CR (2020) Soil quality shapes the composition of microbial community  
757 stress response and core cell metabolism functional genes. *Applied Soil Ecology* 148: 103483
- 758 Follows MJ, Dutkiewicz S (2011) Modeling diverse communities of marine microbes. *Annual*  
759 *review of marine science* 3: 427-451
- 760 Freney J (1986) Forms and reactions of organic sulfur compounds in soils. *Sulfur in agriculture*  
761 (27): 207-232
- 762 Frey SD, Lee J, Melillo JM, Six J (2013) The temperature response of soil microbial efficiency  
763 and its feedback to climate. *Nature Climate Change* 3(4): 395-398
- 764 Frost PC, Evans White MA, Finkel ZV, Jensen TC, Matzek V (2005) Are you what you eat?  
765 Physiological constraints on organismal stoichiometry in an elementally imbalanced world.  
766 *Oikos* 109(1): 18-28



- 767 Garayburu-Caruso VA, Stegen JC, Song H-S, Renteria L, Wells J, Garcia W, Resch CT,  
768 Goldman AE, Chu RK, Toyoda J (2020) Carbon limitation leads to thermodynamic regulation of  
769 aerobic metabolism. *Environmental Science & Technology Letters*:
- 770 Geyer KM, Kyker-Snowman E, Grandy AS, Frey SD (2016) Microbial carbon use efficiency:  
771 accounting for population, community, and ecosystem-scale controls over the fate of  
772 metabolized organic matter. *Biogeochemistry* 127(2-3): 173-188
- 773 Glasl B, Bourne DG, Frade PR, Thomas T, Schaffelke B, Webster NS (2019) Microbial  
774 indicators of environmental perturbations in coral reef ecosystems. *Microbiome* 7(1): 1-13
- 775 Gloor GB, Macklaim JM, Pawlowsky-Glahn V, Egozcue JJ (2017) Microbiome datasets are  
776 compositional: and this is not optional. *Frontiers in microbiology* 8: 2224
- 777 Graham EB, Crump AR, Kennedy DW, Arntzen E, Fansler S, Purvine SO, Nicora CD, Nelson  
778 W, Tfaily MM, Stegen JC (2018) Multi'omics comparison reveals metabolome biochemistry, not  
779 microbiome composition or gene expression, corresponds to elevated biogeochemical function in  
780 the hyporheic zone. *Science of the total environment* 642: 742-753
- 781 Graham EB, Yang F, Bell S, Hofmockel KS (2019) High genetic potential for proteolytic  
782 decomposition in northern peatland ecosystems. *Appl. Environ. Microbiol.* 85(10): e02851-  
783 02818
- 784 Grandy AS, Sinsabaugh RL, Neff JC, Stursova M, Zak DR (2008) Nitrogen deposition effects on  
785 soil organic matter chemistry are linked to variation in enzymes, ecosystems and size fractions.  
786 *Biogeochemistry* 91(1): 37-49
- 787 Grob C, Taubert M, Howat AM, Burns OJ, Dixon JL, Richnow HH, Jehmlich N, von Bergen M,  
788 Chen Y, Murrell JC (2015) Combining metagenomics with metaproteomics and stable isotope

- 789 probing reveals metabolic pathways used by a naturally occurring marine methylotroph.  
790 *Environmental microbiology* 17(10): 4007-4018
- 791 Guo H, Jeong K, Lim J, Jo J, Kim YM, Park J-p, Kim JH, Cho KH (2015) Prediction of effluent  
792 concentration in a wastewater treatment plant using machine learning models. *Journal of*  
793 *Environmental Sciences* 32: 90-101
- 794 Hagerty SB, Allison SD, Schimel JP (2018) Evaluating soil microbial carbon use efficiency  
795 explicitly as a function of cellular processes: implications for measurements and models.  
796 *Biogeochemistry* 140(3): 269-283
- 797 Hall EK, Bernhardt ES, Bier RL, Bradford MA, Boot CM, Cotner JB, del Giorgio PA, Evans SE,  
798 Graham EB, Jones SE (2018) Understanding how microbiomes influence the systems they  
799 inhabit. *Nature microbiology* 3(9): 977-982
- 800 Hengl T, de Jesus JM, Heuvelink GB, Gonzalez MR, Kilibarda M, Blagotić A, Shangguan W,  
801 Wright MN, Geng X, Bauer-Marschallinger B (2017) SoilGrids250m: Global gridded soil  
802 information based on machine learning. *PLoS one* 12(2):
- 803 Herron PM, Stark JM, Holt C, Hooker T, Cardon ZG (2009) Microbial growth efficiencies  
804 across a soil moisture gradient assessed using <sup>13</sup>C-acetic acid vapor and <sup>15</sup>N-ammonia gas. *Soil*  
805 *Biology and Biochemistry* 41(6): 1262-1269
- 806 Heuck C, Weig A, Spohn M (2015) Soil microbial biomass C: N: P stoichiometry and microbial  
807 use of organic phosphorus. *Soil Biology and Biochemistry* 85: 119-129
- 808 Hill BH, Elonen CM, Jicha TM, Kolka RK, Lehto LL, Sebestyen SD, Seifert-Monson LR (2014)  
809 Ecoenzymatic stoichiometry and microbial processing of organic matter in northern bogs and  
810 fens reveals a common P-limitation between peatland types. *Biogeochemistry* 120(1-3): 203-224

- 811 Hungate BA, Dukes JS, Shaw MR, Luo Y, Field CB (2003) Nitrogen and climate change.  
812 Science 302(5650): 1512-1513
- 813 Hungate BA, Mau RL, Schwartz E, Caporaso JG, Dijkstra P, van Gestel N, Koch BJ, Liu CM,  
814 McHugh TA, Marks JC (2015) Quantitative microbial ecology through stable isotope probing.  
815 Appl. Environ. Microbiol. 81(21): 7570-7581
- 816 Hyatt D, Chen G-L, LoCascio PF, Land ML, Larimer FW, Hauser LJ (2010) Prodigal:  
817 prokaryotic gene recognition and translation initiation site identification. BMC bioinformatics  
818 11(1): 119
- 819 Jameson E, Taubert M, Coyotzi S, Chen Y, Eyice Ö, Schäfer H, Murrell JC, Neufeld JD,  
820 Dumont MG (2017) DNA-, RNA-, and protein-based stable-isotope probing for high-throughput  
821 biomarker analysis of active microorganisms. In: Metagenomics. Springer. p 57-74
- 822 Jansson JK, Hofmockel KS (2018) The soil microbiome—from metagenomics to  
823 metaphenomics. Current opinion in microbiology 43: 162-168
- 824 Jardine SL, Siikamäki JV (2014) A global predictive model of carbon in mangrove soils.  
825 Environmental Research Letters 9(10): 104013
- 826 Jehmlich N, Vogt C, Lünsmann V, Richnow HH, von Bergen M (2016) Protein-SIP in  
827 environmental studies. Current opinion in biotechnology 41: 26-33
- 828 Jenkinson DS (1990) The turnover of organic carbon and nitrogen in soil. Philosophical  
829 Transactions of the Royal Society of London. Series B: Biological Sciences 329(1255): 361-368
- 830 Jing X, Chen X, Fang J, Ji C, Shen H, Zheng C, Zhu B (2020) Soil microbial carbon and nutrient  
831 constraints are driven more by climate and soil physicochemical properties than by nutrient  
832 addition in forest ecosystems. Soil Biology and Biochemistry 141: 107657

- 833 Jinyun T, Riley WJ (2020) Linear two-pool models are insufficient to infer soil organic matter  
834 decomposition temperature sensitivity from incubations. *Biogeochemistry* 149(3): 251-261
- 835 Keskin H, Grunwald S, Harris WG (2019) Digital mapping of soil carbon fractions with machine  
836 learning. *Geoderma* 339: 40-58
- 837 Kessell AK, McCullough HC, Auchtung JM, Bernstein HC, Song H-S (2020) Predictive  
838 interactome modeling for precision microbiome engineering. *Current Opinion in Chemical*  
839 *Engineering* 30: 77-85
- 840 Khan KS, Mack R, Castillo X, Kaiser M, Joergensen RG (2016) Microbial biomass, fungal and  
841 bacterial residues, and their relationships to the soil organic matter C/N/P/S ratios. *Geoderma*  
842 271: 115-123
- 843 Killinger BJ, Brandvold KR, Ramos-Hunter SJ, Wright AT (2019) Chemoproteomic Analyses  
844 by Activity-Based Protein Profiling. *Mass Spectrometry-Based Chemical Proteomics*: 67-99
- 845 Kirkby C, Kirkegaard J, Richardson A, Wade L, Blanchard C, Batten G (2011) Stable soil  
846 organic matter: a comparison of C: N: P: S ratios in Australian and other world soils. *Geoderma*  
847 163(3-4): 197-208
- 848 Kirkby CA, Richardson AE, Wade LJ, Batten GD, Blanchard C, Kirkegaard JA (2013) Carbon-  
849 nutrient stoichiometry to increase soil carbon sequestration. *Soil Biology and Biochemistry* 60:  
850 77-86
- 851 Kirkby CA, Richardson AE, Wade LJ, Passioura JB, Batten GD, Blanchard C, Kirkegaard JA  
852 (2014) Nutrient availability limits carbon sequestration in arable soils. *Soil Biology and*  
853 *Biochemistry* 68: 402-409
- 854 Kuzyakov Y, Mason-Jones K (2018) Viruses in soil: Nano-scale undead drivers of microbial life,  
855 biogeochemical turnover and ecosystem functions. *Soil Biology and Biochemistry* 127: 305-317

- 856 Kyker-Snowman E, Wieder WR, Frey SD, Grandy AS (2020) Stoichiometrically coupled carbon  
857 and nitrogen cycling in the Microbial-Mineral Carbon Stabilization model version 1.0  
858 (MIMICS-CN v1. 0). *Geoscientific Model Development* 13(9): 4413-4434
- 859 LaPierre N, Ju CJ-T, Zhou G, Wang W (2019) MetaPheno: A critical evaluation of deep learning  
860 and machine learning in metagenome-based disease prediction. *Methods* 166: 74-82
- 861 Lesnik KL, Liu H (2017) Predicting microbial fuel cell biofilm communities and bioreactor  
862 performance using artificial neural networks. *Environmental science & technology* 51(18):  
863 10881-10892
- 864 Li J, Wang G, Allison SD, Mayes MA, Luo Y (2014) Soil carbon sensitivity to temperature and  
865 carbon use efficiency compared across microbial-ecosystem models of varying complexity.  
866 *Biogeochemistry* 119(1-3): 67-84
- 867 Li Z, Yao Q, Guo X, Crits-Christoph A, Mayes MA, Hervey I, Judson W, Lebeis SL, Banfield  
868 JF, Hurst GB (2019) Genome-Resolved Proteomic Stable Isotope Probing of Soil Microbial  
869 Communities using  $^{13}\text{CO}_2$  and  $^{13}\text{C}$ -methanol. *Frontiers in Microbiology* 10: 2706
- 870 Liaw A, Wiener M (2002) Classification and regression by randomForest. *R news* 2(3): 18-22
- 871 Lieven C, Beber ME, Olivier BG, Bergmann FT, Ataman M, Babaei P, Bartell JA, Blank LM,  
872 Chauhan S, Correia K (2020) MEMOTE for standardized genome-scale metabolic model testing.  
873 *Nature biotechnology* 38(3): 272-276
- 874 Loira N, Dulermo T, Nicaud J-M, Sherman DJ (2012) A genome-scale metabolic model of the  
875 lipid-accumulating yeast *Yarrowia lipolytica*. *BMC systems biology* 6(1): 1-9
- 876 Lu H, Li F, Sánchez BJ, Zhu Z, Li G, Domenzain I, Marcišauskas S, Anton PM, Lappa D,  
877 Lieven C (2019) A consensus *S. cerevisiae* metabolic model Yeast8 and its ecosystem for  
878 comprehensively probing cellular metabolism. *Nature communications* 10(1): 1-13

- 879 Luo W, Friedman MS, Shedden K, Hankenson KD, Woolf PJ (2009) GAGE: generally  
880 applicable gene set enrichment for pathway analysis. *BMC bioinformatics* 10(1): 161
- 881 Malik AA, Martiny JB, Brodie EL, Martiny AC, Treseder KK, Allison SD (2020) Defining trait-  
882 based microbial strategies with consequences for soil carbon cycling under climate change. *The*  
883 *ISME journal* 14(1): 1-9
- 884 Malik AA, Puissant J, Buckeridge KM, Goodall T, Jehmlich N, Chowdhury S, Gweon HS,  
885 Peyton JM, Mason KE, van Agtmaal M (2018) Land use driven change in soil pH affects  
886 microbial carbon cycling processes. *Nature communications* 9(1): 1-10
- 887 Manzoni S (2017) Flexible carbon-use efficiency across litter types and during decomposition  
888 partly compensates nutrient imbalances—results from analytical stoichiometric models. *Frontiers*  
889 *in microbiology* 8: 661
- 890 Manzoni S, Čapek P, Mooshammer M, Lindahl BD, Richter A, Šantrůčková H (2017) Optimal  
891 metabolic regulation along resource stoichiometry gradients. *Ecology letters* 20(9): 1182-1191
- 892 Manzoni S, Jackson RB, Trofymow JA, Porporato A (2008) The global stoichiometry of litter  
893 nitrogen mineralization. *Science* 321(5889): 684-686
- 894 Manzoni S, Taylor P, Richter A, Porporato A, Ågren GI (2012) Environmental and  
895 stoichiometric controls on microbial carbon use efficiency in soils. *New Phytologist* 196(1): 79-  
896 91
- 897 Margalef O, Sardans J, Fernández-Martínez M, Molowny-Horas R, Janssens I, Ciais P, Goll D,  
898 Richter A, Obersteiner M, Asensio D (2017) Global patterns of phosphatase activity in natural  
899 soils. *Scientific reports* 7(1): 1-13

- 900 Mattei M, Frunzo L, D'Acunto B, Esposito G, Pirozzi F (2015) Modelling microbial population  
901 dynamics in multispecies biofilms including Anammox bacteria. *Ecological Modelling* 304: 44-  
902 58
- 903 McClure RS, Lee J-Y, Chowdhury TR, Bottos EM, White RA, Kim Y-M, Nicora CD, Metz TO,  
904 Hofmockel KS, Jansson JK (2020) Integrated network modeling approach defines key metabolic  
905 responses of soil microbiomes to perturbations. *Scientific reports* 10(1): 1-9
- 906 Middelboe M, Søndergaard M (1993) Bacterioplankton growth yield: seasonal variations and  
907 coupling to substrate lability and  $\beta$ -glucosidase activity. *Appl. Environ. Microbiol.* 59(11): 3916-  
908 3921
- 909 Milcu A, Heim A, Ellis RJ, Scheu S, Manning P (2011) Identification of general patterns of  
910 nutrient and labile carbon control on soil carbon dynamics across a successional gradient.  
911 *Ecosystems* 14(5): 710-719
- 912 Min K, Lehmeier CA, Billings SA (2016) Carbon availability modifies temperature responses of  
913 heterotrophic microbial respiration, carbon uptake affinity, and stable carbon isotope  
914 discrimination. *Frontiers in microbiology* 7: 2083
- 915 Moorhead DL, Lashermes G, Sinsabaugh RL (2012) A theoretical model of C-and N-acquiring  
916 exoenzyme activities, which balances microbial demands during decomposition. *Soil Biology*  
917 *and Biochemistry* 53: 133-141
- 918 Moorhead DL, Rinkes ZL, Sinsabaugh RL, Weintraub MN (2013) Dynamic relationships  
919 between microbial biomass, respiration, inorganic nutrients and enzyme activities: informing  
920 enzyme-based decomposition models. *Frontiers in microbiology* 4: 223

921 Mooshammer M, Wanek W, Hämmerle I, Fuchslueger L, Hofhansl F, Knoltsch A, Schnecker J,  
922 Takriti M, Watzka M, Wild B (2014a) Adjustment of microbial nitrogen use efficiency to  
923 carbon: nitrogen imbalances regulates soil nitrogen cycling. *Nature communications* 5: 3694

924 Mooshammer M, Wanek W, Zechmeister-Boltenstern S, Richter AA (2014b) Stoichiometric  
925 imbalances between terrestrial decomposer communities and their resources: mechanisms and  
926 implications of microbial adaptations to their resources. *Frontiers in microbiology* 5: 22

927 Morellos A, Pantazi X-E, Moshou D, Alexandridis T, Whetton R, Tziotziou G, Wiebensohn J,  
928 Bill R, Mouazen AM (2016) Machine learning based prediction of soil total nitrogen, organic  
929 carbon and moisture content by using VIS-NIR spectroscopy. *Biosystems Engineering* 152: 104-  
930 116

931 Nauck D, Klawonn F, Kruse R (1997) *Foundations of neuro-fuzzy systems*. John Wiley & Sons,  
932 Inc.,

933 Naylor D, Sadler N, Bhattacharjee A, Graham EB, Anderton CR, McClure R, Lipton M,  
934 Hofmockel KS, Jansson JK (2020) Soil Microbiomes Under Climate Change and Implications  
935 for Carbon Cycling. *Annual Review of Environment and Resources* 45:

936 Notebaart RA, Van Enkevort FH, Francke C, Siezen RJ, Teusink B (2006) Accelerating the  
937 reconstruction of genome-scale metabolic networks. *BMC bioinformatics* 7(1): 296

938 Orth JD, Thiele I, Palsson BØ (2010) What is flux balance analysis? *Nature biotechnology* 28(3):  
939 245-248

940 Oyetunde T, Di Liu HGM, Tang YJ (2019) Machine learning framework for assessment of  
941 microbial factory performance. *PloS one* 14(1):

942 Papp K, Hungate BA, Schwartz E (2020) Glucose triggers strong taxon-specific responses in  
943 microbial growth and activity: insights from DNA and RNA qSIP. *Ecology* 101(1): e02887



- 944 Parton W (1996) The CENTURY model. In: Evaluation of soil organic matter models. Springer.  
945 p 283-291
- 946 Pasolli E, Truong DT, Malik F, Waldron L, Segata N (2016) Machine learning meta-analysis of  
947 large metagenomic datasets: tools and biological insights. PLoS computational biology 12(7):  
948 Patricelli MP, Szardenings AK, Liyanage M, Nomanbhoy TK, Wu M, Weissig H, Aban A, Chun  
949 D, Tanner S, Kozarich JW (2007) Functional interrogation of the kinome using nucleotide acyl  
950 phosphates. Biochemistry 46(2): 350-358
- 951 Pepe-Ranney C, Campbell AN, Koechli CN, Berthrong S, Buckley DH (2016) Unearthing the  
952 ecology of soil microorganisms using a high resolution DNA-SIP approach to explore cellulose  
953 and xylose metabolism in soil. Frontiers in microbiology 7: 703
- 954 Peterson LE (2009) K-nearest neighbor. Scholarpedia 4(2): 1883
- 955 Porta G, la Cecilia D, Guadagnini A, Maggi F (2018) Implications of uncertain bioreactive  
956 parameters on a complex reaction network of atrazine biodegradation in soil. Advances in water  
957 resources 121: 263-276
- 958 Purcell AM, Dijkstra P, Finley B, Hayer M, Koch BJ, Mau RL, Morrissey E, Papp K, Schwartz  
959 E, Stone BW (2019) Quantitative Stable Isotope Probing with H 2 18 O to Measure Taxon-  
960 Specific Microbial Growth. Methods of Soil Analysis 4(1):
- 961 Qdais HA, Hani KB, Shatnawi N (2010) Modeling and optimization of biogas production from a  
962 waste digester using artificial neural network and genetic algorithm. Resources, Conservation  
963 and Recycling 54(6): 359-363
- 964 Qiao Y, Wang J, Liang G, Du Z, Zhou J, Zhu C, Huang K, Zhou X, Luo Y, Yan L (2019) Global  
965 variation of soil microbial carbon-use efficiency in relation to growth temperature and substrate  
966 supply. Scientific reports 9(1): 1-8

- 967 Rahimian Boogar A, Salehi H, Pourghasemi HR, Blaschke T (2019) Predicting Habitat  
968 Suitability and Conserving Juniperus spp. Habitat Using SVM and Maximum Entropy Machine  
969 Learning Techniques. *Water* 11(10): 2049
- 970 Rana P, Berry C, Ghosh P, Fong SS (2020) Recent advances on constraint-based models by  
971 integrating machine learning. *Current Opinion in Biotechnology* 64: 85-91
- 972 Reed DC, Algar CK, Huber JA, Dick GJ (2014) Gene-centric approach to integrating  
973 environmental genomics and biogeochemical models. *Proceedings of the National Academy of  
974 Sciences* 111(5): 1879-1884
- 975 Reiners WA (1986) Complementary models for ecosystems. *The American Naturalist* 127(1):  
976 59-73
- 977 Riley W, Maggi F, Kleber M, Torn M, Tang J, Dwivedi D, Guerry N (2014) Long residence  
978 times of rapidly decomposable soil organic matter: application of a multi-phase, multi-  
979 component, and vertically resolved model (BAMS1) to soil carbon dynamics. *Geoscientific  
980 Model Development* 7(4): 1335-1355
- 981 Sadler NC, Wright AT (2015) Activity-based protein profiling of microbes. *Current opinion in  
982 chemical biology* 24: 139-144
- 983 Safavian SR, Landgrebe D (1991) A survey of decision tree classifier methodology. *IEEE  
984 transactions on systems, man, and cybernetics* 21(3): 660-674
- 985 Saghatelian A, Jessani N, Joseph A, Humphrey M, Cravatt BF (2004) Activity-based probes for  
986 the proteomic profiling of metalloproteases. *Proceedings of the National Academy of Sciences  
987* 101(27): 10000-10005
- 988 Sanman LE, Bogoyo M (2014) Activity-based profiling of proteases. *Annual review of  
989 biochemistry* 83: 249-273

- 990 Sarle WS (1994) Neural networks and statistical models.
- 991 Schimel J, Becerra CA, Blankinship J (2017) Estimating decay dynamics for enzyme activities in  
992 soils from different ecosystems. *Soil Biology and Biochemistry* 114: 5-11
- 993 Schwartz E, Hayer M, Hungate BA, Koch BJ, McHugh TA, Mercurio W, Morrissey EM,  
994 Soldanova K (2016) Stable isotope probing with <sup>18</sup>O-water to investigate microbial growth and  
995 death in environmental samples. *Current opinion in biotechnology* 41: 14-18
- 996 Seaver SM, Liu F, Zhang Q, Jeffryes J, Faria JP, Edirisinghe J, Mundy M, Chia N, Noor E,  
997 Beber ME (2020) The ModelSEED Database for the integration of metabolic annotations and the  
998 reconstruction, comparison, and analysis of metabolic models for plants, fungi, and microbes.  
999 BioRxiv:
- 1000 Sharifzadeh S, Shirley JD, Carlson EE (2018) Activity-based protein profiling methods to study  
1001 bacteria: the power of small-molecule electrophiles. In: *Activity-Based Protein Profiling*.  
1002 Springer. p 23-48
- 1003 Shen H, Zamboni N, Heinonen M, Rousu J (2013) Metabolite identification through machine  
1004 learning—Tackling CASMI challenge using fingerID. *Metabolites* 3(2): 484-505
- 1005 Sihi D, Gerber S, Inglett PW, Inglett KS (2016) Comparing models of microbial–substrate  
1006 interactions and their response to warming. *Biogeosciences* 13(6): 1733-1752
- 1007 Sinsabaugh RL (2010) Phenol oxidase, peroxidase and organic matter dynamics of soil. *Soil*  
1008 *Biology and Biochemistry* 42(3): 391-404
- 1009 Sinsabaugh RL, Lauber CL, Weintraub MN, Ahmed B, Allison SD, Crenshaw C, Contosta AR,  
1010 Cusack D, Frey S, Gallo ME (2008) Stoichiometry of soil enzyme activity at global scale.  
1011 *Ecology letters* 11(11): 1252-1264

- 1012 Sinsabaugh RL, Manzoni S, Moorhead DL, Richter A (2013) Carbon use efficiency of microbial  
1013 communities: stoichiometry, methodology and modelling. *Ecology letters* 16(7): 930-939
- 1014 Sinsabaugh RL, Shah JJF (2011) Ecoenzymatic stoichiometry of recalcitrant organic matter  
1015 decomposition: the growth rate hypothesis in reverse. *Biogeochemistry* 102(1-3): 31-43
- 1016 Sinsabaugh RL, Turner BL, Talbot JM, Waring BG, Powers JS, Kuske CR, Moorhead DL,  
1017 Follstad Shah JJ (2016) Stoichiometry of microbial carbon use efficiency in soils. *Ecological*  
1018 *Monographs* 86(2): 172-189
- 1019 Six J, Frey S, Thiet R, Batten K (2006) Bacterial and fungal contributions to carbon  
1020 sequestration in agroecosystems. *Soil Science Society of America Journal* 70(2): 555-569
- 1021 Smith EM, Prairie YT (2004) Bacterial metabolism and growth efficiency in lakes: the  
1022 importance of phosphorus availability. *Limnology and Oceanography* 49(1): 137-147
- 1023 Song H-S, Stegen JC, Graham EB, Lee J-Y, Garayburu-Caruso V, Nelson WC, Chen X,  
1024 Moulton JD, Scheibe TD (2020) Representing Organic Matter Thermodynamics in  
1025 Biogeochemical Reactions via Substrate-Explicit Modeling. *bioRxiv*:
- 1026 Spohn M (2016) Element cycling as driven by stoichiometric homeostasis of soil  
1027 microorganisms. *Basic and applied ecology* 17(6): 471-478
- 1028 Starr EP, Shi S, Blazewicz SJ, Probst AJ, Herman DJ, Firestone MK, Banfield JF (2018) Stable  
1029 isotope informed genome-resolved metagenomics reveals that Saccharibacteria utilize  
1030 microbially-processed plant-derived carbon. *Microbiome* 6(1): 122
- 1031 Sterner RW (1995) Elemental stoichiometry of species in ecosystems. In: *Linking species &*  
1032 *ecosystems*. Springer. p 240-252
- 1033 Sterner RW, Elser JJ (2002) *Ecological stoichiometry: the biology of elements from molecules to*  
1034 *the biosphere*. Princeton university press,

- 1035 Strickland MS, Rousk J (2010) Considering fungal: bacterial dominance in soils—methods,  
1036 controls, and ecosystem implications. *Soil Biology and Biochemistry* 42(9): 1385-1395
- 1037 Sullivan BW, Alvarez-Clare S, Castle SC, Porder S, Reed SC, Schreeg L, Townsend AR,  
1038 Cleveland CC (2014) Assessing nutrient limitation in complex forested ecosystems: alternatives  
1039 to large-scale fertilization experiments. *Ecology* 95(3): 668-681
- 1040 Sulman BN, Shevliakova E, Brzostek ER, Kivlin SN, Malyshev S, Menge DN, Zhang X (2019)  
1041 Diverse mycorrhizal associations enhance terrestrial C storage in a global model. *Global*  
1042 *Biogeochemical Cycles* 33(4): 501-523
- 1043 Takriti M, Wild B, Schnecker J, Mooshammer M, Knoltsch A, Lashchinskiy N, Alves RJE,  
1044 Gentsch N, Gittel A, Mikutta R (2018) Soil organic matter quality exerts a stronger control than  
1045 stoichiometry on microbial substrate use efficiency along a latitudinal transect. *Soil Biology and*  
1046 *Biochemistry* 121: 212-220
- 1047 Tang J, Riley WJ (2015) Weaker soil carbon–climate feedbacks resulting from microbial and  
1048 abiotic interactions. *Nature Climate Change* 5(1): 56-60
- 1049 Taylor PG, Townsend AR (2010) Stoichiometric control of organic carbon–nitrate relationships  
1050 from soils to the sea. *Nature* 464(7292): 1178-1181
- 1051 Tfaily MM, Chu RK, Tolić N, Roscioli KM, Anderton CR, Paša-Tolić L, Robinson EW, Hess NJ  
1052 (2015) Advanced solvent based methods for molecular characterization of soil organic matter by  
1053 high-resolution mass spectrometry. *Analytical chemistry* 87(10): 5206-5215
- 1054 Tfaily MM, Chu RK, Toyoda J, Tolić N, Robinson EW, Paša-Tolić L, Hess NJ (2017)  
1055 Sequential extraction protocol for organic matter from soils and sediments using high resolution  
1056 mass spectrometry. *Analytica chimica acta* 972: 54-61

- 1057 Thompson J, Johansen R, Dunbar J, Munsky B (2019) Machine learning to predict microbial  
1058 community functions: an analysis of dissolved organic carbon from litter decomposition. *PloS*  
1059 *one* 14(7):
- 1060 Todd-Brown K, Randerson J, Hopkins F, Arora V, Hajima T, Jones C, Shevliakova E, Tjiputra J,  
1061 Volodin E, Wu T (2014) Changes in soil organic carbon storage predicted by Earth system  
1062 models during the 21st century. *Biogeosciences* 11(8): 2341-2356
- 1063 Todd-Brown K, Randerson J, Post W, Hoffman F, Tarnocai C, Schuur E, Allison S (2013)  
1064 Causes of variation in soil carbon simulations from CMIP5 Earth system models and comparison  
1065 with observations. *Biogeosciences* 10(3): 1717-1736
- 1066 Tsilimigras MC, Fodor AA (2016) Compositional data analysis of the microbiome:  
1067 fundamentals, tools, and challenges. *Annals of epidemiology* 26(5): 330-335
- 1068 Van de Waal DB, Elser JJ, Martiny AC, Sterner RW, Cotner JB (2018) Progress in ecological  
1069 stoichiometry. *Frontiers in microbiology* 9: 1957
- 1070 van Groenigen K-J, Six J, Hungate BA, de Graaff M-A, Van Breemen N, Van Kessel C (2006)  
1071 Element interactions limit soil carbon storage. *Proceedings of the National Academy of Sciences*  
1072 103(17): 6571-6574
- 1073 Von Bergen M, Jehmlich N, Taubert M, Vogt C, Bastida F, Herbst F-A, Schmidt F, Richnow H-  
1074 H, Seifert J (2013) Insights from quantitative metaproteomics and protein-stable isotope probing  
1075 into microbial ecology. *The ISME journal* 7(10): 1877-1885
- 1076 Wang G, Jagadamma S, Mayes MA, Schadt CW, Steinweg JM, Gu L, Post WM (2015)  
1077 Microbial dormancy improves development and experimental validation of ecosystem model.  
1078 *The ISME journal* 9(1): 226-237

- 1079 Wang L (2005) Support vector machines: theory and applications. Springer Science & Business  
1080 Media,
- 1081 Wang L, Long F, Liao W, Liu H (2020) Prediction of anaerobic digestion performance and  
1082 identification of critical operational parameters using machine learning algorithms. *Bioresource*  
1083 *Technology* 298: 122495
- 1084 Wang YP, Houlton BZ (2009) Nitrogen constraints on terrestrial carbon uptake: Implications for  
1085 the global carbon-climate feedback. *Geophysical Research Letters* 36(24):
- 1086 Waring BG, Sulman BN, Reed S, Smith AP, Averill C, Creamer CA, Cusack DF, Hall SJ,  
1087 Jastrow JD, Jilling A (2020) From pools to flow: The PROMISE framework for new insights on  
1088 soil carbon cycling in a changing world. *Global Change Biology* 26(12): 6631-6643
- 1089 Weinbauer MG (2004) Ecology of prokaryotic viruses. *FEMS microbiology reviews* 28(2): 127-  
1090 181
- 1091 Were K, Bui DT, Dick ØB, Singh BR (2015) A comparative assessment of support vector  
1092 regression, artificial neural networks, and random forests for predicting and mapping soil organic  
1093 carbon stocks across an Afromontane landscape. *Ecological Indicators* 52: 394-403
- 1094 Whidbey C, Wright AT (2018) Activity-Based Protein Profiling—Enabling Multimodal  
1095 Functional Studies of Microbial Communities. In: *Activity-Based Protein Profiling*. Springer. p  
1096 1-21
- 1097 Wieder W, Grandy A, Kallenbach C, Bonan G (2014) Integrating microbial physiology and  
1098 physio-chemical principles in soils with the MIMICs (MIMICs) model. *Biogeosciences* 11(14): 3899-3917
- 1100 Wieder WR, Bonan GB, Allison SD (2013) Global soil carbon projections are improved by  
1101 modelling microbial processes. *Nature Climate Change* 3(10): 909-912

- 1102 Wieder WR, Cleveland CC, Smith WK, Todd-Brown K (2015) Future productivity and carbon  
1103 storage limited by terrestrial nutrient availability. *Nature Geoscience* 8(6): 441-444
- 1104 Wilhelm RC, Singh R, Eltis LD, Mohn WW (2019) Bacterial contributions to delignification and  
1105 lignocellulose degradation in forest soils with metagenomic and quantitative stable isotope  
1106 probing. *The ISME journal* 13(2): 413-429
- 1107 Wilhelm SW, Suttle CA (1999) Viruses and nutrient cycles in the sea: viruses play critical roles  
1108 in the structure and function of aquatic food webs. *Bioscience* 49(10): 781-788
- 1109 Wilson RM, Neumann RB, Crossen KB, Raab NM, Hodgkins SB, Saleska SR, Bolduc B,  
1110 Woodcroft BJ, Tyson GW, Chanton JP (2019) Microbial community analyses inform  
1111 geochemical reaction network models for predicting pathways of greenhouse gas production.  
1112 *Frontiers in Earth Science* 7: 59
- 1113 Woolf D, Lehmann J (2019) Microbial models with minimal mineral protection can explain  
1114 long-term soil organic carbon persistence. *Scientific reports* 9(1): 1-8
- 1115 Youngblut ND, Barnett SE, Buckley DH (2018a) HTSSIP: an R package for analysis of high  
1116 throughput sequencing data from nucleic acid stable isotope probing (SIP) experiments. *PLoS*  
1117 *One* 13(1):
- 1118 Youngblut ND, Barnett SE, Buckley DH (2018b) SIPSIm: a modeling toolkit to predict accuracy  
1119 and aid design of DNA-SIP experiments. *Frontiers in microbiology* 9: 570
- 1120 Zakem EJ, Polz MF, Follows MJ (2020) Redox-informed models of global biogeochemical  
1121 cycles. *Nature communications* 11(1): 1-10
- 1122 Zampieri G, Vijayakumar S, Yaneske E, Angione C (2019) Machine and deep learning meet  
1123 genome-scale metabolic modeling. *PLoS computational biology* 15(7): e1007084



- 1124 Zegeye EK, Sadler NC, Lomas GX, Attah K, Jansson JK, Hofmockel KS, Anderton CR, Wright  
1125 A (2020) Activity-based protein profiling of chitin catabolism. *ChemBioChem* 21:  
1126 Zhang Q, Wang Y-P, Matear R, Pitman A, Dai Y (2014) Nitrogen and phosphorous limitations  
1127 significantly reduce future allowable CO<sub>2</sub> emissions. *Geophysical Research Letters* 41(2): 632-  
1128 637  
1129

