

1 **Applying conservation reserve design strategies to define ecosystem monitoring priorities**

2

3 Irene Martín-Forés^{1,2*}, Greg R. Guerin^{1,2}, Samantha E.M. Munroe^{1,2}, Ben Sparrow^{1,2}

4

5 ¹ School of Biological Sciences, The University of Adelaide, Adelaide, South Australia 5005,

6 Australia

7 ² Terrestrial Ecosystem Research Network (TERN), University of Adelaide, Adelaide, South

8 Australia 5005, Australia

9

10 *corresponding author: irene.martin@adelaide.edu.au

11

12

13 **Authors' Contributions:**

14 All authors conceived the ideas; IMF analysed the data with help from GRG and SEMM; IMF

15 drafted the paper. All authors reviewed the final version of the manuscript and approved its

16 submission.

17

18 **Abstract**

19 In an era of unprecedented ecological upheaval, accurately monitoring ecosystem change at
20 large spatial scales and over long-time frames is an essential to effective environmental
21 management and conservation. However, economic limitations often preclude revisiting
22 entire monitoring networks at a high enough frequency to accurately detect ecological
23 changes. Thus, a prioritisation strategy is needed to select a subset of sites that meets the
24 principles of complementarity and representativeness of the whole ecological reality. Here,
25 we applied two well-known approaches for conservation design, the 'minimum set' and the
26 'maximal coverage' problems, to develop a strategic monitoring prioritisation procedure
27 that compares potential monitoring sites using a suite of alpha and beta biodiversity
28 metrics. To accomplish this, we created a novel function for the R environment that easily
29 performs biodiversity metric comparisons and site prioritisation on a plot-by-plot basis. We
30 tested our procedures using plot data provided by the Terrestrial Ecosystem Research
31 Network (TERN) AusPlots, an Australian long-term monitoring network of 774 vegetation
32 and soil monitoring plots. We selected 250 plots and 80% of the total species recorded for
33 the maximal coverage and minimum set problems, respectively. We compared the results of
34 each approach in terms of ecological complementarity (species accumulation) and the
35 spatial and environmental representativeness of the plots selected by the different
36 biodiversity metrics. We repeated the selection process for clusters of plots to incorporate
37 logistic constraints for field expeditions. We found that prioritisation based on species
38 turnover (i.e. selection of the most dissimilar plots in terms of species composition but
39 ignoring species richness) maximised ecological complementarity and spatial
40 representativeness, while also providing high environmental coverage. Species richness was
41 an unreliable metric for spatial representation, whereas plot selection based on corrected

42 weighted endemism failed to capture ecological and environmental variation. Range-rarity-
43 richness was a more balanced metric in terms of complementarity and representativeness.
44 Prioritisation based on species turnover is desirable to cover the maximum variability of the
45 whole network.

46 Synthesis and applications: Our results inform monitoring design and conservation
47 priorities, which should consider changes in the turnover component of the beta diversity
48 instead of being based on univariate metrics.

49

50

51 **Introduction**

52 *Ecological change and monitoring*

53 Monitoring ecological change is of the utmost importance in the face of increasing
54 anthropogenic encroachment on natural systems and staggering rates of biodiversity loss
55 worldwide (Spellerberg, 2005).

56 Ecological monitoring programs are a prerequisite for successful environmental
57 policy and decision making, and the development of effective management and
58 conservation programs (Jeffers, 1989; Parr et al., 2003; Sparrow, Edwards et al., 2020).

59 Monitoring programs systematically measure certain ecological variables and processes (e.g.
60 species richness and abundance, net primary production, etc; Vos et al., 2000) over time to
61 inform ecological status and environmental quality (Wolfe et al., 1987; Haase et al., 2018).

62 Updated and thorough long-term records inform about the change between precedent and
63 current ecological status which will guide policy makers, and subsequent restoration and
64 conservation priorities (Lovett et al., 2007; Jones, 2011).

65 Over the past two decades, monitoring programs have been developed at large
66 scales to incorporate broader ecological processes (Yoccoz et al., 2001; Parr et al., 2003).
67 Field-based sampling of terrestrial ecosystems at multi-scales has therefore been required
68 to build comprehensive ecosystem observation networks (Sparrow, Edwards et al., 2020). As
69 such, there are now several examples of extensive monitoring networks established at the
70 continental scale, including the pan-European Integrated Carbon Observation System (ICOS),
71 the National Ecological Observatory Network (NEON; USA), the Global Ecosystems
72 Monitoring (GEM) network across the tropics (Malhi et al., 2021), and the Terrestrial
73 Ecosystem Research Network (TERN) in Australia (Cleverly et al., 2019).

74 However, long-term monitoring programs are expensive. Large financial investments
75 are required to provide standardized surveying training, fieldwork organisation, sample
76 preservation and storage, as well as data curation, access and promotion. Therefore,
77 ecosystem monitoring programs are limited by strict financial constraints (Lovett et al.,
78 2007). Consequently, monitoring resources and funding need to be carefully weighted,
79 allocated, and prioritised (James et al., 1999).

80

81 *Approaches to optimise representation for reserve design and monitoring prioritisation*

82 When resources and funding are limited, 'conservation prioritisation' style strategies should
83 be implemented to select an optimal subset of sites for monitoring. This includes ensuring
84 high information content (i.e., the largest possible set of species), and meeting the
85 principles of complementarity and representativeness (Bennett et al., 2014; Guerin,
86 Williams, Sparrow et al., 2020; Guerin, Williams, Leitch et al., 2020) to be effective and
87 useful for decision-making. Thus, the monitored sites should constitute a spatially and
88 environmentally representative subset of locations to obtain an ecologically representative

89 and complementary sample of habitats and communities, which will ensure both ecological
90 and statistical validity (Cullen, 1990; Vos et al., 2000; Sparrow, Foulkes et al., 2020). Hence,
91 to detect ecological changes in the long-term at large spatial scales, a cost-effective
92 resampling strategy needs to be designed to define a subset of priority sites to be revisited,
93 and overcome the existing resource-limited trade-offs between effective spatial and
94 temporal monitoring (Hewitt & Thrush, 2007).

95 Little agreement has been reached so far in terms of how to best monitor and
96 quantify ecological change (Hill et al., 2016). Normally, fieldwork locations are determined
97 depending on specific objectives (e.g. monitoring biodiversity hotspots; assessing the most
98 anthropogenically disturbed areas, etc.), or ultimately by logistics, feasibility, and
99 administrative constraints. While species richness has been the most commonly employed
100 biodiversity metrics to prioritise areas to monitor and protect, it is not robust or ideal
101 (Gotelli & Colwell, 2001). First, complementarity rather than richness *per se* should be used
102 for location selection through an iterative process (Kirkpatrick, 1983; Justus & Sarkar, 2002).
103 Second, there may be a mismatch between biodiversity hotspots in terms of species
104 richness and centres of endemism (Godoy-Bürki et al., 2014) and it is well known that site
105 selection based on species richness offers a poor optimisation for both monitoring planning
106 and conservation practices (Hillebrand et al., 2018).

107 The 'minimum set' problem and the 'maximal coverage' problem are two common
108 approaches to design conservation reserves aiming to maximise conservation benefits at
109 minimum costs (McIntosh et al., 2017). The minimum set problem is based on ecological
110 constraints; it identifies a set of plots that meets certain conservation targets (typically
111 species) within the fewest possible number of sites (Margules & Pressey, 2000). In contrast,

112 the maximal coverage problem is based on economic constraints and consists of maximising
113 the number of species in a given number of sites (Alagador & Cerdeira, 2020).

114 While heuristic algorithms are effective at optimizing complex reserve design for
115 both minimum set and maximal coverage problems, they can require complex analytical
116 decisions or be computationally expensive (Pressey et al. 1996; Ball et al. 2009). Alternative
117 approaches based on simpler optimisation strategies have been employed, including nature
118 reserve design in California based on rarity-rarity richness (RRR; i.e. the inverse of the
119 number of sites in which it is present; Albuquerque & Beier, 2015), or a trans-frontier
120 conservation area in Southern Africa based on endemism and threatened species (Smith et
121 al., 2008). The species turnover component (i.e. species replacement) of beta diversity
122 (Baselga, 2010) has been proposed as one of the most robust biodiversity metrics to detect
123 ecological changes over time, as it reflects compositional change within plant communities
124 and is a strong indicator of how those communities respond to global change (Hillebrand et
125 al., 2018). Yet, the use of species turnover to define conservation priorities and
126 complementarity is still scarce (but see Socolar et al., 2016). Given the different approaches
127 employed in the literature it is worth investigating how they perform when applying to site
128 prioritisation in a continental ecological monitoring program.

129

130 *TERN AusPlots as a case-study for the implementation of a practical and efficient*
131 *prioritisation method*

132 Here, we aim to identify which biodiversity metric is the best optimiser in terms of selecting
133 a subset of plots that maximises the number of species accumulated while ensuring
134 environmental and spatial representativeness. To do this we have developed a free and easy
135 tool for the R environment to perform an optimisation process applying the maximal

136 coverage problem. The function we developed is called *optim_species* (included in the
137 *ausplotsR* package; Guerin, Saleeba et al., 2020; see Supplementary material S1 for R code
138 details) and it compares a suite of different optimisers related to the biodiversity metrics
139 that are most commonly used in the scientific literature.

140 We tested this methodology on TERN AusPlots, an Australian long-term monitoring
141 network of ecosystem surveillance and monitoring sites distributed at a continental scale
142 (*sensu* Eyre et al., 2011; Sparrow, Edwards et al., 2020), to select a subset of sites that
143 optimise the complementarity and representativeness of the whole network. We applied
144 both the minimum set and maximal coverage approaches to design a revisiting strategy for
145 the collection of AusPlots. Specifically, we aimed to identify which biodiversity metrics could
146 be most efficiently used to achieve an optimal revisiting strategy that maximises the number
147 of species accumulated when imposing i) an ecological constraint (i.e. minimum set problem
148 –consisting on covering 80% of the total number of species recorded in the dataset–) and ii)
149 an economic constraint (i.e. maximal coverage problem –consisting on selecting a subset of
150 250 plots–).

151 For the results of the maximal coverage problem approach (i.e. those obtained with
152 the *optim_species* function), we subsequently compared the environmental and spatial
153 representativeness of the subsets of sites selected by each of those biodiversity metrics as
154 optimisers to determine which is the preferred option to meet our complementarity and
155 representativeness principles.

156

157 **METHODS**

158 **Case study dataset**

159 Our study uses the AusPlots database, a network systemically surveyed over ten years by
160 TERN's Ecosystem Surveillance Program, a component of Australia's land ecosystem
161 observatory (Cleverly et al., 2019). We used species presence and cover data from 774 one-
162 hectare plots to compare biodiversity metrics and test our site prioritisation analysis
163 approach. TERN plots are established in homogenous areas of terrestrial vegetation to take
164 quantitative measurements of vegetation and soil characterisation (Guerin et al., 2017). In
165 each plot, vegetation structure and composition are recorded using the point-intercept
166 module (Sparrow, Foulkes et al., 2020). Parallel transects (10x100 m long) are laid out in a
167 5x5 grid pattern, spaced 20 meters apart. Species identity, cover and growth form are
168 recorded at each 1 m point along each transect, resulting in 1010 survey points per plot.
169 Data for each plot are available and freely accessible within the AusPlots database, and
170 were extracted from the database using *ausplotsR* (v1.2; Guerin, Saleeba et al., 2020;
171 Munroe et al., 2020; TERN 2020).

172 Some of the sites included in the dataset were revisited (i.e. 99 sites had been
173 revisited, 73 of them twice and 26 three times) and different sets of species were recorded
174 on each revisit. Where repeat visits occurred, each was treated as a sample (hereafter we
175 refer each visit as plots for language simplification).

176

177 **Biodiversity metrics**

178 Using the function *optim_species* from the *ausplotsR* package, we compared a selection of
179 often-used biodiversity metrics to be employed as optimisers to define monitoring
180 priorities. The biodiversity metrics included univariate metrics: i) species richness, ii)
181 weighted species richness or range-rarity richness (RRR; Guerin & Lowe, 2017), iii) corrected
182 weighted endemism (CWE; Crisp et al., 2001), iv) Shannon-Wiener diversity index, (Shannon;

183 Shannon & Weaver, 1949) and v) Simpson diversity index (Simpson; Simpson, 1949). Species
184 turnover-based metrics used included: i) pairwise Simpson dissimilarity (Simpson_Beta;
185 Baselga, 2010) and ii) the most frequent selected plots after simulating pairwise Simpson
186 dissimilarity selection over 1000 iterations (Frequent). See Table 1 and Supplementary
187 material S2 for details of these optimisers.

188

189 **Data analyses**

190 ***Multi-site beta diversity***

191 We carried out all statistical analyses in R (R Core Team 2020). To check to what degree
192 biodiversity differences between plots were due to species replacement or species loss we
193 computed multiple-site Sorenson dissimilarities in beta diversity (β_{sor}) accounting for both
194 the spatial turnover (β_{sim}) and the nestedness (β_{nes}) components of beta diversity ($\beta_{sor} =$
195 $\beta_{sim} + \beta_{nes}$; Koleff et al., 2003; Baselga et al., 2018).

196

197 ***Conservation reserve design applied to optimise monitoring strategies***

198 We applied the maximum coverage and the minimum set problems to optimise monitoring
199 site selection to prioritise sites to revisit. For both the minimum set problem and the
200 maximal coverage problem we performed the analyses by adding individual plots. In
201 addition, we applied the maximal coverage problem to clusters of plots to consider a more
202 realistic and feasible scenario because it is unlikely a field team would go to a remote area
203 to only sample one plot.

204 We developed an R function called *optim_species* as part of this study which
205 combines functionalities from the vegan (Oksanen et al., 2019) and betapart (Balsega et al.,
206 2018) packages. The optimisation analysis is captured in this function which can be accessed

207 in `ausplotsR` (Guerin, Saleeba et al., 2020). The function is thus free and easily accessible and
208 can be run on any similar dataset (see R code as well as another example in the
209 supplementary material S1 and S3 for details). Hence, we performed the analyses employing
210 the `optim_species` function, using as data input the species vs sites matrix in terms of
211 presence/absence, except for Shannon and Simpson, for which we used the matrix including
212 percent cover values.

213

214 *The maximal coverage problem*

215 To address the maximal coverage problem, we set to 250 the number of plots to be selected
216 for future revisits and monitoring. We decided on 250 plots within the AusPlots monitoring
217 network because it is within our ability to revisit over a three to five years time scale.

218

219 *The minimum set problem*

220 To address the minimum set problem, we elucidated how many plots we would need to be
221 revisited using each optimiser to account for at least 80% of the overall species richness
222 (2822 species). The minimum set problem was analysed employing the same optimisers
223 described for the maximal coverage problem (Table 1).

224

225 ***Spatial coverage representativeness***

226 To compare spatial coverage representativeness of the plots selected by different
227 optimisers, we computed the Clark and Evans aggregation index (Clark & Evans, 1954) for
228 the spatial point patterns obtained with each of the optimisers using `spatstat` (Baddeley et
229 al., 2015). We applied the cumulative distribution function `cdf` without edge correction
230 because of corresponding to the mean value of nearest neighbour distance distribution

231 function $G(r)$ from a point pattern within an arbitrary shape. The Clark-Evans test values
232 show whether a spatial point pattern distribution is clustered ($R < 1$), or ordered or regular
233 ($R > 1$). We also mapped the location of the 250 selected plots obtained from the maximal
234 coverage problem to visually support the differences in spatial representativeness when
235 applying each of the biodiversity optimisers.

236

237 ***Environmental coverage representativeness***

238 We compared sets of optimised plots for their climatic representativeness across Australia.
239 We extracted data for 25 climatic variables from Harwood et al. (2016) (Supplementary
240 material S4). We assessed plant species composition data from field plots in the order they
241 were selected by the different optimisers, treating successive plots as additions to a
242 cumulative sample of environmental and ecological space. We computed Euclidean
243 distances for environmental variables with the function *vegdist* from the *vegan* package to
244 assess the environmental representativeness of the subsets of plots selected by different
245 optimisers. We implemented the *betadisper* function to analyse multivariate homogeneity
246 of dispersions (distance to group centroid in principal coordinates space) of the cumulative
247 samples (Anderson et al., 2006) for the different optimisers. We plotted the cumulative
248 mean of environmental variation against the subsets of plots selected and visually
249 compared the representativeness when using each of the biodiversity metrics as optimisers.
250 Finally, we conducted a permutation test for homogeneity of multivariate dispersions with
251 999 permutations to explore pairwise comparisons between optimisers with regards to
252 environmental coverage.

253

254 ***Monitoring strategy optimisation considering logistics***

255 *Spatial clustering*

256 To make the optimisation more realistic in terms of field work feasibility, we clustered the
257 774 plots by geographic distance using a modified version of the *CalcDists* function
258 (<https://gist.github.com/sckott/931445>) in which we estimated the distances among plots
259 with the *distCosine* function from the *geosphere* package (Hijmans, 2019). The final number
260 of clusters was 68, with an average number of eleven plots (nine sites) within each of them.
261 The number of sites within each cluster ranged from three to 24 (Supplementary material
262 S5).

263 We aggregated the species presence/absence data of species in the plots comprising
264 each cluster. For the cover data, we calculated the Shannon and Simpson indices per plot,
265 and then calculated the average value of the index for all the plots. We set to 20 the number
266 of clusters to be selected via the same optimisation process. We then compared the species
267 accumulation in the top 20 clusters when employing each of the biodiversity metrics.

268

269 **RESULTS**

270 ***Multi-site beta diversity across Australia***

271 A total of 3528 species were recorded across all of the sampled plots ($n = 774$ plots). The
272 multi-site Sorenson dissimilarity index was 0.998, the species turnover component (i.e.
273 Simpson dissimilarity) corresponded to 0.997, while the nestedness component was only
274 0.001, indicating a very high rate of species replacement across the distributed plot
275 network.

276

277 ***Conservation reserve design applied to optimise monitoring strategies***

278 When comparing species accumulated with each of the optimisers, we observed that the
279 species turnover-based metrics (i.e. the pairwise Simpson dissimilarity with its three
280 implementation variants: Simpson_Beta, Simpson_Random, and Frequent) were the indices
281 that maximised the cumulative number of species (Fig. 1). In particular, the Frequent variant
282 outperformed the other two, with 3,051 species accumulated (86.5% of the species
283 recorded; Supplementary material S6).

284 For univariate indices, the subsets of plots selected by RRR and species richness
285 when applying the maximal coverage problem, accumulated a greater number of species
286 (2,866 and 2,864, respectively, which accounted for 81.2% of all the species recorded in
287 AusPlots sampling) than the rest of the optimisers. When incorporating species frequencies,
288 the Shannon index outperformed the Simpson index for both the maximal coverage
289 problem and the minimum set problem. CWE was a poor optimiser, with 2,024 species
290 accumulated which accounted for 57.4% of the total number of species recorded (Fig. 1;
291 Supplementary material S6).

292

293 ***Spatial coverage representativeness***

294 All the subsets of plots selected were spatially clustered, but they differed among each
295 other regarding their spatial representativeness. To visually complement the results from
296 the Clark-Evan test, we mapped the subsets of plots selected with different optimisers (Fig.
297 2). Species richness was the optimiser that presented the most clustered spatial distribution
298 ($R = 0.366$), followed by Shannon and Simpson optimisers (both displaying $R = 0.408$). Plots
299 selected with RRR and CWE displayed Clark-Evans values of $R = 0.414$ and $R = 0.428$,
300 respectively. From the species turnover-based metrics, pairwise Simpson dissimilarity

301 (Simpson_Beta) showed better spatial coverage ($R = 0.450$), whereas the best optimiser in
302 terms of spatial representativeness was Frequent ($R = 0.545$).

303 Plot selection based on species richness and RRR was geographically biased towards
304 coastal regions, failing to cover remote areas within the Australian outback. This was also
305 the case for the Shannon and Simpson indices. Contrarily, the opposite trend (i.e. optimised
306 plots located towards central and remote areas) was found when selecting plots based on
307 CWE. This suggests that when selecting plots using univariate diversity metrics the results
308 are geographically biased towards sites located either in biodiversity hotspots and areas
309 with milder environmental conditions (e.g. richness) or in remote centres of endemism (e.g.
310 CWE). Plot selection with pairwise Simpson dissimilarity accounts for the species ID and the
311 turnover component; therefore, the spatial distributions displayed with these indices were
312 more balanced in terms of representation across the whole Australia, than those obtained
313 by univariate biodiversity metrics. This trend was enhanced when selecting the most
314 frequent plots after 1,000 simulations of the pairwise Simpson dissimilarity plot selection.

315

316 ***Environmental coverage representativeness***

317 The permutation test for homogeneity of multivariate dispersions showed significant
318 differences in environmental representativeness among optimisers ($F = 6.49$; p -value \leq
319 0.001 ; Table 2). We found that optimisation with CWE was the least representative in terms
320 of environmental coverage (CWE: average distance to median = 3.41), showing significant
321 differences with the environmental coverage of the subsets selected by all the other
322 optimisers. Richness and RRR were the most representative with regards to environmental
323 coverage (Richness and RRR: average distance to median = 4.75 and 4.64, respectively),
324 followed by Simpson_Beta and Frequent (Simpson_Beta and Frequent: average distance to

325 median = 4.33 and 4.46, respectively; Fig. 3; Table 2), with only marginally significant
326 differences between Richness and Simpson_Beta (Table 2). Shannon and Simpson (Shannon
327 and Simpson: average distance to median = 4.15 and 4.07, respectively) were both
328 significantly less environmentally representative than Richness and RRR, while no significant
329 differences were found between the former two and the results obtained by Simpson_Beta
330 and Frequent. Hence, richness was the biodiversity metric that best covered environmental
331 differences when used as the optimiser. Results of the monitoring strategy optimisation for
332 plot clusters are detailed in Supplementary material S7.

333

334 **Discussion**

335 Large-scale monitoring networks aim for high spatial coverage but resource constraints
336 enforce trade-offs between spatial and temporal sampling. Our results, as applied to the
337 TERN Ausplots dataset, clearly demonstrate that to design monitoring strategies that track
338 the greatest number of species, it is better to focus on the turnover component (i.e. species
339 replacement) through diversity partitioning than on univariate diversity indices.

340 Species turnover best optimised the selection of plots from a monitoring network to
341 be revisited more often when applying both the minimum set and the maximal coverage
342 approaches. Species turnover maximised species complementarity and spatial
343 representativeness, without being significantly worse than the other optimisers regarding
344 environmental representativeness. We obtained more robust results when we ran 1,000
345 random-seed iterations and extracted the most frequently selected plots ('Frequent')
346 compared to using a predetermined, fixed seed ('Simpson_Beta'). These results make sense
347 considering that turnover was the most relevant source of change (accounting for 99% of
348 total multi-site beta diversity) among the Australian vegetation communities sampled.

349 When comparing different univariate biodiversity metrics, our results indicated
350 species richness was the worst performing biodiversity metric optimiser in terms of spatial
351 representativeness. This is consistent with previous findings that have also demonstrated
352 richness-based decisions do not meet the complementarity principle (Gotelli & Colwell,
353 2001; Godoy-Bürki et al., 2014), and are biased towards spatial clustering around more
354 tropical climatic conditions (Veitch et al., 2017). Based on these results, we provide further
355 evidence that species richness is not an efficient measure of biodiversity and its change over
356 time (Hillebrand et al., 2018).

357 Optimisation based on corrected weighted endemism (i.e. CWE) failed to be
358 environmentally, spatially and ecologically representative in terms of biodiversity, with the
359 lowest number of species accumulated across the whole network (worse than random) and
360 the worst environmental representativeness. The poor performance of optimisation based
361 on CWE has implications for monitoring strategies and conservation planning. While
362 conservation reserves could aim to protect endemic species and therefore should consider
363 this metric (Pelletier et al., 2018), monitoring priorities should not be based on CWE, as it
364 will neither meet the principle of complementarity nor representativeness of the whole
365 network and will fail to inform on the ecological reality.

366 Among the univariate biodiversity metrics, RRR was the most balanced, capturing a
367 great number of species and being spatially and environmentally representative. Its
368 estimation is straightforward from incidence datasets; therefore, when seeking a simple but
369 relatively reliable way to select sites for a monitoring program, from the univariate metrics
370 we recommend using RRR as an alternative to species turnover-based prioritisation. Our
371 findings are supported by previous work demonstrating the great ecological representation
372 of this index, as well as its effectiveness as a surrogate for biodiversity when fitted to

373 environmental models to predict biodiversity in the absence of available data (Albuquerque
374 & Beier, 2015, 2016).

375 The Shannon and Simpson optimisers performed comparatively poorly in the three
376 dimensions studied here (i.e. ecological, spatial and environmental representativeness).
377 Hence, plot selection prioritisation processes should preferably not be based on these
378 metrics.

379 Both reserve design approaches (minimum set problem and maximal coverage
380 problem) displayed similar results in terms of species accumulation, but with important
381 consideration of the threshold *a priori* selected regardless of the approach. For example, as
382 observed in Figure 1, species accumulations curves for some metrics crossed-over when
383 reaching approximately either the 70% of the species (minimum set problem) or 150 plots
384 (maximal coverage problem). This suggests the target matters and robust results may be
385 jeopardised if thresholds are set too low.

386 When implementing optimisation for clusters of plots, differences in ecological
387 representativeness were diluted relative to plot-by-plot selection (except in the case of
388 CWE, which still performed significantly worse than the rest of the optimisers).
389 Nevertheless, selection based on species turnover (most specifically employing the Frequent
390 optimiser) performed best, with Frequent and Simpson_Beta approaches the most, and
391 second most, environmentally and spatially representative, respectively. This has
392 implications for hands-on applications of the current findings, since the prioritisation of
393 clusters of plots will need to be carefully supervised to ensure complementarity and
394 representativeness. We therefore suggest that practitioners perform plot-by-plot
395 optimisation to get the ideal subset and then apply logistic principles to determine clusters
396 of plots in a given geographic area.

397 Our results have potential application to conservation reserve design, whereby
398 species turnover metrics could be considered to optimise complementarity and
399 representativeness. Various criteria have been followed to design conservation reserves
400 historically, including maximising species richness or genetic diversity, protecting rare or
401 endemic species or restoring impacted or degraded areas (Margules et al., 1982; Kingsland,
402 2002). In this sense, Simpson pairwise dissimilarity is potentially useful as it selects a
403 representative subset of the habitats and flora within a region.

404 The application of the findings reported here has some limitations. For example,
405 optimisation and therefore reserve design based on species turnover relies on already
406 available ground data and sampled communities and in some cases this information is
407 incomplete or even non-existent. The optimisation process employed in this study (and the
408 tools developed for the analysis) are able to be implemented in a variety of studies, and that
409 can potentially be extended to similar approaches such as site selection based on
410 phylogenetic or functional alpha and beta diversity. Similarly, it could be used to detect
411 change in ecosystem composition over time in the context of a spatial framework; or within
412 a temporal framework to identify sites with the most dissimilar samples among revisits, i.e.
413 sites where vegetation is shifting more rapidly over time. These techniques will enable
414 large-scale monitoring programs to maximise the value of information at a given resourcing
415 level.

416 In summary, monitoring ecological state, function and change over time has become
417 essential across national scales. The selection of sites for regular monitoring based on
418 univariate biodiversity metrics (e.g. richness, CWE) often fails to meet the principles of
419 complementarity and representativeness. We have therefore developed a practical, free
420 and easy-to-use tool that can be used in any species versus sites dataset. The tool uses a set

421 of alpha and beta diversity metrics to optimise species representation in a subset of
422 monitoring sites to maximise species complementarity and spatial and environmental
423 representation. Our results demonstrate that a representative subset of monitoring sites
424 can be selected by finding the most ecologically dissimilar communities. This approach
425 targets differences in composition instead of focusing on univariate metrics such as species
426 richness, while also capturing spatial and environmental diversity. Long-term monitoring
427 sampling strategies need to be carefully planned and designed. Applying reserve design
428 approaches based on spatial vegetation compositional differences to maximise coverage
429 constitutes a cost-effective and easily updated strategy to define monitoring priorities that
430 leverages ground data already collected. This will in turn help policy, decision-making and
431 conservation practices ensuring them to be based on accurate information that meets the
432 complementarity and representativeness principles.

433

434 **Acknowledgements**

435 We acknowledge the TERN Ecosystem Surveillance field team (especially Emrys Leitch), and
436 volunteers as well as support by the Australian Government through the National
437 Collaborative Research Infrastructure Strategy. We have no conflicts of interest to declare.

438

439 **Data availability**

440 The dataset will be published in a public repository upon the acceptance of the manuscript

441 **References**

- 442 Alagador, D., & Cerdeira, J. O. (2020). Revisiting the minimum set cover, the maximal
443 coverage problems and a maximum benefit area selection problem to make
444 climate-change-concerned conservation plans effective. *Methods in Ecology and*
445 *Evolution*, *11*, 1325–1337.
- 446 Albuquerque, F., & Beier, P. (2015). Rarity-weighted richness: a simple and reliable
447 alternative to integer programming and heuristic algorithms for minimum set and
448 maximum coverage problems in conservation planning. *PLoS One*, *10*, e0119905.
- 449 Albuquerque, F., & Beier, P. (2016). Predicted rarity-weighted richness, a new tool to
450 prioritize sites for species representation. *Ecology and Evolution*, *6*, 8107–8114.
- 451 Anderson, M. J., Ellingsen, K. E., & McArdle, B. H. (2006). Multivariate dispersion as a
452 measure of beta diversity. *Ecology Letters*, *9*, 683–693.
- 453 Baddeley, A., Rubak, E., & Turner, R. (2015). Spatial Point Patterns: Methodology and
454 Applications with R. London:Chapman and Hall/CRC Press, 2015. URL
455 [http://www.crcpress.com/Spatial-Point-Patterns-Methodology-and-Applications-](http://www.crcpress.com/Spatial-Point-Patterns-Methodology-and-Applications-with-R/Baddeley-Rubak-Turner/9781482210200/)
456 [with-R/Baddeley-Rubak-Turner/9781482210200/](http://www.crcpress.com/Spatial-Point-Patterns-Methodology-and-Applications-with-R/Baddeley-Rubak-Turner/9781482210200/)
- 457 Ball, I. R., Possingham, H. P., & Watts, M. (2009). Marxan and relatives: software for spatial
458 conservation prioritisation. *Spatial conservation prioritisation: Quantitative*
459 *methods and computational tools*, 185-195.
- 460 Baselga, A. (2010). Partitioning the turnover and nestedness components of beta diversity.
461 *Global Ecology and Biogeography*, *19*, 134–143.
- 462 Baselga, A., Orme, D., Vileger, S., De Bortoli, J. & Leprieur, F. (2018). betapart: Partitioning
463 Beta Diversity into Turnover and Nestedness Components. R package version
464 1.5.1. <https://CRAN.R-project.org/package=betapart>

465 Bennett, J. R., Sisson, D. R., Smol, J. P., Cumming, B. F., Possingham, H. P., & Buckley, Y. M.
466 (2014). Optimizing taxonomic resolution and sampling effort to design cost-
467 effective ecological models for environmental assessment. *Journal of Applied*
468 *Ecology*, 51, 1722–1732.

469 Clark, P. J. & Evans, F.C. (1954). Distance to nearest neighbour as a measure of spatial
470 relationships in populations. *Ecology*, 35, 445–453.

471 Cleverly, J., Eamus, D., Edwards, W., Grant, M., Grundy, M. J., Held, A., Karan, M., Lowe, A.
472 J., Prober, S. M., Sparrow, B. & Morris, B. (2019). TERN, Australia’s land
473 observatory: addressing the global challenge of forecasting ecosystem responses
474 to climate variability and change. *Environmental Research Letters*, 14, 095004

475 Csuti, B., Polasky, S., Williams, P., Pressey, R. L., Camm, J., Kershaw, M., Kierster, A. R.,
476 Downs, B., Hamilton, R., Huso, M., & Sahr, K. (1997). A comparison of reserve
477 selection algorithms using data on terrestrial vertebrates in Oregon. *Biological*
478 *Conservation*, 80, 83–97.

479 Cullen, P. (1990). Biomonitoring and environmental management. *Environmental*
480 *Monitoring Assessment*, 14, 107–114.

481 Godoy-Bürki, A. C., Ortega-Baes, P., Sajama, J. M., & Aagesen, L. (2014). Conservation
482 priorities in the Southern Central Andes: mismatch between endemism and
483 diversity hotspots in the regional flora. *Biodiversity and Conservation*, 23, 81–107.

484 Gotelli, N. J., & Colwell, R. K. (2001). Quantifying biodiversity: procedures and pitfalls in the
485 measurement and comparison of species richness. *Ecology Letters*, 4, 379–391.

486 Guerin, G. R., Sparrow, B., Tokmakoff, A., Smyth, A., Leitch, E., Baruch, Z., Lowe, A.J. (2017)
487 Opportunities for integrated ecological analysis across inland Australia with
488 standardised data from AusPlots Rangelands. *PLoS ONE*, 12, e0170137

489 Guerin, G., Saleeba, T., Munroe, S., Blanco-Martin, B., Martín-Forés, I., Tokmakoff, A. (2020).
490 ausplotsR: TERN AusPlots analysis package. R package version 1.2. [https://CRAN.R-](https://CRAN.R-project.org/package=ausplotsR)
491 [project.org/package=ausplotsR](https://CRAN.R-project.org/package=ausplotsR)

492 Guerin, G. R., Williams, K. J., Sparrow, B., & Lowe, A. J. (2020). Stocktaking the
493 environmental coverage of a continental ecosystem observation network.
494 *Ecosphere*, *11*, e03307.

495 Guerin, G. R., Williams, K. J., Leitch, E., Lowe, A. J., & Sparrow, B. (2020). Using generalised
496 dissimilarity modelling and targeted field surveys to gap-fill an ecosystem
497 surveillance network. *Journal of Applied Ecology*.

498 Haase, P., Tonkin, J. D., Stoll, S., Burkhard, B., Frenzel, M., Geijzendorffer, I. R., Häuser, C.,
499 Klotz, S., Kühn, I., McDowell, W. H., Mirtl, M., Müller, F., Musche, M., Penner, J.,
500 Zacharias, S., & Schmeller, D. S. (2018). The next generation of site-based long-
501 term ecological monitoring: Linking essential biodiversity variables and ecosystem
502 integrity. *Science of the Total Environment*, *613*, 1376–1384.

503 Harwood, T., Donohue, R., Harman, I., McVicar, T., Ota, N., Perry, J., & Williams, K. (2016) 9s
504 climatology for continental Australia 1976-2005: Summary variables with elevation
505 and radiative adjustment. v1. In: CSIRO (ed.) Data Collection. CSIRO,
506 <https://doi.org/10.4225/08/5afa9f7d1a552>.

507 Hewitt, J. E., & Thrush, S. F. (2007). Effective long-term ecological monitoring using spatially
508 and temporally nested sampling. *Environmental Monitoring and Assessment*, *133*,
509 295–307.

510 Hijmans, R. J. (2019). geosphere: Spherical Trigonometry. R package version 1.5-10.
511 <https://CRAN.R-project.org/package=geosphere>

512 Hill, S. L. L., Harfoot, M., Purvis, A., Purves, D. W., Collen, B., Newbold, T., Burgess, N. D. &
513 Mace, G. M. (2016). Reconciling biodiversity indicators to guide understanding and
514 action. *Conservation Letters*, 9, 405–412.

515 Hillebrand, H., Blasius, B., Borer, E. T., Chase, J. M., Downing, J. A., Eriksson, B. K., Filstrup, C.
516 T., Harpole, W.S., Hodapp, D., Larsen, S., Lewandowska, A. M., Seabloom, E. W.,
517 Van de Waal, D. B., & Ryabov, A. B. (2018). Biodiversity change is uncoupled from
518 species richness trends: Consequences for conservation and monitoring. *Journal of*
519 *Applied Ecology*, 55, 169–184.

520 James, A.N., Green, M.J.B., Paine, J.R. (1999). A Global Review of Protected Area Budgets
521 and Staff, WCMC–World Conservation: Cambridge.

522 Jeffers, J. N. R. (1989) Environmental monitoring. *Biologist*, 36, 171

523 Jones, J.P.G. (2011) Monitoring species abundance and distribution at the landscape scale.
524 *Journal of Applied Ecology*, 48, 9–13.

525 Justus, J., & Sarkar, S. (2002). The principle of complementarity in the design of reserve
526 networks to conserve biodiversity: a preliminary history. *Journal of Biosciences*, 27,
527 421–435.

528 Kingsland, S. E. (2002). Creating a science of nature reserve design: perspectives from
529 history. *Environmental Modeling and Assessment*, 7, 61–69.

530 Kirkpatrick, J. B. (1983). An iterative method for establishing priorities for the selection of
531 nature reserves: an example from Tasmania. *Biological Conservation*, 25, 127–134.

532 Koleff, P., Gaston, K. J., & Lennon, J. J. (2003). Measuring beta diversity for presence–
533 absence data. *Journal of Animal Ecology*, 72, 367–382.

534 Lovett, G. M., Burns, D. A., Driscoll, C. T., Jenkins, J. C., Mitchell, M. J., Rustad, L., Shanley, J.
535 B., Likens, G. E., & Haeuber, R. (2007). Who needs environmental monitoring?.
536 *Frontiers in Ecology and the Environment*, 5, 253–260.

537 Malhi, Y., Girardin, C., Metcalfe, D. B., Doughty, C. E., Aragão, L. E., Rifai, S. W., ... & Phillips,
538 O. L. (2021). The Global Ecosystems Monitoring network: Monitoring ecosystem
539 productivity and carbon cycling across the tropics. *Biological Conservation*, 253,
540 108889.

541 Margules, C., Higgs, A. J., & Rafe, R. W. (1982). Modern biogeographic theory: are there any
542 lessons for nature reserve design?. *Biological Conservation*, 24, 115–128.

543 Margules, C.R. & Pressey, R.L. (2000). Systematic conservation planning. *Nature*, 405, 243–
544 253

545 McIntosh, E. J., Pressey, R. L., Lloyd, S., Smith, R., & Grenyer, R. (2017). The impact of
546 systematic conservation planning. *Annual Review of Environment and Resources*,
547 42, 677–697.

548 Moilanen, A., Wilson, K. & Possingham, H.P. (Eds.) (2009) Spatial conservation prioritization.
549 Oxford: Oxford Univ. Press.

550 Munroe, S., Guerin, G., Saleeba, T., Martín-Forés, I., Blanco-Martin, B., Sparrow B. &
551 Tokmakoff, A. (2020) *ausplotsR*: An R package for rapid extraction and analysis of
552 vegetation and soil data collected by Australia’s Terrestrial Ecosystem Research
553 Network. *Ecoevorxiv*. Doi: 1032942/osf.io/25phx

554 Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P. R.,
555 O’Hara, R. B., Simpson, G. L., Solymos, P., Stevens, M. H. H., Szoecs, E., Wagner, H.
556 (2019). *vegan*: Community Ecology Package. R package version 2.5-6.
557 <https://CRAN.R-project.org/package=vegan>

558 Parr, T. W., Sier, A. R. J., Battarbee, R. W., Mackay, A., & Burgess, J. (2003). Detecting
559 environmental change, science and society – perspectives on long-term research
560 and monitoring in the 21st century. *Science of the Total Environment*, 310, 1–8.

561 Pelletier, T. A., Carstens, B. C., Tank, D. C., Sullivan, J., & Espíndola, A. (2018). Predicting
562 plant conservation priorities on a global scale. *Proceedings of the National
563 Academy of Sciences*, 115, 13027–13032.

564 Pressey, R. L., Possingham, H. P., & Margules, C. R. (1996). Optimality in reserve selection
565 algorithms: when does it matter and how much?. *Biological Conservation*, 76, 259–
566 267.

567 R Core Team (2020). R: A language and environment for statistical computing. R Foundation
568 for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.

569 Shannon, C.E. & Weaver, W. (1949) The mathematical theory of communication. The
570 University of Illinois Press, Urbana, 117pp.

571 Simpson, E. H. (1949) Measurement of species diversity. *Nature*, 163, 688.

572 Smith, R. J., Easton, J., Nhancale, B. A., Armstrong, A. J., Culverwell, J., Dlamini, S. D.,
573 Goodman, P. S., Loffler, L., Matthews, W. S., Monadjem, A., Mulqueeny, C. M.,
574 Ngwenya, P., Ntumi, C. P., Soto, B. & Leader-Williams, N. (2008). Designing a
575 transfrontier conservation landscape for the Maputaland centre of endemism
576 using biodiversity, economic and threat data. *Biological Conservation*, 141, 2127–
577 2138.

578 Socolar, J. B., Gilroy, J. J., Kunin, W. E., & Edwards, D. P. (2016). How should beta-diversity
579 inform biodiversity conservation?. *Trends in Ecology and Evolution*, 31, 67–80.

580 Sparrow, B. D., Edwards, W., Munroe, S. E., Wardle, G. M., Guerin, G. R., Bastin, J. F., Morris,
581 B., Christensen, R., Phinn, S. & Lowe, A. J. (2020). Effective ecosystem monitoring
582 requires a multi-scaled approach. *Biological Reviews*, 95, 1706–1719.

583 Sparrow, B., Foulkes, J., Wardle, G., Leitch, E., Caddy-Retalic, S., van Leeuwen, S., Tokmakoff,
584 A., Thurgate, N. Y., Guerin, G. R. & Lowe, A. J. (2020). A vegetation and soil survey
585 method for surveillance monitoring of rangeland environments. *Frontiers in*
586 *Ecology and Evolution*, 8, 157.

587 Spellerberg, I. F. (2005). Monitoring ecological change. Cambridge University Press.

588 TERN (2020) AusPlots ecosystem surveillance monitoring dataset (<http://aekos.org.au/>).
589 Obtained via the `ausplotsR` R package, accessed June 2020.

590 Veach, V., Di Minin, E., Pouzols, F. M., & Moilanen, A. (2017). Species richness as criterion
591 for global conservation area placement leads to large losses in coverage of
592 biodiversity. *Diversity and Distributions*, 23, 715–726.

593 Vos, P., Meelis, E., Ter Keurs, W.J. (2000). A framework for the design of ecological
594 monitoring programs as a tool for environmental and nature management.
595 *Environmental Monitoring and Assessment* 61, 317–344

596 Yoccoz, N. G., Nichols, J. D., & Boulinier, T. (2001). Monitoring of biological diversity in space
597 and time. *Trends in Ecology and Evolution*, 16, 446–453.




Table 1. Optimisers description

Optimiser ID	Optimiser name	Description	Special utilisation / Best used
Richness	Species richness	Count of the number of species present in a given site	Identify biodiversity hotspots
RRR	Range-rarity richness	Inverse of the number of sites in which a species occurs. $RRR = \sum_1^n 1/c_i$, where c_i is the number of map grid cells occupied by species i and n is the number of species	When the goal is to identify areas of high biodiversity and biological uniqueness
CWE	Corrected weighted endemism	Range rarity richness (RRR) divided by species richness. $CWE = (\sum_1^n 1/c_i)/n$, where c_i is the number of map grid cells occupied by species i and n is the number of species	When the goal is to identify centres of endemism highlighting range-restricted species
Shannon	Shannon-Wiener diversity index	Combines species richness and the evenness or equitability by computing the species' relative abundances. $H' = -\sum_{i=1}^S p_i \log_n p_i$, where S is the species richness and p_i is the relative abundance of the species	It assumes that all species are represented in a sample and that they are randomly sampled
Simpson	Simpson diversity index	Combines species richness and the evenness or equitability by computing the species' relative abundances $D = 1 - \sum p_i^2$, where p_i is the proportional abundance of species i	It is the complement of Simpson's original dominance index, and represents the probability that two randomly chosen individuals belong to different species
Simpson_Beta	Pairwise Simpson dissimilarity index	It is based on diversity partitioning, which separates species replacement (i.e. turnover) from species loss (i.e. nestedness). The Simpson dissimilarity corresponds to the turnover component of the Sorensen dissimilarity. Considering two sites, $\beta sim = \min(b,c) / (a + \min(b,c))$, where a is the number of	It is used to maximise species turnover

	species present in both sites, b is the number of species present in the first site, but not in the second, and c is the number of species present in the second site, but not in the first.
Frequent	The most frequent plots selected over 1,000 iterations with a randomised starting seed using the pairwise Simpson dissimilarity index
Simpson_Random	The pairwise Simpson dissimilarity index with a randomised starting seed iterated 1,000 times

DRAFT

Table 2. Pairwise comparisons between optimisers with regards to environmental representativeness when applying maximal coverage problem at plot level. The observed p-value are located in the below diagonal, while the permuted p-value are in the above diagonal. Only significant differences are highlighted in bold. Notice that marginally significant values (p-value ≤ 0.1) are shown although not highlighted.

	Richness	RRR	CWE	Shannon	Simpson	Simpson_Beta	Frequent
Richness		0.68	≤ 0.001	≤ 0.01	≤ 0.05	≤ 0.1	0.25
RRR	0.68		≤ 0.001	≤ 0.1	≤ 0.05	0.20	0.49
CWE	≤ 0.001	≤ 0.001		≤ 0.01	≤ 0.01	≤ 0.001	≤ 0.001
Shannon	≤ 0.05	≤ 0.1	≤ 0.01		0.78	0.48	0.21
Simpson	≤ 0.01	≤ 0.05	≤ 0.01	0.75		0.29	0.12
Simpson_Beta	≤ 0.1	0.22	≤ 0.001	0.48	0.30		0.58
Frequent	0.25	0.48	≤ 0.001	0.21	0.12	0.58	

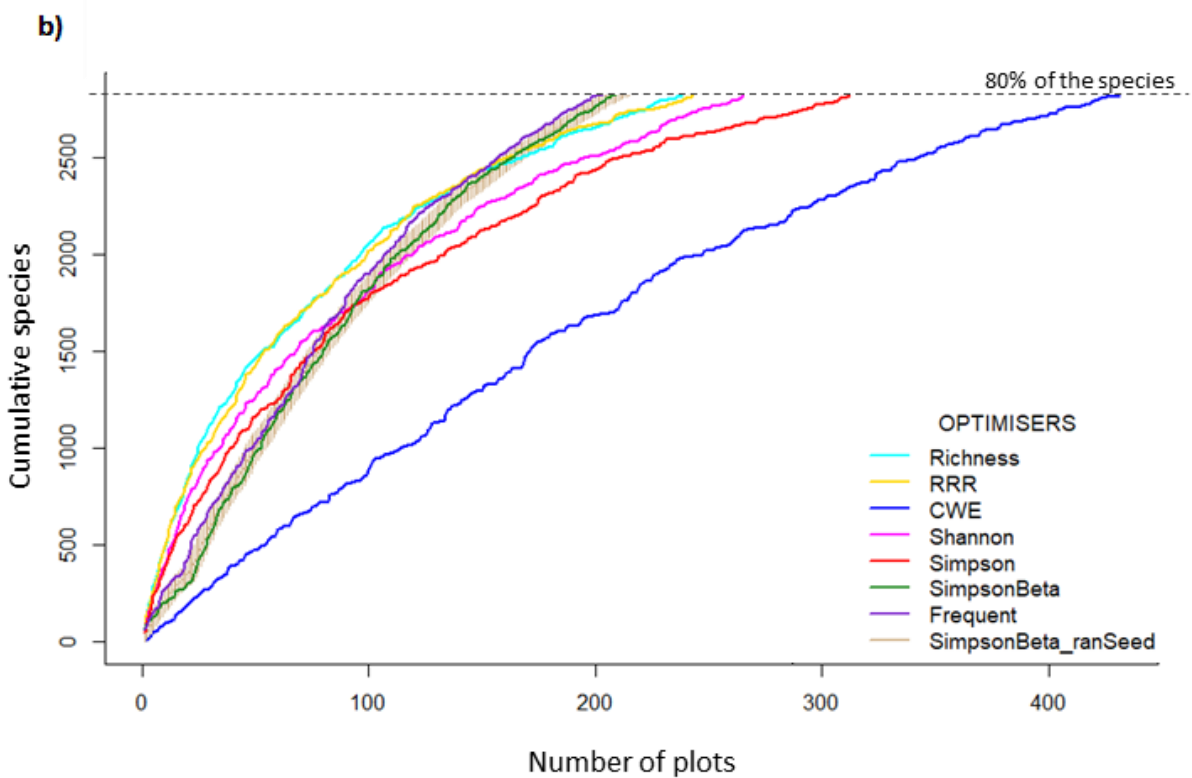
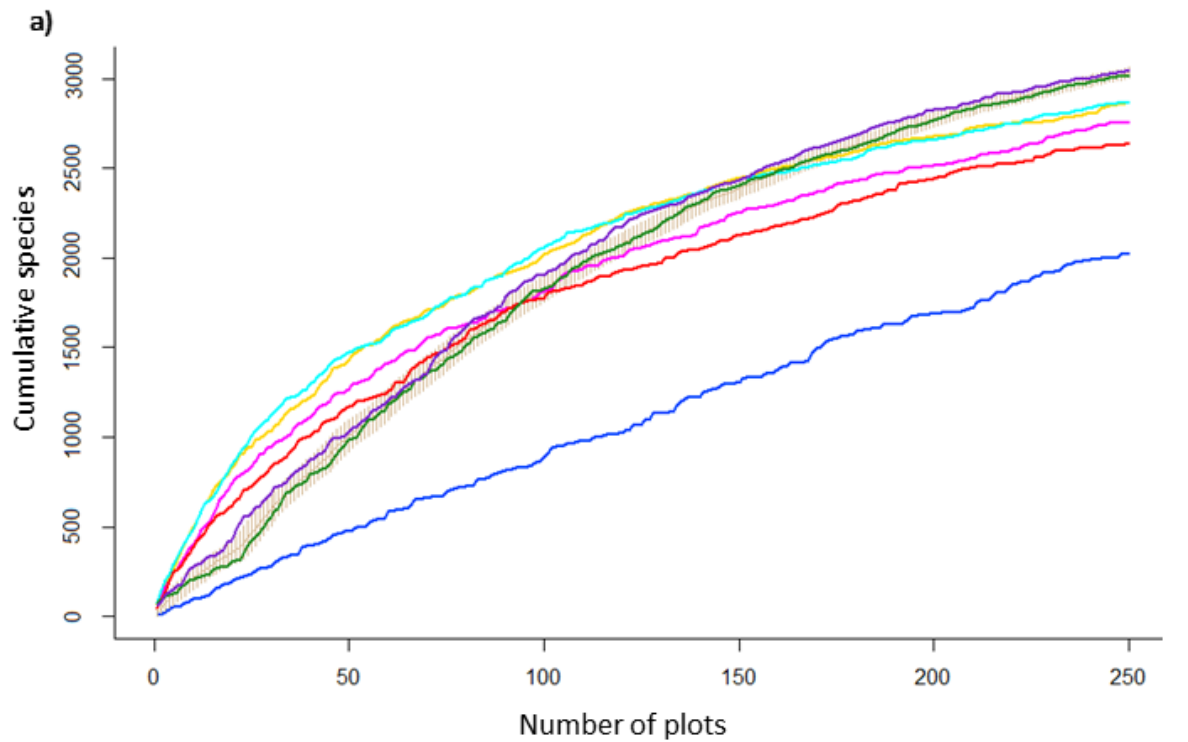


Fig. 1. Site optimisation process applying conservation reserve design strategies based on: a) Maximum coverage problem (selection of 250 sites), and b) Minimum set problem (selection of the minimum number of plots that allow including 80% of the species,

represented by dashed line). Optimisation has been performed in both cases employing different optimisers, including species richness, range rarity richness (RRR), corrected weighted endemism (CWE), Shannon-Wiener diversity index (Shannon), Simpson diversity index (Simpson), the turnover component of beta diversity or pairwise Simpson dissimilarity index (Simpson beta), the most frequent plots selected in 1000 iterations with a randomised starting seed using the pairwise Simpson dissimilarity index (Frequent), and the plots selected with a randomised seed using the pairwise Simpson dissimilarity index (SimpsonBeta_random seed).

DRAFT

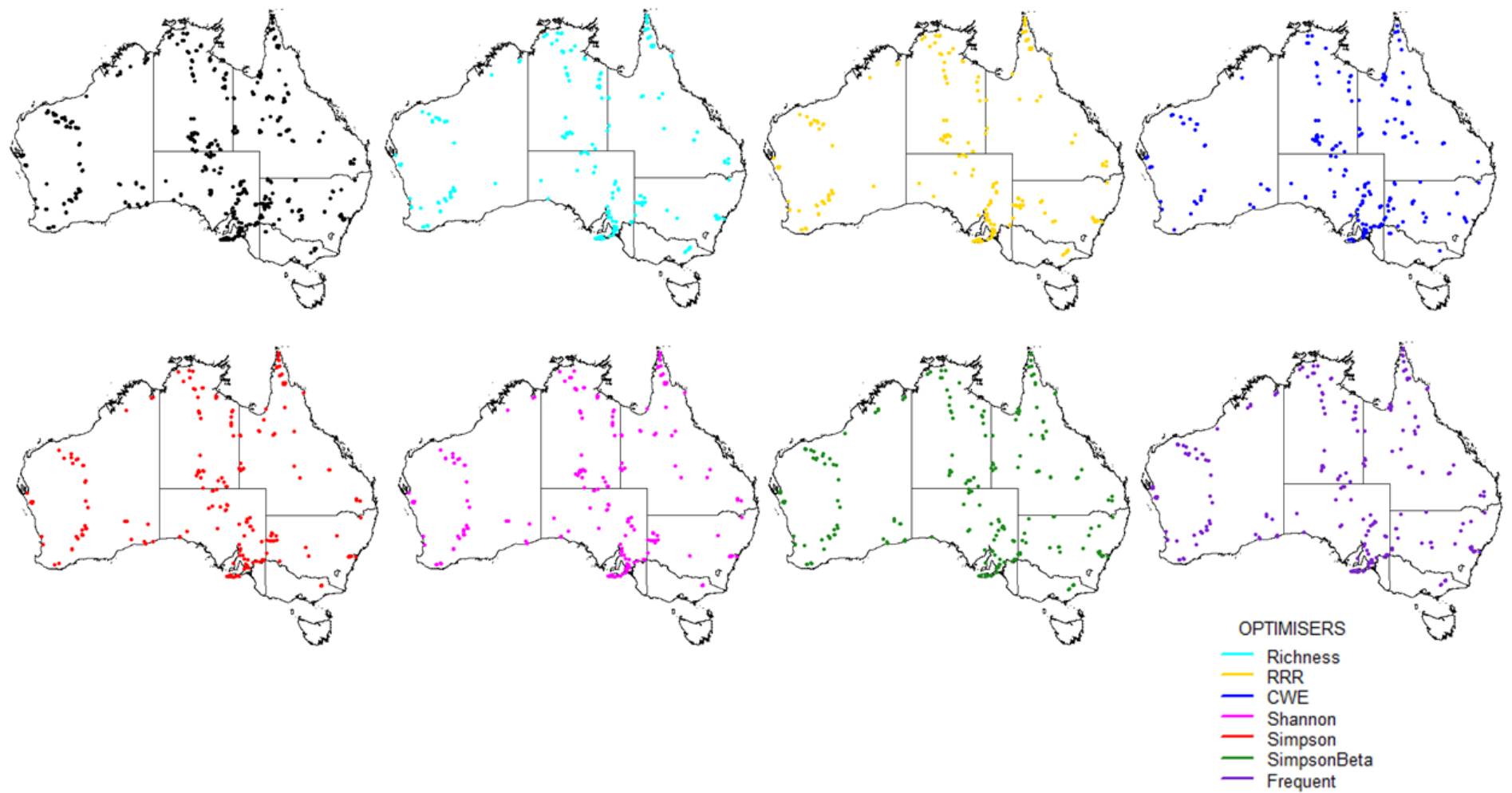


Fig. 2. Geographic location of the selected plots ($N = 250$) applying the maximum coverage problem. Black dots correspond to all the plots established. Colour dots refer to each of the selection employing different optimisers.

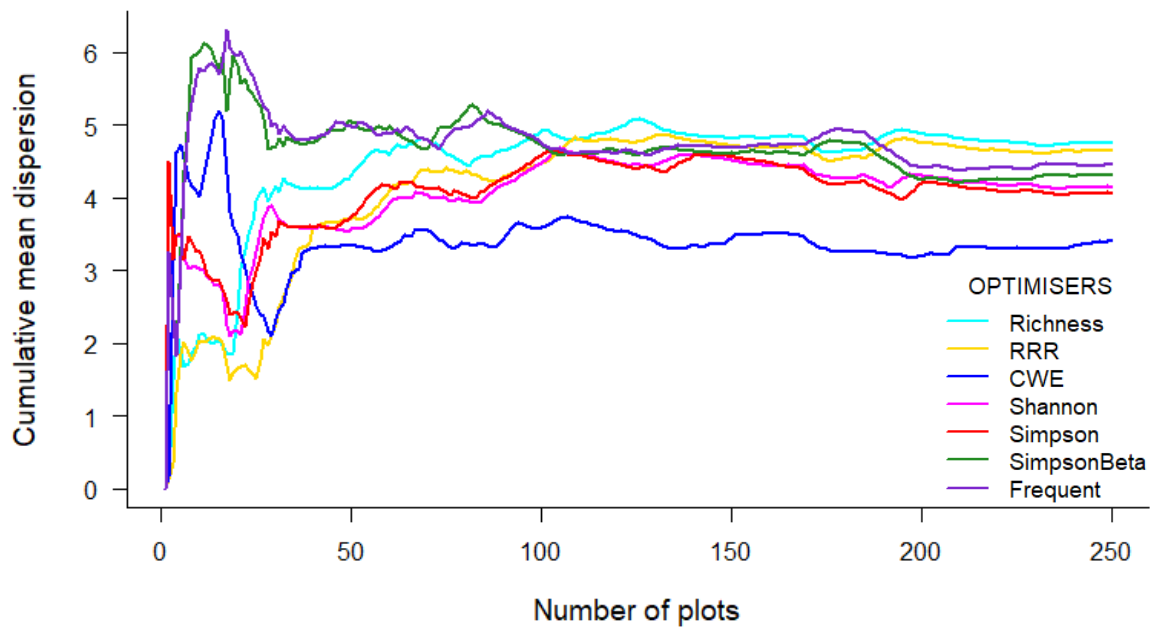


Fig. 3. Environmental representativeness of the 250 selected plots using different optimisers reflected by the cumulative mean dispersion. All environmental variables employed in the analyses are described in the Supplementary material S4.

Supplementary material S1.

Rcode

Not included in the preprint.

DRAFT

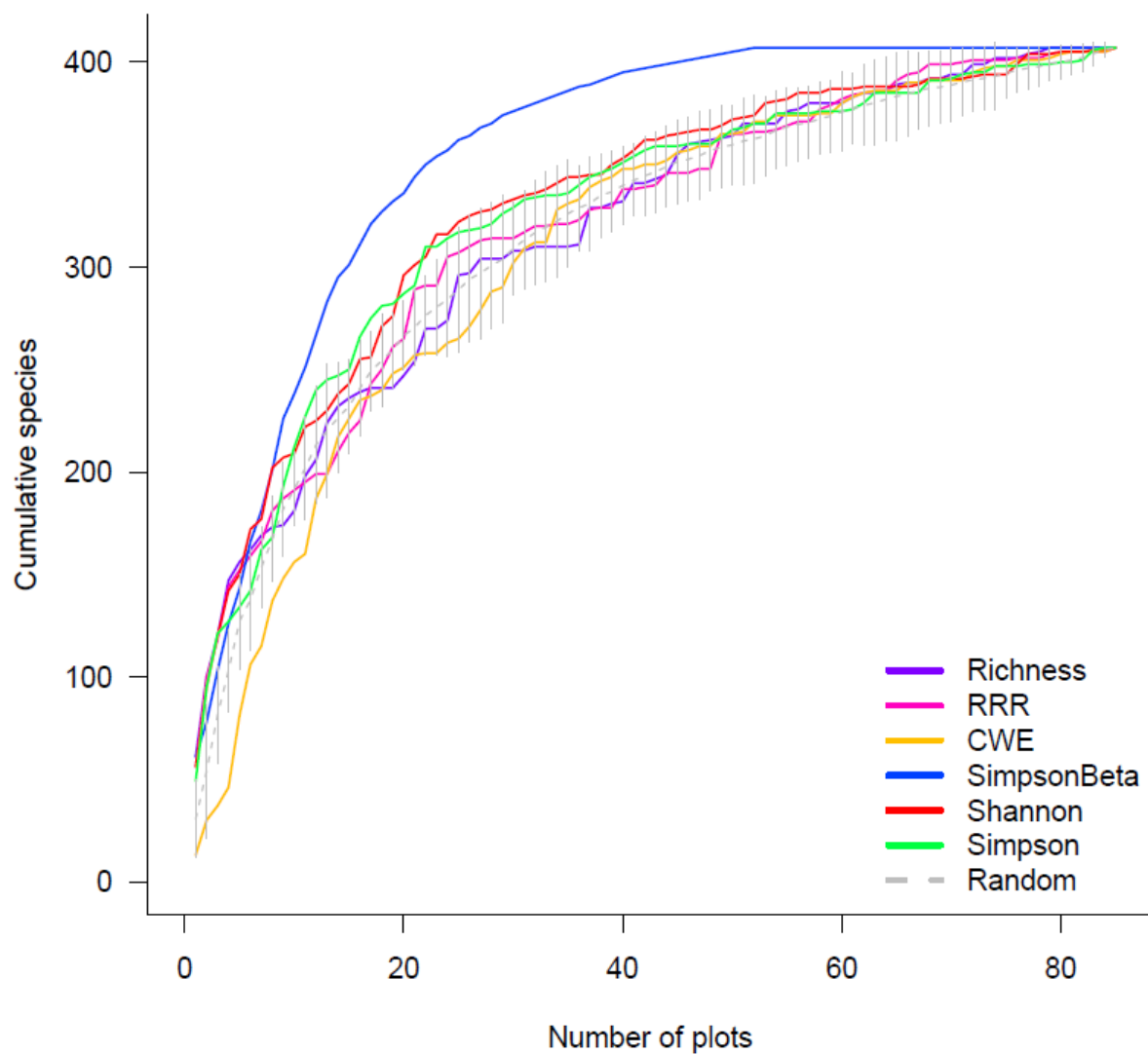
Supplementary material S2. Additional explanation of biodiversity metrics

Species richness is simply the count of the number of species present in a given site and it is best used as an optimiser when the goal is to identify areas containing the greatest number of unique species. RRR is a rarity-weighted richness calculated as the sum of the inverse of the number of sites in which a species occurs and it is used to identify areas of high biodiversity and biological uniqueness. CWE is calculated as RRR corrected by the species richness and is used to identify centres of endemism highlighting range-restricted species. Shannon combines species richness and evenness or equitability by computing the species' relative abundances; it assumes that all species are represented in a sample and that they are randomly sampled. Simpson is the complement of Simpson's original dominance index, and represents the probability that two randomly chosen individuals belong to different species. The species turnover-based metric is based on diversity partitioning, which separates species replacement (i.e. turnover) from species loss (i.e. nestedness); it corresponds to the turnover component of the Sorensen dissimilarity and describes spatial turnover without the influence of richness gradients (because unshared species in the larger species sample are disregarded; Lennon et al., 2001; Baselga, 2010). For the species turnover-based metric, as pairwise Simpson dissimilarity differs depending on the plot that is chosen as seed, we computed three variants: fixed seed (Simpson_Beta), in which we selected the plot with the greatest richness as the starting point; random seed (Simpson_Random), in which we randomly selected the plot at the starting point and we iterated this process 1,000 times; and the most frequent plots (Frequent), by computing the most frequently selected plots in the former 1,000 simulations. When employing the pairwise Simpson dissimilarity, the most dissimilar plot to the one used as seed in terms of pairwise Simpson dissimilarity was added by the function. Subsequently the species present

in both plots are pooled together and the most dissimilar one to the combination of both is added. This process was repeated until reaching the top 250 plots that maximised species turnover.

DRAFT

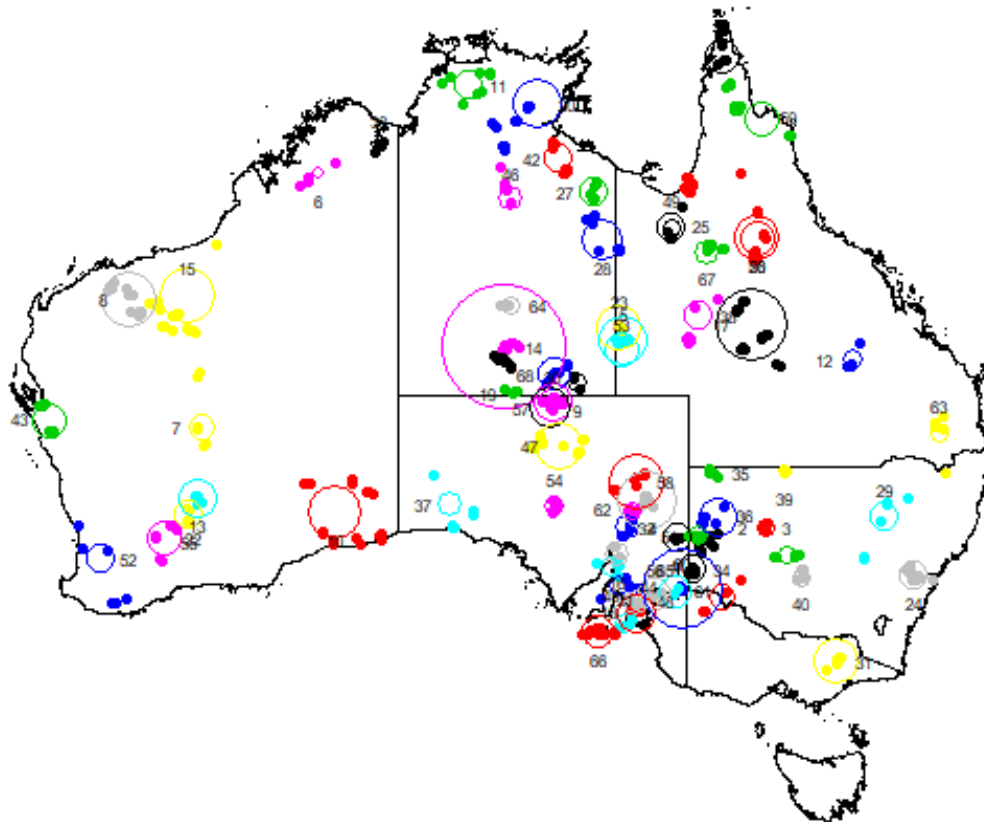
Supplementary material S3. Example of the comparison of different optimisers after applying the *optim_species* function to an alternative dataset. In this case the dataset corresponds to plots located in South Australia within the frame of the project Transect for Environmental Monitoring and Decision Making (TREND). We applied the maximal coverage problem selecting 85 plots to optimise.



Supplementary material S4. Climate variables employed to calculate the environmental representativeness of the selected plots. The variables were obtained from Harwood et al. (2016)

Code	Brief description
<i>Aridity index</i>	
ADI	Minimum monthly aridity index (proportion)
ADM	Mean annual aridity index (annual precipitation/annual potential evaporation) (proportion)
ADX	Maximum monthly aridity index (proportion)
<i>Evaporation</i>	
EPA	Annual potential evaporation (mm)
EPI	Minimum monthly potential evaporation (mm)
EPX	Maximum monthly potential evaporation (mm)
EAA	Annual total actual evapotranspiration terrain scaled using MODIS (mm)
EAAS	Annual total actual evapotranspiration modelled using terrain-scaled water holding capacity (mm)
<i>Maximum temperature</i>	
TXM	Maximum temperature – Annual mean °C
TXI	Maximum temperature - monthly minimum °C
TXX	Maximum temperature - monthly maximum °C
<i>Minimum temperature</i>	
TNM	Minimum temperature – Annual mean °C
TNI	Minimum temperature - monthly minimum °C
TNX	Minimum temperature - monthly maximum °C
<i>Temperature range</i>	
TRI	Minimum monthly mean diurnal temperature range °C
TRX	Maximum monthly mean diurnal temperature range °C
TRA	Annual temperature range (TXX – TNI) °C
<i>Precipitation</i>	
PTA	Annual precipitation mm
PTI	Minimum monthly precipitation mm
PTX	Maximum monthly precipitation mm
PTS1	Precipitation seasonality 1- solstice seasonality composite factor ratio
PTS2	Precipitation seasonality 2- equinox seasonality composite factor ratio
<i>Water deficit</i>	
WDA	Annual atmospheric water deficit (annual precipitation – annual potential evaporation) mm
WDI	Minimum monthly atmospheric water deficit (precipitation - potential evaporation) mm
WDX	Maximum monthly atmospheric water deficit (precipitation - potential evaporation) mm

Supplementary material S5.



S5-Fig. 1. Map representing geographic clusters of plots to translate the findings into a realistic and feasible approach. Colours indicate different clusters, number indicate the cluster ID (see Supplementary material S3) and the size of the circle indicates the number of plots within each cluster.

S5-Table 1. Clusters of plots, coordinates of their centroids and number of plots within**them**

Cluster ID	Number of plots	Number of sites	Latitude	Longitude
clu1	12	12	-11.9637	142.3816
clu2	6	6	-31.4776	144.2586
clu3	6	6	-32.6306	145.1296
clu4	8	5	-31.4964	138.544
clu5	17	6	-23.7562	138.3792
clu6	4	4	-16.8637	125.6894
clu7	9	8	-27.353	120.8142
clu8	19	19	-22.0562	117.8034
clu9	7	7	-25.5573	136.4409
clu10	18	18	-30.8827	126.355
clu11	10	10	-13.1988	131.9348
clu12	7	7	-24.568	147.8653
clu13	13	13	-30.3101	120.6808
clu14	42	18	-24.0693	133.4427
clu15	19	19	-21.8701	120.2695
clu16	8	4	-32.6554	138.0497
clu17	24	24	-23.1361	143.6343
clu18	13	7	-35.0169	138.8766
clu19	4	4	-25.9328	133.86
clu20	17	17	-13.9922	134.7901
clu21	7	4	-35.5107	138.4406
clu22	12	12	-31.9722	119.3232
clu23	15	5	-23.2451	138.2123
clu24	10	9	-33.4671	150.4113
clu25	6	6	-19.079	140.3453
clu26	12	12	-19.5532	143.9305
clu27	10	10	-17.6146	137.0923
clu28	14	14	-19.6014	137.4888
clu29	10	10	-31.0635	149.1966
clu30	13	13	-26.3325	135.3958
clu31	15	15	-37.0249	147.163
clu32	20	10	-30.4005	139.3001
clu33	4	4	-15.7952	128.1739
clu34	9	9	-34.3382	142.4097
clu35	4	4	-29.2578	142.0513
clu36	13	11	-31.0322	142.2075
clu37	8	8	-30.5226	131.1769
clu38	10	10	-22.7827	141.4962
clu39	4	4	-29.2031	145.0439
clu40	6	6	-33.5424	145.6879
clu41	10	6	-31.9441	140.5602

clu42	10	10	-16.27	135.6802
clu43	12	9	-27.0428	114.4813
clu44	7	4	-33.951	138.207
clu45	7	6	-33.1889	137.8196
clu46	8	8	-17.9138	133.6234
clu47	16	16	-28.1194	135.6589
clu48	8	4	-34.5859	138.8827
clu49	10	10	-19.079	140.3453
clu50	15	15	-19.5532	143.9305
clu51	7	7	-31.8375	141.4666
clu52	10	10	-32.7933	116.6572
clu53	12	4	-24.1515	138.2374
clu54	7	7	-30.5942	135.4801
clu55	10	10	-30.9752	120.2899
clu56	11	8	-34.2875	139.6342
clu57	14	14	-26.5941	135.2836
clu58	19	11	-29.5789	138.9052
clu59	12	12	-14.5616	144.045
clu60	27	16	-34.0285	140.7325
clu61	11	7	-34.1447	140.4778
clu62	6	3	-30.7738	138.67
clu63	6	6	-27.5596	151.4464
clu64	6	6	-22.29	133.654
clu65	10	10	-33.2362	141.2395
clu66	11	11	-35.8077	137.3207
clu67	8	8	-20.1054	141.8365
clu68	11	11	-25.1547	135.4541

Supplementary material S6. Species accumulated when applying the maximum coverage problem to select a subset of 250 plots to revisit. For details of selected plots see associated datasets in Dryad (Martin-Fores et al. 2021).

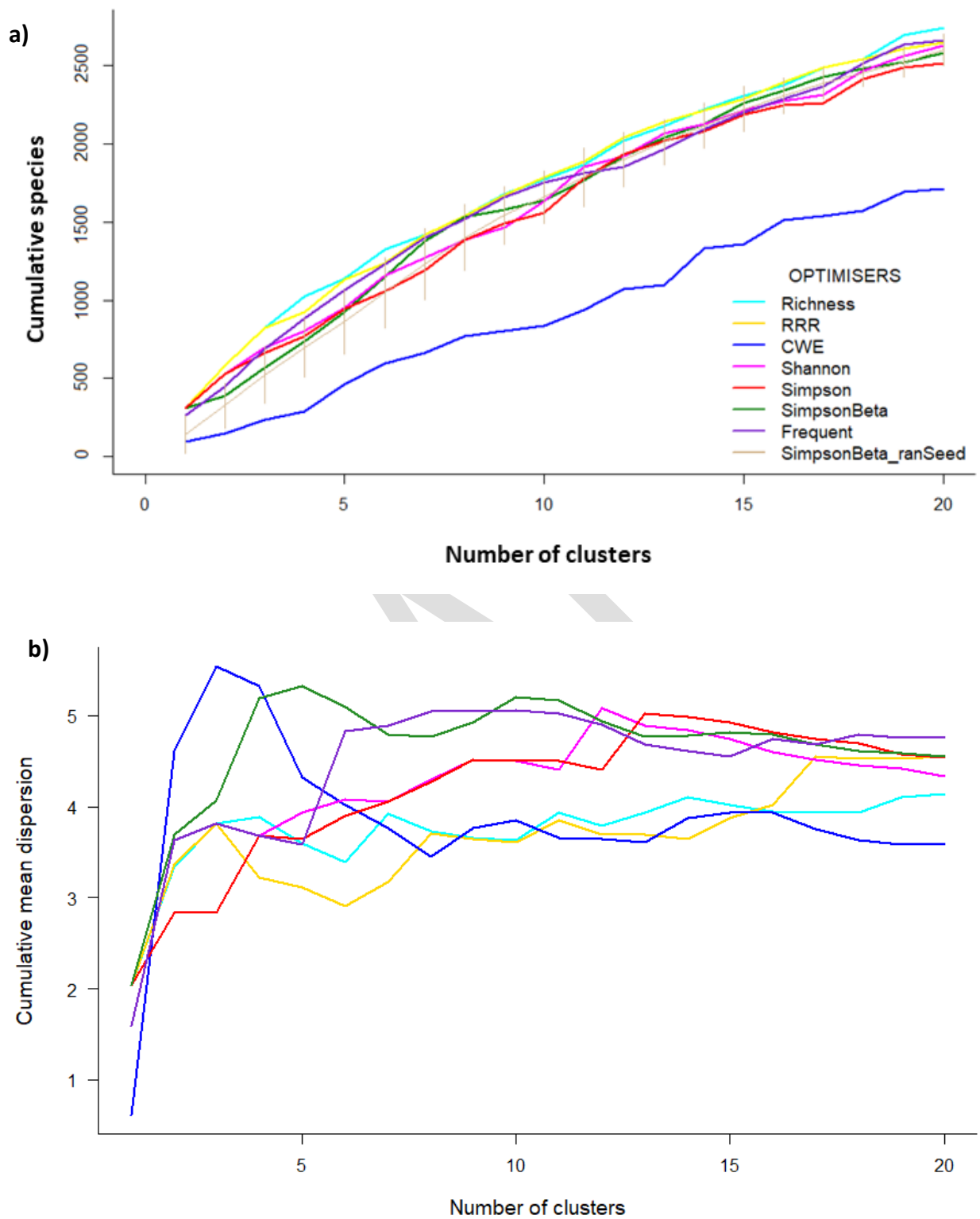
Optimiser	Number of plots	Number of sites	Species accumulated	% of total species
Richness	250	229	2864	81.2%
RRR	250	233	2866	81.2%
CWE	250	235	2024	57.4%
Shannon	250	231	2756	78.1%
Simpson	250	234	2633	74.6%
Simpson_Beta	250	245	3021	85.6%
Frequent	250	245	3051	86.5%
Simpson_Random	250		3030.9 ± 14.7	85.9%

Supplementary material S7. Results of monitoring strategy optimisation for plot clusters

The results obtained when analysing clusters of plots differed from those obtained for single plots. In terms of ecological representativeness, the differences in species accumulation among most of the different optimisers become diluted when selecting clusters of plots instead of single plots. The only biodiversity metric that had a considerably worse performance was CWE (S7-Fig. 1).

Regarding spatial representativeness, when selecting clusters of plots, we obtained opposite trends than when selecting single plots. Species richness was the best optimiser regarding spatial representativeness ($R = 0.218$), followed by RRR, Frequent and CWE optimisers ($R = 0.188$, $R = 0.184$ and $R = 0.172$). Pairwise Simpson dissimilarity (Simpson_Beta) showed Clark-Evans values of $R = 0.134$. Clusters of plots selected with Shannon and Simpson metrics displayed both the most clustered spatial coverages ($R = 0.011$ and $R = 0.009$, respectively; S7-Fig. 2; Supplementary material S8).

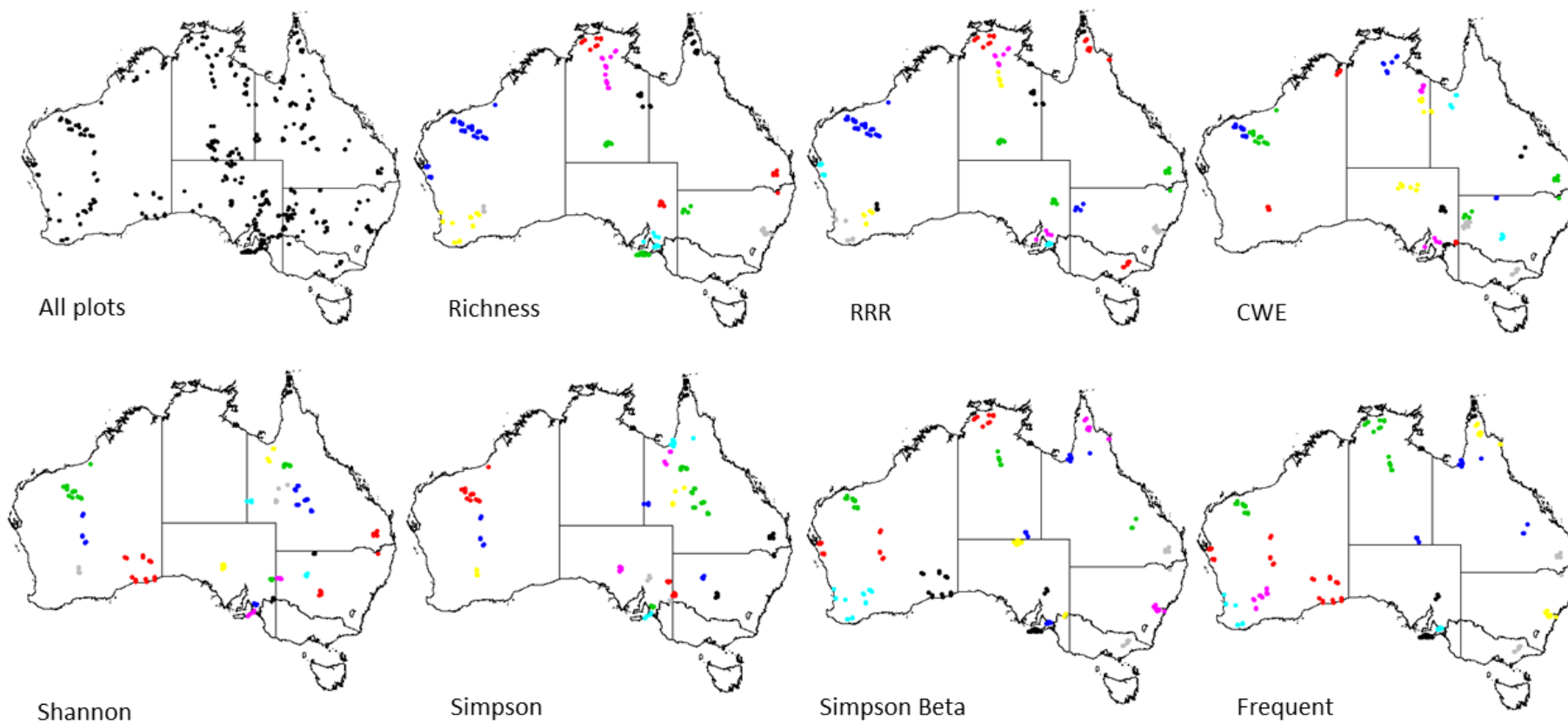
The most environmentally representative subset of selected clusters was obtained using species turnover, with Frequent the best optimiser in terms of environmental representation (Frequent: average distance to median = 4.753, respectively; S7-Fig. 1; S7-Table 1), followed by RRR, Simpson_Beta and Simpson (RRR, Simpson_Beta and Simpson: average distance to median = 4.56, 4.55 and 4.53, respectively), with no significant differences among them. 'Frequent' displayed marginally significantly better environmental representativeness than Shannon (Shannon: 4.34), whereas all the four former biodiversity metrics performed significantly (for Frequent) or marginally significantly (for the later three) better than richness in terms of environmental representativeness (Richness: 4.14). Finally, all the optimisers performed significantly better than CWE in terms of environmental representativeness (CWE: average distance to median = 3.60; S7-Fig. 1; S7-Table 1).



S7-Fig. 1. a) Site optimisation process applying conservation reserve design strategies based on Maximum coverage problem (selection of 20 clusters); b) environmental

representativeness of the 20 selected clusters using different optimisers reflected by the cumulative mean dispersion

DRAFT



S7-Fig. 2. Geographic location of the selected clusters ($n = 20$) applying the maximal coverage problem. Black dots correspond to all the plots established. Colour dots refer to each of the selection employing different optimisers.

S7-Table 1. Pairwise comparisons between optimisers with regards to environmental representativeness when applying maximal coverage problem selecting spatial clusters. The observed p-value are located in the below diagonal, while the permuted p-value are in the above diagonal. Only significant differences are highlighted in bold. Notice that marginally significant values (p-value ≤ 0.1) are shown although not highlighted.

	Richness	RRR	CWE	Shannon	Simpson	SimpsonBeta	Frequent
Richness		≤ 0.1	≤ 0.05	≤ 0.001	≤ 0.001	≤ 0.05	≤ 0.01
RRR	≤ 0.1		≤ 0.01	≤ 0.001	≤ 0.001	0.99	0.42
CWE	≤ 0.05	≤ 0.001		≤ 0.001	≤ 0.05	≤ 0.001	≤ 0.001
Shannon	≤ 0.001	≤ 0.001	≤ 0.01		0.36	≤ 0.001	≤ 0.001
Simpson	≤ 0.001	≤ 0.001	≤ 0.05	0.35		≤ 0.001	≤ 0.001
SimpsonBeta	≤ 0.05	0.99	≤ 0.001	≤ 0.001	≤ 0.001		0.39
Frequent	≤ 0.05	0.42	≤ 0.001	≤ 0.001	≤ 0.001	0.41	

Supplementary material S8. Selected clusters with the different optimisers, number of sites and plots included and species accumulated

Optimiser	Clusters selected	Number of plots	Number of sites	Species accumulation
Richness	clu59, clu52, clu8, clu18, clu15, clu43, clu1, clu14, clu55, clu22, clu24, clu36, clu20, clu11, clu28, clu46, clu63, clu32, clu66, clu44	278	230	2645
RRR	clu59, clu8, clu52, clu15, clu43, clu14, clu18, clu1, clu55, clu36, clu20, clu28, clu63, clu22, clu24, clu11, clu31, clu32, clu46, clu44	282	234	2693
CWE	clu56, clu31, clu63, clu39, clu60, clu12, clu40, clu36, clu8, clu28, clu49, clu27, clu25, clu20, clu33, clu44, clu47, clu13, clu32, clu15	245	216	1698
Shannon	clu59, clu46, clu1, clu66, clu63, clu44, clu11, clu24, clu55, clu43, clu36, clu31, clu51, clu37, clu52, clu18, clu47, clu48, clu22, clu49	210	173	2435
Simpson	clu59, clu46, clu1, clu66, clu11, clu63, clu44, clu55, clu24, clu43, clu37, clu47, clu31, clu25, clu48, clu12, clu9, clu51, clu36, clu52	195	184	2351
Simpson_Beta	clu59, clu61, clu66, clu31, clu22, clu30, clu52, clu24, clu8, clu11, clu63, clu43, clu4, clu9, clu50, clu18, clu46, clu10, clu12, clu7	227	212	2628
Frequent	clu8, clu66, clu52, clu50, clu43, clu31, clu24, clu11, clu46, clu18, clu63, clu7, clu12, clu10, clu55, clu1, clu9, clu59, clu22, clu4	225	213	2706
Simpson_Random				2626.5 ± 49.8