

Sex-specific behavioral syndromes allow the independent evolution of behavioral dimorphism

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Running Title: Sex-specific syndromes and behavioral sexual dimorphism

Keywords: Behavioral syndromes, evolutionary constraint, **G** matrix, personality, behavioral ecology, sexual dimorphism, intralocus sexual conflicts

Author Contributions: All co-authors conceived the project and supervised the gathering of data. R.R conducted behavioral trials and analyzed the data. R.R. wrote the first draft of the manuscript and all authors contributed to subsequent versions.

Data Accessibility Statement: All data, code and analyses are available at: <https://osf.io/pnug5/>

2 **ABSTRACT**

3 When selection differs by sex, the capacity for sexes to reach optimal phenotypes can be
4 constrained by the shared genome of males and females. Because phenotypic traits are
5 often correlated, this difference extends across multiple traits and underlying genetic
6 correlations can further constrain evolutionary responses. Behaviors are frequently
7 correlated as behavioral syndromes, and these correlations often have a genetic basis.
8 However, whether cross-sex and across behavior correlations lead constrained evolution
9 remains unknown. Here, we show that a boldness-activity syndrome is strongly sex-
10 specific at the genetic level in the western field cricket (*Gryllus integer*) and that emergence
11 from a shelter is genetically independent between males and females. However, male
12 activity is strongly related to female shelter emergence, creating the potential for biased
13 responses to selection. Our results show that the sex-specific genetic architecture of
14 behavioral syndromes can shape the evolution of behavioral phenotypes.

15

16 INTRODUCTION

17 Males and females share the same genome but often rely on different strategies to increase
18 fitness. This can lead to intralocus sexual conflicts where selection acting on a shared trait
19 displaces one sex from its optimum (Hedrick and Temeles 1989; Bonduriansky and
20 Chenoweth 2009). This sexual conflict is further complicated by the constraining potential
21 of genetic correlations (Lande 1980; Walsh and Blows 2009). For example, behaviors are
22 often correlated as components of a behavioral syndrome (Sih et al. 2004) and these
23 behavioral syndromes have the potential to alter evolutionary outcomes (Dochtermann
24 and Dingemanse 2013).

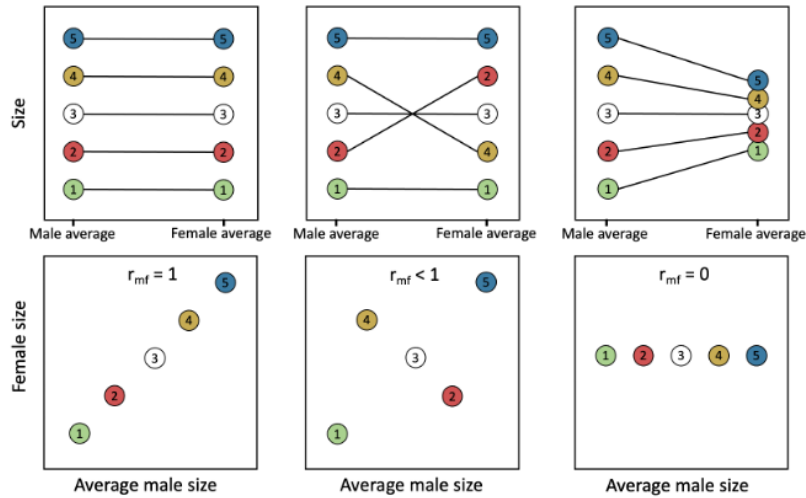
25 In general, any sustained selection favoring different optima between sexes, i.e.
26 sexually discordant selection, will eventually resolve the sexual conflict and attenuate
27 genetic constraints over time, thus allowing sexes to evolve independently (Delph et al.
28 2011). This constraint can be formally measured as the cross-sex correlation coefficient r_{mf} ,
29 with values < 1 increasing the rapidity at which sexual dimorphism can evolve. Cross-sex
30 correlations are generally large but tend to decrease in species with stronger sexual
31 dimorphism (Poissant et al. 2010). Additional constraints can emerge when generalizing to
32 multiple phenotypes expressed in males and females and their interactions (Lande 1980).
33 By decomposing the additive genetic covariance matrix into its sex-specific (\mathbf{G}_m , \mathbf{G}_f) and
34 cross-sex sub-matrices (\mathbf{B}), one can estimate if genetic correlations across sexes and traits
35 create constrained evolutionary outcomes (Fig. 1).

36 This decomposition approach has been used for morphological traits, revealing that
37 cross-sex covariances (i.e. the \mathbf{B} matrix, (Lande 1980)) can profoundly alter the evolution
38 of sexual dimorphism (Gosden et al. 2012; Berger et al. 2014; Gosden and Chenoweth
39 2014), especially when selection favors different average trait values for each sex (Long
40 and Rice 2007; Berger et al. 2014). However, the \mathbf{G}_m , \mathbf{G}_f , and \mathbf{B} have rarely been estimated
41 for behaviors. How much the genetic architecture of behavioral syndromes can affect
42 evolutionary outcomes under intralocus sexual conflict remains unclear. This is an
43 important omission because behavior frequently impacts both survival and fitness (Moiron
44 et al. 2020) and sexual dimorphism is common for behaviors (Blanckenhorn 2005; Aragón

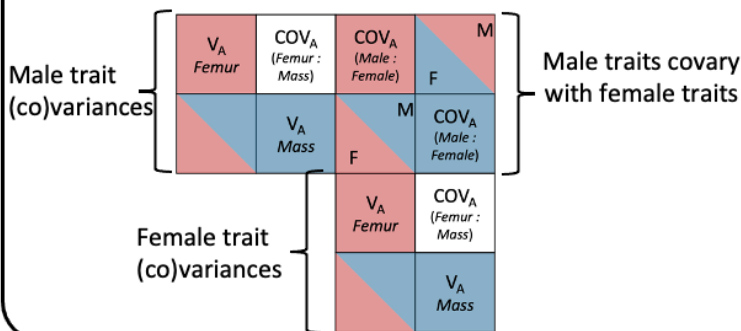
45 2011; Mainwaring et al. 2011; Kokras et al. 2012). Despite many conceptual arguments for
46 why behavioral syndromes should differ by sexes (Schuett et al. 2010; Hämäläinen et al.
47 2018; Immonen et al. 2018), sex-differences in the expression of behavioral correlations
48 are frequently ignored in practice. As a result, the degree to which cross-sex genetic
49 correlations might be shaped by sexual conflict for behavioral phenotypes remains
50 unknown and the importance of these correlations for the evolution of behaviors is
51 similarly unknown.

A) Origin of cross-sex correlations

Cross-sex correlations occur when families with high male trait average also have high averages for females

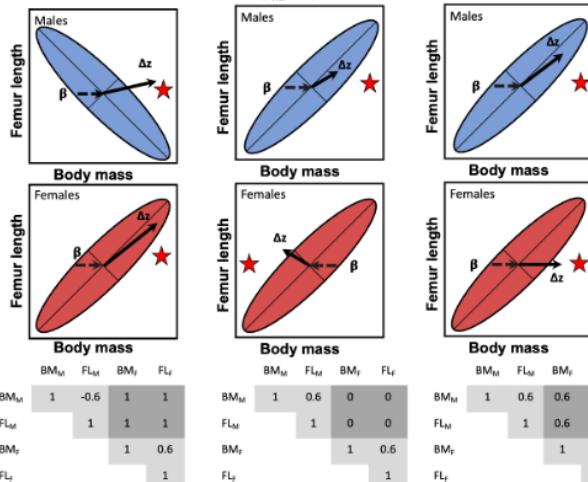


B) Anatomy of a cross-sex genetic covariance matrix (G_{mf})



C) Cross-sex genetic structure and evolutionary responses

C.1) Opposite sex-specific covariances ($r_m \neq r_f$) C.2) Sexes are uncoupled ($r_{mf} = 0$) C.3) Cross-sex cross-trait covariances are asymmetric ($r_{mf} \neq r_{fm}$)



Response to selection:

$$\begin{bmatrix} \Delta \bar{z}_m \\ \Delta \bar{z}_f \end{bmatrix} = \frac{1}{2} \begin{bmatrix} \mathbf{G}_m & \mathbf{B}' \\ \mathbf{B} & \mathbf{G}_f \end{bmatrix} \begin{bmatrix} \beta_m \\ \beta_f \end{bmatrix}$$

53 **Figure 1.** The genetic architecture of cross-sex correlations can have non-intuitive consequences on
54 evolutionary responses and sexual dimorphism. A) High values of cross-sex correlations for the
55 same traits (here, body size) indicate that male and female averages are positively correlated
56 among families. B) The cross-sex covariance matrix (\mathbf{G}_{mf}) allows to compare patterns of genetic
57 covariances within sexes (here between body mass and body size) as well as understand the
58 strength of cross-trait correlations within and among traits. C) Cross-sex cross-trait correlation
59 (r_{mf}) can produce non-intuitive responses to selective pressures. Here we show 3 scenarios
60 indicative of the variety of responses to selection (Δz) that can occur depending on the direction of
61 selection (β) and the magnitude and orientation of r_{mf} . The ellipses represent the bivariate
62 (co)variation in femur length and mass, with the population average at the vertex of the (thin solid)
63 lines indicating the directions in bivariate space with the most variation. Dashed lines represent the
64 direction selection (β) is pushing a population's average and the solid thick arrows show the
65 direction and magnitude (arrow tip) of selection responses. In both scenarios C.1 and C.2, selection
66 is concordant—i.e. acting the same—between sexes and males and females have the same fitness
67 optimum. In scenario C.3, selection is sexually discordant and favors increased sexual dimorphism.
68 Scenario C.1 shows that strong cross-sex cross trait correlations can bias sexes away from their
69 optimum. In scenario C.2, null r_{mf} results in independent trajectories by sexes. In scenario C.3, the
70 effect of fathers on their daughters' phenotype is opposite of that of mother's on their sons',
71 resulting in asymmetric r_{mf} and biased responses to selection in males.

72 Intralocus sexual conflict should be particularly strong in the field crickets
73 (Gryllidae). In many field crickets, males and females differ in their behaviors and
74 reproductive strategies (Hedrick and Kortet 2012). For example, females explore their
75 environments to sample mates while males remain at burrows from which they signal
76 (French and Cade 1987). Further, a boldness-activity syndrome has a conserved genetic
77 basis in at least one species, the Western stutter-trilling field cricket (*Gryllus integer*),
78 which seems to have constrained behavioral divergence (Royauté et al. 2020). Whether this
79 syndrome is sex-specific is currently unknown but the combination of conserved
80 syndromes together with a lack of sex-specificity would further constrain the ability of
81 populations to respond to local selective pressures.

82 Here we used behavioral measurements of over 960 *Gryllus integer* individuals to
83 estimate the influence of cross-sex genetic correlations on evolutionary responses. We
84 predicted that crickets would exhibit dimorphism in average activity given the mate-
85 sampling behavior of females and that genetic variance for this trait would be lower for
86 female crickets due to selection favoring increased traveling. We also predicted sex-
87 specificity in the behavioral syndrome, with a stronger positive genetic correlation
88 between antipredator response and activity in females. Since exit from refugia and activity
89 should have a stronger influence on female fitness, we also expected a stronger correlation
90 between shelter emergence with activity and antipredator response in females. Although
91 there are few estimates for cross-sex correlations of behaviors, those available suggest
92 behaviors are under similar constraints as other phenotypes (Poissant et al. 2010). We
93 therefore expected that cross-sex genetic correlations (r_{mf}) would not depart significantly
94 from 1. We tested these predictions by estimating the cross-sex covariance matrix, i.e. \mathbf{G}_{mf} ,
95 decomposed into its sex-specific and cross-sex sub-matrices (\mathbf{G}_m , \mathbf{G}_f and \mathbf{B} submatrices,
96 Figure 1). We compared the strength of covariances among sexes and the effects of these
97 covariances on responses to selection using random skewer analysis.

98

99 **METHODS**

100 The full details of our breeding design and behavioral trials can be found in Royauté et al.
101 2020. Briefly, we collected adult female crickets from four populations throughout the
102 southwestern and western US: Socorro, NM; Las Cruces, NM; Aguila, AZ; and Dunnigan, CA
103 (Figure 2) during the summer of 2017 and housed them and in our laboratory facilities at
104 North Dakota State University. Females were housed individually in 0.71 L containers and
105 provided with ad libitum food (Purina Chick Starter) and water (water was provided in
106 glass vials capped with cotton). Each cricket was also provided with a small piece of
107 cardboard egg carton for shelter. The cricket housing room was maintained ~27C on a
108 12:12 dark:light cycle reversed such that the room was dark during daytime hours. We run
109 offspring of this parental generation through multiple behavioral trials before mating
110 individuals at random within each population. We repeated this process for two additional
111 generations.

112 Behavioral testing

113 *Latency to emerge from shelter*

114 Gryllid crickets use small burrows and natural cracks as refuges to which they retreat when
115 under threat. The time taken to emerge from a shelter after disturbance can therefore be
116 considered a proxy for risk-taking behavior or “boldness” (Kortet and Hedrick 2007). We
117 transferred individuals from their home containers into small artificial burrows (40 cm³)
118 placed within a 34.6 × 21 cm arena. We left the crickets to rest for two minutes after which
119 we removed the cap from the burrow and let individuals emerge. We then recorded how
120 long it took for an individual to emerge (in seconds) for up to six minutes and thirty
121 seconds. Individuals that did not emerge were given a maximum latency of 390 seconds.

122 *Open field exploratory behavior*

123 We used open field tests to measure activity and exploratory propensity in a 30 × 30 cm
124 plexiglass arena. These tests are classic behavioral assay across taxa (Walsh and Cummins
125 1976) and can reveal strong among-individual differences in exploration patterns,
126 including in crickets (Royauté et al. 2015, 2019; Royauté and Dochtermann 2017).

127 Individuals that move through more of the arena are considered more thorough explorers
128 (Réale et al. 2007). We introduced individuals into the arena and left them to rest under a
129 small container for 30 seconds. At the end of this 30 seconds, we removed the container
130 and the cricket was allowed to explore the arena for 3 minutes and 40 seconds. The arena
131 was cleaned with isopropyl alcohol between trials to remove any chemosensory cues from
132 the arena. We used Ethovision XT to record the total distance the individual moved during
133 the trial (cm).

134 *Response to cues of predator presence*

135 We used a behavioral assay to measure response to cues of predator presence also
136 previously used with another Gryllid species (Royauté and Dochtermann 2017; Royauté et
137 al. 2019). Specifically, individuals were introduced into a 15 cm diameter circular arena
138 (7.5 cm height), the floor of which was covered with dry filter paper that had been soaked
139 with diluted excreta from leopard geckos (*Eublepharis macularius*). Crickets respond to
140 exposure to leopard gecko cues by increasing activity and individuals with higher distance
141 moved are considered more responsive to the cue (Royauté and Dochtermann 2017;
142 Royauté et al. 2019). We introduced crickets to a portion of the arena without predator cue
143 and left them to rest under a small shelter for 30 seconds. We then removed the shelter and
144 allowed the individual allowed to freely move throughout the arena for 3 minutes and 40
145 seconds. We then used Ethovision XT to record the total distance an individual moved
146 during the trial (cm).

147

148 Statistical analyses

149 All analyses were performed using R version 4.0.3 (R citation) using the MCMCglmm
150 package (Hadfield 2010).

151 *Estimation of cross-sex genetic covariances (\mathbf{G}_{mf})*

152 We used a multi-response mixed effect animal models (Kruuk 2004; Wilson et al. 2010) to
153 estimate genetic variances and covariances (i.e. the \mathbf{G}_{mf} matrix). We included the effects of

154 temperature, day and time of testing in the behavioral arena room along with sex, life-stage
155 and mass of the individual as fixed effects. We used the individual relatedness matrix
156 (based on the known pedigree) as a random effect and the following behavioral traits were
157 included as response variables: (i) the latency that an individual emerged from the shelter
158 during the trial (modeled as censored Gaussian), (ii) the distance moved during the open
159 field trial (Gaussian), (iii) the distance an individual moved during the predator cue
160 response trial (Gaussian). To estimate both sex-specific and cross-sex covariances, we
161 treated the behavior of each sex as a separate trait - resulting in the estimation of a 6×6
162 covariance matrix. We ran the multi-response model with an MCMC chain of 4.8×10^6
163 iterations, with an 800,000 burn-in period and a thinning interval of 4,000 and we used a
164 parameter expanded prior that was minimally informative for both variances and
165 covariances. All variances and covariances were estimated at the additive genetic level and
166 on the latent scale.

167

168 *Estimation of behavioral dimorphism*

169 We tested for the existence of sexual dimorphism in behavioral expression by comparing
170 linear coefficient for the sex fixed effect included in our multivariate animal model and base
171 our statistical inference on their Bayesian probability (Pmcmc). This metric varies between
172 0.5 and 1 and indicates the probability of a significant difference based on the number of
173 posterior estimates overlapping with 0. Pmcmc values > 0.95 were judged as significant.

174

175 *Comparison of sex-specific covariances (\mathbf{G}_m and \mathbf{G}_f matrices)*

176 We used a two-step approach to compare the intensity of the difference in genetic
177 covariances among sexes. First, we calculated the difference in covariance between males
178 and females (ΔCOV_A) for each pair of behaviors and their associated Bayesian probabilities
179 (Pmcmc). Next, we tested whether sex-specific covariances were similarly oriented. To do
180 so, we calculated the vector correlation (r°) between axes containing the highest amount of
181 genetic variation using eigenvalue decomposition. We then estimated whether the

182 resulting vector correlations among eigenvectors of \mathbf{G} differed substantially from
183 expectations of 0 (no alignment of genetic variation among sexes) and 1 (perfect
184 alignment). Because vector correlations are bounded by 0 and 1, we estimated the Region
185 of Posterior Equivalence (ROPE) which we defined as the intervals [0.0; 0.1] and [0.9; 1.0].
186 Estimates falling within these ROPE regions are judged “practically equivalent” to vector
187 correlations of 0 and 1 respectively. We converted the proportion of estimates falling
188 outside these ROPE regions into Pmcmc values to infer significance, with Pmcmc > 0.95
189 indicating significant departure from these null hypotheses.

190

191 *Estimating the genetic constraint imposed by the cross-sex covariance matrix (\mathbf{B})*

192 We first tested whether cross-sex covariances within traits differed significantly from 0 and
193 1. Cross-sex covariances are represented on the diagonal elements of the \mathbf{B} matrix. A
194 covariance of 0 indicate complete genetic independence among sexes, which we
195 determined using Bayesian probabilities (Pmcmc). To test for a departure from complete
196 genetic coupling among sexes, we converted the covariances to correlation coefficients
197 (r_{mf}) and used the ROPE test described above. We then investigated whether cross-sex
198 cross-traits covariances – represented on the off-diagonal elements of \mathbf{B} – differed
199 significantly from one another by calculating the posterior difference in covariance as
200 described (ΔCOV_A).

201 Next, we compared the evolutionary trajectories of sexes with a scenario where sexes
202 evolved independently with one where sexes were fully constrained (Cox et al. 2017). To do
203 so, we simulated 500 selection gradients based on a multivariate normal distribution that
204 we scaled to unit length. We generated concordant selection gradients by assigning the
205 same values to male and female gradients ($\beta_f = \beta_m$) and discordant selection by setting $\beta_f = -$
206 β_m . We applied each selection gradient to all 1,000 posterior covariance matrices of \mathbf{G}_{mf}
207 estimated from our multivariate animal model, thus ensuring we took the uncertainty in
208 estimates forward. We then calculated the resulting response to selection ($\Delta \bar{z}$) by applying
209 the multivariate breeder’s equation for cross-sex covariance (Lande 1980):

210
$$\begin{bmatrix} \Delta \bar{z}_f \\ \Delta \bar{z}_m \end{bmatrix} = \frac{1}{2} \begin{bmatrix} \mathbf{G}_f & \mathbf{B}' \\ \mathbf{B} & \mathbf{G}_m \end{bmatrix} \begin{bmatrix} \beta_f \\ \beta_m \end{bmatrix} \quad (\text{equation 1})$$

211 We contrasted these responses to selection to cases where we set all cross-sex covariances
 212 0 and by fully constraining cross-sex correlations to 1. To achieve this last step, we
 213 converted \mathbf{G}_{mf} to a correlation matrix and replaced \mathbf{B} elements by 1. We then back-
 214 converted this modified matrix into covariances by replacing the diagonal elements of \mathbf{B} by
 215 the geometric mean of male and female genetic variances (i.e. the diagonal elements of \mathbf{G}_f
 216 and \mathbf{G}_m respectively). We then estimated the vector correlation between the response
 217 calculated from our estimated matrix and those estimated with unconstrained ($r_{B=0}$) and
 218 constrained matrices ($r_{B=1}$). Next we compared how consistent male and female response
 219 was when evolving toward the same optimum (concordant selection) and when selection
 220 was discordant. This was achieved by calculating the vector correlation between the
 221 direction of selection for each sex and the corresponding response ($r_{\beta \times \Delta z}$).

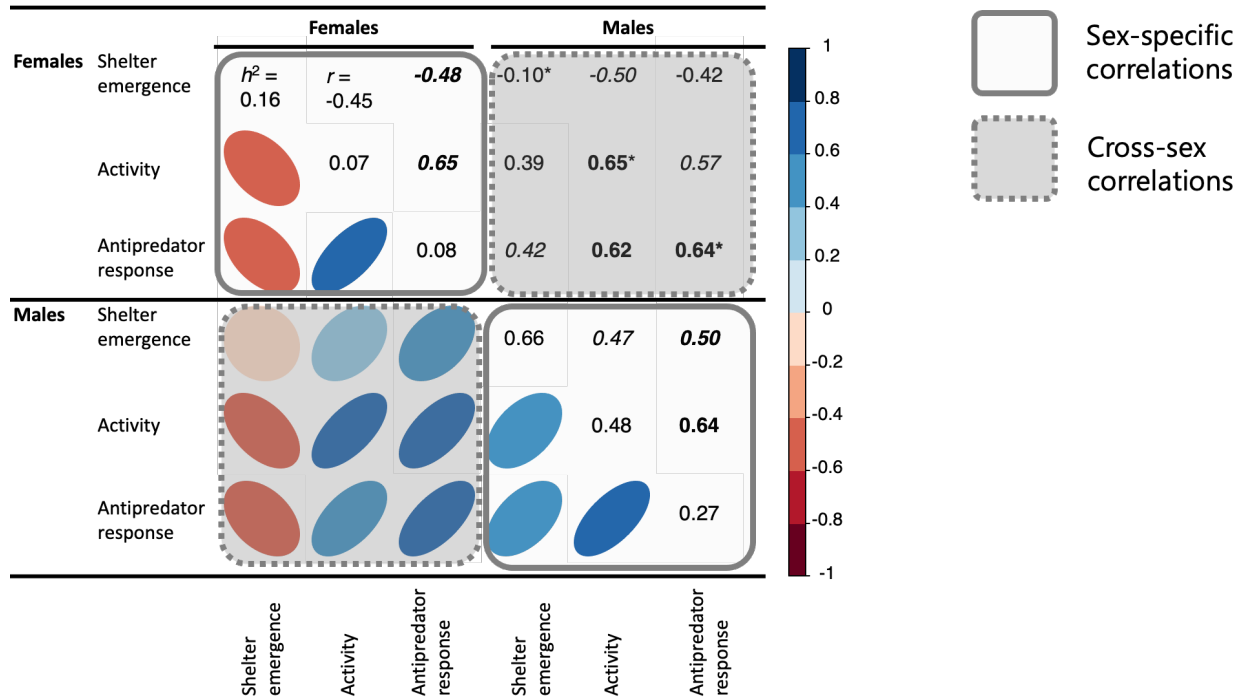
222 RESULTS

223 Males and females showed little evidence for sexual dimorphism in average behavior (all
 224 $\text{Pmcmc} < 0.57$, Table S1, S2). We did find evidence of a $\mathbf{G} \times \mathbf{Sex}$ interaction, and females had
 225 lower heritability and evolvability compared to males (mean female $h^2 = 0.10$; mean male
 226 $h^2 = 0.47$; mean female $I = 3.04\%$; mean male $I = 15.73\%$). This is confirmed by the fact
 227 that the additive genetic variance was lower in females compared to males for all three
 228 behaviors (posterior median [89% CI]; shelter emergence: $\Delta V_A (\text{females} - \text{males}) = -50.12$ [-
 229 110.18; 8.18], $\text{Pmcmc} = 0.92$; activity: $\Delta V_A = -43.38$ [-89.01; -9.19], $\text{Pmcmc} = 0.99$;
 230 antipredator response: $\Delta V_A = -17.63$ [-44.47; 5.45], $\text{Pmcmc} = 0.92$) (Figure 2, 3; Table S3).

231 We also found strong evidence for the sex-specific expression of behavioral
 232 syndromes, with weaker genetic correlations between behaviors in females. Males and
 233 females differed primarily in how shelter emergence related to open-field activity and
 234 antipredator response (Figure 2), while the relationship between open-field activity and
 235 antipredator response was stable between sexes (females: $r_{mf} = 0.65$ [0.20; 0.98], $\text{Pmcmc} =$
 236 0.94 , $\text{Pmcmc} = 0.91$; males: $r_{mf} = 0.64$ [0.27; 0.92], $\text{Pmcmc} = 0.98$). Females that stayed
 237 longer in the shelter had lower antipredator response and travelled further in the open-

238 field (shelter emergence \times open-field: $r_{mf} = -0.45 [-0.92; 0.11]$, $P_{mcmc} = 0.86$; shelter
239 emergence \times antipred. : $r_{mf} = -0.48 [-0.90; -0.01]$, $P_{mcmc} = 0.91$). In contrast, males with
240 slow shelter emergence had higher activity and antipredator response (shelter emergence
241 \times open-field: $r_{mf} = 0.47 [0.06; 0.87]$, $P_{mcmc} = 0.93$; shelter emergence \times antipred. : $r_{mf} =$
242 $0.50 [-0.01; 0.84]$, $P_{mcmc} = 0.92$). In addition, the correlation between major axes of
243 genetic variation (\mathbf{g}_{max}) was significantly < 1 , providing another line of evidence for the
244 presence of a sex-specific syndrome (vector correlation $r^o = 0.36 [0.00; 0.76]$, $P_{mcmc\#1} =$
245 0.97) (Table S2, S3).

246



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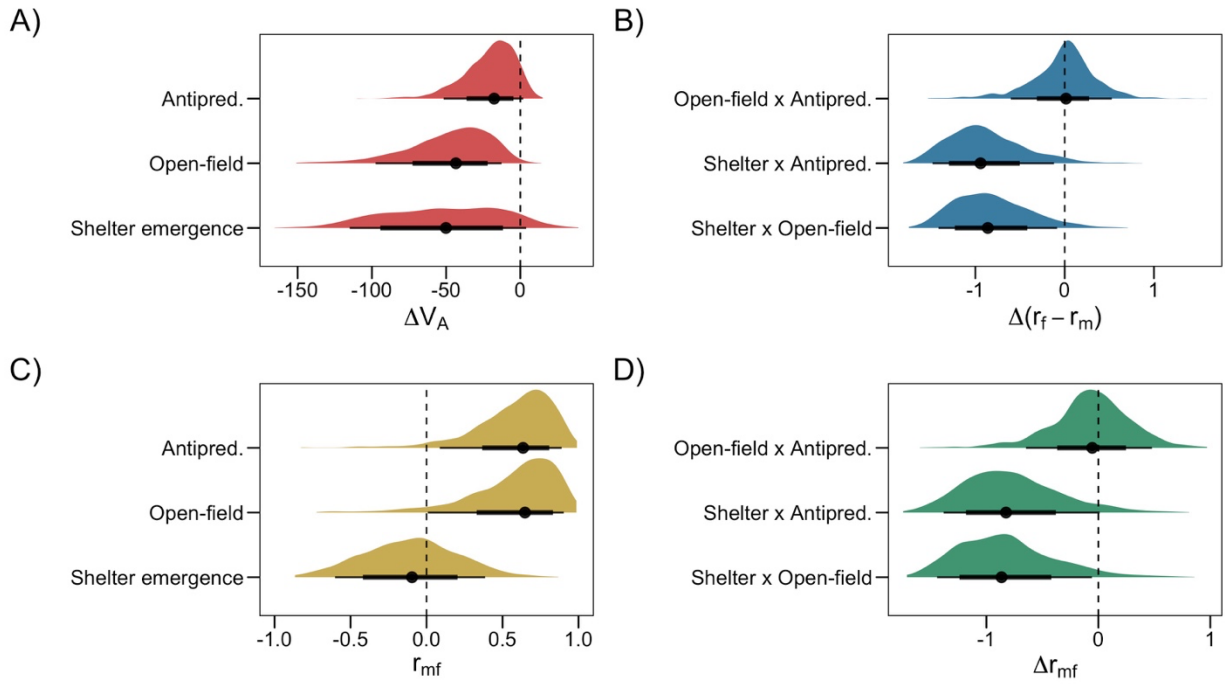
248 **Figure 2.** Genetic correlation matrix (G_{mf}) indicating sex-specific and cross-sex genetic
 249 correlations. Heritabilities (h^2) are indicated on the main diagonal and genetic correlations
 250 (r) on the off-diagonal elements. Off-diagonal elements represent either the sex-specific
 251 genetic correlations (r_f and r_m in light grey) or the cross-sex genetic correlations (r_{mf} in
 252 dark grey). Bold indicate significant correlations based on $P_{mcmc} > 0.95$. Correlations with
 253 $P_{mcmc} > 0.90$ are indicated in italics. Cross-sex correlations significantly different from 1
 254 are indicated with an asterisk.

255

256 All behaviors showed weak to moderate cross-sex correlations and were
257 significantly below 1 (shelter emergence: $r_{mf} = -0.09 [-0.59; 0.39]$, $P_{mcmc\neq 1} = 1.00$; activity:
258 $r_{mf} = 0.65 [0.22, 0.97]$, $P_{mcmc\neq 1} = 0.94$; antipredator response: $r_{mf} = 0.64 [0.26; 0.94]$,
259 $P_{mcmc\neq 1} = 0.96$) (Table S3). In addition, sexes had biased expression of different trait
260 combinations, as indicated by the high proportion of asymmetry in **B**—the cross-sex
261 covariance matrix (proportion of skew-symmetry = $0.25 [0.04, 0.42]$). This degree of
262 asymmetry was most pronounced for the cross-sex correlations between shelter
263 emergence and activity ($\Delta COV_A = -21.39 [-43.72; -0.56]$, $P_{mcmc} = 0.97$) and between
264 shelter emergence and antipredator response ($\Delta COV_A = -14.8 [-32.33; 1.46]$, $P_{mcmc} =$
265 0.95). In contrast, the correlation between activity and antipredator response did not differ
266 among sexes ($\Delta COV_A = -3.02 [-11.73; 6.73]$, $P_{mcmc} = 0.71$) (Figure 3).

267 This means that highly active fathers produced daughters with faster shelter
268 emergence ($r_{mf} = -0.50 [-0.85; -0.06]$, $P_{mcmc} = 0.94$) and higher antipredator response (r_{mf}
269 $= 0.57 [0.12; 0.95]$, $P_{mcmc} = 0.93$). In contrast, active mothers only weakly contributed to
270 their sons' shelter emergence ($r_{mf} = 0.39 [-0.14; 0.85]$, $P_{mcmc} = 0.85$) and antipredator
271 response ($r_{mf} = 0.42 [-0.01; 0.85]$, $P_{mcmc} = 0.90$).

272

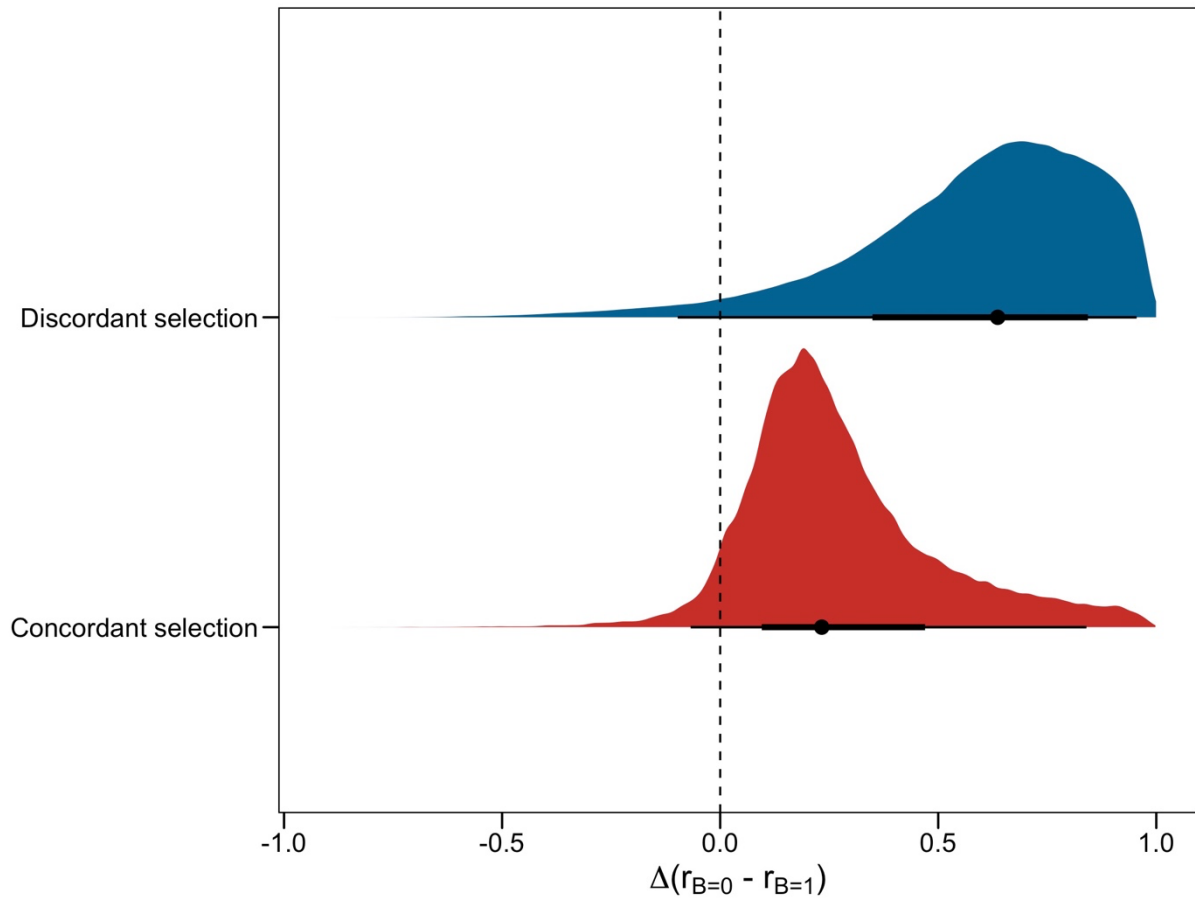


273

274 **Figure 3.** The genetic structure of the boldness-activity syndrome differed both in terms of
 275 its sex-specific genetic variances (A) and correlations (B) as well as its cross-sex
 276 correlations within traits (C) and cross-sex cross-traits correlations (D).

277

278 Finally, by simulating responses to selection, we found that evolutionary trajectories
279 were more consistent with behaviors being sexually independent than with a constrained
280 expression of behaviors. This was the case regardless of whether selection favored sex-
281 specific optima (i.e. discordant selection: $r_{B=0} = 0.95$ [0.73; 1.00]; $r_{B=1} = 0.28$ [0.00; 0.56]; Δr
282 $= 0.64$ [0.24; 0.98], $P_{mcmc} = 0.96$) or when both sex had the same optimum (i.e. concordant
283 selection: $r_{B=0} = 0.95$ [0.74; 1.00], $\Delta r = 0.23$ [-0.04; 0.66], $P_{mcmc} = 0.95$) (Figure 4). We next
284 compared the agreement between simulated selection gradients and predicted response to
285 selection ($r_{\beta \times \Delta z}$) between sexes. Males and females had equally consistent responses to
286 concordant selection (female $r_{\beta \times \Delta z} = 0.72$ [0.29; 1.00]; male $r_{\beta \times \Delta z} = 0.75$ [0.47; 1.00]; $\Delta r_{\beta \times \Delta z} =$
287 -0.19 [-0.67; 0.07], $P_{mcmc} = 0.88$). However, female response tended to be biased away
288 from selection compared to males when selection was discordant (female $r_{\beta \times \Delta z} = 0.50$ [0.06;
289 0.95]; male $r_{\beta \times \Delta z} = 0.77$ [0.46; 1.00], $\Delta r_{\beta \times \Delta z} = -0.27$ [-0.75; 0.11], $P_{mcmc} = 0.86$). This could
290 mean that the magnitude of the genetic constraint is expressed more strongly in males,
291 which is consistent with the asymmetric nature of the cross-sex covariance matrix.
292 However, this last result should be interpreted with caution given the wide credible
293 intervals around these estimates.



294

295 **Figure 4.** The response to selection is more consistent with models where sexes evolve
 296 independently ($\mathbf{B} = 0$) than models where sexes are fully constrained ($\mathbf{B} = 1$). This trend is
 297 most pronounced when selection favours opposite optima among sexes (discordant
 298 selection). Positive values indicate that the observed response to selection (Δz) is more
 299 strongly correlated with responses to selection where elements of the cross-sex covariance
 300 matrix are set to 0 ($\mathbf{B} = 0$) compared to responses to selection where the cross-sex
 301 covariance is fully constrained ($\mathbf{B} = 1$), based on 1,000 random selection gradients applied
 302 to each 1,000 posterior covariance matrices.

303 **DISCUSSION**

304 Our results show that males and females differ substantially in their behavioral syndromes
305 at the genetic level. Shelter emergence was genetically independent between males and
306 females, whereas the genetic constraint for activity and antipredator response was
307 stronger but still departed from 1. The absence of a strong genetic constraint linking the
308 sexes is surprising given that the behaviors we measured are not typically considered
309 distinct sex-specific traits. In the case of shelter emergence, a cross-sex correlation
310 approaching zero implies that the same behavior is underpinned by completely
311 independent sets of genes in males and females. Such genetic uncoupling means that this
312 behavior can fulfill different functions in each sex and can evolve independently. Our
313 results suggest that sexual conflict may have been resolved in this species even in absence
314 of observable behavioral dimorphism.

315 By applying a quantitative genetic approach, we were able to uncover multiple ways
316 in which sexes differed in their behaviors. While males and females did not differ in mean
317 behaviors, we uncovered a signature of behavioral dimorphism in the amount of genetic
318 variation expressed by each sex. This **G × Sex** effect was characterized by female behavior
319 being less heritable than observed for males. In our case, traits with lower heritabilities
320 also had lower evolvabilities, indicating that females are less responsive to selective
321 pressures than males in this species. This strong difference in genetic variance between
322 sexes could result from stronger stabilizing selection in females eroding genetic variation
323 in traits related to exploration and risk taking while maintaining a similar optimum in each
324 sex. Multivariate stabilizing selection has indeed been shown to be an important driver of
325 trait evolution, for example in genital morphology in arthropods (Arnqvist 1997; House et
326 al. 2020). Another possibility is that rearing individuals in laboratory conditions with ad-lib
327 access to food and shelter could alleviate the resource acquisition and allocation trade-offs
328 that may be negotiated differently by each sex. This explanation is especially plausible if
329 males and females modify their phenotypes to different degree in response to captivity.
330 Note too that the environmental contribution to females' behavioral phenotype was much
331 greater than that of males, leaving space for differential adjustment of behavior by sexes to
332 occur.

333 Males and females differed substantially in how they expressed a boldness-activity
334 syndrome. Contrary to our predictions, in both sexes active genotypes were also more
335 sensitive to predatory cues, with no change to the magnitude of the genetic correlation. The
336 primary difference was therefore in how shelter emergence related to activity and
337 antipredator response. “Bold” females – i.e. females with fast emergence from the shelter –
338 had higher activity and antipredator response. In contrast, active males were more
339 sensitive to predator cues and behaved cautiously when emerging from the shelter. This
340 result provides important insight into how male and female crickets handle risky
341 situations. Female must travel through risky environments in order to locate mates.
342 Therefore, bold and active genotypes may need to compensate for these risky behaviors by
343 being more reactive to the presence of predator cues. Males, in contrast, stay close to their
344 shelter but produce courtship signals that make them the target of predators and parasites.
345 As a result, bolder males may ignore the presence of predatory cues if this strategy yields
346 higher frequency of encounters with females. This type of risk compensation strategy has
347 already been shown in previous studies where males with more attractive songs stay
348 longer in shelters (Hedrick 2000).

349 Our findings that males and females differed in the genetic expression behavioral
350 syndromes and that cross-sex correlations are uncoupled suggest that risk-taking is
351 regulated by different physiological pathways in each sex. In crickets, several monoamine
352 neurohormones are involved in the regulation of multiple behaviors tied to a “fight or
353 flight” response, including aggression, courtship, dispersal and response to simulated
354 predation exposure (Adamo et al. 1995, 2013; Stevenson and Rillich 2016; Adamo 2017;
355 Lundgren et al. 2021). However, experiments tend to either focus on males exclusively or
356 are not designed to address sex-differences in physiological mechanisms. However, sex-
357 differences in a similar axis of variation, the proactive/reactive axis, have also been
358 described in several vertebrate taxa (Kokras et al. 2012; Immonen et al. 2018).

359 We showed previously that the genetic structure of behavioral syndromes is highly
360 conserved in this species, even across isolated populations (Royauté et al. 2020). This
361 indicates that behavioral syndromes may not vary much when exposed to different
362 selective pressures. Instead, genetic constraints resulting from pleiotropy was the more

363 likely explanations for the presence of behavioral syndrome in this species. Here, we show
364 that behavioral syndromes differed more strongly between sexes than among populations.
365 A likely explanation is that males and females express sex-specific behaviors regardless of
366 the population of origin. As a result, selection may be more likely to differ between sexes
367 than between populations. While comparisons of differences in selection among sexes and
368 populations are scarce, previous studies suggest that sexually discordant selection is
369 common in the wild and tends to be stronger in traits in which sexual dimorphism is
370 pronounced (Cox et al. 2009).

371 While rarely investigated, sex specific differences in behavioral syndromes have
372 been demonstrated in some species. However, most of these studies have focused on
373 comparisons of behavioral repeatabilities between males and females in single traits
374 (Jenkins 2011; Debeffe et al. 2015) rather providing a broader exploration of multivariate
375 patterns of cross-sex correlation in behavioral traits, as we did here. Interestingly, (Hedrick
376 and Kortet 2012) previously identified sex differences in the repeatability of shelter
377 emergence in a separate population of *G. integer*. Contrary to what we detected; males had
378 lower repeatability compared to females. Note that repeatability and heritability are not
379 directly comparable because repeatability also includes sources of variation due to the
380 “permanent environment” while heritability only includes additive genetic variation. More
381 recently several studies have also compared syndrome structure among sexes with mixed
382 results. Some studies reported large differences in behavioral syndromes between males
383 and females (Fresneau et al. 2014; Han et al. 2015; Royauté 2015; Way et al. 2015) while
384 others support a conserved syndrome structure between sexes (Michelangeli et al. 2016;
385 Goulet et al. 2021). Our results supply an additional line of evidence in favor of sex-specific
386 syndromes.

387 There is only limited exploration of multivariate patterns of cross-sex correlation in
388 behavioral traits. Most studies reported from (Poissant et al. 2010) concern single traits
389 and point to strong cross-sex correlations with relatively weak dimorphism for behavior.
390 The majority of these estimates were, however, from studies interested in sexual selection,
391 with < 4 % of estimates coming from explicitly behavioral studies. Because we also
392 estimated the cross-sex correlation at the additive genetic level, we were able to determine

393 that behavioral traits may be less constrained by sex than previously thought. Indeed, our
394 mean estimate for r_{mf} was much weaker than observed in previous studies evaluating intra-
395 locus sexual conflicts in behavior (mean r_{mf} = 0.46 vs. 0.77, Poissant et al. 2010). Our
396 results suggest that behavioral traits may commonly fulfill different functions between
397 sexes and should therefore be analyzed as separate traits between the sexes in behavioral
398 syndrome and animal personality studies.

399 Very few studies have been able to estimate the genetic contribution of sexes to
400 behavioral syndromes as we have done here. In the orb-weaving spider *Nuctenea*
401 *umbratical*, cross-sex correlations for activity and aggression were in the same range as our
402 estimates for activity and antipredator response (r_{mf} = 0.50) (Kralj-Fišer et al. 2019). In
403 *Gryllus bimaculatus*, which is closely related to *G. integer*, exploration and aggression are
404 more strongly correlated in females and these traits have asymmetric contributions across
405 sexes (Han et al. 2019). However, cross-sex correlations within traits were qualitatively
406 stronger than observed in our study (mean r_{mf} = 0.70 Lande 1980 vs mean r_{mf} = 0.46),
407 pointing to a stronger genetic constraint between sexes than we observed in *Gryllus*
408 *integer*. Finally, White et al. (2019) showed differences between sex-specific covariances in
409 guppies to similar *G. integer*. However, they reported a much weaker signal of asymmetry
410 in **B**. The fact that these few studies differ widely in conclusions shows that there is no
411 consensus yet on how behavioral syndromes differ between sexes and even closely related
412 species may show strong differences in the expression of behavioral dimorphism.

413 Other studies, conducted primarily on morphological traits, point to strong
414 constraints imposed by genetic cross-sex covariances (Gosden et al. 2012; Gosden and
415 Chenoweth 2014; Sztepanacz and Houle 2019). In our case, the constraining effect of cross-
416 sex covariances was weak at best and responses to selection were more consistent with
417 independent evolutionary trajectories for each sex. This is similar to a previous study on
418 sexually dimorphic ornaments in brown anoles which showed that the cross-sex
419 covariance among ornaments did not substantially constrain the evolution of dimorphism
420 (Cox et al. 2017).

421 We found a high degree of asymmetry in the cross-sex cross-traits components of
422 the genetic covariance matrix. In particular, male activity had a stronger contribution to
423 female shelter emergence than female activity on male shelter emergence. This type of
424 asymmetric contribution to the phenotype is expected to bias responses away from
425 selection. In *Drosophila* for example, wing shape evolution is primarily shaped by cross-sex
426 covariances for the same trait (i.e. the diagonal elements of **B**) rather than cross-sex cross-
427 trait covariances (Sztepanacz and Houle 2019). This was the case even though upper and
428 lower elements of the **B** matrix frequently had opposite signs and despite a general, though
429 weaker, signal of asymmetry such as we report here (5 % vs. 25 %). Despite strong
430 asymmetry in our estimate of cross-sex covariances, we found only weak evidence for
431 biased responses to selection. Instead, our simulations of the responses to selections
432 showed that the evolution of behavioral dimorphisms was more consistent with scenarios
433 in which sexes are genetically uncoupled and allowed to evolve independently compared to
434 a fully constrained cross-sex genetic architecture. This means that the independent
435 evolutionary trajectories by sexes we report are likely a result of the weak cross-sex
436 correlations within traits we detected (mean $r_{mf} < 0.50$).

437 We show that traits involved in studies of animal personality have a sex-specific
438 genetic architecture. These behaviors are genetically uncoupled between sexes and allow
439 sexes to follow independent evolutionary trajectories. Our results are intriguing because
440 theory predicts that cross-sex covariances will act as constraints and limit the abilities of
441 sexes to reach their optimum. The type of genetic independence we found suggests a past
442 history of discordant selection that has strongly shaped behavioral dimorphism and has
443 resolved sexual conflict in this species.

444 **ACKNOWLEDGEMENTS**

445 We thank Monica Berdal, Katelyn Cannon, Jeremy Dalos, Sarah Felde, Brady Klock, Ishan Joshi,
446 Hannah Lambert, Jenna LaCoursiere and Alondra Neunsinger for assistance in conducting
447 behavioral trials and in rearing and care of the crickets and Martori Farms, David Lightfoot, Scott
448 Bundy, Nico Franz, Sangmi Lee, Cameron Jones, Kenny Chapin, Ti Eriksson, Meranda Feagins,
449 Charlotte Mulloney, Melody Martinez, Allyson Richins, Mauriel Rodriguez, Helen Vessels and David
450 Wikman for assistance in collecting the crickets. We thank for J. Sztepanacz and A.J. Wilson for their

451 advice on statistical analyses. This work was supported by US NSF IOS grants 1557951 and
452 1558069 to N.A.D. and A.H. respectively.

453 **LITTERATURE CITED**

- 454 Adamo, S. A. 2017. The stress response and immune system share, borrow, and reconfigure
455 their physiological network elements: Evidence from the insects. *Hormones and Behavior,*
456 *Neuroendocrine-Immune Interactions: Implications for Integrative and Comparative*
457 *Physiologists* 88:25–30.
- 458 Adamo, S. A., I. Kovalko, and B. Mosher. 2013. The behavioural effects of predator-induced
459 stress responses in the cricket (*Gryllus texensis*): the upside of the stress response. *Journal*
460 *of Experimental Biology* 216:4608–4614.
- 461 Adamo, S. A., C. E. Linn, and R. R. Hoy. 1995. The role of neurohormonal octopamine during
462 “fight or flight” behaviour in the field cricket *Gryllus bimaculatus*. *Journal of Experimental*
463 *Biology* 198:1691–1700.
- 464 Aragón, P. 2011. The response to the social environment reveals sex-dependent
465 behavioural syndromes in the Bosca’s newt (*Lissotriton boscai*). *Journal of Ethology* 29:79–
466 83.
- 467 Arnqvist, G. 1997. The evolution of animal genitalia: distinguishing between hypotheses by
468 single species studies. *Biological Journal of the Linnean Society* 60:365–379.
- 469 Berger, D., E. C. Berg, W. Widegren, G. Arnqvist, and A. A. Maklakov. 2014. Multivariate
470 intralocus sexual conflict in seed beetles. *Evolution* 68:3457–3469.
- 471 Blanckenhorn, W. U. 2005. Behavioral Causes and Consequences of Sexual Size
472 Dimorphism. *Ethology* 111:977–1016.
- 473 Bonduriansky, R., and S. F. Chenoweth. 2009. Intralocus sexual conflict. *Trends in Ecology &*
474 *Evolution* 24:280–288.
- 475 Cox, R. M., R. A. Costello, B. E. Camber, and J. W. McGlothlin. 2017. Multivariate genetic
476 architecture of the *Anolis dewlap* reveals both shared and sex-specific features of a sexually
477 dimorphic ornament. *Journal of Evolutionary Biology* 30:1262–1275.
- 478 Debeffe, L., J. F. Lemaître, U. A. Bergvall, A. J. M. Hewison, J. M. Gaillard, N. Morellet, M.
479 Goulard, et al. 2015. Short- and long-term repeatability of docility in the roe deer: sex and
480 age matter. *Animal Behaviour* 109:53–63.
- 481 Delph, L. F., J. C. Steven, I. A. Anderson, C. R. Herlihy, and E. D. B. Iii. 2011. Elimination of a
482 Genetic Correlation Between the Sexes Via Artificial Correlational Selection. *Evolution*
483 65:2872–2880.
- 484 Dochtermann, N. A., and N. J. Dingemanse. 2013. Behavioral syndromes as evolutionary
485 constraints. *Behavioral Ecology* 24:806–811.
- 486 French, B. W., and W. H. Cade. 1987. The timing of calling, movement, and mating in the
487 field crickets *Gryllus veletis*, *G. pennsylvanicus*, and *G. integer*. *Behavioral Ecology and*
488 *Sociobiology* 21:157–162.
- 489 Fresneau, N., E. Kluehn, and J. E. Brommer. 2014. A sex-specific behavioral syndrome in a
490 wild passerine. *Behavioral Ecology* 25:359–367.

491 Gosden, T. P., and S. F. Chenoweth. 2014. The Evolutionary Stability of Cross-Sex, Cross-
492 Trait Genetic Covariances. *Evolution* 68:1687–1697.

493 Gosden, T. P., K.-L. Shastri, P. Innocenti, and S. F. Chenoweth. 2012. The B-Matrix Harbors
494 Significant and Sex-Specific Constraints on the Evolution of Multicharacter Sexual
495 Dimorphism. *Evolution* 66:2106–2116.

496 Goulet, C. T., W. Hart, B. L. Phillips, J. Llewelyn, B. B. M. Wong, and D. G. Chapple. 2021. No
497 behavioral syndromes or sex-specific personality differences in the southern rainforest
498 sunskink (*Lampropholis similis*). *Ethology* 127:102–108.

499 Hadfield, J. D. 2010. MCMC methods for multi-response generalized linear mixed models:
500 the MCMCglmm R package. *Journal of Statistical Software* 33:1–22.

501 Hämäläinen, A., E. Immonen, M. Tarka, and W. Schuett. 2018. Evolution of sex-specific pace-
502 of-life syndromes: causes and consequences. *Behavioral Ecology and Sociobiology* 72:50.

503 Han, C. S., T. P. Gosden, and N. J. Dingemans. 2019. Protein deprivation facilitates the
504 independent evolution of behavior and morphology. *Evolution* 73:1809–1820.

505 Han, C. S., P. G. Jablonski, and R. C. Brooks. 2015. Intimidating courtship and sex differences
506 in predation risk lead to sex-specific behavioural syndromes. *Animal Behaviour* 109:177–
507 185.

508 Hedrick, A. V., and R. Kortet. 2012. Sex differences in the repeatability of boldness over
509 metamorphosis. *Behavioral Ecology and Sociobiology* 66:407–412.

510 Hedrick, A. V., and E. J. Temeles. 1989. The evolution of sexual dimorphism in animals:
511 Hypotheses and tests. *Trends in Ecology & Evolution* 4:136–138.

512 House, C., P. Tunstall, J. Rapkin, M. J. Bale, M. Gage, E. del Castillo, and J. Hunt. 2020.
513 Multivariate stabilizing sexual selection and the evolution of male and female genital
514 morphology in the red flour beetle*. *Evolution* 74:883–896.

515 Immonen, E., A. Hämäläinen, W. Schuett, and M. Tarka. 2018. Evolution of sex-specific pace-
516 of-life syndromes: genetic architecture and physiological mechanisms. *Behavioral Ecology*
517 and *Sociobiology* 72:60.

518 Jenkins, S. H. 2011. Sex differences in repeatability of food-hoarding behaviour of kangaroo
519 rats. *Animal Behaviour* 81:1155–1162.

520 Kokras, N., C. Dalla, A. C. Sideris, A. Dendi, H. G. Mikail, K. Antoniou, and Z. Papadopoulou-
521 Daifoti. 2012. Behavioral sexual dimorphism in models of anxiety and depression due to
522 changes in HPA axis activity. *Neuropharmacology, Anxiety and Depression* 62:436–445.

523 Kortet, R., and A. Hedrick. 2007. A behavioural syndrome in the field cricket *Gryllus integer*:
524 intrasexual aggression is correlated with activity in a novel environment. *Biological Journal*
525 of the *Linnean Society* 91:475–482.

526 Kralj-Fišer, S., K. L. Laskowski, and F. Garcia-Gonzalez. 2019. Sex differences in the genetic
527 architecture of aggressiveness in a sexually dimorphic spider. *Ecology and Evolution*
528 9:10758–10766.

529 Kruuk, L. E. B. 2004. Estimating genetic parameters in natural populations using the ‘animal
530 model.’ *Philosophical Transactions of the Royal Society of London. Series B: Biological*
531 *Sciences* 359:873–890.

532 Lande, R. 1980. Sexual Dimorphism, Sexual Selection, and Adaptation in Polygenic
533 Characters. *Evolution* 34:292–305.

534 Long, T. A. F., and W. R. Rice. 2007. Adult locomotory activity mediates intralocus sexual
535 conflict in a laboratory-adapted population of *Drosophila melanogaster*. *Proceedings of the*
536 *Royal Society B: Biological Sciences* 274:3105–3112.

537 Lundgren, K. A., R. N. Abbey-Lee, L. C. Garnham, A. Kreshchenko, S. Ryding, and H. Løvlie.
538 2021. Manipulating monoamines reduces exploration and boldness of Mediterranean field
539 crickets. *Behavioural Processes* 183:104298.

540 Mainwaring, M. C., J. L. Beal, and I. R. Hartley. 2011. Zebra finches are bolder in an asocial,
541 rather than social, context. *Behavioural Processes* 87:171–175.

542 Michelangeli, M., D. G. Chapple, and B. B. M. Wong. 2016. Are behavioural syndromes sex
543 specific? Personality in a widespread lizard species. *Behavioral Ecology and Sociobiology*
544 70:1911–1919.

545 Moiron, M., K. L. Laskowski, and P. T. Niemelä. 2020. Individual differences in behaviour
546 explain variation in survival: a meta-analysis. *Ecology Letters* 23:399–408.

547 Poissant, J., A. J. Wilson, and D. W. Coltman. 2010. Sex-Specific Genetic Variance and the
548 Evolution of Sexual Dimorphism: A Systematic Review of Cross-Sex Genetic Correlations.
549 *Evolution* 64:97–107.

550 Réale, D., S. M. Reader, D. Sol, P. T. McDougall, and N. J. Dingemanse. 2007. Integrating
551 animal temperament within ecology and evolution. *Biological Reviews* 82:291–318.

552 Royauté, C., R. , Buddle, C. M. , Vincent. 2015. Under the influence: Sublethal exposure to an
553 insecticide affects personality expression in a jumping spider 29.

554 Royauté, R., and N. A. Dochtermann. 2017. When the mean no longer matters:
555 Developmental diet affects behavioral variation but not population averages in the house
556 cricket (*Acheta domesticus*). *Behavioral Ecology* 28:337–345.

557 Royauté, R., C. Garrison, J. Dalos, M. A. Berdal, and N. A. Dochtermann. 2019. Current energy
558 state interacts with the developmental environment to influence behavioural plasticity.
559 *Animal Behaviour* 148:39–51.

560 Royauté, R., K. Greenlee, M. Baldwin, and N. A. Dochtermann. 2015. Behaviour, metabolism
561 and size: phenotypic modularity or integration in *Acheta domesticus*? *Animal behaviour*
562 110:163–169.

563 Royauté, R., A. Hedrick, and N. A. Dochtermann. 2020. Behavioural syndromes shape
564 evolutionary trajectories via conserved genetic architecture. *Proceedings of the Royal*
565 *Society B: Biological Sciences* 287:20200183.

566 Schuett, W., T. Tregenza, and S. R. X. Dall. 2010. Sexual selection and animal personality.
567 *Biological Reviews* 85:217–246.

- 568 Sih, A., A. Bell, and J. C. Johnson. 2004. Behavioral syndromes: an ecological and
569 evolutionary overview. *Trends in Ecology & Evolution* 19:372–378.
- 570 Stevenson, P. A., and J. Rillich. 2016. Controlling the decision to fight or flee: the roles of
571 biogenic amines and nitric oxide in the cricket†. *Current Zoology* 62:265–275.
- 572 Sztepanacz, J. L., and D. Houle. 2019. Cross-sex genetic covariances limit the evolvability of
573 wing-shape within and among species of *Drosophila*. *Evolution* 73:1617–1633.
- 574 Walsh, B., and M. W. Blows. 2009. Abundant Genetic Variation + Strong Selection =
575 Multivariate Genetic Constraints: A Geometric View of Adaptation. *Annual Review of*
576 *Ecology, Evolution, and Systematics* 40:41–59.
- 577 Way, G. P., A. L. Kiesel, N. Ruhl, J. L. Snekser, and S. P. McRobert. 2015. Sex differences in a
578 shoaling-boldness behavioral syndrome, but no link with aggression. *Behavioural*
579 *Processes* 113:7–12.
- 580 Wilson, A. J., D. Réale, M. N. Clements, M. M. Morrissey, E. Postma, C. A. Walling, L. E. B.
581 Kruuk, et al. 2010. An ecologist’s guide to the animal model. *Journal of Animal Ecology*
582 79:13–26.
- 583