

Phylogenetic conservation of behavioral variation and behavioral syndromes

Jeremy Dalos ^{a, b}
Raphael Royauté ^{c, d}
Ann V. Hedrick ^{e, f}
Ned A. Dochtermann ^{a, g}

^a Department of Biological Sciences; North Dakota State University

^b jeremy.dalos@ndsu.edu

^c Movement Ecology Group, Senckenberg Biodiversity and Climate Research Centre (SBIK-F), Frankfurt, Germany

^d raphael.royaute@gmail.com

^e Department of Neurobiology, Physiology, and Behavior; University of California, Davis

^f avhedrick@ucdavis.edu

^g **corresponding author:** ned.dochtermann@gmail.com

Author to whom correspondence should be addressed:

Ned Dochtermann
1340 Bolley Drive, 201 Stevens Hall
Department of Biological Sciences
North Dakota State University
Fargo, ND 58102

Running Title: Conservation of behavioral (co)variation

Keywords: behavioral syndrome, personality, field crickets, comparative method

Acknowledgements

We thank Jonathan Albers, Monica Berdal, Katelyn Cannon, Sarah Felde, Brady Klock, Ishan Joshi, Hannah Lambert, Jenna LaCoursiere, Alondra Neunsinger, Katie Pnewski, Maddi Rick, Teresa Tesarek, and Amanda Wilson for assistance in conducting behavioral trials and in rearing and care of the crickets and Martori Farms, David Lightfoot, Scott Bundy, Nico Franz, Sangmi Lee, Cameron Jones, Kenny Chapin, Ti Eriksson, Meranda Feagins, Charlotte Mulloney, Melody Martinez, Allyson Richins, Mauriel Rodriguez, Helen Vessels and David Wikman for assistance in collecting *Gryllus integer*, *Gryllus lineaticeps*, and *Gryllus assimilis*. We also thank Scott Sakaluk for providing us with populations of *Gryllodes sigillatus*. This work was supported by US NSF IOS grants 1557951 and 1558069 to N.A.D. and A.H. respectively.

Conflicts of Interest

The authors have no conflicts of interest.

Data Availability Statement

All data and code are available at: <https://osf.io/q23ud/>
DOI 10.17605/OSF.IO/Q23UD

Phylogenetic conservation of behavioral variation and behavioral syndromes

Abstract

1
2 Individuals frequently differ consistently from one another in their average behaviors (i.e.
3 “animal personality”) and in correlated suites of consistent behavioral responses (i.e.
4 “behavioral syndromes”). However, understanding the evolutionary basis of this
5 (co)variation has lagged behind demonstrations of its presence. This lag partially stems
6 from comparative methods rarely being used in the field. Consequently, much of the
7 research on animal personality has relied on “adaptive stories” focused on single species
8 and populations. Here we used a comparative approach to examine the role of phylogeny in
9 shaping patterns of average behaviors, behavioral variation, and behavioral correlations. In
10 comparing the behaviors and behavioral variation for five species of Gryllid crickets we
11 found that phylogeny shaped average behaviors and behavioral (co)variation. Despite
12 differences among species, behavioral responses and variation were most similar among
13 more closely related species. These results suggest that phylogenetic constraints play an
14 important role in the expression of animal personalities and behavioral syndromes and
15 emphasize the importance of examining evolutionary explanations within a comparative
16 framework.

Introduction

17
18 Behavioral syndromes, correlations between behaviors at the among-individual
19 level (Dingemanse et al. 2012), have been documented across taxa (Brommer and Class
20 2017). Behavioral syndromes can conceptually be thought of as correlations between
21 individual averages and stem from underlying genetic correlations and correlations due to
22 developmental plasticity and other sources of permanent environmental covariance
23 (Dingemanse et al. 2012, Dingemanse and Dochtermann 2014). Among-individual variation
24 in behavior, often referred to as “personality variation”, has been found to be similarly
25 ubiquitous (Bell et al. 2009). Similar to behavioral syndromes, this personality variation
26 can be thought of as variation across individuals in their average behaviors and likewise
27 stem from genetic and permanent environmental variation (Dingemanse and Dochtermann

28 2013, Dochtermann et al. 2015). Attempts to infer whether general taxonomic patterns
29 exist for both personality variation and behavioral syndromes have generally been
30 conducted via literature reviews and meta-analyses (Bell et al. 2009, Dochtermann 2011,
31 Garamszegi et al. 2012, 2013, Dochtermann et al. 2015, Brommer and Class 2017). These
32 synthesis efforts have shown that among-individual variation is common (average
33 repeatability ~ 0.37 , Bell et al. 2009), that the magnitude of behavioral syndromes is
34 generally weak (average $r \sim 0.19$, Garamszegi et al 2012, 2013), and that there is general
35 alignment between nested patterns of correlations at the phenotypic, among-individual,
36 within-individual, and genetic levels (Dochtermann 2011, Brommer and Class 2017).

37 Although the observation that both among-individual variation and behavioral
38 syndromes are common, we have a poor understanding of the evolution of either. This gap
39 in our understanding is partly because comparative approaches have rarely been used in
40 studies of among-individual behavioral variation and behavioral syndromes (White et al.
41 2020). Comparative approaches allow for direct comparison across species of behavioral
42 (co)variation and are necessary for a proper understanding of the importance of phylogeny
43 in shaping “personality” and behavioral syndromes (Royauté et al. 2020, White et al. 2020).

44 Direct assessment of evolutionary hypotheses can also be extended to the study of
45 personality and behavioral syndromes: both among-individual variation and behavioral
46 syndromes have clear connections to quantitative genetic parameters; specifically, additive
47 genetic variation, and additive genetic covariances (Dochtermann and Roff 2010,
48 Dingemanse and Dochtermann 2014). The mathematical relationships between among-
49 individual (co)variances and additive genetic (co)variances (Boake 1989, Dingemanse and
50 Dochtermann 2014, Dochtermann et al. 2015) allows the extension of predictions from
51 quantitative genetics to among-individual variation and behavioral syndromes.

52 One such prediction is that differences in the magnitude of variation present for a
53 trait might be attributable to differences in selection between populations or species.
54 Specifically, Mousseau and Roff (1987) argued that traits with low heritability might be
55 indicative of strong selection having eroded genetic variation. Likewise, because among-
56 individual variation represents the sum of additive genetic variation, dominance (and other
57 epistatic) genetic variation, and permanent environmental variation (e.g. irreversible and
58 developmental plasticity), selection is expected to deplete this variation. Note, however,

59 that drift is often also expected to reduce genetic and, therefore, among-individual
60 variation.

61 Selection is likewise expected to shape additive genetic covariances and
62 correlations, both by the loss of variation in single traits and changes to the magnitude and
63 directions of covariances (Roff 1997). For example, correlational selection is expected to
64 produce genetic correlations (Phillips and Arnold 1989, Armbruster and Schwaegerle
65 1996). As in the case of among-individual variances, these effects on genetic correlations
66 are expected to carry over to behavioral syndromes. In other words, behavioral syndromes
67 are expected to reflect the effects of selection on genetic correlations. Therefore, if
68 behavioral syndromes differ across populations, species, or other groupings, then this
69 suggests differences in genetic correlations and correlational selection (i.e. the "adaptive"
70 hypothesis, Bell 2005). In contrast, if behavioral syndromes are conserved across groups
71 then this would suggest that either behavioral syndromes stem from pleiotropic effects (i.e.
72 the "constraints" hypothesis, Bell 2005) or that selection is similar across groups.

73 While these topics have been addressed for other types of traits, particularly
74 morphological and chemical characteristics (Aguirre et al. 2014, Hine et al. 2014,
75 McGlothlin et al. 2018), addressing them for behavior remains important for several
76 reasons. First, considerable behavioral research assumes an adaptive framework for both
77 among-individual variation and behavioral syndromes, thereby minimizing the importance
78 of phylogeny and minimizing the potential role of phylogenetic constraints. Second,
79 behaviors, life-history, and physiological traits exhibit substantially lower heritabilities
80 than do morphological traits (Mousseau and Roff 1987, Stirling et al. 2002, Dochtermann et
81 al. 2019). Consequently, the role of phylogeny and selection in constraining and shaping
82 morphology may not generalize to traits with lower heritabilities and thus greater
83 plasticity. Failure to address alternative explanations, like phylogenetic constraints, can
84 lead to the uncritical acceptance of adaptive arguments (Gould and Lewontin 1979) and the
85 perpetuation of "zombie ideas" (sensu Quiggin 2012). For example, recent meta-analyses
86 have cast doubt on general organizing frameworks for behavior and behavioral
87 correlations (Niemelä and Dingemanse 2018, Royauté et al. 2018).

88 Here, we compared the behavior of five closely related cricket species: *Gryllus*
89 *integer*, *Gryllus assimilis*, *Gryllus lineaticeps*, *Gryllodes sigillatus*, and *Acheta domesticus*. For

90 each species we measured exploratory behavior and response to cues of predator
91 presence. By working with the same behavioral assays in five closely related species we
92 were able to assess the importance of phylogeny for average behaviors and to evaluate
93 predictions about trait (co)variation. Specifically, we addressed the following questions:

94 1. Does the average expression of behavior differ among species?

95 We predicted that species would differ but do so in a manner constrained by
96 phylogeny. Put another way, more closely related species will have more similar
97 average behaviors.

98 2. Do among-individual variances differ among species?

99 We did not have species level predictions but, because selection and drift should
100 both reduce among-individual variance, we predicted that among-individual
101 variation would differ across species independent of phylogeny.

102 3. Do within-individual variances differ among species? Within-individual variation,

103 typically disregarded as residual variation, includes phenotypic plasticity—

104 specifically reversible plasticity or “phenotypic flexibility” not captured by factors

105 and covariates of a statistical model (Piersma and Drent 2003, Whitman and

106 Agrawal 2009, Piersma and Van Gils 2011, Westneat et al. 2015, Berdal and

107 Dochtermann 2019). Differences across groups in the magnitude of within-

108 individual variation therefore are, in part, differences in the magnitude of plasticity.

109 We did not have *a priori* expectations as to species differences or phylogenetic

110 signal for within-individual variances.

111 4. Do behavioral syndromes differ among species?

112 Because behavioral syndrome structure has been conserved at the genetic level

113 across cricket populations of *G. integer* (Royauté et al. 2020), we predicted that

114 syndromes would similarly be phylogenetically conserved and shared across

115 species.

116

Methods

117 *Cricket Acquisition, Housing, and Rearing Conditions*

118 Data used in this study were originally collected for various studies investigating the
119 effects of development on behavioral variation and the presence of behavioral constraints
120 and behavioral syndromes (Royauté et al. 2019, Royauté et al. 2020). *A. domesticus* males
121 and females were obtained as nymphs (~ 1 mm in size) from a commercial supplier
122 (Fluker's Cricket Farm, Port Allen, LA, U.S.A.) and were measured once mature. *G. integer*
123 females were captured in Aguila, AZ, *G. lineaticeps* males and females were caught in
124 Dunnigan, CA, and the *G. assimilis* males and females were caught in Maricopa County, AZ.
125 These species were all captured during the summer of 2017. *G. sigillatus* individuals were
126 taken from an outbred population established by S. Sakaluk with crickets collected from
127 California and currently maintained in Fargo, ND. For *G. lineaticeps* and *G. assimilis*, the
128 same individuals that were caught in the field were measured, while lab reared offspring of
129 *G. integer* were measured. All species were reared under a 12:12 light: dark photoperiod at
130 a temperature of 25-28°C. All individuals were housed in 0.71-liter containers with
131 transparent covers that included food, shelter, and water filled glass vials plugged with
132 cotton balls. *A. domesticus* were exposed to a mixture of high and low quality diets
133 described in Royauté et al. (2019), while all other species included in this study were fed ad
134 libitum food (commercially purchased chicken feed).

135 *Behavior Trials*

136 To measure exploratory behavior and anti-predator responses we repeatedly
137 recorded individuals' activity levels in an open field arena, followed by their responses to
138 cues of predator presence created from diluted *Eublepharis macularius* excreta (see details
139 below). *A. domesticus* were measured between March 2015 and October 2016, *G. lineaticeps*
140 were measured from August 2017 to September 2017, *G. assimilis* were measured between
141 September 2017 and October 2017, *G. integer* were measured between May 2018 and June
142 2018, and *G. sigillatus* were measured in May 2019. All trials were conducted in a plastic
143 arena (60 cm x 60 cm and 15 cm high) with a Plexiglas lid. The arena was split into four 30
144 cm x 30 cm arenas separated by a divider, allowing up to four crickets to be tested at one
145 time. Open field trials were always conducted first followed by antipredator response trials
146 either immediately after or on another day to minimized potential carryover effects from

147 exposure to cues of predator presence. After each behavioral assay, arenas were
 148 thoroughly cleaned with 70% ethanol wipes to avoid accumulation of any chemical traces
 149 of conspecifics. Mass at the time of behavioral trials was recorded to the nearest 1 mg. All
 150 individuals were measured in each assay for a maximum of three repetitions, with some
 151 individuals measured fewer times due to escape or natural mortality (Table 1). In total, we
 152 conducted 2478 behavioral assays across a total of 460 individuals (Table 1).

153 By measuring behavior in the same manner across species we reduce the likelihood
 154 of naming fallacies—i.e. jingle fallacies (where the same name is used for the different
 155 behaviors) and jangle fallacies (where different names are used for the same behavior).
 156 The standardized protocols also allow us to assume similar measurement error across
 157 species.

Table 1. Number of individuals, by species, for which behavior was assayed in a first, second, and third repetition.

Species	Behavioral Assay	Repetition 1	Repetition 2	Repetition 3	Total Trials
<i>Acheta domesticus</i>	Open field	281	263	225	769
	Antipredator	262	235	220	717
<i>Gryllus assimilis</i>	Open field	16	16	16	48
	Antipredator	16	16	15	47
<i>Gryllus integer</i>	Open field	92	91	74	257
	Antipredator	88	88	72	248
<i>Gryllus lineaticeps</i>	Open field	21	17	11	49
	Antipredator	21	13	11	45
<i>Gryllodes sigillatus</i>	Open field	50	50	49	149
	Antipredator	50	50	49	149
Total		896	837	743	2478

158 *Open field behavior*

159 Individual crickets were left to rest for 30 seconds under a 5 cm diameter cup after
 160 being introduced into the lower right section of the arena (Figure S1). After these 30
 161 seconds we allowed the individuals to move freely through the arena for 220 seconds. We
 162 measured each individual’s exploratory propensity by digitally overlaying a 6 × 6 grid over
 163 the arena (Figure S1) and calculating the number of *unique zones* visited (UZ) by the cricket
 164 with Ethovision X (Noldus Information Technology, Wageningen, The Netherlands). This
 165 behavioral protocol has previously been used with *A. domesticus* and *G. integer* to evaluate

166 genetic and individual differences in activity and exploratory behaviors (Royauté et al.
167 2015, Royauté and Dochtermann 2017, Royauté et al. 2019, Royauté et al. 2020).

168 *Predator cue response*

169 To measure responses to cues of potential predator presence, we collected excreta
170 from three adult leopard geckos, *Eublepharis macularius*, that were fed a mixed diet of *A.*
171 *domesticus*, *G. sigillatus*, *G. lineaticeps*, *G. integer*, and *G. assimilus*. Leopard geckos were
172 housed according to the standards of the Institutional Animal Care and Use Committee of
173 North Dakota State University (Protocol A14006, A17015, and A19067) and the Animal
174 Behavior Society (2020). Collected excreta was frozen and then finely ground and diluted
175 with deionized water (1 ml H₂O: 5 mg of excreta). This solution was then applied to 15 cm
176 diameter filter paper disks with a 5 cm diameter central cutout that allowed crickets to be
177 left to rest unexposed to the predator cues (Royauté and Dochtermann 2017, Royauté et al.
178 2019, Royauté et al. 2020). Each predator cue disk was left to dry for a minimum of 2 hours
179 then stored at -23°C until needed for trials. Predator cue disks were allowed to warm to
180 room temperature before use in antipredator trials and discarded after a single use.
181 Between each trial thawed cue disks were stored at 4°C for a maximum of 14 days. After 14
182 days any unused disks were discarded.

183 We placed the predator cue disk at the bottom of a 15 cm diameter arena and left
184 the cricket to rest for a minimum of 30 seconds under a 5 cm diameter cup in the
185 nontreated central cutout. We then removed the cup and allowed the cricket to move freely
186 for 220 seconds and estimated the distance travelled in cm (AP distance) using Ethovision
187 X (Figure S1). Previous studies with this protocol show that crickets had heightened
188 activity levels in the presence of this diluted gecko excreta compared to water controls
189 (Royauté and Dochtermann 2017). Consistent with this, *G. sigillatus* crickets have been
190 found to increase their activity after direct exposure to predators (Bucklaew and
191 Dochtermann 2020). Greater activity during these antipredator response assays, i.e.
192 greater *AP distance*, was therefore interpreted as a greater responsiveness to predator
193 cues.

194 *Data Analysis: Univariate Models*

195 To assess differences in behavioral responses between species for means and
196 variances we analyzed behavioral data using separate univariate mixed-effects models for
197 unique zones visited and AP distance (square root transformed). We included species,
198 temperature (Celsius, mean centered), mass (using among- and within-individual centering
199 (Van de Pol and Wright 2009)), and sex as fixed effects. Individual ID was included as a
200 random effect. We compared the fit of four univariate mixed models structured as follows:

- 201 1) Model 1: $V_i = & V_w = A$ A null model where the among- (V_i) and within-individual (V_w)
202 variances were kept constant between species.
- 203 2) Model 2: $V_i \neq & V_w = A$ A model where the among-individual variance differed between
204 species, but the within-individual variance was kept constant.
- 205 3) Model 3: $V_i = & V_w \neq$ The within-individual variance differs between species, but the
206 among-individual variance was kept constant.
- 207 4) Model 4: $V_i \neq & V_w \neq$ Both the among and within-individual variances were allowed
208 to vary between species.

209 These models were specified using the `MCMCglmm` package for Bayesian mixed models
210 (Hadfield 2010) using Markov-chain Monte Carlo (MCMC) with 1.3 million iterations,
211 300,000 iteration burn-in, a thinning interval of 1000, and an inverse-Wishart prior. AP
212 distance and unique zone models were fit with Gaussian and Poisson error distributions,
213 respectively.

214 To determine whether species differed in average behavior, Models 1 and 4 were run
215 with and without species as a fixed effect and compared based on deviance information
216 criterion (DIC) values. If species differ in average behavior, models with species included as
217 a fixed effect would be expected to have lower (DIC) values. Average behavioral differences
218 among species reported in the Results section were then qualitatively assessed using
219 posterior-modal estimates for each species (Congdon 2006).

220 We then compared DIC values among models 1 through 4 to determine whether either
221 among- or within-individual variances differed among species following Royauté et al.
222 (2019) and Royauté and Dochtermann (2021). The model with the lowest DIC value was
223 considered the best model and models with $\Delta \text{DIC} > 5$ were considered to have a

224 substantively poorer fit (Barnett et al. 2010). Models with $\Delta\text{DIC}<5$ were considered as
225 having comparable support relative to the best model (Barnett et al. 2010). All models
226 were specified with the same fixed effect structure as specified above to prevent biased
227 estimates of variance components and repeatability (Spiegelhalter et al. 2003, Nakagawa
228 and Schielzeth 2010b, Westneat et al. 2011).

229 *Data Analysis: Phylogenetic Signal*

230 As our primary questions were about differences in behavioral averages and
231 variances, our results and discussion focus on the above model comparisons. However, we
232 also calculated the variation in behavior directly attributable to phylogeny. To do so, we fit
233 mixed effects models with the same fixed effects, prior structure, and chain parameters as
234 above but omitting Species as a fixed effect. Species was instead incorporated as a random
235 effect, along with individual ID, with the relationship among species modeled according to
236 the current phylogeny (Figure 1, (Gray et al. 2020, Yang et al. 2021)). All nodes in our
237 phylogeny had high support ($\geq 95\%$) in prior phylogenetic analyses (Gray et al. 2020, Yang
238 et al. 2021). While Gray et al. (2020) provided divergence times, these were estimated with
239 high uncertainty and unavailable across genera. Therefore, we did not include branch
240 lengths in our analyses. From these models we then estimated the strength of phylogenetic
241 signal as the proportion of variation attributable to the hierarchical pattern of relatedness
242 among species (i.e. λ , Pagel 1999, Hadfield and Nakagawa 2010, Nakagawa and Santos
243 2012). From the same models we also estimated the proportion of variation attributable to
244 among-individual differences (i.e. τ , repeatability, Dingemanse and Dochtermann 2013). We
245 estimated both phylogenetic signal and repeatability as unadjusted values; that is, we
246 included the variation attributable to fixed effects in the ratio denominator (Nakagawa and
247 Schielzeth 2010a).

248 Importantly, because we only had data for five species, our estimation of
249 phylogenetic signal must be interpreted with caution despite high within-species
250 replication (Table 1). As stated above, our inferences are therefore primarily based on the
251 model comparisons and qualitative comparison of values over the phylogeny.

252 *Data Analysis: Bivariate Models*

253 Behavioral syndromes were estimated using bivariate mixed-effects models with
254 unique zones traveled and AP distance as response variables, also using the `MCMCglmm`
255 library (Hadfield 2010), and analyzed separately for each individual species. We fit models
256 using temperature (Celsius, mean centered), mass (using among- and within-individual
257 centering on subjects (Van de Pol and Wright 2009)), and sex as fixed effects and individual
258 ID was fit as a random effect. These models were fit with 2.6 million iterations, a 600,000
259 burn-in period, a thinning interval of 2000, and a prior that was flat for correlations.
260 Among-individual correlations were estimated for all species, while within-individual
261 correlations were only assessed when individuals were measured for unique zones
262 traveled and antipredator activity during the same testing period (Dingemanse and
263 Dochtermann 2013). Consequently, we were unable to assess within-individual covariation
264 of *G. lineaticeps* and *G. assimilis* due to the fact that these species were not measured for
265 each behavior in immediate succession. Because model comparisons as used above for
266 single traits could not be conducted for correlations (due to software imposed model
267 limitations), differences in behavioral correlations across species were assessed based on
268 whether 95% HPD intervals overlapped. Overlap of 95% intervals is an over-conservative
269 comparison metric (Royauté and Dochtermann 2021), with overlap of 83% intervals more
270 closely approximating an alpha of 0.5 (Austin and Hux 2002, MacGregor-Fors and Payton
271 2013), but this did not affect our species comparison results here. All analyses were
272 conducted in R 3.4.4 (Team 2018).

273 **Results**

274 *Differences in average behavior among species*

275 Species differed in average behaviors: the inclusion of species as a fixed effect
276 substantially improved model fit for both behaviors (Table 2, Table S1). The monophyletic
277 group of *G. assimilis*, *G. integer*, and *G. lineaticeps* exhibited the lowest number of unique
278 zones visited (Figure 1a) but differences in AP distance were less obviously associated with
279 phylogenetic structure (Figure 1b). Consistent with this, phylogenetic signal was stronger
280 for unique zones visited ($\lambda : 0.27$) than for AP distance ($\lambda : 0.16$; Table S2).

Table 2. DIC values for statistical models with and without the inclusion of species as a fixed effect. The effect of species was evaluated in a model where variances did not (Model 1) or did (Model 4) differ by species. For both behaviors and both models, the inclusion of species substantially improved model fit, as indicated by the lower DIC values for models with species included as a fixed effect.

	Behavior	DIC with species	DIC without species	DIC(without) - DIC(with)
Model 1 ($V_i =$ & $V_w =$)	AP Distance	8025.51	8058.71	32.2
	Unique Zones Visited	8982.97	8456.17	526.8
Model 4 ($V_i \neq$ & $V_w \neq$)	AP Distance	7763.82	7780.94	17.12
	Unique Zones Visited	8338.21	8344.88	6.67

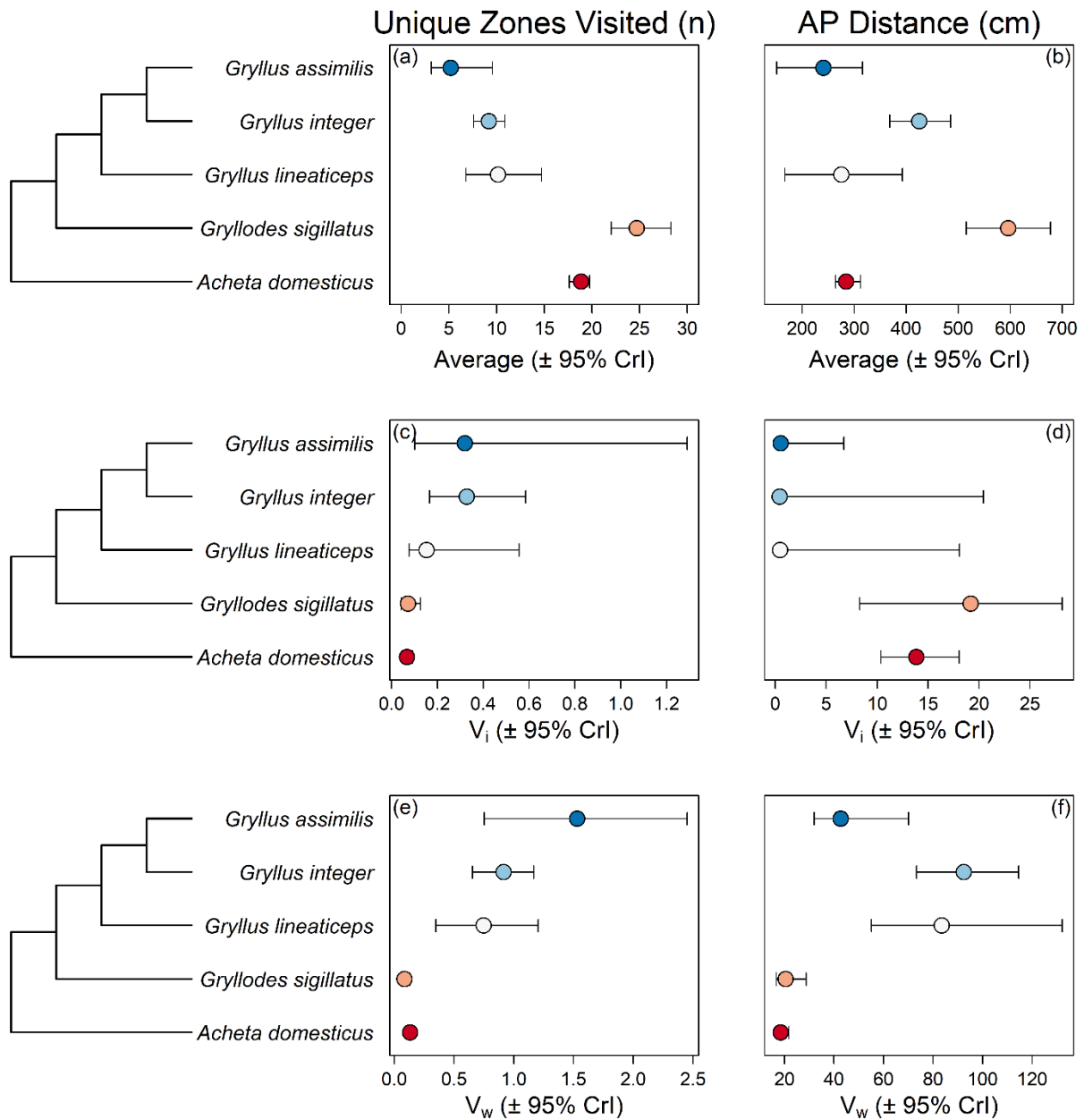


Figure 1. Species posterior-modal values with 95% HPD credibility intervals. (a) Average unique zones visited. (b) Average AP distance in centimeters. (c) Among-individual variances in unique zones traveled. (d) Among-individual variances in AP distance. (e) Within-individual variances in unique zones traveled. (f) Within-individual variances in AP distance.

281 *Differences in variances among species*

282 The best fit model for unique zones visited was Model 4, which allowed both among
 283 and within-individual variances to vary across species. All other models were poorly
 284 supported ($\Delta\text{DIC}>8$; Table 3). This indicates that both among- and within-individual
 285 variances differed among species in open field trials. For AP distance, Models 3 and 4 fit

286 comparably well (Table 3). Both of these models support differences among species in
 287 within-individual variances for AP distance. The difference between the models therefore
 288 suggests mixed support for species differences in among-individual variances for AP
 289 distance.

290 The monophyletic group of *G. assimilis*, *G. integer*, and *G. lineaticeps* exhibited higher
 291 among-individual variation for unique zones visited and lower among-individual variation
 292 for AP distance (Figure 1c & d; Table S3). This monophyletic group also exhibited higher
 293 within-individual variation for both unique zones visited and AP distance than observed for
 294 *A. domesticus* and *G. sigillatus* (Figure 1e & f; Table S3).

Table 3. DIC and Δ DIC values of model fit for AP distance and unique zones visited.

Model (variance constraints)	Behavior	DIC	Δ DIC
Model 1 ($V_i =$ & $V_w =$)	Unique Zones Visited	8982.97	644.76
Model 2 ($V_i \neq$ & $V_w =$)	Unique Zones Visited	8420.69	82.48
Model 3 ($V_i =$ & $V_w \neq$)	Unique Zones Visited	8346.44	8.23
Model 4 ($V_i \neq$ & $V_w \neq$)	Unique Zones Visited	8338.21	0
Model 1 ($V_i =$ & $V_w =$)	AP Distance	8025.51	263.31
Model 2 ($V_i \neq$ & $V_w =$)	AP Distance	8010.04	247.84
Model 3 ($V_i =$ & $V_w \neq$)	AP Distance	7762.20	0
Model 4 ($V_i \neq$ & $V_w \neq$)	AP Distance	7763.82	1.62

295 *Differences in behavioral correlations among species*

296 Among-individual behavioral correlations were of similar magnitude for *A.*
 297 *domesticus*, *G. assimilis*, *G. lineaticeps*, and *G. sigillatus* (0.3 : 0.5, Figure 2a, Table S4) while
 298 the correlation for *G. integer* was estimated to be slightly higher (0.66, Figure 2a, Table S4).
 299 Importantly, the lower bounds of the HPD intervals for *G. assimilis*, *G. integer*, *G. lineaticeps*,
 300 and *G. sigillatus* also overlapped with 0 (Figure 2, Table S3). This is perhaps unsurprising
 301 given the small sample sizes for *G. assimilis* and *G. lineaticeps*.

302 Behavioral correlations at the within-individual level ranged from 0.1 to 0.35 for *A.*
 303 *domesticus*, *G. sigillatus*, and *G. integer*, with *G. integer* having the lower bound of its HPD

304 interval overlapping with 0 (Figure 2b). The overlapping of 0 indicates that behavioral
 305 plasticity might not be correlated in this species. Behavioral correlations at either level did
 306 not show obvious patterns relative to phylogeny and were not statistically different across
 307 species (Figure 2).

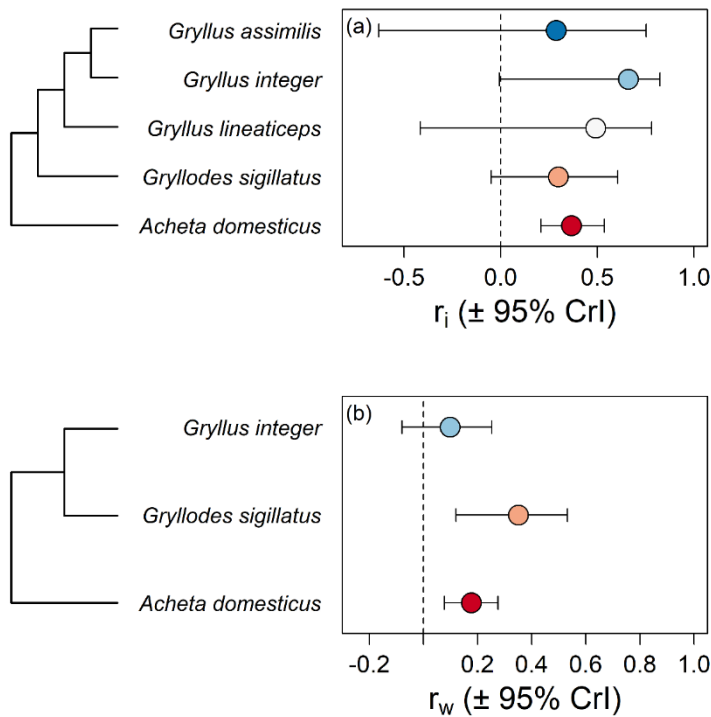


Figure 2. Species posterior-modal values with 95% HPD credibility intervals. (a) Among-individual behavioral correlations of unique zones visited and AP distance. (b) Within-individual differences of behavioral correlations of unique zones traveled and AP distance. Within-individual correlations for *G. assimilis* and *G. lineaticeps* were not calculated as behavior trials were not performed in close succession.

308

Discussion

309 Our results demonstrate that species differed in their exploratory behavior and
 310 response to cues of predator presence at all levels of variation but that behavioral variation
 311 and syndromes were conserved across species. These results suggest an important
 312 influence of phylogenetic constraints on how behaviors evolve.

313 Species differed from one another in their average behaviors (Table 2), and in a
 314 manner consistent with phylogenetic relationships. Specifically, the monophyletic group of
 315 *G. assimilis*, *G. integer*, and *G. lineaticeps* were generally similar in average unique zones

316 visited (Figure 1a). In contrast, while average AP distance differed by species, it did not do
317 so in a manner clearly concordant with phylogeny (Figure 1b). Consistent with this,
318 phylogenetic signal, the proportion of variation attributable to the hierarchical pattern of
319 relatedness among species, was higher for unique zones visited than for AP distance (Table
320 S2). Interestingly, and relevant for future research, phylogeny explained considerably more
321 variation in our measure of exploratory behavior—unique zones visited—than did among-
322 individual variation, i.e. “animal personality” ($\tau = 0.15$ versus $\lambda = 0.27$; Table S2). However,
323 given that phylogenetic signal was estimated with only five species, this finding should be
324 interpreted with caution. Further, while the patterns we detected are consistent with
325 phylogenetic constraint, it is also possible that similarity among species in behaviors stems
326 from niche similarity rather than phylogenetic constraint on the observed behaviors.

327 The species we examined also differed in among-individual variation in exploratory
328 (unique zones visited) and predator response (AP distance) behaviors, again in a manner
329 consistent with phylogenetic relationships (Figures 1c, d). Unfortunately, phylogenetic
330 methods have been developed primarily with the goal of understanding differences in trait
331 averages rather than trait (co)variances. Our comparisons of “personality” variation and
332 syndromes among species are therefore based on the model comparison methods
333 identifying the presence of species differences and subsequent qualitative comparisons of
334 species level estimates. Nonetheless, the concordance between patterns of the magnitude
335 of among-individual variation and the currently described phylogeny suggests
336 phylogenetic constraints on the magnitude of “personality” variation. Of the five total
337 species, the monophyly of *G. assimilis*, *G. integer*, and *G. lineaticeps* exhibited the highest
338 among-individual variation in unique zones visited and the lowest among-individual
339 variation in AP distance (Figure 1c and d). While the expression of average behaviors and
340 behavioral syndromes might be expected to exhibit phylogenetic signal, we did not expect
341 this to be the case for among-individual variances. One possible explanation would be
342 bottlenecks at more basal phylogenetic nodes led to reduced genetic variation present in
343 subsequent groupings. While this could explain the lower among-individual variation in AP
344 distance for *G. assimilis*, *G. integer*, and *G. lineaticeps*, it does not explain that those same
345 species exhibit higher within-individual variation for unique zones visited.

346 Differences observed in among-individual variation could also be attributable to
347 selection differentially acting upon these species by reducing the additive genetic variation
348 present in a population or species (Mousseau and Roff 1987). Our results therefore suggest
349 the possibility that exploratory behavior, for which unique zones visited is a proxy, has
350 been under stronger selection for *A. domesticus* and *G. sigillatus* than for the other species.
351 Importantly, because we do not know the strength and direction of selection acting on
352 these phenotypes, the data presented here only suggests this possibility and cannot be used
353 to distinguish between the effects of selection and drift for either behavior.

354 Alternative explanations for the observed differences in among-individual variances
355 stem from differences across source populations and sampling of these populations. For
356 example, individual *A. domesticus* used in this study were from a captive population where
357 inbreeding could have reduced genetic variation over generations. This potentially explains
358 the low among-individual variation the species shows for unique zones visited (Figure 1c)
359 but is contradicted by the high among-individual variation in AP distance (Figure 1d). In
360 contrast, *G. assimilis* and *G. lineaticeps* behavior was measured for field-caught individuals.
361 If individuals of these species experienced different developmental environments from one
362 another, we would predict higher among-individual variation in behavior because
363 permanent environmental variation contributes, on average, 50% of the observed among-
364 individual variation in behavior present in populations (Dochtermann et al. 2015). This
365 explanation is not, however, supported: while *G. assimilis* and *G. lineaticeps* indeed showed
366 high relative among-individual variation in unique zones visited, the same was not the case
367 for AP distance (Figure 1c and d). Moreover, for both behaviors, *G. assimilis* and *G.*
368 *lineaticeps* were very similar to *G. integer*, for which lab reared individuals were measured.
369 Another potential confound in our results is that species were measured at distinct times
370 (*A. domesticus* first, *G. lineaticeps*, *G. assimilis*, and *G. integer* second through fourth
371 respectively, and *G. sigillatus* last). However, there is no clear relationship between this
372 sequence of sampling and behavioral averages or variance. To summarize, the conflicting
373 patterns of among-individual variation observed between AP distance and unique zones
374 visited prevents clear interpretation.

375 Estimated within-individual variances include variation from a variety of sources,
376 including plasticity in response to short-term environmental variation and measurement

377 error (Dingemanse et al. 2012, Berdal and Dochtermann 2019). In comparing species, if we
378 assume measurement error is similar among species, differences in within-individual
379 variation will primarily represent differences in plasticity. This short-term plasticity, also
380 referred to as phenotypic flexibility (Piersma and Van Gils 2011), allows individuals to
381 respond flexibly to an environment (Westneat et al. 2015). As was the case for among-
382 individual variation and average unique zones visited, *G. assimilis*, *G. integer*, and *G.*
383 *lineaticeps* were grouped together and exhibited similar magnitudes of within-individual
384 variation (Figure 1e and f). For both behaviors, this group exhibited considerably higher
385 within-individual variation than observed for *A. domesticus* and *G. sigillatus*, differences
386 supported by our model comparison results (Table 3). In other words, the *Gryllus* genus
387 exhibited greater behavioral plasticity.

388 One possible explanation for this pattern is that our sample of *G. assimilis*, *G. integer*,
389 and *G. lineaticeps* were of individuals either caught from the field or the direct offspring of
390 field inseminated and subsequently captured individuals. In contrast, the population of *G.*
391 *sigillatus* we sampled had been in captivity for around 75 generations and the population of
392 *A. domesticus* was reared for production purposes for some undetermined but large
393 number of generations. Consequently the differences in within-individual variation could
394 be attributable to exposure to a frequently changing environment (Relyea 2001) in the case
395 of *G. assimilis*, *G. integer*, and *G. lineaticeps* and the loss of plasticity in *A. domesticus* and *G.*
396 *sigillatus*. This possibility could be assessed for crickets via experimental evolution with
397 populations experiencing different levels of environmental heterogeneity.

398 With regard to behavioral correlations, Bell (2005) proposed two hypotheses for the
399 expression of behavioral syndromes within a population relevant to the species level
400 comparisons we performed. The first of these, the constraints hypothesis, chiefly attributes
401 behavioral syndromes to the presence of pleiotropy, with the expression of genes affecting
402 multiple behaviors. This hypothesis can be extended to other mechanistic connections
403 constraining independent trait expression. Second, the adaptive hypothesis states that
404 behavioral syndromes are the adaptive outcome of correlated selection. While pleiotropy
405 and other mechanistic connections can evolve and be adaptive, syndromes attributable to
406 the adaptive hypothesis are expected to respond more quickly to changes in selection (Roff
407 1997). Consequently, phylogenetic similarity in behavioral syndromes provides indirect

408 support for the constraints hypothesis. Due to among-individual correlations not
409 substantively differing among species (Figure 2), our results are consistent with the
410 constraints hypothesis, despite species differing in variances and average expressions of
411 behaviors (Figure 1). Unfortunately, this conclusion is not strongly supported given the
412 large uncertainties around the estimates of among-individual correlations. Nonetheless, at
413 least the direction of correlations is likely consistent across species.

414 While a comparative approach has only rarely been used for examining behavioral
415 variation, three particular studies are relevant to the interpretation of our results here.
416 First, Blankers et al. (2017) compared the phenotypic variances and (co)variances of seven
417 calling traits of multiple cricket species (including *G. lineaticeps*, which was included in our
418 study). These authors found that the phenotypic covariance matrices differed among
419 cricket species. One of the major differences among species was in the magnitude of
420 variation present in single traits (Blankers et al. 2017). This is consistent with our findings
421 that variances of behaviors differed across species (Figure 1c-f). Unfortunately, these
422 authors compared phenotypic (co)variances, which conflate among- and within-individual
423 (co)variation (Dingemanse et al. 2012). Second, compared the among-individual
424 covariance matrices of seven species of fish. Comparable to our results, these authors
425 detected differences in the magnitude of among-individual behavioral variability and also
426 found overall phylogenetic signal and similarity in how variation was expressed across
427 multiple behaviors (White et al. 2020). Finally, compared the expression of additive
428 genetic (co)variance (i.e. **G** matrices) in behavior among four populations of *G. integer*.
429 Similar to White et al. (2020) and our results presented here, found differences in single
430 trait variances and covariances but the overall structure of trait covariance was generally
431 conserved across populations—indicating support for the constraints hypothesis.

432 An additional area of research with which our results can be compared is
433 comparative psychology. While the operational definitions of animal personality and
434 behavioral syndrome used by animal behaviorists have greatly diverged from the definition
435 of personality used in comparative psychology, both areas examine patterns of behavioral
436 (co)variation. Many of the same challenges in attempting comparative work appear in both
437 fields. In particular, naming fallacies hinder broad descriptions (Gosling 2001, Carter et al.
438 2013). Nonetheless, comparative psychology has found that similar patterns of trait

439 correlations are observed in many species. Specifically, behavioral axes characterizing
440 responses to novelty are common across taxa (Gosling 2001). The frequency with which
441 this reactivity (Koolhaas et al. 1997, Koolhaas et al. 1999) or shy-bold (Sih et al. 2004a, Sih
442 et al. 2004b, Smith and Blumstein 2008) axis is observed across taxa is consistent with our
443 finding of behavioral (co)variation being evolutionarily conserved.

444 More generally, our findings here suggest that behavioral correlations are
445 phylogenetically conserved. Conserved trait correlations like those observed here
446 constrain the divergence of populations and species (Schluter 1996). While the potential
447 for such constraints has been speculated about for behaviors (Dochtermann and
448 Dingemanse 2013), prior demonstrations of such have primarily focused on morphological
449 traits (McGlothlin et al. 2018, Sztepanacz and Houle 2019) and chemical traits (Blows et al.
450 2004, Aguirre et al. 2014).

451 Jointly, our approach allowed us to determine whether there were differences in
452 average behavior, “personality”, behavioral plasticity, and behavioral syndromes among
453 species. Our results demonstrate phylogenetic conservation of behavioral averages,
454 behavioral variation, and behavioral syndromes. This finding is potentially surprising given
455 that behavior is often assumed to be more flexible and labile than other types of traits (but
456 see Zuk and Spencer 2020) and suggests an important role for phylogenetic constraints as
457 an alternative to the dominant adaptive explanations commonly employed when discussing
458 animal personality and behavioral syndromes.

459 **Literature Cited**

- 460
461 2020. Guidelines for the treatment of animals in behavioural research and teaching. *Animal*
462 *Behaviour* **159**:I-XI.
- 463 Aguirre, J., E. Hine, K. McGuigan, and M. Blows. 2014. Comparing **G**: multivariate analysis of
464 genetic variation in multiple populations. *Heredity* **112**:21-29.
- 465 Armbruster, W., and K. Schwaegerle. 1996. Causes of covariation of phenotypic traits
466 among populations. *Journal of Evolutionary Biology* **9**:261-276.
- 467 Austin, P. C., and J. E. Hux. 2002. A brief note on overlapping confidence intervals. *Journal of*
468 *vascular surgery* **36**:194-195.

- 469 Barnett, A. G., N. Koper, A. J. Dobson, F. Schmiegelow, and M. Manseau. 2010. Using
470 information criteria to select the correct variance–covariance structure for
471 longitudinal data in ecology. *Methods in Ecology and Evolution* **1**:15-24.
- 472 Bell, A. M. 2005. Behavioural differences between individuals and two populations of
473 stickleback (*Gasterosteus aculeatus*). *Journal of Evolutionary Biology* **18**:464-473.
- 474 Bell, A. M., S. J. Hankson, and K. L. Laskowski. 2009. The repeatability of behaviour: a meta-
475 analysis. *Animal Behaviour* **77**:771-783.
- 476 Berdal, M. A., and N. A. Dochtermann. 2019. Adaptive alignment of plasticity with genetic
477 variation and selection. *Journal of Heredity* **110**.
- 478 Blankers, T., D. A. Gray, and R. M. Hennig. 2017. Multivariate phenotypic evolution:
479 Divergent acoustic signals and sexual selection in *Gryllus* field crickets. *Evolutionary*
480 *Biology* **44**:43-55.
- 481 Blows, M. W., S. F. Chenoweth, and E. Hine. 2004. Orientation of the genetic variance-
482 covariance matrix and the fitness surface for multiple male sexually selected traits.
483 *American Naturalist* **163**:329-340.
- 484 Boake, C. R. B. 1989. Repeatability - Its Role in Evolutionary Studies of Mating-Behavior.
485 *Evolutionary Ecology* **3**:173-182.
- 486 Brommer, J. E., and B. Class. 2017. Phenotypic correlations capture between-individual
487 correlations underlying behavioral syndromes. *Behavioral Ecology and Sociobiology*
488 **71**:50.
- 489 Bucklaew, A., and N. Dochtermann. 2020. The effects of exposure to predators on
490 personality and plasticity. *Ethology*.
- 491 Carter, A. J., W. E. Feeney, H. H. Marshall, G. Cowlshaw, and R. Heinsohn. 2013. Animal
492 personality: what are behavioural ecologists measuring? *Biological Reviews* **88**:465-
493 475.
- 494 Congdon, P. 2006. Bayesian model choice based on Monte Carlo estimates of posterior
495 model probabilities. *Computational statistics & data analysis* **50**:346-357.
- 496 Dingemanse, N. J., and N. A. Dochtermann. 2013. Quantifying individual variation in
497 behaviour: mixed-effect modelling approaches. *Journal of Animal Ecology* **82**:39-54.
- 498 Dingemanse, N. J., and N. A. Dochtermann. 2014. Individual behaviour: behavioural ecology
499 meets quantitative genetics. *in* A. Charmantier, D. Garant, and L. E. B. Kruuk, editors.
500 *Quantitative genetics in the wild*. Oxford University Press.

- 501 Dingemanse, N. J., N. A. Dochtermann, and S. Nakagawa. 2012. Defining behavioural
502 syndromes and the role of "syndrome" deviation in understanding their evolution.
503 Behavioral Ecology and Sociobiology **66**:1543-1548.
- 504 Dochtermann, N. A. 2011. Testing Cheverud's conjecture for behavioral correlations and
505 behavioral syndromes. Evolution **65**:1814-1820.
- 506 Dochtermann, N. A., and N. J. Dingemanse. 2013. Behavioral syndromes as evolutionary
507 constraints. Behavioral Ecology **24**:806-811.
- 508 Dochtermann, N. A., and D. A. Roff. 2010. Applying a quantitative genetics framework to
509 behavioural syndrome research. Philosophical Transactions of the Royal Society B-
510 Biological Sciences **365**:4013-4020.
- 511 Dochtermann, N. A., T. Schwab, M. A. Berdal, J. Dalos, and R. Royaute. 2019. The heritability
512 of behaviour: a meta-analysis. Journal of Heredity **in press**.
- 513 Dochtermann, N. A., T. Schwab, and A. Sih. 2015. The contribution of additive genetic
514 variation to personality variation: heritability of personality. Proceedings Of The
515 Royal Society B-Biological Sciences **282**:20142201.
- 516 Garamszegi, L. Z., G. Marko, and G. Herczeg. 2012. A meta-analysis of correlated behaviours
517 with implications for behavioural syndromes: mean effect size, publication bias,
518 phylogenetic effects and the role of mediator variables. Evolutionary Ecology
519 **26**:1213-1235.
- 520 Garamszegi, L. Z., G. Marko, and G. Herczeg. 2013. A meta-analysis of correlated behaviors
521 with implications for behavioral syndromes: relationships between particular
522 behavioral traits. Behavioral Ecology **24**:1068-1080.
- 523 Gosling, S. D. 2001. From mice to men: What can we learn about personality from animal
524 research? Psychological Bulletin **127**:45-86.
- 525 Gould, S. J., and R. C. Lewontin. 1979. Spandrels Of San-Marco And The Panglossian
526 Paradigm - A Critique Of The Adaptationist Program. Proceedings of the Royal
527 Society of London Series B-Biological Sciences **205**:581-598.
- 528 Gray, D. A., D. B. Weissman, J. A. Cole, and E. M. Lemmon. 2020. Multilocus phylogeny of
529 Gryllus field crickets (Orthoptera: Gryllidae: Gryllinae) utilizing anchored hybrid
530 enrichment. Zootaxa **4750**:zootaxa. 4750.4753. 4752-zootaxa. 4750.4753. 4752.
- 531 Hadfield, J., and S. Nakagawa. 2010. General quantitative genetic methods for comparative
532 biology: phylogenies, taxonomies and multi-trait models for continuous and
533 categorical characters. Journal of Evolutionary Biology **23**:494-508.
- 534 Hadfield, J. D. 2010. MCMC methods for multi-response generalized linear mixed models:
535 the MCMCglmm R package. Journal of Statistical Software **33**:1-22.

- 536 Hine, E., K. McGuigan, and M. W. Blows. 2014. Evolutionary Constraints in High-
537 Dimensional Trait Sets. *The American Naturalist* **184**:119-131.
- 538 Koolhaas, J. M., S. F. deBoer, and B. Bohus. 1997. Motivational systems or motivational
539 states: Behavioural and physiological evidence. *Applied Animal Behaviour Science*
540 **53**:131-143.
- 541 Koolhaas, J. M., S. M. Korte, S. F. De Boer, B. J. Van Der Vegt, C. G. Van Reenen, H. Hopster, I.
542 C. De Jong, M. A. W. Ruis, and H. J. Blokhuis. 1999. Coping styles in animals: current
543 status in behavior and stress-physiology. *Neuroscience and Biobehavioral Reviews*
544 **23**:925-935.
- 545 MacGregor-Fors, I., and M. E. Payton. 2013. Contrasting diversity values: statistical
546 inferences based on overlapping confidence intervals. *Plos One* **8**:e56794.
- 547 McGlothlin, J. W., M. E. Kobiela, H. V. Wright, D. L. Mahler, J. J. Kolbe, J. B. Losos, and E. D.
548 Brodie III. 2018. Adaptive radiation along a deeply conserved genetic line of least
549 resistance in *Anolis* lizards. *Evolution Letters* **2**:310-322.
- 550 Mousseau, T. A., and D. A. Roff. 1987. Natural selection and the heritability of fitness
551 components. *Heredity* **59**:181-197.
- 552 Nakagawa, S., and E. S. Santos. 2012. Methodological issues and advances in biological
553 meta-analysis. *Evolutionary Ecology* **26**:1253-1274.
- 554 Nakagawa, S., and H. Schielzeth. 2010a. Repeatability for Gaussian and non-Gaussian data: a
555 practical guide for biologists. *Biological Reviews* **85**:935-956.
- 556 Nakagawa, S., and H. Schielzeth. 2010b. Repeatability for Gaussian and non-Gaussian data:
557 a practical guide for biologists. *Biological Reviews* **85**:935-956.
- 558 Niemelä, P. T., and N. J. Dingemanse. 2018. Meta-analysis reveals weak associations
559 between intrinsic state and personality. Page 20172823 *in* *Proc. R. Soc. B. The Royal*
560 *Society*.
- 561 Pagel, M. 1999. Inferring the historical patterns of biological evolution. *Nature* **401**:877-
562 884.
- 563 Phillips, P. C., and S. J. Arnold. 1989. Visualizing multivariate selection. *Evolution* **43**:1209-
564 1222.
- 565 Piersma, T., and J. Drent. 2003. Phenotypic flexibility and the evolution of organismal
566 design. *Trends in Ecology & Evolution* **18**:228-233.
- 567 Piersma, T., and J. A. Van Gils. 2011. *The flexible phenotype: a body-centred integration of*
568 *ecology, physiology, and behaviour*. Oxford University Press.
- 569 Quiggin, J. 2012. *Zombie economics*. Princeton University Press.

- 570 Relyea, R. A. 2001. Morphological and behavioral plasticity of larval anurans in response to
571 different predators. *Ecology* **82**:523-540.
- 572 Roff, D. A. 1997. *Evolutionary Quantitative Genetics*. Chapman and Hall, New York.
- 573 Royauté, R., M. A. Berdal, C. R. Garrison, and N. A. Dochtermann. 2018. PACELESS life? A meta-
574 analysis of the pace-of-life syndrome hypothesis. *Behavioral Ecology and*
575 *Sociobiology* **72**:64.
- 576 Royauté, R., and N. A. Dochtermann. 2017. When the mean no longer matters:
577 developmental diet affects behavioral variation but not population averages in the
578 house cricket (*Acheta domesticus*). *Behavioral Ecology* **28**:337-345.
- 579 Royauté, R., and N. A. Dochtermann. 2021. Comparing ecological and evolutionary
580 variability within datasets. *Behavioral Ecology and Sociobiology*.
- 581 Royauté, R., C. Garrison, J. Dalos, M. A. Berdal, and N. A. Dochtermann. 2019. Current energy
582 state interacts with the developmental environment to influence behavioural
583 plasticity. *Animal behaviour* **148**:39-51.
- 584 Royauté, R., K. Greenlee, M. Baldwin, and N. A. Dochtermann. 2015. Behaviour, metabolism
585 and size: phenotypic modularity or integration in *Acheta domesticus*? *Animal*
586 *Behaviour* **110**:163-169.
- 587 Royauté, R., A. Hedrick, and N. A. Dochtermann. 2020. Behavioural syndromes shape
588 evolutionary trajectories via conserved genetic architecture. *Proceedings of the*
589 *Royal Society B* **287**:20200183.
- 590 Schluter, D. 1996. Adaptive radiation along genetic lines of least resistance. *Evolution*
591 **50**:1766-1774.
- 592 Sih, A., A. Bell, and J. C. Johnson. 2004a. Behavioral syndromes: an ecological and
593 evolutionary overview. *Trends in Ecology & Evolution* **19**:372-378.
- 594 Sih, A., A. M. Bell, J. C. Johnson, and R. E. Ziemba. 2004b. Behavioral syndromes: An
595 integrative overview. *Quarterly Review of Biology* **79**:241-277.
- 596 Smith, B. R., and D. T. Blumstein. 2008. Fitness consequences of personality: a meta-
597 analysis. *Behavioral Ecology* **19**:448-455.
- 598 Spiegelhalter, D., A. Thomas, N. Best, and D. Lunn. 2003. WinBUGS user manual. version.
- 599 Stirling, D. G., D. Reale, and D. A. Roff. 2002. Selection, structure and the heritability of
600 behaviour. *Journal of Evolutionary Biology* **15**:277-289.
- 601 Sztepanacz, J. L., and D. Houle. 2019. Cross-sex genetic covariances limit the evolvability of
602 wing-shape within and among species of *Drosophila*. *Evolution* **73**:1617-1633.

- 603 Team, R. C. 2018. R: A language and environment for statistical computing.
- 604 Van de Pol, M., and J. Wright. 2009. A simple method for distinguishing within-versus
605 between-subject effects using mixed models. *Animal behaviour* **77**:753.
- 606 Westneat, D. F., M. I. Hatch, D. P. Wetzel, and A. L. Ensminger. 2011. Individual variation in
607 parental care reaction norms: integration of personality and plasticity. *The*
608 *American Naturalist* **178**:652-667.
- 609 Westneat, D. F., J. Wright, and N. J. Dingemanse. 2015. The biology hidden inside residual
610 within-individual phenotypic variation. *Biological Reviews* **90**:729-743.
- 611 White, S. J., D. J. Pascall, and A. J. Wilson. 2020. Towards a comparative approach to the
612 structure of animal personality variation. *Behavioral Ecology* **31**:340-351.
- 613 Whitman, D., and A. Agrawal. 2009. What is phenotypic plasticity and why is it important?
614 Phenotypic plasticity of insects: mechanisms and consequences:1-63.
- 615 Yang, J., H. Dong, M. He, and J. Gao. 2021. Mitochondrial genome characterization of
616 *Grylloides sigillatus* (Orthoptera: Gryllidae) and its phylogenetic implications.
617 *Mitochondrial DNA Part B* **6**:1056-1058.
- 618 Zuk, M., and H. G. Spencer. 2020. Killing the Behavioral Zombie: Genes, Evolution, and Why
619 Behavior Isn't Special. *Bioscience* **70**:515-520.
- 620

Supplemental Materials

Table S1. Fixed effects coefficients for Model 4 (Table 3). The intercept estimate is for *Acheta domesticus* females (fixed effect coefficients are contrasts versus these values).

AP Distance (square-root transformed)					
	posterior mean	95% credibility interval		effective sample size	pMCMC
		lower	upper		
Intercept	16.97	16.25	17.66	1000	<0.001
<i>Gryllus assimilis</i>	-1.44	-4.18	1.22	1000	0.298
<i>Gryllus integer</i>	3.67	2.31	5.18	1000	<0.001
<i>Gryllus lineaticeps</i>	-0.52	-4.00	2.95	1000	0.782
<i>Grylloides sigillatus</i>	7.43	5.69	9.17	1000	<0.001
Temp2	0.65	0.34	0.96	1000	<0.001
SexM	-0.40	-1.44	0.72	1000	0.454
Mass (w/in individual centered)	0.72	-0.77	2.29	1000	0.344
Mass (b/w individual centered)	0.02	-0.76	0.98	800.8	0.972
Unique zones visited					
	posterior mean	95% credibility interval		effective sample size	pMCMC
		lower	upper		
Intercept	2.929	2.869	2.984	1000	<0.001
<i>Gryllus assimilis</i>	-1.227	-1.757	-0.628	1000	<0.001
<i>Gryllus integer</i>	-0.728	-0.891	-0.526	1000	<0.001
<i>Gryllus lineaticeps</i>	-0.593	-0.979	-0.219	1000	0.004
<i>Grylloides sigillatus</i>	0.282	0.160	0.411	1000	<0.001
Temp2	0.043	0.016	0.070	815.1	0.002
SexM	0.111	0.010	0.201	1000	0.022
Mass (w/in individual centered)	-0.022	-0.175	0.135	1000	0.792
Mass (b/w individual centered)	0.120	0.043	0.212	899.8	0.006

Table S2. Variance estimates (posterior modes with 95% credibility intervals) for models including phylogenetic structure as a random effect. Models were fit with temperature (centered), sex, and mass (within and between individual centered) as fixed effects. Phylogeny was modeled according to the trees shown in Figure 1 and with uniform branch lengths. Subject was also included as a random effect. Variance ratios are presented as unadjusted ratios; that is, variance due to fixed effects is included in the denominator. Ratios for unique zones include the distribution specific variance (DSV) in the denominator. λ and τ correspond to unadjusted phylogenetic signal and unadjusted repeatabilities respectively.

	Variance estimate (95% CrI)	Variance ratios* (95% CrI)
AP Distance		
Phylogeny	12.73 (2.10 : 106.61)	λ : 0.16 (0.05 : 0.68)
Subject	13.34 (10.21 : 17.66)	τ : 0.19 (0.07 : 0.27)
Fixed Effects	1.10 (0.32 : 2.60)	0.01 (0 : 0.04)
Residual	38.34 (33.96 : 41.50)	0.50 (0.24 : 0.71)
Unique Zones		
Visited		
Phylogeny	0.12 (0.03 : 1.07)	λ : 0.27 (0.09 : 0.72)
Subject	0.11 (0.07 : 0.13)	τ : 0.15 (0.05 : 0.23)
Fixed Effects	0.01 (0 : 0.02)	0.01 (0 : 0.03)
Residual	0.26 (0.23 : 0.31)	0.48 (0.16 : 0.58)
DSV**	0.06 (0.03 : 0.11)	NA

* while the ratios for any single MCMC estimate will sum to 1, the posterior modes can sum to other values due to uncertainty across the MCMC chain

** estimated as $\ln\left(\frac{1}{\exp(\beta_0)} + 1\right)$ following Nakagawa & Schielzeth (2010)

622

623

Table S3. Species level estimates of among-individual variance, within-individual variances, and repeatability for each behavior. Repeatabilities (τ) are presented as adjusted ratios; that is, variance due to fixed effects is not included in the denominator. Repeatabilities for unique zones include the distribution specific variance (DSV) in the denominator*.

	Among-individual Variance (95% CrI)	Within-individual Variance (95% CrI)	Repeatability (95% CrI)
AP Distance			
<i>Acheta domesticus</i>	13.85 (10.38 : 18.04)	18.52 (16.66 : 21.78)	0.418 (0.35 : 0.51)
<i>Gryllus assimilis</i>	0.54 (0.09 : 6.73)	42.72 (32.98 : 70.03)	0.007 (0 : 0.12)
<i>Gryllus integer</i>	0.43 (0.10 : 20.43)	92.45 (73.33 : 114.54)	0.004 (0 : 0.20)
<i>Gryllus lineaticeps</i>	0.47 (0.12 : 18.09)	83.57 (55.14 : 132.18)	0.004 (0 : 0.18)
<i>Gryllodes sigillatus</i>	19.18 (8.30 : 28.17)	20.51 (16.78 : 28.74)	0.441 (0.26 : 0.60)
Unique Zones Visited			
<i>Acheta domesticus</i>	0.07 (0.05 : 0.09)	0.133 (0.11 : 0.16)	0.254 (0.20 : 0.35)
<i>Gryllus assimilis</i>	0.32 (0.10 : 1.29)	1.53 (0.75 : 2.45)	0.190 (0.06 : 0.51)
<i>Gryllus integer</i>	0.33 (0.17 : 0.59)	0.92 (0.65 : 0.17)	0.271 (0.15 : 0.41)
<i>Gryllus lineaticeps</i>	0.15 (0.08 : 0.56)	0.75 (0.35 : 1.21)	0.200 (0.08 : 0.46)
<i>Gryllodes sigillatus</i>	0.07 (0.05 : 0.13)	0.08 (0.06 : 0.13)	0.355 (0.23 : 0.50)

* estimated as $\ln\left(\frac{1}{\exp(\beta_0)} + 1\right)$ following Nakagawa & Schielzeth (2010)

Table S4. Among- and within-individual correlations by species. Correlation estimates are posterior modes and are presented along with 95% credibility intervals (CrI).

Species	Among-individual correlation (95% CrI)	Within-individual correlation (95% CrI)
<i>Gryllus assimilis</i>	0.37 (0.21 : 0.54)	NA
<i>Gryllus integer</i>	0.29 (-0.63 : 0.75)	0.10 (-0.08 : 0.25)
<i>Gryllus lineaticeps</i>	0.66 (-0.01 : 0.82)	NA
<i>Gryllodes sigillatus</i>	0.49 (-0.42 : 0.78)	0.35 (0.12 : 0.53)
<i>Acheta domesticus</i>	0.3 (-0.05 : 0.6)	0.18 (0.08 : 0.27)

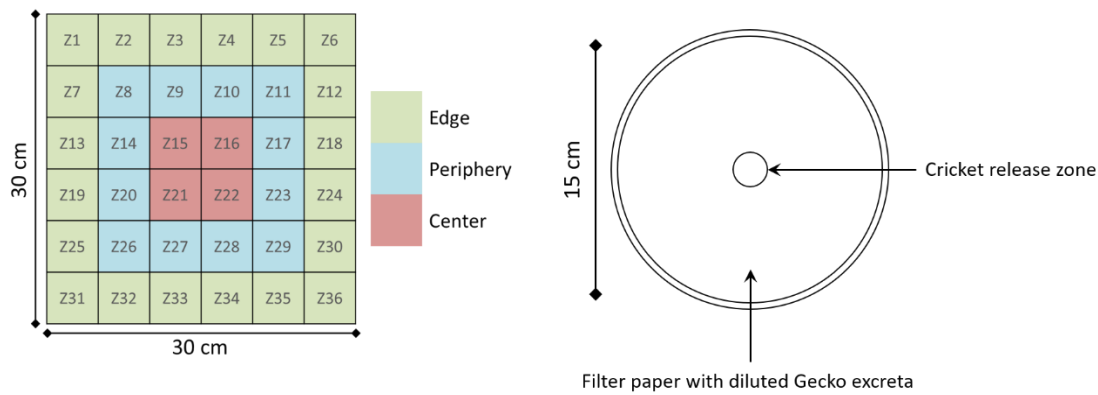


Figure S1. Schematics of the open field (left) and predator cue arenas (right). The open field arenas were subdivided into 36 unique “zones” during video processing. For the anti-predator response trials the cricket was introduced, under a container, to the center point. This cricket release zone did not have predator cues present.