

Invasive goby larvae: First evidence as stowaways in small watercraft motors.

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Abstract

Aquatic invasive species (AIS) are a major threat to freshwater and marine ecosystems worldwide. Despite management efforts, human assisted dispersal continues to distribute AIS within and across waterbodies. An understudied translocation vector for AIS, especially for invasive fish, are the cooling systems of small watercraft motors. Here, we investigate the contents of boat motor cooling systems for the presence of invasive goby larvae in a collaboration with local boat owners. Because of the exclusively nocturnal drift activity of goby larvae, we drove boats in the first hours after sunset. For an estimate of the translocation potential, we quantified drift density of goby larvae as well as boat traffic after sunset. We found a goby larva in a boat motor once in 30 drives of 1-2 hours duration each. Peak drift densities of goby larvae were 2.5 per 100 m³, which is comparable to previously reported data. Recreational boats were active after sunset throughout the reproductive season of invasive gobies and are therefore a realistic translocation vector for goby larvae. Additionally, evidence of fish and other animals inside boat motor cooling systems, gathered from online boating forums, speaks for the potential of AIS transport in small watercraft. Translocation inside motors is especially likely for in-water transport of boats, which should be a management focus in interconnected aquatic systems.

Keywords:

anthropogenic transport, biosecurity, invasive fish, *Neogobius melanostomus*, non-indigenous species, translocation, vector

Introduction

The role of recreational boats in secondary translocation of aquatic invasive species (AIS) has been well established for plants, invertebrates or microbes (Ashton et al. 2014; Murray et al. 2011). Most studies focus on biofouling organisms as potential hitchhikers on the outside of recreational boats (Rothlisberger et al. 2010; Ulman et al. 2019). Residual waters on recreational boats are of concern as well, because of their vector potential for mobile species – for example, Campbell et al. (2016) and Darbyson et al. (2009) confirmed the presence of AIS in bilge waters. However, to fully estimate and manage the risk of translocation by small watercraft, investigating motor cooling water systems in addition to bilge water or other standing water compartments is important. Bilge water originates from passive processes (water swashing on the deck and drain into the bilge),

45 while cooling water is actively sucked into the motor from the water column. The potential for
46 uptake and survival of propagules should therefore differ between the two types of residual waters.
47 Problematically, the insides of boat motors are often hardly accessible and therefore poorly
48 characterized. While some studies indeed investigated the residual cooling water contents
49 (Johnson et al. 2001, Minchin et al. 2006, Montz and Hirsch 2016), all of them took their samples
50 when the boats were out of the water and a lot of the residual water had already drained. This
51 sampling strategy might be realistic if one only focuses on overland-transport, but it neglects
52 potential translocation within a connected system of waterbodies, or in a marine system.

53
54 Transport of small watercraft within systems of interconnected waterbodies is an important
55 secondary transport mechanism, as it can help AIS to overcome migration barriers like dams or
56 waterfalls via locks or boat lifts (Rahel 2007). Organisms inside of dead volumes of boat motors
57 might have increased chances of survival than those in standing waters, because e.g. aeration and
58 temperature could be more favourable in cooling systems while the motor is still submerged, than
59 they are in small volumes of residual waters during overland transport (Havel and Stelzleni-
60 Schwent 2000; Johnson et al. 2001).

61
62 Sampling cooling systems while the boat is inside of the water can be challenging, because most
63 openings for draining water are below the water surface or require tools to reach. For example,
64 Kelly et al. (2012) investigated AIS presence in standing waters of boats that were still in the water,
65 but did or could not access cooling systems. In a transdisciplinary project about effective boat
66 cleaning practices with local environmental authorities and boat mechanics, we discovered
67 numerous living organisms inside detached lower units of boat motors, which did not drain with
68 the residual water (AUE 2019). Furthermore, boat mechanics reported that they have encountered
69 fish inside boat motors on occasion. The reports of fish in residual water in boats are relevant,
70 because they indicate that the water-intake of near-surface vessels might suffice for pick-up,
71 transport and viable release of propagules within connected water systems – as it has been shown
72 for ballast water of transoceanic ships (Wonham et al. 2000). For this sequence of events to occur
73 there must be a spatial and temporal overlap with vector activity and propagule.

74
75 One of the most prominent aquatic invasive fish in Europe and North America, the round goby
76 *Neogobius melanostomus* (Pallas, 1814), keeps spreading outside of navigable waters
77 (Bronnenhuber et al. 2011). Active migration within connected water systems is increasingly
78 possible for fish due to efforts to make hydroelectric dams passable (Silva et al. 2018) – however,
79 for bottom-dwelling fish like the round goby, the high flow rates within fish stairs might still hinder
80 or slow down upstream migration (Wiegler et al. 2020). Secondary transport mechanisms are
81 therefore likely to promote their dispersal (Bronnenhuber et al. 2011). Round gobies have pelagic
82 larval and juvenile stages (diel vertical migration or larval drift), which are present in the water
83 column throughout the reproductive season of the gobies from dusk until dawn (Borcherding et al.
84 2016; Ramler et al. 2016). The drifting life stages are considered propagules for translocation via
85 ballast water of commercial ships (Hensler and Jude 2007). The typical time of larval drift is
86 between April and August and overlaps with the peak boating season in Europe (Hirsch et al.
87 2016). The possibility of goby larvae transport in residual waters of recreational boats, however,
88 is so far unexplored, and it is unclear how much recreational boat traffic happens during the high-
89 risk hours after sunset.

90

91 Here, we investigate the role of motor cooling water of recreational boats in assisted dispersal of
92 invasive ponto-caspian gobies. We examine boat motor contents after drives after sunset in the
93 river Rhine. Additionally, we determine drift densities of goby larvae at the water surface to
94 estimate the chances for an uptake into boat cooling systems. Importantly, we sampled boat motors
95 in close collaboration with local boat owners, resulting in a mutually informative exchange of
96 knowledge and experiences about invasive species translocation that furthered our understanding
97 of boater behaviour.

98

99 **Methods**

100 *Study location*

101 Our study took place at four locations along the High Rhine between Ryburg-Moehlin (km 144)
102 and Basel (km 170, Figure 1). Round goby populations are established at all locations investigated,
103 bighead gobies *Ponticola kessleri* (Günther, 1861) are only documented in Basel.

104

105 *Contact to boat owners*

106 We sent an inquiry for collaboration to 33 contacts from a stakeholder network that our working
107 group has established in the context of the ponto-caspian goby invasion in the High Rhine area
108 since 2012. The network is based on outreach and collaborative projects, and includes boat owners,
109 fishermen, government employees, and other people professionally or privately connected to the
110 Rhine across Switzerland, Germany and France. The inquiry contained the question if they a) had
111 a suitable boat available, and b) would be willing to either lend us the boat, or drive with us
112 (complete inquiry presented in Document S1). We defined a suitable boat as a boat with either an
113 outboard or an inboard motor/stern drive with a minimum power of 50-75 HP, as those motors
114 have large enough water inlets to take up particles in the size range of drifting goby larvae (6 – 10
115 mm, Borcherdig et al. 2016; Ramler et al. 2016), as well as sizeable lower unit volumes.

116

117 After evaluating all replies to our inquiry, we ended up collaborating with five boat owners at four
118 locations who offered to drive with us, or lend us a boat (specifications in Table S1). We conducted
119 14 drives in Basel, five drives in Kaiseraugst, three in Möhlin, and eight in Rheinfelden (Figure 1,
120 Table S1).

121

122 *Sampling of boat motor cooling systems and control sampling*

123 In total, we conducted 30 drives after sunset over the reproductive season of round gobies (June-
124 August 2020), adding up to 43 hours with running engine and covered a total distance of 202 km
125 (Table S1). After clearing the initial contents of the boat motor residual waters or raw water
126 strainers, we drove the boat for 1-2 h on the river with speeds < 10 km/h above water, so that we
127 drove half the way each up- and downstream, while keeping the boat close (< 10 m) to shore.
128 According to the collaborating boat owners, this drive profile is realistic for boaters enjoying
129 sunset-drives, or anglers on nocturnal fishing-trips.

130

131 After each drive, we emptied the cooling water system of the motor again and preserved the
132 contents in 100% ethanol. For outboard motors, the boat owners lifted the motor slowly, while we
133 caught the contents exiting the water inlets using a landing net (mesh size 500 µm, Figure 2 B).
134 For boats with a stern drive or an inboard motor and a raw water strainer, we opened the strainer
135 before the drive and collected the content (Figure 2 C). We installed a second layer of filter by
136 wrapping a stocking around the raw water strainer (mesh size 75 µm, Figure 2 D), as smaller goby

137 larvae might be flushed through the raw water strainer (mesh size 1 mm). The typical size of a
138 round goby larva during their drifting stages is 6 – 10 mm (Borcherding et al. 2016; Ramler et al.
139 2016), the gape width in that size range is 0.5 – 1 mm (Olson and Janssen 2017). After each drive,
140 we collected the contents of the raw water strainer, and then removed the stocking and emptied it
141 in a sampling vial filled with 100 % ethanol.

142
143 Additionally, we quantified the drift density of goby larva and native fish larvae in the upper water
144 layers during the boat drives, assuming that the likelihood of a boat motor to take up goby larva is
145 dependent on their occurrence per volume of water. To catch goby larvae drifting in the upper
146 water layers, we used a plankton net (manta trawl, HYDRO-BIOS Apparate Bau GmbH,
147 Altenholz, Germany), which was towed along the water surface 15 m behind the boat (Figure S1
148 A, B). The manta trawl consisted of a metal frame with a mouth opening of 30 x 15 cm, two lifting
149 bodies attached on both sides of the frame to keep the trawl at the surface, a net length of 2 m, a
150 mesh size of 300 μm , and a removable soft net bucket to empty the contents (Figure S1 A, B). A
151 flow meter was attached on the inside of the metal frame to collect data on the amount of water
152 filtered during each drive. After each drive, we removed the net bucket at the lower end of the
153 plankton net and preserved the contents in sample containers (100 ml), filled 100% ethanol.

154
155 Furthermore, we wanted to find out if goby larvae are also present in the shallow waters right next
156 to shore (< 5 m). This information is important, because the shoreline is often the part of the river
157 in which boats stay for longer times to load/unload the boat, or warm up the engine. We sampled
158 shallow waters using a so-called “fishing-noose”, a traditional local fishing device equipped with
159 a fishing net, which is lowered into the water parallel to the riverbed (Figure S1 C, D). We installed
160 a plankton net with a mesh size of 650 μm and dimensions of 3.5 x 3.5 m on the noose. We lowered
161 the net into the water at sunset until it just touched the river bottom (depth 0.5 – 1.5 m) and pulled
162 it to the shore after half an hour to search for fish and fish larvae. We euthanized all larvae and
163 stored them in 100% ethanol. We repeated this procedure four times every night starting at sunset
164 with an interval of 30 minutes each. In total, we used the fishing noose on 19 nights across the
165 sampling season. Because the noose was located in Basel, we used the noose primarily during the
166 same nights in which boat drives in Basel took place. This ensured that we had trawl net and motor
167 content data of the same location on the same dates to compare the results of the noose fishing to.

168

169 *Quantification of boat traffic after sunset*

170 To evaluate whether the uptake of goby larvae into boat motors after sunset was a realistic option,
171 we collected data on boat traffic at the same time as when our sampling took place. Starting on
172 June 24th, we recorded the presence of other boats active on the sampled stretch of the Rhine during
173 the time of sampling whenever possible. On some dates, we were not able to count boats because
174 of rainy weather conditions and therefore poor visibility.

175

176 *Analysis of samples*

177 In the lab, we searched all collected samples for fish larvae. We looked at every fish larva under a
178 dissecting microscope, separated goby larvae and larvae of native fish, and counted them. Gobies
179 were identified by the presence of a fused ventral fin, a trait that no native species exhibits.

180

181 We calculated the drift density per 100 m³ of goby larvae and native fish caught in the manta trawl
182 using the formula $DD = n \cdot 100 / r \cdot 0.3 \cdot A$, where DD = drift density, n = number of larvae caught,
183 r = number of revolutions of the flowmeter, A = area of net opening (constant: $0.3 \cdot 0.1 = 0.03$).
184 We counted all goby larvae caught close to shore with the fishing noose and compared the numbers
185 to the DD measured on the same dates with the manta trawl. We used the DD to calculate the
186 expected number of gobies in the volume of water sampled by the fishing noose in one pull (ca. 9
187 m³) and compared it to the actually caught number of goby larvae in the noose averaged per pull.
188

189 *Collection of anecdotal evidence for organisms in cooling systems*

190 To further support our finding of fish in boat motor cooling systems and to increase the
191 geographical scope with potentially available anecdotal evidence from international boat owners,
192 we conducted an online search using the search engine Google. On 29. January 2021, we entered
193 the keywords “raw water strainer” OR “outboard motor lower unit” AND “fish” OR “animal” OR
194 “critter” OR “crab” OR “shrimp”. Within the results, we focused on boating forums or grey
195 literature in the area of aquatic invasive species, and compiled reports, pictures and anecdotes of
196 live and dead animals in water circuits in a supplementary document (Document S2).
197

198 **Results**

199 *Goby larvae in boat motor cooling systems*

200 We detected one goby larva in the raw water strainer of a sterndrive once, after a drive on August
201 19th in Basel. The goby larva caught measured 7 mm total length (Figure 3). It was located in the
202 fine-meshed filter, indicating that it moved through the raw water strainer.
203

204 *Quantification of propagule density and vector activity*

205 The plankton net, which was towed behind the boat, filtered a total of 6050 m³ of water and caught
206 50 larvae of native fish and 23 larvae of invasive gobies in total. Native fish larvae in the surface
207 waters were present predominantly in June and were not present in the plankton net after mid-July,
208 while goby larvae occurred from early June until late August (Figure 3 A). However, also for
209 gobies the number of larvae caught was higher in June than in July or August.
210

211 We only caught goby larvae with the fishing noose between mid-June and mid-July (Figure 3 B),
212 reflecting the peak of larval drift. The number of larvae caught with the fishing noose did not match
213 the expected goby larvae occurrence as determined by the calculated drift densities, if we assume
214 an equal distribution of larvae density across the whole stream. Either we did not catch any goby
215 larvae with the fishing noose, or we caught more than expected (Figure 3 B).
216

217 During all but two drives on which we quantified boat traffic, there were other boats present and
218 active after sunset (Figure 3 C). Identifiable categories of boat rides were pleasure rides (people
219 eating/drinking), water sports (swimming, water ski, wakeboarding), or angling trips (fishing rods
220 visible).
221

222 *Collection of anecdotal evidence for organisms in cooling systems*

223 We found eleven websites with documented evidence of fish and other creatures inside of boat
224 cooling systems and other boat compartments. Especially in boat forums there were often several
225 reports by different users reporting findings, so that the total number of reports exceeds that of

226 the number of websites found: There were three reports of live fish, five reports of dead fish, and
227 eight reports of fish for which it was unclear whether they were alive or dead. Additionally, we
228 found reports of live and dead other animals inside of cooling systems, among those were crabs,
229 shrimp, jellyfish and even a snake. The complete results of the online search are presented in
230 Document S1.

231

232 **Discussion**

233 The presence of a goby larva in the cooling system of a boat motor is an important proof of
234 principle for the hypothesis that invasive fish in general and specifically invasive gobies can be
235 translocated by small watercraft. This mode of translocation has long been assumed but is still
236 lacking empirical evidence (Ahnelt et al. 1998; Moskal'kova 1996, but see Bussmann and
237 Burkhardt-Holm 2020). Moreover, the only mode considered for goby translocation via
238 recreational boats so far is transport of their eggs on boat hulls (Adrian-Kalchhauser et al. 2017).
239 Goby larvae as propagules transported by recreational boats have, to the best of our knowledge,
240 never been considered in the scientific literature.

241

242 The density of round goby larvae in the uppermost layers of water in the sampled area of the river
243 Rhine during the reproductive season 2020 was never higher than 2.5 individuals per 100 m³,
244 which is in a similar range as found in some other drift-net studies (Hensler and Jude 2007; O'Brien
245 et al. 2019). However, the long duration of round goby reproductive season makes the pick-up and
246 viable transport and release of larvae within recreational boat engines possible at any point of time
247 during the European summer months. Additionally, the density of round goby larvae can be much
248 higher and vary between locations and years (Borcherding et al. 2016; Hayden and Miner 2009).
249 Data collected with the fishing noose shows that goby larvae are present in shallow waters along
250 the shore of rivers, sometimes even in higher numbers than expected considering the drift density
251 data. This has ramifications for assessing the probability of larvae uptake by boats: boats often
252 remain running for a long time to warm up the engine or load/unload the boat close to the shoreline,
253 which might increase uptake probability.

254

255 The observed numbers of recreational boats active after sunset confirm that the intake of goby
256 larvae is not just a hypothetical risk. Especially angling is a popular night-time activity for boat
257 owners in the area (personal communication of various boat club members and anglers). This
258 connection highlights how important it is to gain insight into the behavioural patterns of
259 stakeholders, who might unintentionally translocate invasive species. For example, fishing trips
260 can be used to infer invasion risk of round goby released as baitfish (Drake and Mandrak 2014).
261 Our study demonstrates how invasive species could be translocated not only as baitfish but also
262 within motor engines of fishing boats. This information can inform and improve risk models
263 evaluating translocation probabilities (Acosta and Forrest 2009; Parretti et al. 2020), which might
264 so far drastically underestimate the vector potential of boat cooling systems.

265

266 The lack of scientific studies reporting macrofauna in general, and especially fish, in cooling water
267 systems of small watercraft might reflect missing awareness of the potential of those systems
268 holding larger organisms, and/or unsuitable methodologies to sample them. We think that part of
269 the explanation for the low number of studies looking at cooling water content of recreational boats
270 and the lack of macrofauna reported therein are difficulties in sampling, as there are:

271

- 272 - The locations of water circuits and their contents are less accessible than the boat hull and
273 bilge. For many drive types it is impossible to fully sample the cooling water systems
274 without taking the motor apart or at least actively opening a latch/hood/valve.
275 - Sampling of boats often takes place on land, when large parts of the cooling water residuals
276 have already drained from the motor – however, sediments and organisms might still be
277 inside (Johnson et al. 2001; Montz and Hirsch 2016).
278 - Boat owners spontaneously asked for samples of organisms on or in their boats might agree
279 to a check from the outside, but hesitate to give access to the inside of the boat, or lack
280 knowledge of all the spaces where residual waters are.
281 - Sampling typically takes place during the day, while many organisms show diel vertical
282 migration with high abundances at the water surface only during the night (Ringelberg
283 2009). For a complete estimate of potentially translocated organisms in cooling water
284 systems, sampling would need to take place at times before as well as after sunset.
285

286 *Limitations*

287 We cannot be certain that the goby larva that we found in the raw water strainer was still alive in
288 the motor, because we only found it during analysis in the laboratory. To prove the actual
289 translocation potential of boat motors, the documentation of a live fish larva would be necessary.
290 We have ample reason to believe that survival in motor cooling systems is possible: The goby
291 larva we found looked externally unharmed upon examination of the sample. Fletcher et al. (2017)
292 documented the successful transport of a living juvenile fish through an impeller bilge pump,
293 which is a similar pumping system as used in boat motors. Furthermore, some of the collected
294 anecdotal evidence (see Document S1) speaks for the possible survival of organisms in cooling
295 water circuits and raw water strainers. Additionally, we found numerous other live organisms in
296 the boat motor cooling systems and strainers over the course of this project, as well as during
297 former projects looking at motor interiors (Amt für Umwelt und Energie Kanton Basel Stadt 2019).
298

299 *Minimizing translocation risk of cooling systems*

300 The risk of taking up a round goby larvae at surface drift densities as determined in our study might
301 seem low, as we only found a single goby larvae in one out of 30 drives. However, the uptake risk
302 of an individual boat is determined by multiple factors. For example, cooling water throughput of
303 a motor is dependent on engine power, as well as speed. Larger engines with more power therefore
304 have a higher probability of taking up fish larvae, as well as boats that are driving faster. Larger
305 motors also generally have larger water inlets and larger volumes in the lower unit. Furthermore,
306 the motor type in combination with the cooling system type differs largely in the amount of
307 standing water inside the water circuits. An outboard motor for example has a short water circuit
308 compared to an inboard motor, which sits amidships. Knowledge of the motor type configuration
309 and how to properly empty and flush it should ideally be part of every boater's education, e.g. as
310 part of boating license training. After all, motor maintenance also increases the service life of boat
311 engines, which would be in the interest of all boat owners. Finally, the cumulative risk of
312 translocation in cooling systems depends on the number of boats active after sunset, and the
313 number of those boats moving to uninvaded areas afterwards. This cumulative risk remains to be
314 assessed, taking regional and ecosystem-dependent circumstances into account.
315

316 Further potential measures against the translocation of invasive species in the inside of boat motors
317 and residual waters could include: motor flushing devices (“muffs”) as mandatory boat equipment,

318 check-points with information and instructions for boats at harbours or locks, or (mobile) boat
319 cleaning stations (Horvath 2008). We forwarded the outcomes of this study to relevant authorities
320 in Switzerland, resulting e.g. in adjustments in a newly launched information campaign about
321 translocation risks of recreational boats and measures for prevention to specifically include the
322 insides of boat motors (AWEL Zuerich 2020).

323

324 **Conclusions**

325 The finding of an invasive goby larvae inside the cooling water system of a recreational boat motor
326 is important to develop effective measures against translocation of AIS, especially during in-water
327 transport of boats. For organisms exhibiting diel vertical migration, nocturnal boat use increases
328 the chances for uptake into residual waters. Anecdotal evidence confirms the repeated occurrence
329 and, in parts, the viability of organisms inside the cooling systems. The hard-to-access insides of
330 boat motors are often neglected in studies, and underrepresented in management recommendations
331 for boat cleaning. However, there are simple methods to avoid unintentional translocation of AIS
332 in cooling systems: tilting the motor to remove most residual water whenever stationary, flushing
333 the motor with hot water, and educating boat owners about the hidden organisms inside the motors
334 and effective measures for the different types of motors could help preventing the further spread
335 of AIS, especially across biogeographical barriers.

336

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345

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351

352 **Author`s contributions**

353 KB: research conceptualization, sample design and methodology, investigation and data
354 collection, data analysis and interpretation; writing - original draft, writing – review and editing
355 PEH: research conceptualization, sample design and methodology, writing – review and editing
356 PBH: research conceptualization, sample design and methodology, funding provision, writing –
357 review and editing

358

359 **Declaration of interest**

360 Declarations of interest: none.

361

362 **Ethics and permits**

363 Ethics approval was not required according to Swiss law. The offices for environment and
364 energy of the cantons Basel-Stadt and Aargau granted permits for using the manta trawl and
365 fishing noose to catch fish.

366
367

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Invasive gobies as stowaways in boat motors

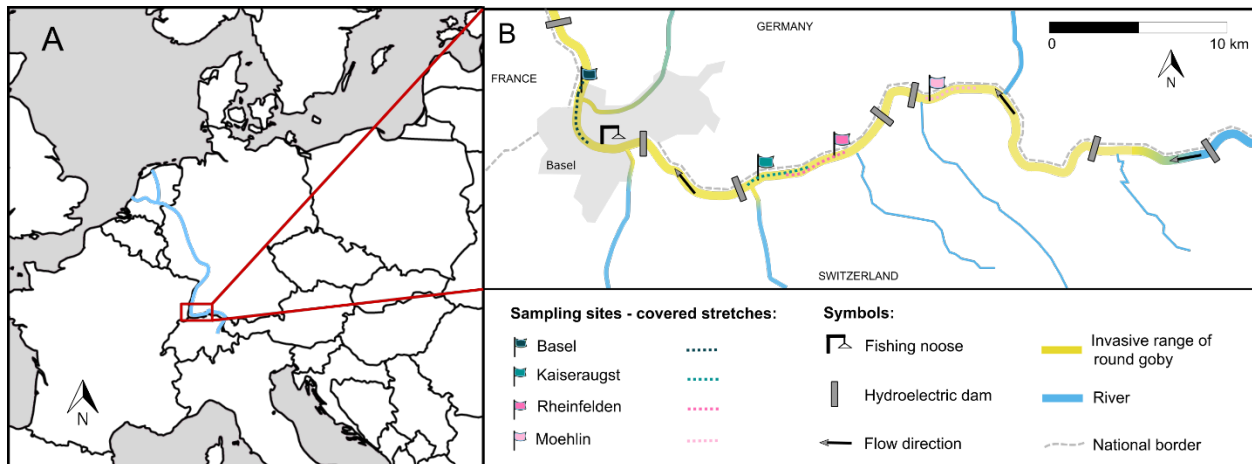
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469 **Figures**

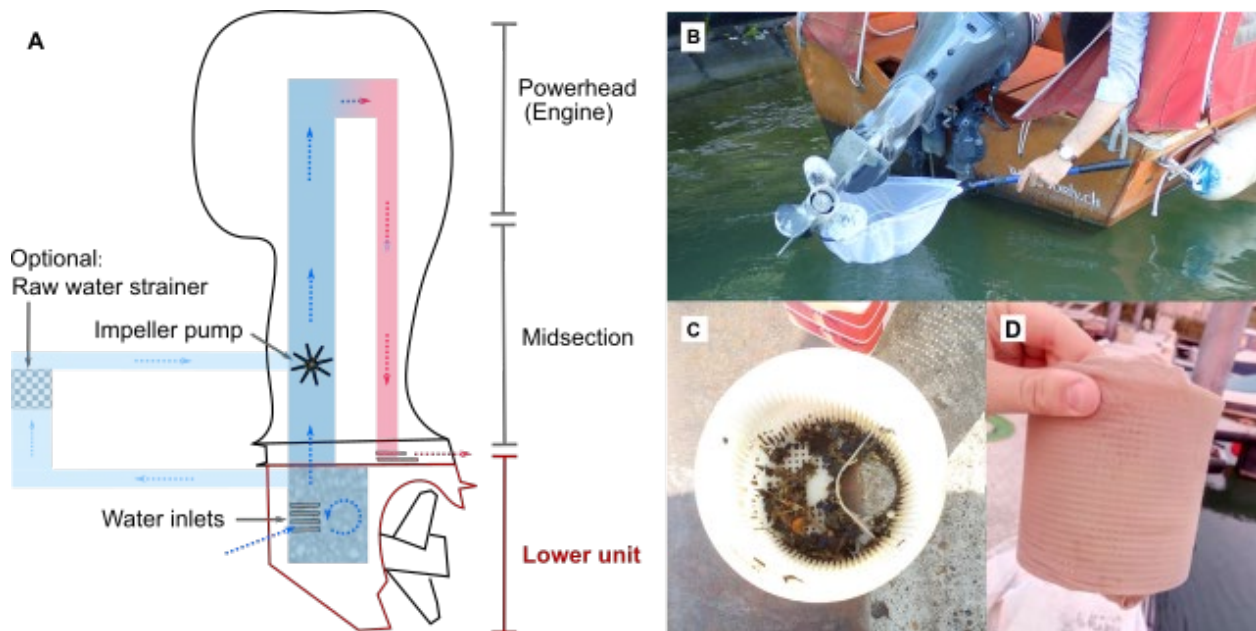
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471

472 **Figure 1.** Map of the study locations. **A** Political map of Europe showing the river Rhine (blue)
 473 and the location of the studied area (red box). **B** Map of the river Rhine and its tributaries in the
 474 studied area. Regular night drives took place at the sampling sites on the covered stretches between
 475 June and August 2020. Control fishing for drifting larvae took place with the fishing noose.
 476 Commercial shipping including ballast water exchange stops at Rheinfelden. Hydroelectric dams
 477 act as dispersal barriers for invasive goby species, slowing down their natural spread.

478

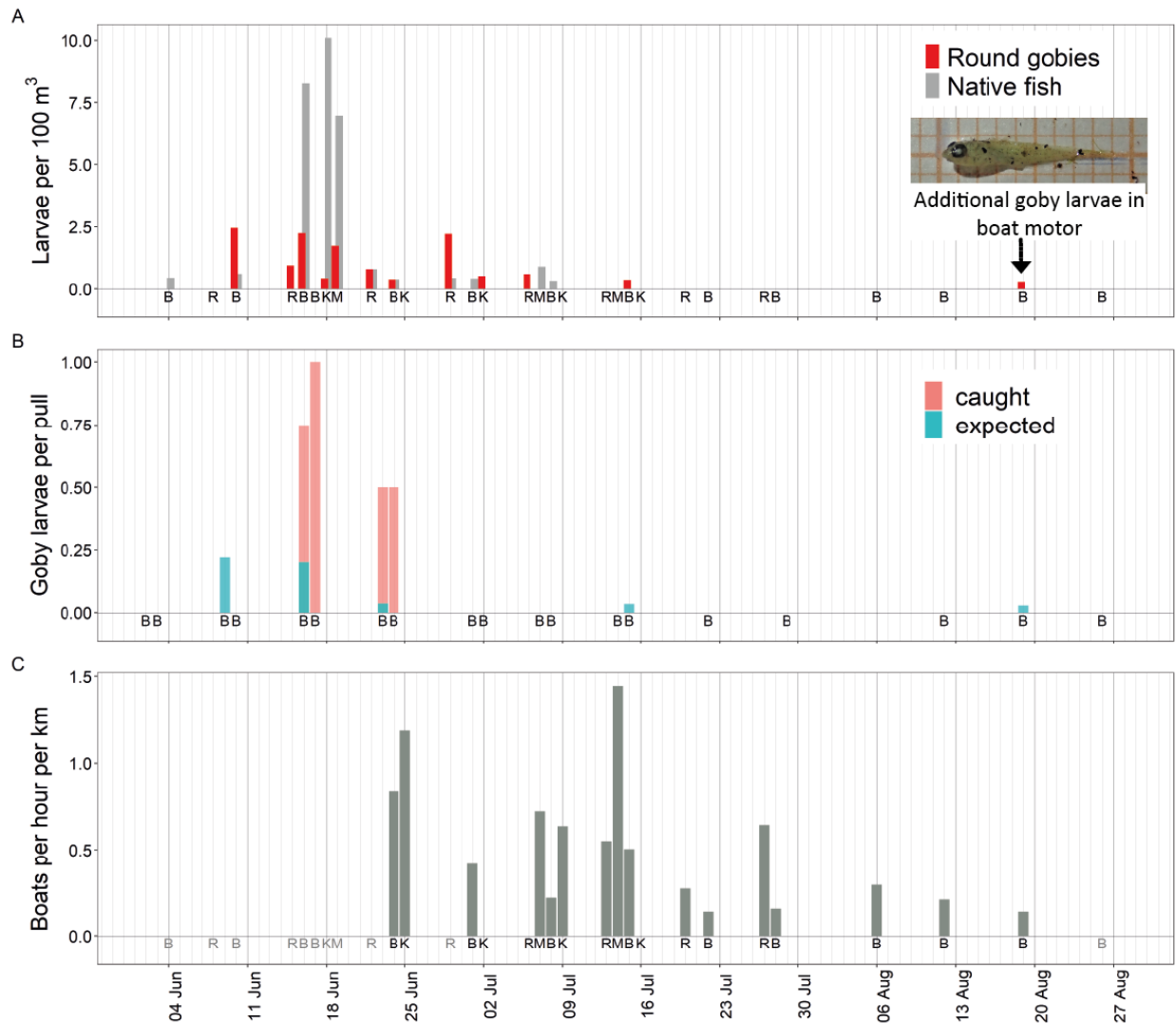


479

480 **Figure 2.** Boat motor water circuits and sampling. **A.** Simplified schematic drawing of an outboard
 481 boat motor and the water flow through the cooling system. Arrows represent flow direction. In the
 482 cavity of the lower unit (shaded), sediment and organisms accumulate if left under water. For
 483 inboard motors/stern drives (optional circuit, light blue), the engine is located inside the boat and
 484 the water circuits are therefore longer and hold more water. Additionally, they optionally include
 485 a raw water strainer to filter larger solids from the cooling water before they reach the impeller

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486 pump and the engine. **B** Sampling of an outboard motor in the water using a landing net. **C** Raw
 487 water strainer with contents (mesh size 1 mm). **D** Additional fine-meshed filter material wrapped
 488 around the strainer for boat drives in this study.
 489



490

491 **Figure 3.** Results of invasive goby larval sampling and boating activity after sunset. **A** Drift
 492 density of fish larvae caught in the plankton net towed behind recreational boats after sunset at
 493 different locations along the river Rhine (B = Basel, R = Rheinfelden, K = Kaiseraugst, M =
 494 Moehlin). Boat drives only took place on dates that are indicated with a study location. Round
 495 goby larvae are presented in red, native fish larvae (species not identified) are presented in grey.
 496 On 19th August, an additional goby larva was caught in a boat motor. **B** Expected and actual catch
 497 numbers of goby larvae per pull with the fishing noose. The location of the noose was in Basel
 498 (B). **C** Number of recreational boats active during the 1-2 h after sunset per kilometre at different
 499 locations along the river Rhine. Quantification of boats only took place on dates with letters printed
 500 in black, not on those printed in grey. Vertical grid lines represent days. A value of zero on a date
 501 at which a sampling took place means that no fish larvae were caught on that date, respectively no
 502 boats were seen.

503 **Supplementary information**

504

505 Document S1: Inquiry sent to boat owners to ask for collaboration on the project.

506

507 Document S2: Anecdotal evidence for organisms in boat motors from online boat forums.

508

509 Figure S1: Figure showing control sampling methods.

510

511 Table S1: Data on sampling locations, sampling dates, and boat types used for sampling.

512