

1 Many defense systems in microbial genomes, but which is defending 2 whom from what?

3
4 Eduardo P.C. Rocha^{1,*}, David Bikard²

5
6 ¹Microbial Evolutionary Genomics, Institut Pasteur, CNRS, UMR3525, Paris, France

7 ²Synthetic Biology, Department of Microbiology, Institut Pasteur, Paris, France

8
9 * corresponding author: erocha@pasteur.fr

10
11
12 **Abstract:** Prokaryotes have numerous mobile genetic elements (MGE) that mediate
13 horizontal gene transfer between cells. These elements can be costly, even deadly, and cells
14 use numerous defense systems to filter, control or inactivate them. Surprisingly, many
15 phages, conjugative plasmids, and their parasites, phage satellites or mobilizable plasmids,
16 encode defense systems homologous to those of bacteria. They constitute a significant
17 fraction of the systems found in bacterial genomes. As components of MGEs, they have
18 presumably evolved to provide them, not the cell, adaptive functions that may be defensive,
19 offensive, or both. This sheds new light on the role, effect, and fate of the so called “cellular
20 defense systems”, whereby they are not merely microbial defensive weapons in a two-
21 partner arms race, but tools of intragenomic conflict between multiple genetic elements
22 with divergent interests. It also raises many intriguing questions.

23 24 25 Introduction: mobile genetic elements drive gene flow at a (sometimes hefty) cost

26
27 Horizontal gene transfer (HGT) allows Bacteria and Archaea to rapidly match novel
28 ecological challenges and opportunities. HGT is most frequently mediated by self-
29 mobilizable mobile genetic elements (MGE) like bacteriophages (phages) and conjugative
30 elements that are present in most genomes, often in multiple copies. These elements can
31 autonomously transfer themselves from one cell to another using viral particles or
32 conjugative pilus, processes that also contribute to the exchange of chromosomal DNA.
33 Besides their ability to drive HGT, many MGEs encode traits adaptive to the host genome.
34 For example, key virulence factors in human pathogens are encoded in prophages and
35 antibiotic resistance genes are often transferred by conjugative elements [1, 2]. By
36 increasing the host fitness, these traits contribute to increase the frequency of MGEs in
37 communities, *i.e.*, they directly contribute to MGE fitness.

38
39 Conjugative elements and phages have their own molecular parasites that take advantage
40 of their mechanisms of horizontal transmission to transfer between cells. For example, viral
41 particles produced by phages can be hijacked by phage satellites [3] and conjugative pili can
42 be used by so-called mobilizable elements. The latter are at least as abundant as conjugative
43 plasmids, and possibly much more [4]. Recent data suggests that satellite phages are also
44 very common [5]. Many other MGEs lack known mechanisms of horizontal transmission and
45 may transfer between cells by exploiting phages and conjugative elements [6]. Importantly,

46 the presence of a MGE affects the frequency of other MGEs in the cell. This is the case for
47 mobilizable plasmids and phage satellites that co-transfer in synchrony with self-mobilizable
48 elements. It is also the case of phages that use the conjugative pilus as a receptor for cell
49 infection [7] and of plasmids capable of retro-transfer [8], a process by which a plasmid in
50 the recipient cell uses the incoming pilus to transfer to the donor cell. Finally, MGE infection
51 may spur the transfer of other elements. Phage infection favors the transfer of SXT-like
52 integrative conjugative elements (ICE) [9] and conjugation-induced SOS response activates
53 MGEs in the recipient cells [10]. The cellular genome thus harbors a cosmos of MGEs
54 establishing complex interactions among each other and with the host cell.

55

56 The association between the host and its MGEs lays on a gradient from pure parasitism to
57 intimate mutualism because vertical and horizontal transmission of MGEs impose fitness
58 costs to the cell that may eventually be compensated by the accessory traits encoded by
59 them. The replication of virulent phages implicates cell death, and they are at the edge of
60 maximal virulence in this gradient. The fitness effects of the remaining MGEs are more
61 diverse and vary with the physiological state of the cell and the presence of competing
62 MGEs. Temperate phages provide striking examples of such ambiguity. Their integration in
63 the genome can provide novel adaptive traits [11], but their subsequent excision from the
64 genome usually ends in host death [12]. MGEs that are parasites of other MGEs impact the
65 fitness of the latter. If this impact is very high and the parasitized MGE is deleterious to
66 bacteria, then the parasite of the parasite may end up benefiting the host cell. For example,
67 some satellites can abolish phage transmission resulting in cell death by release of viral
68 particles exclusively packaged with the satellite genome [13]. Although this process still
69 ends in cell death, the inhibitory effect of the satellite on phage reproduction blocks its
70 epidemic growth thereby protecting the microbial population. Since most genomes contain
71 MGEs, and virulent phages are extremely abundant in the environment [4, 14, 15], the fate
72 of cells often hangs in the outcome of their interaction with MGEs and that of MGEs among
73 themselves.

74

75 The wavering nature of interactions between MGEs and the host led to the evolution of
76 defense mechanisms to filter, control or inactivate these elements [16, 17]. Some defenses
77 are part of core cellular systems and provide protection from MGEs as part of a broader set
78 of cellular functions. For example, RecBCD is a powerful exonuclease involved in the repair
79 of double strand breaks by homologous recombination. It degrades linear double stranded
80 DNA until it meets a Chi site beyond which DNA is cut and RecA is loaded. Phages lacking Chi
81 sites are rapidly degraded by the enzyme [18]. Yet, phages can overcome this cell defense
82 by either blocking the host RecBCD enzymes or by evolving chi sites that trick RecBCD in
83 recognizing them as self [19]. In response to phage-encoded anti-RecBCD systems, retrons
84 that induce cell death when the RecBCD function is compromised have a protective anti-
85 anti-RecBCD function [20]. Contrary to RecBCD, many defense systems are not involved in
86 core cellular processes. Instead, they are specialized in providing innate or adaptive
87 immunity. Restriction-modification (R-M) systems, by far the most abundant [21], provide
88 excellent illustrations of the evolutionary processes resulting in the evolution of defense
89 and counter-defense systems (Figure 1). They imprint epigenetically the cellular genome
90 and inactivate (restrict) infecting MGEs lacking the adequate DNA modifications. As a
91 response, some phages counteract the activity of R-M systems by either producing anti-
92 restriction proteins or by extensively modifying their DNA [22, 23]. Anti-restriction functions

93 can in turn be recognized by anti-anti-restriction systems that provide a second layer of
94 resistance when R-M fails [24, 25]. As a complement, phages with extensive modifications in
95 their DNA can be recognized by specific anti-methylation restriction systems [26]. Some
96 host-phage interactions can be very complex, revealing long successions of tit-for-tat
97 strategies. For example, phage T4 encodes an anti-restriction system that can be recognized
98 by an anti-anti-restriction system inducing cell death in *E. coli* by tRNA cleaving, which can
99 be repaired by T4 using a pair of proteins that constitute an anti-anti-anti-restriction system
100 [27].

101

102 The function and evolution of defense and counter-defense systems are often studied in the
103 light of the antagonistic interaction between one host and one MGE, often a virulent phage.
104 But the reality is much more complex, and interesting. Genomes have many different MGEs
105 [4, 14] and results are piling up to show that many of the systems found in bacterial
106 genomes and once thought to be dedicated to the defense of the cell are actually encoded
107 in these MGEs. This includes systems encoded in temperate phages [28-31], satellites [24,
108 32, 33], conjugative elements [9, 34, 35] or mobilizable plasmids [21]. It has also been
109 pointed out that some components of MGEs, such as site-specific nucleases, are often
110 shuttled between MGEs and defense systems [36]. These observations raise intriguing
111 questions concerning the role, function and evolution of the co-called cellular defense
112 systems (Figure 1.1).

113

114 [Why are there so many defense systems in each genome?](#)

115

116 Most bacterial genomes encode several R-Ms, but often also CRISPR-Cas, retrons, and many
117 other defense systems [37]. For example, the two first sequenced genomes of *Helicobacter*
118 *pylori* encoded a total of more than 20 putative R-M systems [38], and genomes with
119 multiple CRISPR arrays and Cas systems are frequent [39]. The fast pace of discovery of
120 novel defense and counter-defense systems suggests they may account for a significant
121 number of the unknown function genes in genomes. The abundance of defense systems
122 could allow cells to be protected from a broad range of MGEs, counteracting the latter's
123 tendency to evolve counter-defenses. Yet, defense systems can be costly [40], because of
124 production costs when they are required at high concentration [41], because their activity
125 can be energetically costly [42], or because they may be incompatible with other cellular
126 mechanisms [43]. They can also kill the cell by auto-immunity [44]. Hence, the number of
127 defense systems in a genome is expected to depend on the balance between these costs
128 and advantages of extensive protection against MGEs.

129

130 The observations that genomes have many MGEs and that these encode many defense
131 systems provide an alternative or complementary explanation for why genomes contain so
132 many such systems: they acquire the systems when they are infected by the MGEs. This
133 does not exclude selection for a multiplicity of systems by each cell, but it brings to the fore
134 that to understand their frequency in cellular genomes one must also account for the
135 infectivity of MGEs. This means that the multiplicity of systems in cellular genomes might be
136 a consequence of the high transmissibility and abundance of MGEs, not (only) the result of
137 natural selection for protection of the cell.

138

139 Why are defense systems very diverse within species?

140

141 Defense systems tend to be diverse across strains of a species [16]. This could be explained
142 by several factors. First, the MGEs may differ between habitats and locally adapted
143 populations may need to select for different systems. Second, genetic diversity favors the
144 emergence of resistance in populations [45] and antagonistic co-evolution between MGEs
145 and their hosts could be result in a diverse repertoire of defenses in populations. For
146 example, individuals with rare alleles are favored by negative frequency dependent
147 selection because most antagonists lack the response mechanisms to tackle them. As these
148 individuals rise in frequency in the population, antagonists with the ability to tackle them
149 also rise in frequency, thereby decreasing the advantage of the initial clone. This may result
150 in diversification of the population into many different systems or in the rapid cyclic turn-
151 over of a few defense systems [46]. In addition, the presence of various systems providing
152 immunity against different phages within a single community of bacteria is expected to
153 provide better defense at the population level without requiring each individual genome to
154 encode a very large number of defense systems, in what has been described as distributed
155 (pan) immunity [37].

156

157 The existence of multiple different MGEs across strains of a species contributes to explain
158 why defense systems are so diverse: they are brought by different MGEs. To understand
159 why different MGEs carry different defenses systems it will be necessary to dissect the
160 complex networks of interactions between the host and its multiple MGEs.

161

162 How is immunity gained?

163

164 The repertoire of defenses in genomes can vary as the result of dedicated molecular
165 mechanisms of variation. Many systems include mechanisms driving their own
166 diversification, e.g., CRISPR arrays can acquire spacers to target novel elements [17]. Some
167 R-M systems are also capable of rapidly change their sequence specificity [47]. These
168 mechanisms allow the host to fine-tune its defenses very rapidly. Yet, the available evidence
169 suggests that HGT and gene loss have key roles in the diversification of defense repertoires
170 at the species level [48]. Accordingly, pseudogenes of defense systems have been observed
171 for many R-M [38] and CRISPR-Cas systems [49].

172

173 The abundance of defense systems in MGEs suggests a very straightforward mechanism for
174 the acquisition of novel mechanisms of immunity: systems are transferred across strains by
175 the MGEs encoding them. Mechanisms of transfer of MGEs between cells are well-known
176 and their epidemiological patterns are being described in detail. Furthermore, MGEs are
177 gained at high rates because of their infectiousness, and they are frequently lost from
178 populations because of their cost. The genetic linkage between the defense systems and the
179 MGEs thereby contributes to explain the acquisition of novel defense systems. It may also
180 offer some clues on how entirely novel defense strategies emerge. The recent discovery of
181 many anti-phage systems shows that they frequently consist in an assemblage of protein
182 domains also implicated in other cellular processes such as nucleases, kinases, deaminases,
183 proteases, or ATPases [36]. For instance, the Stk2 defense kinase is part of a family of
184 kinases whose members are implicated in various cellular process such as the control of the
185 cell cycle or the exit of dormancy [50]. The anti-phage Viperins are close homologues to GTP

186 cyclases involved in other functions [51]. The cooption of proteins, or protein domains, with
187 other functions, and the creation of novel assemblages leading to genetic innovation by
188 recombination and mutation is likely facilitated by the horizontal transfer of defense
189 systems across genetic backgrounds [52]. While the probability of a functional innovation
190 resulting from the co-option of each these systems is low, their very frequent transfer and
191 rapid evolution may result in such a high rate of novel combinations of domains that some
192 will eventually result in adaptive novel defense systems. MGEs will then eventually capture
193 such innovations for their own use and spread them across species.

194

195 [Defending whom from what?](#)

196

197 Experimental verification of the function of defense systems usually involves testing the
198 success of infection by virulent phages. As a result, the role of defense systems tends to be
199 discussed in the light of phage-bacteria interactions. It does seem reasonable to assume
200 that systems present in a microbial genome for a long time are protecting the cell from
201 MGEs, and especially against virulent phages given their lethality for the cell. Yet, systems
202 encoded in MGEs are more likely to be selected because they benefit the MGE. Sometimes
203 the two objectives, cell defense and MGE fitness, coincide. Defense systems encoded in P4-
204 like satellites were shown experimentally to protect the cell from several phages [53]. In this
205 case the satellite and the cell have the same interest in preventing infection by phages that
206 can kill the cell and cannot be exploited by the satellite to propagate. But sometimes, the
207 interests of the MGE and the cell are not so well aligned. This is exemplified by expensive
208 exclusion systems encoded by conjugative systems to fend off closely related plasmids [54].
209 For example, the surface exclusion system of plasmid F prevents infection by similar
210 plasmids thanks to the production of thousands of copies of an outer membrane protein
211 that accounts for a large part of the plasmid carrier cost [55]. A plasmid encoding an
212 expensive defense system against another plasmid is engaging in an antagonistic interaction
213 whose cost to the cell may be much larger than the expected reward. Temperate phages
214 encoding defense systems against virulent phages seem very common [29, 31, 56]. While
215 they may provide temporary relief to a cell, they may also have little long-term impact in
216 bacterial fitness when the victorious phage is eventually induced and lyses the cell. Finally,
217 phages encoding defense or anti-defense systems against satellites are engaging in an
218 interaction with their parasites in a way that resembles their own interaction with the cell
219 (but with their own position reversed as they are now the host) [33]. Such prophage-
220 encoded defense systems could be highly deleterious to the cell because they remove a
221 protective satellite and favor a phage that will eventually kill the host.

222

223 The existence of defense systems in MGEs and the interactions between them raise two key
224 questions. Who is encoding the system? The identification of the defense system as MGE-
225 associated depends on the precise delimitation of the latter, which may be difficult both
226 computationally and experimentally. Genomes encode many defective MGEs and it may
227 also be unclear if a defense system is part of a functional MGE, is being co-opted by the cell,
228 or is non-functional. Such distinctions may be key to understand their role. Which genetic
229 elements are being targeted by the defense system? While many systems are effective
230 against virulent phages, it is often unclear which other elements are being targeted, i.e.
231 which target elements lead to the selection for the conservation of the defense system.
232 Systems encoded by MGEs may be targeting other competing MGE that are not costly to the

233 cell. They may even be targeting elements that are adaptive to the cell, or targeting the cell
234 itself (*e.g.*, addictive systems or anti-defense systems), thereby lowering bacterial fitness.
235 The role of the defense system in the MGE can thus be intimately associated with the
236 positive or negative fitness effect of the MGE to the cell. Knowing which genetic elements
237 are being targeted in nature will require a better understanding of the ecological contexts
238 where such systems are selected for.

239

240 [How do defense systems affect gene flow?](#)

241

242 Selection for cell defense systems depends on a trade-off between costs and benefits [40].
243 In addition to the costs of production and the cost of auto-immunity, there is an
244 evolutionary cost to restricting adaptive gene flow, including allelic recombination and
245 acquisition of novel genes by HGT. For example, epidemic *V. cholerae* strains depend on a
246 prophage for a key virulence factor (the cholera toxin). When they are infected by SXT-like
247 conjugative elements carrying defense systems they are hampered in their ability to acquire
248 the toxin [9]. More generally, a computational analysis of ca. 80 species showed that gene
249 flow is decreased when strains of a species have incompatible R-M systems [57]. Hence,
250 when R-M systems diversify within populations, their DNA exchanges become more
251 frequent between strains with similar systems (Figure 1.II). Beyond R-M, other defense
252 systems including BREX, DISARM, CRISPR or Wadjet might also restrict gene flow. As a rule,
253 one would expect that very generic systems, like R-M, would have an important effect on
254 restricting gene flow, whereas more targeted systems, like CRISPR-Cas, would tend to affect
255 gene flow driven by a few particularly deleterious elements. Accordingly, the impact of
256 CRISPR-Cas in restricting gene flow has remained controversial [58, 59]. The effect of
257 defense systems on gene flow is however not always straightforward. Transduction, the
258 transfer of bacterial DNA in viral particles, is favored by the existence of CRISPR-Cas systems
259 in recipient cells when they target the phage DNA. In this case, bacterial DNA in viral
260 particles is transferred into the cell whereas the phage DNA is excluded, resulting in cells
261 that receive exogenous cellular DNA while being protected from phages [60].

262

263 The negative impact of defense systems on gene flow has been regarded as a costly by-
264 product of selection for protection of the cell. But defense systems in MGEs may be selected
265 because they block HGT to prevent the cell from acquiring competitor MGEs. The resulting
266 sexual isolation is advantageous for the MGE but can be deleterious to the cell. Further
267 work is needed to quantify the impact of different systems in gene flow and how they affect
268 the evolvability of microbes.

269

270 [Is it defense, attack, or something else?](#)

271

272 While many systems have been called defensive relative to their ability to defend bacteria,
273 they may be attack systems when part of MGEs. A striking example is provided by phage-
274 satellite interactions. The reproduction of virulent phages of the ICP1 family in *Vibrio*
275 *cholerae* is abolished by the PLE satellite elements [13]. In response, ICP1 phages have
276 evolved the ability to encode a CRISPR-Cas system or specific nucleases that allow to
277 eliminate the satellite [53]. In this context they could be regarded as attack systems, since
278 their success results in cell death. Some systems may even have multiple roles. R-M systems
279 contribute to the stabilization of plasmids in the cell by acting as poison-antidote addictive

280 systems [61]. In such cases, loss of the plasmid and its R-M system prevents further
281 expression of the latter. Since endonucleases have longer half-lives than methylases, this
282 eventually results in genomes that are restricted because they are insufficiently methylated.
283 R-Ms are thus part of the offensive arsenal of plasmids that allows them to be maintained in
284 cells. Yet, these R-M systems can also protect the consortium (cell and plasmid) from
285 infection by other MGEs, thereby acting as cell defense systems. Plasmids also frequently
286 encode toxin-antitoxin systems that behave as addiction systems [62], some of which are
287 implicated in phage defense. Homologs of cell defense systems encoded in MGEs can thus
288 be offensive tools with positive side-effects in cell defense. The relative contribution of such
289 systems to the different types of ecological interactions, defense or offense, remains to be
290 explored.

291

292 Are MGEs at the origin of “defense islands”?

293

294 It was observed a decade ago that defense systems are often clustered in a few loci in
295 microbial chromosomes [63]. This characteristic was leveraged into a systematic method to
296 discover novel systems by co-localization with known ones [64]. Interestingly, recent data
297 has revealed that anti-defense systems, both anti-R-M and anti-CRISPR-Cas, tend to cluster
298 in bacterial genomes, often in recognizable MGEs [65]. The clustering of defense systems,
299 and that of anti-defense systems, could be selected to facilitate their co-regulation and
300 interaction for a more effective function against MGEs. For the moment, there is very little
301 evidence available of that.

302

303 The presence of such systems in MGEs provides a simple explanation for their co-
304 localization in bacterial genomes (Figure 1.III). Genes acquired by HGT, and MGEs in
305 particular, tend to integrate a small number of chromosome hotspots [6]. These elements
306 may degenerate by the accumulation of mutations, deletions, and insertions. Chromosome
307 hotspots are thus littered with remnants of previous events of transfer. As MGEs are
308 integrated and eventually degrade in the hotspot, some genes may remain functional
309 because they are adaptive for the cell [52]. Since MGEs often carry defense and anti-
310 defense systems, the rapid turnover of the former in hotspots may be accompanied by
311 selection for the conservation of the latter. As rounds of MGE integration/degradation
312 succeed in natural history, the remnant defense systems form clusters in the chromosome.
313 The clustering of these systems may facilitate the evolution of functional interactions
314 between them or co-regulation of gene expression. For example, it has been observed that
315 type I and type III CRISPR-Cas systems provide two integrated levels of defense against
316 phages [66]. These systems are often co-localized [39], but the functional advantages of
317 their co-localization in the genome are yet unclear.

318

319 Outlook

320

321 MGEs of Bacteria and Archaea encode many accessory functions that can have adaptive
322 value for the host. They are also units of selection that prosper if they manage to increase
323 their population size by horizontal or vertical transmission. To understand the roles of
324 defense and/or counter-defense systems, given their abundance in MGEs, one must attain a
325 better understanding of the complex networks of interactions between these semi-
326 autonomous agents. Their study will shed novel light on the function, evolution, and ecology

327 of defense systems. In terms of function, many novel molecular mechanisms of interference
328 may yet remain to be discovered. From the evolutionary point of view, these observations
329 suggests that the existence of many defense systems in genomes may be more directly
330 related to selection for survival and reproduction of the MGEs than of the cell. In terms of
331 ecology, given the impact of MGEs in adaptation and regulation of bacterial populations, a
332 better understanding of the defense systems may be key to understand and manipulate
333 microbial population dynamics.

334
335

336 Acknowledgements

337 Aude Bernheim and Frédérique Le Roux for comments and suggestions and Marie Touchon
338 for discussions and graphical elements for the figure.

339
340

341 Bibliography

342

- 343 1. Taylor VL, Fitzpatrick AD, Islam Z, Maxwell KL. The diverse impacts of phage morons
344 on bacterial fitness and virulence. *Advances in virus research*. 2019;103:1-31.
- 345 2. Bennett P. Plasmid encoded antibiotic resistance: acquisition and transfer of
346 antibiotic resistance genes in bacteria. *Br J Pharmacol*. 2008;153(S1):S347-S57.
- 347 3. Penadés JR, Christie GE. The phage-inducible chromosomal islands: a family of highly
348 evolved molecular parasites. *Annual review of virology*. 2015;2:181-201.
- 349 4. Smillie C, Pilar Garcillan-Barcia M, Victoria Francia M, Rocha EPC, de la Cruz F.
350 Mobility of Plasmids. *Microbiol Mol Biol Rev*. 2010;74(3):434-52.
- 351 5. de Sousa JM, Rocha EP. To catch a hijacker: abundance, evolution and genetic
352 diversity of P4-like bacteriophage satellites. *bioRxiv*. 2021.
- 353 6. Oliveira PH, Touchon M, Cury J, Rocha EPC. The chromosomal organization of
354 horizontal gene transfer in bacteria. *Nature communications*. 2017;8(1):841.
- 355 7. Harb L, Chamakura K, Khara P, Christie PJ, Young R, Zeng L. ssRNA phage penetration
356 triggers detachment of the F-pilus. *Proceedings of the National Academy of Sciences*.
357 2020;117(41):25751-8.
- 358 8. Szpirer C, Top E, Couturier M, Mergeay M. Retrotransfer or gene capture: a feature
359 of conjugative plasmids, with ecological and evolutionary significance. *Microbiology*.
360 1999;145 (Pt 12):3321-9.
- 361 9. LeGault K, Hays SG, Angermeyer A, McKitterick AC, Johura F-t, Sultana M, et al.
362 Temporal shifts in antibiotic resistance elements govern virus-pathogen conflicts. *Science*.
363 2021;373:eabg2166.
- 364 10. Baharoglu Z, Bikard D, Mazel D. Conjugative DNA transfer induces the bacterial SOS
365 response and promotes antibiotic resistance development through integron activation. *PLoS*
366 *Genet*. 2010;6(10):e1001165.
- 367 11. Wagner PL, Waldor MK. Bacteriophage control of bacterial virulence. *Infect Immun*.
368 2002;70(8):3985-93.
- 369 12. Paul JH. Prophages in marine bacteria: dangerous molecular time bombs or the key
370 to survival in the seas? *Isme J*. 2008;2(6):579-89.
- 371 13. Seed KD, Lazinski DW, Calderwood SB, Camilli A. A bacteriophage encodes its own
372 CRISPR/Cas adaptive response to evade host innate immunity. *Nature*. 2013;494(7438):489-
373 91.

- 374 14. Touchon M, Bernheim A, Rocha EP. Genetic and life-history traits associated with the
375 distribution of prophages in bacteria. *Isme J.* 2016;10:2744–54.
- 376 15. Wigington CH, Sonderegger D, Brussaard CPD, Buchan A, Finke JF, Fuhrman JA, et al.
377 Re-examination of the relationship between marine virus and microbial cell abundances.
378 *Nature Microbiol.* 2016;1:15024.
- 379 16. van Houte S, Buckling A, Westra ER. Evolutionary ecology of prokaryotic immune
380 mechanisms. *Microbiology and Molecular Biology Reviews.* 2016;80(3):745-63.
- 381 17. Hampton HG, Watson BN, Fineran PC. The arms race between bacteria and their
382 phage foes. *Nature.* 2020;577(7790):327-36.
- 383 18. Cheng K, Wilkinson M, Chaban Y, Wigley DB. A conformational switch in response to
384 Chi converts RecBCD from phage destruction to DNA repair. *Nature structural & molecular*
385 *biology.* 2020;27(1):71-7.
- 386 19. Bobay L-M, Touchon M, Rocha EP. Manipulating or superseding host recombination
387 functions: a dilemma that shapes phage evolvability. *PLoS Genet.* 2013;9(9):e1003825.
- 388 20. Millman A, Bernheim A, Stokar-Avihail A, Fedorenko T, Voichek M, Leavitt A, et al.
389 Bacterial retrons function in anti-phage defense. *Cell.* 2020;183(6):1551-61. e12.
- 390 21. Oliveira PH, Touchon M, Rocha EP. The interplay of restriction-modification systems
391 with mobile genetic elements and their prokaryotic hosts. *Nucleic Acids Res.*
392 2014;42(16):10618-31.
- 393 22. Isaev A, Drobiazko A, Sierro N, Gordeeva J, Yosef I, Qimron U, et al. Phage T7 DNA
394 mimic protein Ocr is a potent inhibitor of BREX defence. *Nucleic acids research.*
395 2020;48(10):5397-406.
- 396 23. Krüger DH, Bickle TA. Bacteriophage survival. Multiple mechanisms for avoiding the
397 deoxyribonucleic acid restriction systems of their hosts. *Microbiol Rev.* 1983;47:345-60.
- 398 24. Rousset F, Dowding J, Bernheim A, Rocha E, Bikard D. Prophage-encoded hotspots of
399 bacterial immune systems. *bioRxiv.* 2021.
- 400 25. Fukuda E, Kaminska KH, Bujnicki JM, Kobayashi I. Cell death upon epigenetic genome
401 methylation: a novel function of methyl-specific deoxyribonucleases. *Genome Biol.*
402 2008;9(11):R163.
- 403 26. Loenen WA, Raleigh EA. The other face of restriction: modification-dependent
404 enzymes. *Nucleic Acids Res.* 2014;42(1):56-69.
- 405 27. Penner M, Morad I, Snyder L, Kaufmann G. Phage T4-coded Stp: double-edged
406 effector of coupled DNA and tRNA-restriction systems. *J Mol Biol.* 1995;249(5):857-68.
- 407 28. Bondy-Denomy J, Qian J, Westra ER, Buckling A, Guttman DS, Davidson AR, et al.
408 Prophages mediate defense against phage infection through diverse mechanisms. *Isme J.*
409 2016;10(12):2854-66.
- 410 29. Dedrick RM, Jacobs-Sera D, Bustamante CA, Garlena RA, Mavrigh TN, Pope WH, et al.
411 Prophage-mediated defence against viral attack and viral counter-defence. *Nat Microbiol.*
412 2017;2:16251.
- 413 30. Piel D, Bruto M, Labreuche Y, Blanquart F, Chenivresse S, Lepanse S, et al. Genetic
414 determinism of phage-bacteria coevolution in natural populations. *bioRxiv.* 2021.
- 415 31. Hussain FA, Dubert J, Elsherbini J, Murphy M, VanInsberghe D, Arevalo P, et al. Rapid
416 evolutionary turnover of mobile genetic elements drives microbial resistance to viruses.
417 *bioRxiv.* 2021.
- 418 32. McKitterick AC, Seed KD. Anti-phage islands force their target phage to directly
419 mediate island excision and spread. *Nature communications.* 2018;9(1):1-8.

- 420 33. Fillol-Salom A, Miguel-Romero L, Marina A, Chen J, Penadés JR. Beyond the CRISPR-
421 Cas safeguard: PICI-encoded innate immune systems protect bacteria from bacteriophage
422 predation. *Current Opinion in Microbiology*. 2020;56:52-8.
- 423 34. Klaenhammer TR. Plasmid-directed mechanisms for bacteriophage defense in lactic
424 streptococci. *FEMS Microbiology Reviews*. 1987;3(3):313-25.
- 425 35. Pinilla-Redondo R, Mayo-Muñoz D, Russel J, Garrett RA, Randau L, Sørensen SJ, et al.
426 Type IV CRISPR–Cas systems are highly diverse and involved in competition between
427 plasmids. *Nucleic acids research*. 2020;48(4):2000-12.
- 428 36. Koonin EV, Makarova KS, Wolf YI, Krupovic M. Evolutionary entanglement of mobile
429 genetic elements and host defence systems: guns for hire. *Nature Reviews Genetics*. 2019:1-
430 13.
- 431 37. Bernheim A, Sorek R. The pan-immune system of bacteria: antiviral defence as a
432 community resource. *Nature Reviews Microbiology*. 2019:1-7.
- 433 38. Lin LF, Posfai J, Roberts RJ, Kong H. Comparative genomics of the restriction-
434 modification systems in *Helicobacter pylori*. *Proc Natl Acad Sci USA*. 2001;98:2740-5.
- 435 39. Bernheim A, Bikard D, Touchon M, Rocha EP. Atypical organizations and epistatic
436 interactions of CRISPRs and cas clusters in genomes and their mobile genetic elements.
437 *Nucleic acids research*. 2020;48(2):748-60.
- 438 40. Bondy-Denomy J, Davidson AR. To acquire or resist: the complex biological effects of
439 CRISPR-Cas systems. *Trends Microbiol*. 2014;22(4):218-25.
- 440 41. Vale PF, Lafforgue G, Gatchitch F, Gardan R, Moineau S, Gandon S. Costs of CRISPR-
441 Cas-mediated resistance in *Streptococcus thermophilus*. *Proceedings of the Royal Society B:*
442 *Biological Sciences*. 2015;282(1812):20151270.
- 443 42. Seidel R, Bloom JG, Dekker C, Szczelkun MD. Motor step size and ATP coupling
444 efficiency of the dsDNA translocase EcoR124I. *The EMBO journal*. 2008;27(9):1388-98.
- 445 43. Bernheim A, Calvo-Villamanan A, Basier C, Cui L, Rocha EPC, Touchon M, et al.
446 Inhibition of NHEJ repair by type II-A CRISPR-Cas systems in bacteria. *Nature*
447 *communications*. 2017;8(1):2094.
- 448 44. Rollie C, Chevallereau A, Watson BN, Chyou T-y, Fradet O, McLeod I, et al. Targeting
449 of temperate phages drives loss of type I CRISPR–Cas systems. *Nature*. 2020;578(7793):149-
450 53.
- 451 45. Chabas H, Lion S, Nicot A, Meaden S, van Houte S, Moineau S, et al. Evolutionary
452 emergence of infectious diseases in heterogeneous host populations. *PLoS biology*.
453 2018;16(9):e2006738.
- 454 46. Woolhouse ME, Webster JP, Domingo E, Charlesworth B, Levin BR. Biological and
455 biomedical implications of the co-evolution of pathogens and their hosts. *Nat Genet*.
456 2002;32(4):569-77.
- 457 47. Dybvig K, Sitaraman R, French CT. A family of phase-variable restriction enzymes
458 with differing specificities generated by high-frequency gene arrangements. *Proc Natl Acad*
459 *Sci USA*. 1998;95:13923-8.
- 460 48. Jeltsch A, Pingoud A. Horizontal gene transfer contributes to the wide distribution
461 and evolution of type II restriction-modification systems. *J Mol Evol*. 1996;42:91-6.
- 462 49. Stern A, Keren L, Wurtzel O, Amitai G, Sorek R. Self-targeting by CRISPR: gene
463 regulation or autoimmunity? *Trends Genet*. 2010;26(8):335-40.
- 464 50. Depardieu F, Didier J-P, Bernheim A, Sherlock A, Molina H, Duclos B, et al. A
465 eukaryotic-like serine/threonine kinase protects *Staphylococci* against phages. *Cell host &*
466 *microbe*. 2016;20(4):471-81.

467 51. Bernheim A, Millman A, Ofir G, Meitav G, Avraham C, Shomar H, et al. Prokaryotic
468 viperins produce diverse antiviral molecules. *Nature*. 2020:1-5.

469 52. Touchon M, Bobay LM, Rocha EP. The chromosomal accommodation and
470 domestication of mobile genetic elements. *Curr Opin Microbiol*. 2014;22:22-9.

471 53. Barth ZK, Nguyen MH, Seed KD. A chimeric nuclease substitutes a phage CRISPR-Cas
472 system to provide sequence-specific immunity against subviral parasites. *eLife*.
473 2021;10:e68339.

474 54. Garcillan-Barcia MP, de la Cruz F. Why is entry exclusion an essential feature of
475 conjugative plasmids? *Plasmid*. 2008;60(1):1-18.

476 55. Achtman M, Kennedy N, Skurray R. Cell-cell interactions in conjugating *Escherichia*
477 *coli*: role of traT protein in surface exclusion. *Proc Natl Acad Sci U S A*. 1977;74(11):5104-8.

478 56. Medvedeva S, Liu Y, Koonin EV, Severinov K, Prangishvili D, Krupovic M. Virus-borne
479 mini-CRISPR arrays are involved in interviral conflicts. *Nature communications*.
480 2019;10(1):1-10.

481 57. Oliveira PH, Touchon M, Rocha EP. Regulation of genetic flux between bacteria by
482 restriction-modification systems. *Proc Natl Acad Sci U S A*. 2016;113(20):5658-63.

483 58. Gophna U, Kristensen DM, Wolf YI, Popa O, Drevet C, Koonin EV. No evidence of
484 inhibition of horizontal gene transfer by CRISPR-Cas on evolutionary timescales. *Isme J*.
485 2015.

486 59. Wheatley RM, MacLean RC. CRISPR-Cas systems restrict horizontal gene transfer in
487 *Pseudomonas aeruginosa*. *Isme J*. 2021;15(5):1420-33.

488 60. Watson BNJ, Staals RHJ, Fineran PC. CRISPR-Cas-Mediated Phage Resistance
489 Enhances Horizontal Gene Transfer by Transduction. *MBio*. 2018;9(1):e02406-17.

490 61. Naito T, Kusano K, Kobayashi I. Selfish behavior of restriction-modification systems.
491 *Science*. 1995;267:897-9.

492 62. Dy RL, Przybilski R, Semeijn K, Salmond GP, Fineran PC. A widespread bacteriophage
493 abortive infection system functions through a Type IV toxin-antitoxin mechanism. *Nucleic*
494 *Acids Res*. 2014;42(7):4590-605.

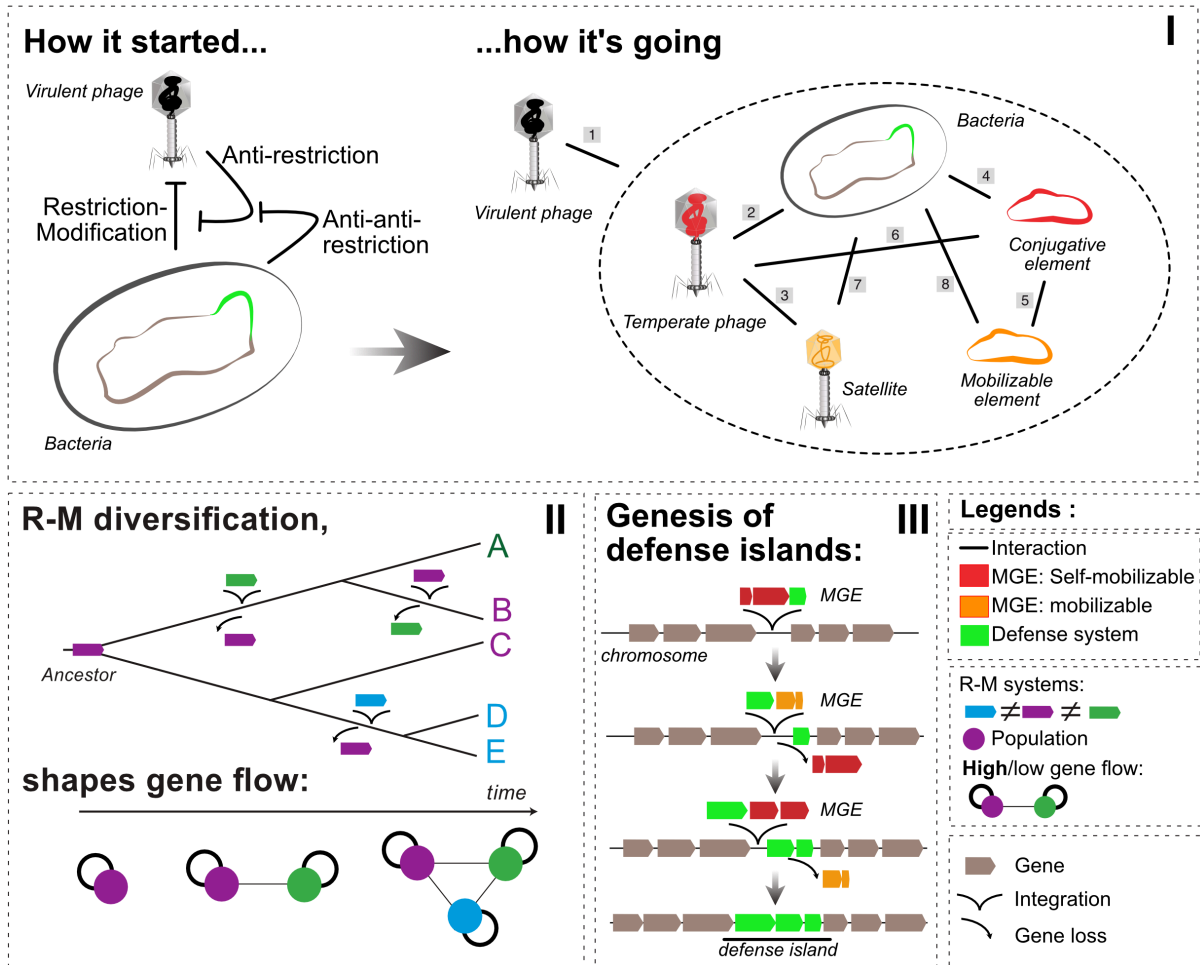
495 63. Makarova KS, Wolf YI, Snir S, Koonin EV. Defense islands in bacterial and archaeal
496 genomes and prediction of novel defense systems. *J Bacteriol*. 2011;193(21):6039-56.

497 64. Doron S, Melamed S, Ofir G, Leavitt A, Lopatina A, Keren M, et al. Systematic
498 discovery of antiphage defense systems in the microbial pangenome. *Science*.
499 2018;359(6379).

500 65. Pinilla-Redondo R, Shehreen S, Marino ND, Fagerlund RD, Brown CM, Sorensen SJ, et
501 al. Discovery of multiple anti-CRISPRs highlights anti-defense gene clustering in mobile
502 genetic elements. *Nature communications*. 2020;11(1):5652.

503 66. Silas S, Lucas-Elio P, Jackson SA, Aroca-Crevillen A, Hansen LL, Fineran PC, et al. Type
504 III CRISPR-Cas systems can provide redundancy to counteract viral escape from type I
505 systems. *eLife*. 2017;6:e27601.

506
507
508



510
 511
 512
 513
 514
 515
 516
 517
 518
 519
 520
 521
 522
 523
 524
 525
 526
 527
 528
 529
 530

Figure 1. I. Defense and anti-defense systems are often studied in the context of the interaction between one host and one MGE, usually a virulent phage (left). Yet, the presence of numerous MGEs in populations and their ability to encode their own systems renders the picture more complex (right). Virulent phages establish antagonistic interactions with the other MGEs and the cell (1). But the associations between the other MGEs and the cell can be more diverse (2-7). Temperate phages and conjugative plasmids exploit their cellular host (2,4) and can be exploited by other MGEs (3, 5). Plasmids often encode systems that are effective barriers to phages, e.g. R-M (6). Phages are a threat to plasmids when they kill the host cell (6). Satellites may benefit the host by diminishing phage infection (7). Most of these interactions (2-8) can at times be beneficial to both partners, e.g. when a conjugative plasmid provides a nosocomial bacterium with antibiotic resistance. **II.** Diversification of R-M systems changes gene flow within species. As the diversity of systems increases, cells preferentially exchange genes with those carrying the same R-M. Hence, diversification of R-M systems may result in the fragmentation of gene flow in populations. **III.** MGEs tend to integrate the chromosome at a few hotspots and may become inactivated by mutations resulting in the loss of genes that are not adaptive to the host. The succession of MGE integration and co-option of their defense systems in the hotspots may result in clusters (or islands) of defense systems.