

1 **Misinterpretation of genomic data matters for endangered species listing: The subspecific**
2 **status of the Peñasco least chipmunk (*Neotamias minimus atristriatus*)**

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11 Abstract.—Puckett et al. (2021. *Ecology and Evolution*, 11, 12114-12128) evaluated the
12 subspecies status of the Peñasco least chipmunk (*Neotamias minimus atristriatus*) using genomic
13 approaches and concluded that their results did not support the distinction of this taxon as a
14 subspecies and recommended it be synonymized with *N. m. operarius*. We refute the
15 interpretations, conclusions, and taxonomic recommendations of Puckett et al. (2021), and
16 explain in clearer terms how to interpret genomic analyses for applied management. We identify
17 six conceptual issues that led to misinterpretations and errant recommendations: 1) error in
18 hypothesis testing, 2) overlooking statistical support (or lack thereof) of lineages, 3)
19 inappropriate use of reciprocal monophyly as a criterion for subspecies, 4) importance of
20 geographic isolation and inferences from historical biogeography, 5) diagnosable criteria, and 6)
21 importance of phenotype. We conclude that the data of Puckett et al. (2021) add to information
22 from prior studies providing strong support for recognition of *N. m. atristriatus* as a subspecies.
23 This finding has important and immediate implications for the proposed listing of *N. m.*
24 *atristriatus* as an endangered species under the U.S. Endangered Species Act.

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27 Keywords: conservation genomics, Distinct Population Segment, Endangered Species Act,
28 independent evolutionary trajectory, integrative taxonomy, reciprocal monophyly

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31 Recent decades have seen a surge in technological development of genomic sequencing methods
32 for non-model organisms, along with associated bioinformatic data processing, and downstream
33 evolutionary analyses. In parallel with these advances, genomic data are increasingly being
34 applied to questions of taxonomic validity among wildlife, and towards systematic
35 rearrangements within and among species of conservation concern. These applications may have
36 consequential repercussions for regulatory legislation. It is therefore increasingly critical to
37 bridge the “conservation genomics gap” for more effective exchange of knowledge among
38 genomic researchers, conservation managers, and public stakeholders (Shafer et al. 2015; Taylor
39 et al. 2017; Kadykalo et al. 2020). This will necessitate an improvement in how scientists
40 translate technical jargon for knowledge-users, and will ensure that the scientific interpretation of
41 results accurately reflects the limitations of the genomic data or analyses. As an example of these
42 persistent issues, we discuss the results, interpretation and conservation-related recommendations
43 from a recently published phylogenomic study of chipmunks, which bear on a pending decision
44 of U.S. federal protections under the Endangered Species Act (ESA; Puckett et al. 2021).

46 Least chipmunks (*Neotamias minimus*) are the most widespread species within a diverse
47 mammalian genus collectively distributed throughout western North America (Piaggio and
48 Spicer 2001; Reid et al. 2012). There are currently 21 subspecies of least chipmunks that reflect a
49 complex history of differentiation and diverse ecological and biogeographical associations (Verts
50 and Carraway 2001). The main focus of Puckett et al. (2021) was to evaluate the taxonomic
51 validity of the Peñasco least chipmunk (*N. m. atristriatus*), a geographically isolated subspecies
52 at the southernmost extent of the species' range, and which has been extirpated from most of its
53 historical distribution (Hope and Frey 2000; Frey and Boykin 2007; McKibben and Frey 2020).
54 The taxon was described as a species by Vernon Bailey, a seminal mammalian taxonomist and
55 naturalist, on basis of cranial and pelage characteristics (Bailey 1913). Bailey stated of *Eutamias*
56 *atristriatus* (= *N. m. atristriatus*), that “In cranial characters this chipmunk shows so little
57 similarity to *E. operarius* (= *N. m. operarius*), its apparently nearest relative, that I have given it
58 full specific rank. A thorough revision of the genus may show some other species to which it is
59 more nearly related, but its range is widely separated from that of any other small species”
60 (Bailey 1913:130). This taxon is currently listed as Endangered within the State of New Mexico
61 (NMDGF 2016) and has been proposed to be listed as endangered under the ESA at the
62 taxonomic level of subspecies (USFWS 2021).

63
64 Puckett et al. (2021) performed a series of genomic data analyses of target-captured nuclear
65 exons (DNA that codes for gene functions and which may or may not be subject to selection
66 Luikart et al. 2018). They reported their data as 513 single nucleotide polymorphisms (SNPs) or
67 259 concatenated exon sequence loci, depending on the analysis. Their analyses also considered
68 a mitochondrial genome dataset (maternally inherited haploid DNA). Taxonomic coverage
69 included individuals representing 6 to 12 of the 21 recognized subspecies of least chipmunks
70 (depending on analysis), as well as samples from several other species of chipmunk occurring in
71 the southwestern U.S. Puckett et al. (2021:9-10) stated that their “... genetic data do not support
72 the current distinct subspecies designation for *N. m. atristriatus*.” From this conclusion they
73 made the taxonomic recommendation to synonymize *N. m. atristriatus* with two other subspecies
74 (*N. m. caryi* and *N. m. operarius*), for which *N. m. operarius* has nomenclatural seniority. Their
75 interpretations of results that led to this recommendation included: 1) “...the clustering analyses,
76 nuclear phylogenomic tree, and mitogenome haplotype network *unequivocally* grouped *N. m.*
77 *atristriatus* with *N. m. operarius* and *N. m. caryi* in the southern clade”; and 2) “Neither
78 mitochondrial nor nuclear datasets identified *reciprocally monophyletic* diversity between *N. m.*
79 *atristriatus* and the geographically proximate *N. m. operarius* and *N. m. caryi*” (Puckett et al.
80 2021:10; 13; emphases added by us). We refute the interpretation, principal conclusions, and
81 taxonomic recommendations of Puckett et al. (2021; as outlined above). In this paper we identify
82 six conceptual issues that led to these faulty interpretations and recommendations: 1) error in
83 hypothesis testing, 2) overlooking statistical support (or lack thereof) of lineages, 3)
84 inappropriate use of reciprocal monophyly as a criterion for subspecies, 4) importance of
85 geographic isolation and inferences from historical biogeography, 5) diagnosable criteria, and 6)
86 importance of phenotype. We conclude that the data generated by Puckett et al. (2021) support
87 recognition of *N. m. atristriatus* at the subspecies rank. In addition, we highlight that the
88 conceptual oversights result in incorrect and misleading information for decision makers that can
89 have profound impact on the conservation of taxa. Our primary intention here is to focus on
90 furthering our collective understanding of how genomic analyses and evolutionary relationships
91 should be interpreted, and their limitations for governing changes in infraspecific taxonomy.

92
93 Puckett et al. (2021) accepted a lack of supported evidence for the genetic distinctness of *N. m.*
94 *atristriatus* (e.g., a lack of strong nodal support of evolutionary relationships recovered from a
95 phylogenetic tree) as conclusive evidence for synonymy of the three southern subspecies of least
96 chipmunks (*N. m. atristriatus*, *N. m. operarius*, and *N. m. caryi*; hereafter Southern group). We
97 agree with Puckett et al. (2021) that the evidence indicates that *N. m. atristriatus* is genetically
98 aligned as a member of the Southern group. The relationship between *N. m. atristriatus* and other
99 populations in the Southern Rocky Mountains is expected based on biogeography (Sullivan
100 1985). However, this relationship has no bearing on the subspecific status of *N. atristriatus*. It
101 simply reflects that these individuals share a more recent common ancestor than other
102 populations of *Neotamias*. Second, even if the recovered phylogenetic pattern was consistently
103 well-supported paraphyly or polyphyly among subspecies within the Southern group (i.e.,
104 evolutionary non-independence that suggests either that interbreeding is still occurring or that not
105 enough time has passed for populations to exhibit fixed genetic differences), it would still not be
106 appropriate to invalidate subspecies status. Subspecies are well-established as potentially
107 interbreeding units of analysis and represent taxa on the continuum of the formation of species
108 (Wilson and Brown 1953; Padial et al. 2010; Patton and Conroy 2017). Lack of strong support
109 for a relationship does not signal strong support for the alternative (unless the alternative is
110 strongly supported). The authors did not provide hypotheses or predictions to be tested, but the
111 implicit null hypothesis they tested was that *N. m. atristriatus* is not a valid subspecies. Thus,
112 their interpretation that *N. m. atristriatus* is not distinct from the other members of the Southern
113 group opens them to a classic type II statistical error, wherein they accepted the null hypothesis
114 as true based on the absence of information that the subspecies are different (Patten 2010; Patten
115 and Remsen 2017).

116
117 None of the analyses used to assess distinction of *N. m. atristriatus*, including the mitogenome
118 haplotype network, clustering analyses, and nuclear phylogenomic tree, provide any statistical
119 support for independence or for non-independence. The Splitstree method for mitogenome
120 haplotype network construction does not provide any statistical support for groups (Puckett et al.
121 2021 – Fig. 2), and is therefore only representative of the genetic distance between individuals
122 (Huson et al. 2008); specimens of *N. m. atristriatus* appear to be grouped more closely to each
123 other than to any other individuals of the Southern group, although distance values are not
124 provided. The principal components clustering analyses do not provide K-values for number of
125 clusters or 95% ellipses around discrete groups (Puckett et al. 2021 – Fig. 4). The first two
126 components of this ordination within *N. minimus* only account for 9.3% of the observed genetic
127 variation, indicating considerable variation among these taxa was not reported. The nuclear
128 concatenated phylogenomic tree provides no bootstrap support for any relationship within the
129 Southern group clade or even for monophyly of the Southern group (Puckett et al. 2021 – Fig. 6).
130 Lack of support values means we can draw no conclusions about the strength of relationships
131 among individuals within this clade. Given this ambiguity, we cannot conclude that *N. m.*
132 *atristriatus* is *not* distinct.

133
134 Puckett et al. (2021) viewed the unequivocal grouping of the three Southern subspecies as
135 support for their primary conclusion that lack of reciprocal monophyly between *N. m. atristriatus*
136 and other subspecies justifies synonymy with *N. m. operarius*. However, reciprocal monophyly
137 is an inappropriate criterion for validating subspecies (Braby et al. 2012). Reciprocal monophyly

138 occurs when two or more clades are each monophyletic (genetically unique) with respect to the
139 other, and given the genetic data being analyzed. This condition forms the basis of both the
140 genealogical and phylogenetic species concepts (Wheeler and Meier 2000; de Quieroz 2007). At
141 the genome scale, reciprocal monophyly would be indicative of a lack of gene flow between
142 biological species. By extension, reciprocal monophyly is explicitly not an acceptable criterion
143 for defining subspecies (Patten 2015). Subspecies are characterized by heritable diagnostic traits
144 including morphological or molecular differences that vary in frequency between geographically
145 discrete but potentially interbreeding units of analysis (Hennig 1966; Patten 2010; Patton and
146 Conroy 2017). Thus, gene flow is expected among subspecies, which would result in a lack of
147 reciprocal monophyly (Patten 2010, Patten and Remsen 2017). Put simply, subspecies are not
148 expected to be reciprocally monophyletic. However, interpretation becomes more complicated
149 when considering that the extent of genetic differentiation between closely related taxa depends
150 on multiple factors. The length of time that taxa have diverged from one another is ultimately
151 reflected by how resolved genetic relationships are, but this also depends on both functional and
152 stochastic processes that cause some parts of a genome to resolve as reciprocally monophyletic
153 faster than others (Funk and Omland 2012). The choice of data is therefore consequential for the
154 power to resolve relationships. Phylogeny estimation might recover well-supported reciprocal
155 monophyly between two recognized subspecies from a given genetic locus. Conversely, even
156 fully reproductively isolated species may exhibit a lack of reciprocal monophyly at a given locus
157 due to processes that include incomplete lineage sorting and ancient hybridization, both common
158 phenomena among mammals, and in particular among western chipmunks (Sullivan et al. 2014).
159 As an example, Puckett et al. (2021) did not recover well supported reciprocal monophyly for *N.*
160 *alpinus*, based on their exon data, although this is a recognized species based on more rapidly
161 evolving loci coupled with other diagnostic characters (Rubidge et al. 2014). As such, although
162 reciprocal monophyly is commonly used to indicate evolutionary independence of species under
163 several species concepts, it is not a relevant criterion for assessing the validity of infraspecific
164 taxonomy (Braby et al. 2012, Patten 2015).

165
166 A lack of gene flow between populations can also be achieved simply through strict geographic
167 isolation, and isolation is a key criterion for diagnosing independent evolutionary trajectories
168 (Franklin 1980; Sobel et al. 2010). From its most basic perspective, strict geographic isolation
169 means that inheritance of genotypes from generation to generation, along with epigenetic factors
170 (genotype-environment interactions) and any local adaptive pressures, is not influenced by any
171 immigration and subsequent reproduction of related individuals from separate populations.
172 Given isolation and local environmental conditions, the phenotype of a population will diverge
173 through various evolutionary mechanisms including neutral genetic drift (particularly in small,
174 declining, or demographically unstable populations) and the adaptive processes of natural
175 selection in response to unique and particularly extreme environments. All of these dynamics are
176 reflected by the ecology of *N. m. atristriatus* (Frey and Boykin 2007). Geography and
177 geographic isolation are inextricably linked to the concept and delineation of subspecies
178 (Vignieri et al. 2006, Patten 2010). As such, geographic isolation of *N. m. atristriatus* for an
179 extended timeframe, with evidence from both the divergence time estimates of Puckett et al.
180 (2021) and by the relatively well-resolved zoogeographic history of isolation and connectivity
181 among the southwestern sky islands (e.g., Patterson 1982; Frey et al. 2007; Hope et al. 2016; not
182 discussed by Puckett et al. 2021) constitute primary lines of evidence for uniqueness of this
183 subspecies. *Neotamias m. atristriatus* diverged from other subspecies of the Southern group

184 between 190 thousand years ago (kya; Puckett et al. 2019) and 824 kya (Puckett et al. 2021), two
185 mean divergence estimates based on nuclear species-tree analysis and mitogenome phylogeny
186 reconstruction (under a Yule tree prior), respectively. The predicted distribution of *N. minimus*
187 during the Last Glacial Maximum (~18 kya) also demonstrates isolation of *N. m. atristriatus*
188 from other Southern group subspecies (Puckett et al. 2021 – Fig. 8). All of the evidence
189 presented supports prolonged isolation of *N. m. atristriatus* on an independent evolutionary
190 trajectory. It then may be considered a matter of philosophical differences as to whether such a
191 taxon represents a distinct subspecies (e.g., King et al. 2006; Ramey et al. 2007; Weckworth et
192 al. 2015; Cronin et al. 2015), or indeed a distinct species (de Queiroz 2020). In our view, the
193 question of taxonomic validity of *N. m. atristriatus* therefore becomes: Have extended isolation,
194 unique population ecology, and an associated independent evolutionary trajectory resulted in
195 diagnosability of this subspecies?
196

197 There exists an extended literature on the various definitions of subspecies (for thorough reviews
198 see Haig et al. 2006; Remsen 2010; Patton and Conroy 2017; and citations therein). Puckett et al.
199 (2021) used a definition of subspecies devised specifically for cetaceans by Taylor et al.
200 (2017:174): “...a population, or collection of populations, that appears to be a separately
201 evolving lineage with discontinuities resulting from geography, ecological specialization, or
202 other forces that restrict gene flow to the point that the population or collection of populations is
203 diagnosably distinct.” With regards to diagnosability, Taylor et al. (2017) invoked the definition
204 of Archer et al. (2017:104) as “...a measure of the ability to correctly determine the taxon of a
205 specimen of unknown origin based on a set of distinguishing characteristics”. Subsequently,
206 Puckett et al (2021:11) stated (including emphasis) that “it is unclear what diagnosable, heritable
207 character could be used to correctly determine that a least chipmunk specimen of *unknown origin*
208 was *N. m. atristriatus*.” We argue that origin matters, given that subspecies are geographic
209 variants, and particularly among mammals, subspecies are described based on their geographic
210 distribution coupled with diagnosable characters (Hall 1981; Remsen 2010). Archer et al. (2017)
211 did not provide insight to why diagnosis of cetaceans must be based on a specimen of unknown
212 origin, but we presume that it reflects whale migration, and this is supported by their citation of
213 Brambilla et al. (2010) with reference to diagnosability of migratory bird subspecies. This
214 definition is not relevant for terrestrial non-migratory small mammals. The geographic origin of
215 *N. m. atristriatus*, coupled with morphological or molecular characters, would allow for
216 recognition of this subspecies. For instance, even photographs taken with remote cameras allow
217 for accurate diagnosis of *N. m. atristriatus* when paired with information about location
218 (McKibben and Frey 2021). Origin aside, the statement of unclear diagnosability by Puckett et
219 al. (2021) is misleading to decision-makers. This is coupled by a statement in the introduction
220 that “...considerable scientific uncertainty surrounds the validity of *N. m. atristriatus* as a
221 subspecies” (Puckett et al. 2021:2), which is dismissive of the statistical support for
222 morphological and ecological distinction of this taxon, based on previously published data. We
223 contend that *N. m. atristriatus* is diagnosable via multiple characters investigated since its
224 description: 1) significant genetic differentiation measured by the fixation index (F_{ST} values)
225 between *N. m. atristriatus* and other Southern group subspecies (Puckett et al. 2021:Supporting
226 Information Appendix S1); 2) unique mitochondrial diversity (Puckett et al. 2021:11); 3) unique
227 pelage, unique cranial and bacular morphology, and unique allozymes (genetic variants; Sullivan
228 1985); and 4) unique ecological habitat associations (Sullivan 1985). For context, we also point

229 out that subspecies are based on population level diagnosability, not diagnosability of each
230 individual in a population (Patten 2015). This taxon *is* diagnosable.

231
232 Given the lack of phylogenetic resolution recovered from exon capture data, Puckett et al. (2021)
233 may have benefited by reporting additional analyses with their data, or minimally by discussing
234 shortcomings, leaving the door open for further future analyses that might more accurately test
235 hypotheses of uniqueness for *N. m. atristriatus* (Padiál and Riva 2021). For instance, exon data
236 are known to evolve more slowly than intron data and may not be most suitable for resolving the
237 tips of the tree of life (Bi et al. 2012). Exon data are most beneficial for quantifying adaptive
238 processes (Luikart et al. 2018), including divergence among taxa, through analysis of non-neutral
239 outlier loci, but assessments of this variation were not presented. Finally, from an explicit
240 conservation standpoint, methods have recently been developed for hierarchical assessment and
241 designation of conservation units including not just evolutionary significant units but also
242 management units based on neutral loci and adaptive units based on loci under selection (Funk et
243 al. 2012; Barbosa et al. 2018; Hohenlohe et al. 2021). Although none of these units are
244 considered for mammalian listing under the ESA, they would surely bolster the importance of a
245 recognized subspecies such as *N. m. atristriatus* in the context of the entire species.

246
247 We conclude that, rather than invalidating *N. m. atristriatus*, the results of Puckett et al. (2021)
248 actually augment prior research demonstrating the validity of *N. m. atristriatus* as a subspecies. It
249 has experienced long-term geographic isolation, and it is morphologically, genetically, and
250 ecologically distinctive. We therefore recommend that *N. m. atristriatus* be considered for listing
251 under the ESA at the subspecies level. The misinterpretation of genomic data as we have
252 described can matter for endangered species listing. In some cases taxonomic disputes have
253 ostensibly even been used in attempt to thwart or cast doubt on ESA listings (Vignieri et al.
254 2006). The proposal to list *N. m. atristriatus* as a subspecies under the ESA is currently under a
255 60-day public review period (USFWS 2021), which makes the discussion about validity of its
256 taxonomic status of critical importance. Since its inception, the ESA has always allowed listing
257 of species and subspecies as these are formally recognized taxonomic entities (Haig et al. 2006).
258 More recently, policy has also allowed the listing of Distinct Population Segments (DPSs) of
259 vertebrates. DPSs are defined based on discreteness and importance relative to the remainder of
260 the taxon, which means that interpretation of taxonomy can influence recognition of a DPS (Haig
261 and D'Elia 2010). Thus, although Puckett et al. (2021) promoted their results as strong support
262 for the Sacramento Mountains population forming a unique DPS, their overarching conclusion
263 that *N. m. atristriatus* is an invalid taxon casts doubt on the current evidence presented as a basis
264 for listing. Our account of the various misinterpretations of Puckett et al. (2021) reflects many of
265 the same issues noted from other molecular genetic studies that have tested the validity of
266 subspecies (e.g., Vignieri et al. 2006, Patten 2015). *Neotamias m. atristriatus* is a Linnean
267 trinomial taxon that was described by a professional taxonomist (Bailey 1913) and has been
268 validated by many subsequent analyses of its genetics, morphology, and ecology (Sullivan 1985;
269 Sullivan and Peterson 1988). It therefore has higher priority for listing; DPSs are more prone to
270 litigation and prolonged interpretation that could stall conservation efforts (Haig and D'Elia,
271 2010). Importantly, subspecies are a formal biological rank that may be contested on biological
272 grounds, but cannot be rescinded through legal legislation, as can DPSs (Haig et al. 2006).

273

274 We present this case study in response to a more general rapid expansion of genomic methods
275 for assessing imperiled taxa associated with ESA listing. Such studies are inherently “applied
276 research” and reach multiple stakeholders with variable levels of expertise for interpreting these
277 complex datasets. Importantly, for those stakeholders not accustomed to translating genomic
278 jargon, such data and analyses are not easily associable with their relevance to the ecology,
279 biogeographic history, and contemporary demographic trends of the taxon of interest. Thus,
280 some may rely on the conclusions presented without the knowledge of theory and molecular
281 methods necessary to rigorously decipher data and results. Greater integration among disciplines
282 is imperative (Godfray and Knapp 2004; Padial et al. 2010). Molecular ecologists that have
283 adopted genomic methods should invest in more comprehensive understanding of the biology of
284 the study taxon and system. Studies focused on taxonomic assessments would benefit from
285 collaboration with taxonomists; molecular ecology and taxonomy are not equivalent disciplines
286 (Pruett and Winker 2010). And, extra care should be made to clearly explain what each analysis
287 can or cannot confirm about the question of interest. Decisions by management agencies based
288 on academic interpretations of complex datasets can be consequential for the maintenance of
289 biodiversity. It is therefore equally important that decision makers have the information they
290 need to accurately assess the findings of genomic analyses. We suggest that second opinions
291 from unbiased experts of both the ecology and evolution of the focal taxon could save time and
292 resources towards conservation legislation. Finally, journal editors should assure that data and
293 comprehensive methods associated with taxonomic studies that relate to listing decisions be
294 made available so that they can be evaluated for reproducibility (e.g., Fanelli 2018; Gilbert et al.
295 2021). In addition, journal editors considering these types of studies should not accept for
296 publication interpretations of reciprocal monophyly for qualifying subspecies status. Although
297 we vigorously disagree with their conclusions, Puckett et al. (2021) have provided the first
298 focused genomic assessment of relationships among Southern subspecies of least chipmunks.
299 Their study has quantified divergence of *N. m. atristriatus* based on two genomic datasets and
300 limited sampling of this imperiled subspecies. Undoubtedly, future studies will benefit from their
301 contributions for appropriate protections of declining wildlife.

302
303 Acknowledgements

304
305 We are grateful for critical comments and suggestions provided by K. C. Bell, F. E. McKibben,
306 N. Herrera, B. D. Patterson, and X anonymous reviewer(s).
307

- 308 Archer, F. I., Martien, K. K., & Taylor, B. L. (2017). Diagnosability of mt DNA with Random
309 Forests: Using sequence data to delimit subspecies. *Marine Mammal Science*, 33(S1), 101-
310 131.
- 311 Bailey, V. (1913). Ten new mammals from New Mexico. *Proceedings of the Biological Society*
312 *of Washington*.
- 313 Barbosa, S., Mestre, F., White, T. A., Paupério, J., Alves, P. C., & Searle, J. B. (2018).
314 Integrative approaches to guide conservation decisions: Using genomics to define
315 conservation units and functional corridors. *Molecular Ecology*, 27(17), 3452-3465.
- 316 Bi, K., Vanderpool, D., Singhal, S., Linderoth, T., Moritz, C., & Good, J. M. (2012).
317 Transcriptome-based exon capture enables highly cost-effective comparative genomic data
318 collection at moderate evolutionary scales. *BMC genomics*, 13(1), 1-14.
- 319 Braby, M. F., Eastwood, E., & Murray, N. (2012). The subspecies concept in butterflies: has its
320 application in taxonomy and conservation biology outlived its usefulness? *Biological Journal*
321 *of the Linnean Society*, 106, 699-716.
- 322 Brambilla, M., Vitulano, S., Ferri, A., Spina, F., Fabbri, E., & Randi, E. (2010). What are we
323 dealing with? An explicit test reveals different levels of taxonomical diagnosability in the
324 *Sylvia cantillans* species complex. *Journal of Ornithology*, 151(2), 309-315.
- 325 Cronin, M. A., Cánovas, A., Bannasch, D. L., Oberbauer, A. M., & Medrano, J. F. (2015). Wolf
326 subspecies: Reply to Weckworth et al. and Fredrickson et al. *Journal of Heredity*, 106(4),
327 417-419.
- 328 de Quieroz, K. (2007). Species concepts and species delimitation. *Systematic Biology*, 56, 879-
329 886.
- 330 de Queiroz, K. (2020). An updated concept of subspecies resolves a dispute about the taxonomy
331 of incompletely separated lineages. *Herpetological Review*.
- 332 Fanelli, D. (2018). Opinion: Is science really facing a reproducibility crisis, and do we need it
333 to?. *Proceedings of the National Academy of Sciences*, 115(11), 2628-2631.
- 334 Franklin, I. R. (1980). Evolutionary change in small populations. *Conservation Biology: An*
335 *Evolutionary-Ecological Perspective*, 395.
- 336 Frey, J. K., & Boykin, K. (2007). Status assessment of the Peñasco least chipmunk (*Tamias*
337 *minimus atristriatus*). *Final report submitted to New Mexico Department of Game and Fish,*
338 *Santa Fe, New Mexico*.
- 339 Frey, J. K., Bogan, M. A., & Yates, T. L. (2007). Mountaintop island age determines species
340 richness of boreal mammals in the American Southwest. *Ecography*, 30(2), 231-240.
- 341 Funk, D. J., & Omland, K. E. (2012). Species-level paraphyly and polyphyly: Frequency, casus
342 and consequences, with insights from animal mitochondrial DNA. *Annual Review of Ecology*
343 *and Systematics*, 34, 397-423.
- 344 Funk, W. C., McKay, J. K., Hohenlohe, P. A., & Allendorf, F. W. (2012). Harnessing genomics
345 for delineating conservation units. *Trends in ecology & evolution*, 27(9), 489-496.
- 346 Gilbert, K. J., Andrew, R. L., Bock, D. G., Franklin, M. T., Kane, N. C., Moore, J. S., ... &
347 Vines, T. H. (2012). Recommendations for utilizing and reporting population genetic
348 analyses: the reproducibility of genetic clustering using the program STRUCTURE.
349 *Molecular ecology*, 21(20), 4925-4930.
- 350 Godfray, H. C. J., & Knapp, S. (2004). Introduction. Taxonomy for the twenty-first century.
351 *Philosophical Transactions of the Royal Society B: Biological Sciences*, 359(1444), 559.

- 352 Haig, S. M., Beever, E. A., Chambers, S. M., Draheim, H. M., Dugger, B. D., Dunham, S., ... &
353 Sheffield, L. M. (2006). Taxonomic considerations in listing subspecies under the US
354 Endangered Species Act. *Conservation Biology*, 20(6), 1584-1594.
- 355 Haig, S. M., & D'Elia, J. (2010). Avian subspecies and the U.S. Endangered Species Act.
356 *Ornithological Monographs*, 67 (1), 24-34.
- 357 Hall, E. R. (1981). The mammals of North America. 2nd ed. John Wiley & Sons, Inc., New York
358 1, 1–600.
- 359 Hennig, W. 1966. Phylogenetic systematics. Translated by D. D. David and R. Zangerl.
360 University of Illinois Press, Urbana.
- 361 Hohenlohe, P. A., Funk, W. C., & Rajora, O. P. (2021). Population genomics for wildlife
362 conservation and management. *Molecular Ecology*, 30(1), 62-82.
- 363 Hope, A. G., & Frey, J. K. (2000). Survey for the Peñasco least chipmunk (*Tamias minimus*
364 *atristriatus*) in the Lincoln National Forest with notes on rodent community assemblages.
365 *Final report submitted to Lincoln National Forest, Alamogordo, New Mexico*.
- 366 Hope, A. G., Malaney, J. L., Bell, K. C., Salazar-Miralles, F., Chavez, A. S., Barber, B. R., &
367 Cook, J. A. (2016). Revision of widespread red squirrels (genus: *Tamiasciurus*) highlights the
368 complexity of speciation within North American forests. *Molecular phylogenetics and*
369 *evolution*, 100, 170-182.
- 370 Huson, D. H., Kloepper, T., & Bryant, D. (2008). SplitsTree 4.0-Computation of phylogenetic
371 trees and networks. *Bioinformatics*, 14, 68-73.
- 372 Kadykalo, A. N., Cooke, S. J., & Young, N. (2020). Conservation genomics from a practitioner
373 lens: Evaluating the research-implementation gap in a managed freshwater fishery. *Biological*
374 *Conservation*, 241, 108350.
- 375 King, T. L., Switzer, J. F., Morrison, C. L., Eackles, M. S., Young, C. C., Lubinski, B. A., &
376 Cryan, P. (2006). Comprehensive genetic analyses reveal evolutionary distinction of a mouse
377 (*Zapus hudsonius preblei*) proposed for delisting from the US Endangered Species Act.
378 *Molecular Ecology*, 15(14), 4331-4359.
- 379 Luikart, G., Kardos, M., Hand, B. K., Rajora, O. P., Aitken, S. N., & Hohenlohe, P. A. (2018).
380 Population genomics: Advancing understanding of nature. In *Population genomics* (pp. 3-79).
381 Springer, Cham.
- 382 McKibben, F. E., & Frey, J. K. (2020). Distribution and habitat selection by the Peñasco least
383 chipmunk (*Neotamias minimus atristriatus*). *Final report submitted to New Mexico*
384 *Department of Game and Fish, Santa Fe, New Mexico*.
- 385 McKibben, F. E., & Frey, J. K. (2021). Linking camera traps to taxonomy: identifying
386 photographs of morphologically similar chipmunks. *Ecology and Evolution*, 11, 9741-9764.
- 387 NMDGF (New Mexico Department of Game and Fish). (2016). State Wildlife Action Plan for
388 New Mexico. *New Mexico Department of Game and Fish*.
- 389 Padial, J. M., & De la Riva, I. (2021). A paradigm shift in our view of species drives current
390 trends in biological classification. *Biological Reviews*, 96(2), 731-751.
- 391 Padial, J. M., Miralles, A., De la Riva, I., & Vences, M. (2010). The integrative future of
392 taxonomy. *Frontiers in Zoology*, 7(1), 1-14.
- 393 Patten, M. A. (2010). Chapter 3: Null expectations in subspecies diagnosis. *Ornithological*
394 *Monographs*, 67(1), 35-41.
- 395 Patten, M. A. (2015). Subspecies and the philosophy of science. *The Auk: Ornithological*
396 *Advances*, 132(2), 481-485.

- 397 Patten, M. A., & Remsen Jr, J. V. (2017). Complementary roles of phenotype and genotype in
398 subspecies delimitation. *Journal of Heredity*, 108(4), 462-464.
- 399 Patton, J. L., & Conroy, C. J. (2017). The conundrum of subspecies: morphological diversity
400 among desert populations of the California vole (*Microtus californicus*, Cricetidae). *Journal*
401 *of Mammalogy*, 98(4), 1010-1026.
- 402 Patterson, B. D. (1982). Pleistocene vicariance, montane islands, and the evolutionary
403 divergence of some chipmunks (genus *Eutamias*). *Journal of Mammalogy*, 63(3), 387-398.
- 404 Piaggio, A. J., & Spicer, G. S. (2001). Molecular phylogeny of the chipmunks inferred from
405 mitochondrial cytochrome b and cytochrome oxidase II gene sequences. *Molecular*
406 *Phylogenetics and Evolution*, 20(3), 335-350.
- 407 Pruett, C. L., & Winker, K. (2010). Chapter 13: Alaska song sparrows (*Melospiza melodia*)
408 demonstrate that genetic marker and method of analysis matter in subspecies assessments.
409 *Ornithological Monographs*, 67(1), 162-171.
- 410 Puckett, E. E., Murphy, S. M., & Bradburd, G. (2019). A population genomics assessment of
411 subspecies status and range stability of Peñasco least chipmunk (*Tamias minimus atristriatus*)
412 within the context of range-wide demographic history. *Final report submitted to New Mexico*
413 *Department of Game and Fish, Santa Fe, New Mexico.*
- 414 Puckett, E. E., Murphy, S. M., & Bradburd, G. (2021). Phylogeographic analysis delimits three
415 evolutionary significant units of least chipmunks in North America and identifies unique
416 genetic diversity within the imperiled Peñasco population. *Ecology and Evolution*, 11, 12114-
417 12128.
- 418 Ramey, R. R., Wehausen, J. D., Liu, H. P., Epps, C. W., & Carpenter, L. M. (2007). How King
419 et al. (2006) define an ‘evolutionary distinction’ of a mouse subspecies: A response.
420 *Molecular Ecology*, 16(17), 3518-3521.
- 421 Reid, N., Demboski, J. R., & Sullivan, J. (2012). Phylogeny estimation of the radiation of
422 western North American chipmunks (*Tamias*) in the face of introgression using reproductive
423 protein genes. *Systematic Biology*, 61(1), 44.
- 424 Remsen, J. V. (2010). Subspecies as a meaningful taxonomic rank in Avian classification. Pages
425 *Ornithological Monographs*, 67, 62-78.
- 426 Rubidge, E. M., Patton, J. L., & Moritz, C. (2014). Diversification of the Alpine Chipmunk,
427 *Tamias alpinus*, an alpine endemic of the Sierra Nevada, California. *BMC evolutionary*
428 *biology*, 14(1), 1-16.
- 429 Shafer, A. B., Wolf, J. B., Alves, P. C., Bergström, L., Bruford, M. W., Brännström, I., ... &
430 Zieliński, P. (2015). Genomics and the challenging translation into conservation practice.
431 *Trends in Ecology & Evolution*, 30(2), 78-87.
- 432 Sobel, J. M., Chen, G. F., Watt, L. R., & Schemske, D. W. (2010). The biology of speciation.
433 *Evolution*, 64, 295-315.
- 434 Sullivan, J., Demboski, J. R., Bell, K. C., Hird, S., Sarver, B., Reid, N., & Good, J. M. (2014).
435 Divergence with gene flow within the recent chipmunk radiation (*Tamias*). *Heredity*, 113(3),
436 185-194.
- 437 Sullivan, R. M. (1985). Phyletic, biogeographic, and ecologic relationships among montane
438 populations of least chipmunks (*Eutamias minimus*) in the southwest. *Systematic Zoology*,
439 34(4), 419-448.
- 440 Sullivan, R. M., & Petersen, K. E. (1988). Systematics of southwestern populations of least
441 chipmunks (*Tamias minimus*) reexamined: A synthetic approach.
442 <https://digitalrepository.unm.edu/occasionalpapers/10>.

- 443 Taylor, B. L., Archer, F. I., Martien, K. K., Rosel, P. E., Hancock-Hanser, B. L., Lang, A. R., ...
444 & Baker, C. S. (2017). Guidelines and quantitative standards to improve consistency in
445 cetacean subspecies and species delimitation relying on molecular genetic data. *Marine*
446 *Mammal Science*, 33(S1), 132-155.
- 447 Taylor, H. R., Dussex, N., & van Heezik, Y. (2017). Bridging the conservation genetics gap by
448 identifying barriers to implementation for conservation practitioners. *Global Ecology and*
449 *Conservation*, 10, 231-242.
- 450 USFWS (2021). Endangered and threatened wildlife and plants; Endangered species status for
451 the Peñasco least chipmunk and designation of critical habitat. Federal Register, 86(185),
452 53583-53609.
- 453 Verts, B. J., & Carraway, L. N. (2001). *Tamias minimus*. *Mammalian Species*, 653, 1-10.
- 454 Vignieri, S.N. E.M. Hallerman, B.J. Bergstrom, D.J. Hafner, A.P. Martin, P. Devers, P. Grobler,
455 and N. Hitt. (2006). Mistaken view of taxonomic validity undermines conservation of an
456 evolutionarily distinct mouse: a response to Ramey et al. (2005). *Animal Conservation*, 9,
457 237-243.
- 458 Weckworth, B. V., Dawson, N. G., Talbot, S. L., & Cook, J. A. (2015). Genetic distinctiveness
459 of Alexander Archipelago wolves (*Canis lupus ligoni*). *Journal of Heredity*, 106(4), 412-414.
- 460 Wheeler, Q. D., & Meier, R. (Eds.). (2000). *Species concepts and phylogenetic theory: A debate*.
461 Columbia University Press.
- 462 Wilson, E. O., & Brown Jr, W. L. (1953). The subspecies concept and its taxonomic application.
463 *Systematic Zoology*, 2(3), 97-111.
464