

The effect of dominance rank on female reproductive success in social mammals

Shivani^{1,2}

Elise Huchard³





Dieter Lukas^{2*}

2021-10-09

Affiliations

1. Indian Institute of Science Education and Research Kolkata
2. Department of Human Behavior, Ecology & Culture; Max Planck Institute for Evolutionary Anthropology Leipzig
3. Institut des Sciences de l'Evolution de Montpellier, Centre National de la Recherche Scientifique, Université de Montpellier

Correspondence: * dieter_lukas@eva.mpg.de

Open...  access;  code;  peer review;  data

The background, objectives, predictions, and methods are unchanged from the preregistration that has been pre-study peer reviewed and received an In Principle Recommendation by:



Matthieu Paquet (2020) The effect of dominance rank on female reproductive success in social mammals. *Peer Community in Ecology*, 100056. [10.24072/pci.ecology.100056] (<https://doi.org/10.24072/pci.ecology.100056>) (Reviewers: Bonaventura Majolo and one anonymous reviewer)

The preregistration for this article can be found here: Shivani, Huchard E., Lukas D. 2020. Preregistration - The effect of dominance rank on female reproductive success in social mammals In principle acceptance by *PCI Ecology* of the version 1.2 on 07 July 2020

https://github.com/dieterlukas/FemaleDominanceReproduction_MetaAnalysis/blob/trunk/Preregistration_MetaAnalysis_RankSuccess.Rmd

Abstract

Life in social groups, while potentially providing social benefits, inevitably leads to conflict among group members. In many social mammals, such conflicts lead to the formation of dominance hierarchies, where high-ranking individuals consistently outcompete other group members. Given that competition is a fundamental tenet of the theory of natural selection, it is generally assumed that high-ranking individuals have higher reproductive success than lower-ranking individuals. Previous reviews have indicated large variation across populations on the potential effect of dominance rank on reproductive success in female mammals. Here, we perform a meta-analysis based on 444 effect sizes from 187 studies on 86 mammal species to investigate how life-history, ecology and sociality modulate the relationship between female dominance rank and fitness. We show that (1) dominance rank is generally positively associated with reproductive success, independent of the approach different studies have taken to answer this question; (2) life-history mechanisms mediate the relationship between rank and reproductive success, with higher effects of dominance rank on reproductive output than on survival, particularly in species with high reproductive investment; (3) the fitness benefits to high-ranking females appear consistent across ecological conditions, and (4) instead the social environment consistently mitigates rank differences on reproductive success by modulating female competition.

Background

In order for social groups to persist, group members need to find strategies to deal with the conflicts that inevitably occur (Ward and Webster (2016)). In many female social mammals, conflicts and aggressive interactions are associated with the formation of different types of hierarchies. In singular cooperative breeders, a single dominant breeding female suppresses reproduction in subordinate group members, who rarely fight amongst each other until an opportunity to become dominant opens (Solomon, French, and others (1997)). In many species where multiple breeding females form stable groups, females can be arranged in stable linear hierarchies, where mothers help their daughters to inherit their rank in their matriline (Holekamp and Smale (1991)). In another set of species, hierarchies are more flexible as a female's rank depends on her body size, condition, or availability of coalition partners (Pusey (2012)). Given that, in species in which dominance hierarchies structure social groups, females can always be attributed either a low or a high rank, it has remained unclear whether and when there is selection on females to compete for a high rank or whether selection is on finding a place in the hierarchy.

The prevailing assumption is that high ranking females benefit from their dominant status because outcompeting other females is expected to provide them with priority of access to resources (Ellis (1995), Pusey (2012)). Subordinates are expected to accept their status, because despite having lower reproductive success than dominants, they have few outside options and would presumably face high costs, or have even lower success if they tried to challenge for the dominant status or to reproduce independently (Alexander (1974), Vehrencamp (1983)). An alternative assumption however is that both dominants and subordinates gain from arranging themselves in a hierarchy to avoid the overt fighting that occurs whenever differentially aggressive individuals repeatedly interact (West (1967)). All individuals make a compromise, such that they all balance the potential benefits of their respective positions with the potential costs (Williams (1966)).

Previous reviews have found that while high ranking female mammals frequently appear to have higher reproductive success, there are many populations where such an association has not been found (Pusey

(2012), T. Clutton-Brock and Huchard (2013)). Most studies that brought together the evidence have focused on primates and generally only provided qualitative summaries of the evidence (Fedigan (1983), Ellis (1995), Stockley and Bro-Jørgensen (2011)). One meta-analysis across primates investigated whether life history might mediate the strength of the association between dominance and reproductive success and found that high-ranking females had higher fecundity benefits in species with a longer lifespan (Majolo et al. (2012)). However, there is no systematic assessment of the many potential factors that have been suggested to mitigate the relationship between rank and reproductive success when high rank might not be associated with higher reproductive success.

Objective

In this study, we will perform a quantitative assessment of the strength of the relationship between dominance rank and reproductive success in female social mammals and explore factors that might mediate this relationship. Our objective is to identify the sources and ranges of variation in the relationship between rank and reproductive success and predict that the relationship will be influenced by differences in life-history, ecology, and sociality. We address our objective through the following questions, by testing the corresponding predictions:

1) Does high rank generally lead to higher reproductive success for females in social mammals?

We expect that, overall, high dominance rank has a positive effect on reproductive success.

2) What are the life history traits that mediate the benefits of rank on reproductive success?

We expect that dominants have higher reproductive success predominantly in species in which females have the ability to quickly produce large numbers of offspring.

3) What are the ecological conditions that mediate the benefits of rank on reproductive success?

We expect that differences in reproductive potential will be particularly marked if resources are limited and monopolizable.

4) What are the social circumstances that mediate the benefits of rank?

We expect that the association between dominance rank and reproduction is stronger in species living in more stable and structured social groups.

Predictions

To answer these questions, we assessed the following predictions. All our predictions consider the potential direct influence of a specific variable on the size of the effect of dominance rank on reproductive success. The predictions present the direction of the influence we consider a-priori most likely. We will report all results, but in instances where influences are opposite to what we predict further studies will be necessary to place these results in context. In addition, several of the variables we will include are likely to influence each other. Accordingly, analyses with single variables might not necessarily show the predicted direct influence even if it is present (e.g. there might not be a positive relationship between a social system and

the size of the effects if species with this particular social system primarily occur in environments where the size of the effect is expected to be smaller). While deciphering all the potential relationships among the variables we include is beyond the scope of this study, we will also perform analyses accounting for these potential interactions among variables by performing path analyses. We focus on instances where we expect that one variable might remove or change the direction of the influence of another variable, and present these at the end of the predictions.

1) Does high rank generally lead to higher reproductive success for females in social mammals?

P1.1: Publication bias does not influence our sample of effect sizes.

We do not predict a publication bias but that our sample will include studies showing small effect sizes with small sample sizes. Most studies set out to test if high dominance might lead to both benefits and costs, and previous meta-analyses did not detect signals of publication bias (e.g. Majolo et al. (2012)).

P1.2: Overall, high dominance rank will be associated with higher reproductive success.

We predict that, taking into account the power of the different studies, the combined effect of high rank on reproductive success will be positive. Previous studies that summarized existing evidence (e.g. Majolo et al. (2012), Pusey (2012)) found support for the consensual framework in socio-ecology which argues that high ranking females generally have higher reproductive success than low ranking females.

P1.3 Effect sizes from the same population and the same species will be similar.

We predict that studies that have been conducted on the same species, and in particular at the same site, will report similar effects of dominance rank on reproductive success. For some long-term studies, multiple studies have been performed using slightly different methods and/or data from different years which might include the same set of individuals leading to very similar effect size estimates. For studies of the same species from different sites, we expect similarities because many aspects of the life-history and social system that will shape the relationship between rank and reproductive success will be conserved.

P1.4: Closely related species will show similar effects of dominance rank on reproductive success.

We predict that effect sizes of the relationship between dominance rank and reproductive success will be more similar among closely related species (Chamberlain et al. (2012)) because methodological approaches can be specific to specific Orders (e.g. ungulates are studied differently than primates) and because closely related species share life history, social and ecological traits that might shape the influence of rank on reproductive success.

P1.5: Effect sizes depend on the approach used.

We expect that some of the variation in effect size across studies arises from methodological differences:

- (i) we predict lower effect sizes for studies of captive populations compared to wild populations: while the absence of stochastic events in captivity might mean that dominance is more consistently associated with certain benefits, the effects of high dominance rank on reproductive success will be reduced because of lower competition over resources;
- (ii) we predict lower effect sizes for studies where rank was measured based on agonistic interactions rather than on size or age because size and age are frequently directly associated with differences in female reproduction and clear differences between dominants and subordinates may indicate the

existence of castes that tend to be associated with strong reproductive monopolization (Lukas and Clutton-Brock (2018)); and

- (iii) we predict different effect sizes for studies classifying individuals into two or three rank categories compared to linear ranking depending on the social system. In cases where there is usually a single dominant female (singular cooperative breeders, such as meerkats), using a linear regression between each individuals' rank and its reproductive success will likely estimate a lower effect size because such an approach assumes differences in rank or reproductive success among the subordinates when there are none. In contrast, grouping individuals into categories to compare dominants to subordinates will capture actual differences more accurately. In cases where several females breed (plural breeders, such as hyenas) and are ordered in a linear hierarchy, a linear regression will exploit the full information available on individual differences in rank and reproductive success, whereas grouping individuals will lead to a loss of resolution, at a risk of underestimating the differences between highest and lowest ranking individuals. We performed simulations to determine the extent to which this choice of approach skews the effect sizes and found that it can lead to differences of more than 35% between the true and the estimated effect sizes. For illustration, we include this simulation in our code.

2) What are the life history traits that mediate the benefits of rank on reproductive success?

P2.1: High dominance rank will benefit females more than their offspring.

We predict that high rank is more likely to be associated with higher reproductive success in studies that measured female age at first reproduction, number of offspring born per year or across a lifetime, or female survival rather than the survival of their offspring. While in cooperatively breeding species reproductive suppression might impact offspring survival, in plural breeders offspring survival is more likely to be influenced by factors that are outside of the control of females, such as infanticide by new males (Cheney et al. (2004)).

P2.2: Dominance will have stronger effects on immediate reproductive success in species in which females produce many offspring over a short time period.

One key mechanism that has been proposed is that females with high dominance rank have priority of access to resources during periods when these resources are limited, which in turn can increase their reproductive success. Accordingly, we predict stronger effects of rank on measures of immediate reproductive success (offspring production, offspring survival) in species in which females have higher energetic investment into reproduction, with larger litter sizes and shorter interbirth intervals (Lukas and Huchard (2019)). In contrast, in long-lived species in which females produce only single offspring at long intervals, high-ranking females are expected to have less opportunity to translate short-term resource access into immediate reproductive success but might store energy to potentially increase their own survival or lifetime reproductive success.

3) What are the ecological conditions that mediate the benefits of rank on reproductive success?

P3.1: Positive effects of high dominance rank on reproductive success will be stronger in populations in which females feed on resources that are more monopolizable.

We predict that high rank will have stronger effects on reproductive success in fruit- and meat-eaters compared to herbivores or omnivores. One of the main expected benefits of high rank is priority of access to

resources, which should be more relevant in populations in which resources can be monopolized (Fedigan (1983)).

P3.2: Effects of dominance rank on reproductive success will be more pronounced in populations living in harsh environments.

We predict that the effect of rank on reproductive success will be stronger in populations in which resources are limited because they live in harsh and unpredictable environments. Previous studies have shown that cooperatively breeding species are more likely to occur in such environments (Lukas and Clutton-Brock (2017)), but we also expect stronger effects among plural breeding populations living in harsh environments.

P3.3: Effects of dominance rank on reproductive success will be more pronounced in populations with high densities of individuals.

We predict that the effect of rank on reproductive success will be stronger in populations in which more individuals share a limited amount of space. At higher population densities, social groupings and interactions are more likely and competition over resources is expected to be stronger.

4) What are the social circumstances that mediate the benefits of rank?

P4.1: Benefits of rank will be most pronounced in cooperatively breeding species.

We predict that rank effects on reproduction will be higher in cooperative breeders, where the dominant female is often the only breeding female because she suppresses the reproduction of subordinate females (Digby, Ferrari, and Saltzman (2006)), compared to plural breeders, where aggressive behaviour is more targeted and limited to access over specific resources.

P4.2: For plural-breeders, the time-scales at which the reproductive benefits of dominance accrue depend on how individuals achieve high rank.

We predict that in populations of plural breeders in which groups contain multiple breeding females, the way in which these females compete over dominance will influence the potential benefits of high rank. In populations in which female rank depends primarily on age, high ranking females will have higher reproductive success for short periods of time because changes in rank are expected to occur regularly, and because high rank may only be reached towards the end of their reproductive life (Thouless and Guinness (1986)). In societies in which female rank depends primarily on size or condition, rank effects on reproductive success are expected to be expressed on intermediate time frames, as individuals may not be able to maintain a larger relative size or condition over lifetime but they are expected to acquire rank relatively early in their reproductive life (Giles et al. (2015), Huchard et al. (2016)). In societies in which female rank primarily depends on nepotism, and ranks are often inherited and stable across a female's lifetime, we predict that effects of rank on reproductive success will be strongest when measured over long periods because small benefits might add up to substantial differences among females (Frank (1986)) whereas stochastic events might reduce differences between females on shorter time scales (Cheney et al. (2004)).

P4.3: Dominance rank will have stronger effects on reproductive success in populations in which females are philopatric in comparison to populations where females disperse to breed.

We predict that effects of rank on reproductive success will be lower in populations in which adult females are able to leave their group and join other groups compared to populations in which females cannot breed

outside their natal group. In populations in which females are philopatric, they are likely to have support from female kin which can strengthen dominance differences (Lukas and Clutton-Brock (2018)). In addition, in species where females can change group membership easily, females are expected to join those groups where they have the best breeding option available to them (Vehrencamp (1983)).

P4.4: In plural breeding species, dominance will have stronger effects on reproductive success when the number of females in the group is smaller.

We predict that the effect of rank on reproductive success will be stronger in plural breeding populations in which there are fewer females per group, because dominant females will be more likely to interfere in reproductive attempts when there are fewer subordinates (T. H. Clutton-Brock et al. (2010) and because increased competition in larger groups is expected to reduce reproductive success even among dominants (Van Noordwijk and Van Schaik (1988)).

P4.5 Dominance rank will be more strongly associated with reproductive success in populations in which average relatedness among female group members is high.

We predict that the relationship between dominance rank and reproductive success will be more pronounced in species in which social groups primarily consist of close kin compared to groups composed of unrelated females. Groups with high levels of average kinship among females are those where groups are small, females remain philopatric (Lukas et al. (2005)), and females have support to establish their positions (Lukas and Clutton-Brock (2018)), which all are expected to lead to higher benefits of high rank.

P4.6 Dominance rank will be more strongly associated with reproductive success in populations in which variance in relatedness among female group members is high.

In addition to levels of average relatedness among group females, we also predict that the relationship between dominance rank and reproductive success will be more pronounced in species in which there is high variance in relatedness, with females being closely related to some group members but not to others, as compared to species in which group females are either all related or all unrelated. In several species with female philopatry, groups are structured into matriline (Fortunato (2019)). Members of the same matriline tend to support each other in interactions with unrelated females, likely reinforcing differences among females.

P4.7 The effect of dominance on reproductive success will be less pronounced in populations in which females regularly form coalitions.

We predict that high ranking females will have less pronounced reproductive benefits in species in which females form strategic coalitions with others (Bercovitch (1991)). Individuals have been suggested to form strategic coalitions to level the reproduction of others (Pandit and Schaik (2003)) and these coalitions are less likely in cooperatively breeding species (Lukas and Clutton-Brock (2018)).

P4.8 Dominance rank will have less effect on reproductive success in populations in which there is intense inter-sexual conflict.

We predict that the association between high dominance rank and increased reproductive success of females will be lower in populations in which males compete intensively over reproductive opportunities because this leads to intersexual conflict that harms female fitness (Swedell et al. (2014)). In such populations, males tend to be aggressive towards females and males taking up tenure in a group tend to kill offspring indiscriminately or might even target offspring of high-ranking females (Fedigan and Jack (2013)), reducing any potential differences between high- and low-ranking females. We will assess whether high ranking

females benefit less from their positions in populations in which groups show strong female-biased sex composition, or in which males regularly commit infanticide, or with strong sexual size dimorphism with males being much larger than females.

5) Potential interactions among predictor variables

We expect potential interactions among the predictor variables because some of them might influence each other while others might potentially modulate the influence of another predictor variable on the dominance effects. The following six predictions were those we added in the preregistration. We added further analyses based on the outcome of the single-factor analyses. These are listed in the changes from the preregistration section.

Studies performed on wild versus captive individuals and using different measures of reproductive success might not only differ in the overall strength of the effect of rank on reproductive success, but also in how other variables influence this effect.

Higher population density [predicted to lead to larger effect sizes] might be associated with larger group sizes [smaller effect sizes predicted], leading to an interactive influence on the strength of the effect sizes of dominance rank on reproductive success.

Smaller group sizes [larger effect sizes predicted] might be associated with more intense intersexual conflict [smaller effect sizes predicted], leading to an interactive influence on the strength of the effect sizes of dominance rank on reproductive success.

Monopolizable resources [larger effect sizes predicted] might be associated with reduced population density [smaller effect sizes predicted]), leading to an interactive influence on the strength of the effect sizes of dominance rank on reproductive success.

Environmental harshness [larger effect sizes predicted] might be associated with reduced population density [smaller effect sizes predicted]), leading to an interactive influence on the strength of the effect sizes of dominance rank on reproductive success.

Female philopatry [larger effect sizes predicted] might be associated with increased group sizes [smaller effect sizes predicted]), leading to an interactive influence on the strength of the effect sizes of dominance rank on reproductive success.

Methods

Literature search The literature search was performed by S & DL. We started with the references in the previous major reviews and meta-analyses on the association between dominance and reproduction in female mammals (see below for inclusion criteria): Fedigan (1983) (8 studies on female primates entered), Ellis (1995) (16 studies entered / 5 studies not entered on female non-primates, 38 studies entered / 22 studies not entered on female primates), Brown and Silk (2002) (28 studies entered / 7 studies not entered on female primates), Stockley and Bro-Jørgensen (2011) (12 studies entered / 2 studies not entered on female non-primates, 11 studies entered / 1 study not entered on female primates), Majolo et al. (2012) (26

studies entered / 2 studies not entered on female primates), Pusey (2012) (45 studies entered / 2 studies not entered on female primates), and T. Clutton-Brock and Huchard (2013) (8 studies entered / 1 study not entered on female primates, 6 studies entered / 1 study not entered on female non-primates). Next, we performed database searches in Google Scholar and Pubmed, first by identifying articles citing these major reviews and next by searching with the terms “dominance, reproductive success/reproduction, female, mammal,” and “rank, reproductive success/reproduction, female, mammal,” “sex ratio, dominance, female, mammal” (searches performed July 2019-January 2020). We limited our checks to the first 1000 results for all searches.

We checked the titles and abstracts to identify studies that observed dominance interactions and reproductive success in social groups of interacting female non-human mammals. We selected studies that measured the association between dominance rank and at least one aspect of female reproductive success and reported the data or a test-statistic. For both dominance and reproductive success, we only included studies that had direct measures, not secondary indicators. For dominance, we excluded studies where authors did not explicitly determine dominance relationships and only assumed that traits such as size, presence in core areas, or reproductive success itself indicate dominance. We did however include studies where authors established dominance hierarchies, found that they are associated with some other trait such as size or condition, and subsequently used the other trait to measure dominance. For reproductive success, we excluded studies that measured traits such as mating frequency or access to food resources which were assumed but not known to influence reproductive success (excluding studies that: measured the size of individuals to argue about dominance; assumed that females in core areas are dominant; assigned dominance to females based on how successful they are; recorded mating success not reproductive success; linked dominance to behaviour assumed to potentially link to reproductive success). We included all kinds of academic publications, from primary articles published in peer-reviewed journals through reviews, books and book chapters, and unpublished PhD theses.

Variables, their definitions, and their sources

Variables coded directly from the relevant publications:

All data from the literature search on publications reporting the effect of dominance rank on reproductive success was entered prior to the first submission of the preregistration. S and DL performed the data extraction. We initially coded eight papers independently, for which we both extracted the same values and classified the approaches in the same way. We extracted the relevant information to calculate the effect size and its associated variance. In addition, we coded a set of variables to characterize the methodological approach. The dataset contains 444 effect sizes from 187 studies on 86 mammalian species. A copy of the datafile is available on github

Z-transformed effect size: we converted all effect sizes to Z-transformed correlation coefficients (Z_r). In cases where articles reported a pairwise correlation coefficient, we directly use this value. In cases where authors had used alternative statistical approaches (e.g. t-test comparison between two groups of individuals), the test statistics were converted to the statistic ‘r’ using formulas provided by Lakens (2013), Lajeunesse et al. (2013), and Wilson (2019). In cases where authors reported individual-level data reflecting dominance rank and reproductive success (for example in the form of a table that listed for groups of dominants and subordinates their mean and deviation of reproductive success or for every individual their rank and reproductive success), we calculated correlation coefficients directly from a 2-by-2 frequency table (when comparing classes of high- to low-ranking individuals) or from linear regressions (when individuals had continuous ranks). In cases where studies simply stated that “all dominants bred but none of the subordinates”

we assumed an error of 0.5% for both dominants not breeding and subordinates breeding to obtain the sampling variance estimates. We extracted separate effect sizes for each reported analysis: for example, if authors reported separately associations between dominance rank and mortality of offspring to 1 year and to independence, we obtained two effect sizes from this population reflecting infant survival. We Z-transformed all correlation coefficients to control for the asymptotic distribution of these values. We changed the sign of the effect sizes to make them consistent across studies. This was necessary because dominance rank was coded differently across studies, for example sometimes studies assigned dominant individuals the lowest value by starting a count from 1, whereas in other cases they were assigned the highest value to reflect the proportion of other females they are dominant over. We set the sign of effect sizes such that positive values mean that higher ranking individuals have shorter interbirth intervals, higher survival as adults and of their infants, higher infant production (e.g. larger litter sizes, higher probability of breeding), and higher lifetime reproductive success (e.g. higher total number of offspring weaned).

Sample size: we recorded the sample size for the relevant statistical comparison (number of females, number of offspring, number of matriline etc.).

Sampling variance: we calculated the sampling variance of the effect sizes based on the correlation coefficient r and the sample size, using the formulas provided by Wilson (2019). The standard error, which is alternatively used in some approaches, is the square root of the sampling variance (Viechtbauer (2010)).

Species identity: we recorded the common name and the latin species name as listed by the authors. We referred to the Mammal Diversity Database (Burgin et al. (2018)) to resolve instances where species attributions had been changed since the publication of the original study.

Study site: we recorded the name of the study site as listed by the authors in the method section. The focus of this variable is to determine whether multiple observations are from the same species from the same study population, and we accordingly assigned different names for the study site label in case two or more different species had been studied at the same site.

Measure of reproductive success: we recorded which aspect of reproduction dominance rank was associated with. We classified reproductive traits into six classes: - age at first reproduction (includes age at first birth, age at first conception, age at first menstrual cycle); - infant survival (includes rates of mortality of offspring prior to their independence; proportion of pregnancies carried to birth); - survival (includes rates of mortality of females per year, age at death); - infant production (includes litter size, offspring weight, litter mass, number of offspring per year, probability of birth in a given year, number of surviving infants per year); - interbirth interval (includes time between life births, number of cycles to conception, number of litters per year); - lifetime reproductive success (includes total number of offspring born or surviving to independence for females who had been observed from first reproduction to death).

Classification of rank: we recorded the approach the authors had used to assign dominance positions to individuals, distinguishing between those based on aggressive/submissive interactions between pairs of individuals and those based on other traits such as age, size, or which female was the first to reproduce.

Scoring of rank: we recorded whether in the analyses individuals were assigned a specific, continuous rank position or whether individuals were classified into rank categories (dominant versus subordinates, high- versus middle- versus low-ranking).

Duration of study: we recorded the number of years that authors had observed the individuals (anything less than one year was assigned a value of 1).

Population type: we recorded whether the population was free-living, provisioned, or captive based on the authors descriptions.

Social group size: we recorded the average number of adult females per group in the study population, based on the information provided in the manuscripts. We relied on the definition of a social group as used by the respective authors, which might include associations of females in: singular-breeder cooperative groups (as in wolves or meerkats); stable groups of multiple breeding females (as in baboons or hyenas); or breeding associations defined by physical proximity (as in bighorn sheep or antelopes). We will have a separate coding of the social system (see below). Where available, we also coded the average number of adult males associated with each group of females to determine the sex ratio in social groups as a proxy for intersexual conflict.

Variables extracted from the broader literature for each species/population:

The following data were added prior to the analyses. For most of these, we extracted information from the relevant papers or publications reporting on the same population. For some of these, we used previously published species' averages, because records from each population for each specific period during which the effect of dominance rank on reproductive success were measured were not available for a large enough sample. We list sources we used to obtain these data.

Litter size: the number of offspring per birth; data available for each population, we used the average as reported by the authors (based on the data in Jones et al. (2009)).

Interbirth interval: the time in months between consecutive births; data available for a limited set of populations, we used the average as reported by the authors. Given that population specific data was available for only a very limited subset, we added species-level averages (based on the data in Jones et al. (2009)).

Maximum lifespan: the maximum time in months that an individual of that species has been recorded to live for (based on the data in Jones et al. (2009)).

Cooperative breeding group: whether social groups usually contain a single breeding female and additional non-breeding adult females that help to raise the offspring of the breeding female. Group membership for females is usually closed and changes occur through birth and death or fissioning of existing groups. This classification is in contrast to plural breeding groups and breeding associations (see below); data available for each population, we used the description of the social system in the population as reported by the authors.

Plural breeding group: whether social groups usually contain multiple breeding females that remain together for extended periods of time. It includes both groups in which females are philopatric or disperse. Females form differentiated relationships with other group members. This classification is in contrast to cooperative breeding groups and breeding associations (see above/below); data available for each population, we used the description of the social system in the population as reported by the authors.

Breeding association: whether social groups consist of multiple breeding females that associate either in space or by mutual attraction. Group membership is fluid and associations among individuals can rapidly change. This classification is in contrast to cooperative breeding groups and plural breeding groups (see above); data available for each population, we will use the description of the social system in the population as reported by the authors.

Dominance system: whether dominance rank of females appears to depend primarily on (i) their age, (ii) their physical attributes such as body size, (iii) support from their mother, or (iv) coalitionary support from same-aged group members. Data available from a subset of populations, to which we added data

from primary reports of species-level classifications from other populations assuming that this trait is usually stable across populations within species (references listed in the data file).

Philopatry: whether females have the majority of their offspring in the same social groups or in the same location in which they have been born or whether females disperse to other groups or locations to reproduce; data from species-level descriptions of female behaviour (based on the data in Barsbai, Lukas, and Ponderfer (2021)).

Monopolizable resources: whether the gross dietary category of a species is based on monopolizable resources (carnivory, frugivory), or non-monopolizable resources (herbivory, or omnivory) (based on the data in Wilman et al. (2014)).

Environmental harshness: whether the average climatic conditions experienced by the species are characterized by cold temperatures, low rainfall, and unpredictability (based on the data and principal components summarizing climate data in Botero et al. (2014)).

Population density: the average number of individuals per square kilometer for the species (based on the data in Jones et al. (2009)).

Average and variance in relatedness among group females: the average and variance in relatedness measured using genetic approaches among adult females within the same group as reported for this species; data available from a subset of the populations (references listed in the data file).

Coalition formation: whether adult females form coalitions with other female group members to support each other during within-group aggressive interactions; data from species-level descriptions of female behaviour (based on the data in Lukas and Clutton-Brock (2018)).

Sexual dimorphism in body weight: we calculated sexual dimorphism following the two step approach of Smith (1999) as the average weight of males divided by average weight of females if males are heavier than females and as 2 minus the average weight of females divided by the average weight of males otherwise (based on data in: Jarman (1983), Loison et al. (1999), Smith and Cheverud (2002), Isaac (2005), and Kappeler et al. (2019))

Male infanticide: whether adult males in that species kill offspring (based on the data in Lukas and Huchard (2014)).

Adult sex ratio: the ratio of the average number of adult males divided by the sum of the average number of females and males per social group of that species. We took species' averages to reflect adaptation to likely levels of potential sexual conflict because several of the studies from which we extracted effect sizes had captive or experimental settings or only reported the number of females that were included in the study (based on the data in Barsbai, Lukas, and Ponderfer (2021)).

Phylogeny We generated a single consensus phylogeny for the mammalian species in our sample from the most recent complete mammalian time-calibrated phylogeny (Upham, Esselstyn, and Jetz (2019)). We downloaded a credible set of 1000 trees of mammalian phylogenetic history from vertlife.org/phylosubsets/ (July 2020) and used TreeAnnotator (version 1.8.2 in BEAST: Drummond et al. (2012)) to generate a maximum clade credibility (MCC) tree (median node heights and a burn in of 250 trees). We trimmed the tree to match the species in our sample (in one instance using a close relative, */Canis lupus/* instead of */Canis familiaris/*) and converted branch lengths using functions of the package ape (Paradis and Schliep (2019)).

Analyses We performed all analyses in the statistical software R (R Software Consortium 2019). We built separate models for each prediction. To assess the robustness of the findings and whether modeling decisions might have an influence on our results, we used a frequentist and a Bayesian approach to build the statistical models. We first estimated all models using functions in the package *metafor* (Viechtbauer (2010)). We fit meta-analytic multilevel mixed-effects models with moderators via linear models, including models that account for the potential correlations among effect sizes due to shared phylogenetic history among species (Nakagawa and Santos (2012)). Second, we estimated relationships with Bayesian approaches as implemented in the package *rethinking* (McElreath (2020)). For the Bayesian models, we fit multilevel models that include the sampling variance as measurement error (Kurz (2019)) and the shared phylogenetic history as a covariance matrix. Weakly regularizing priors are used for all parameters. The models are implemented in Stan. We drew 8000 samples from four chains, checking that for each the Gelman-Rubin convergence diagnostic 'R-hat' values are less than 1.01 indicating that the Markov chains have converged towards the final estimates. Visual inspection of trace plots and rank histograms were performed to ensure that they indicated no evidence of divergent transitions or biased posterior exploration. Posteriors from the model were used to generate estimates of the overall effect size and the influence of potential moderators. We detail model construction in the following: we first assess whether species and population identity create dependencies amongst the measured effect sizes. If so, we include these factors through covariance matrices reflecting the dependence across measurements. We determined whether a variable had a relationship with the variation in the effect of dominance rank on reproductive success when the compatibility interval of the estimated association did not cross zero (continuous variable) or the contrast between levels does not cross zero (categorical variable), indicating that the model estimates that our data shows a consistent positive/negative association. We provide all code showing the setup of the various models and the plots, the input files containing the data and phylogeny, as well as a simulated dataset with the same structure as the actual data on which we assessed our models in the preregistration in the linked github repository

Preregistration

We preregistered our hypotheses, methods, and analysis plans: https://dieterlukas.github.io/Preregistration_MetaAnalysis_RankSuccess.html

The literature search was completed before the first submission of the preregistration. All variables that were coded directly from the source publications (Z transformed effect size, variance, sample size, species identity, aspect of reproductive success, classification of rank, duration of study, population type, and social group size) were also entered prior to the first submission. In July 2019, S worked with a preliminary subset of the data (143 effect sizes), and investigated publication bias, the overall mean and variance in effect sizes, and whether effect sizes differed according to which reproductive output was measured. We added the data on the explanatory variables and started analyses in July 2020 after the preregistration passed pre-study peer review at *Peer Community In Ecology*: Paquet (2020) *Peer Community in Ecology*, 100056. [10.24072/pci.ecology.100056] (<https://doi.org/10.24072/pci.ecology.100056>)

We collected data on the additional explanatory variables: * litter size, litters per year, and population density for the respective species * cooperative vs plural vs associate breeding from the descriptions in the respective population from the articles from which we obtained the effect sizes * dominance system from additional references on the species * philopatry of the respective species * diet category of the respective species * environmental harshness across the range of the respective species * coalition formation in the

respective species * sexual dimorphism in body weight * male infanticide * sex ratio among adult group members * average relatedness from the articles from which we obtained the effect sizes or additional references matching the exact population * we did not collect data on variance in relatedness because it was not possible to extract this information from most studies reporting relatedness levels

Changes from preregistration

Additional variables: We added data on the maximum lifespan of species to address Prediction 4.2. We realized that, whether a study should be considered short- or long-term, depends on the lifespan of the species. We used the information on the number of years a study had been conducted together with the maximum lifespan data to calculate the relative duration of a study.

We added data on the dominance style of macaque species after noting that a large proportion of our sample reflects these species. Across macaque species, dominance interactions among females in a group have been assigned into one of four grades, ranging from egalitarian species in Grade 1 to highly despotic species in Grade 4. We predicted that effect sizes of dominance rank on reproductive success would be larger in species characterized as more despotic, with steeper dominance hierarchies and more asymmetries in social interactions (Prediction 4.9). We extracted the data for the species in our sample from Balasubramaniam et al. (2012)

We changed how we calculated sexual dimorphism in body weight.

Outlier check: Before running the analyses, we made a funnel plot of the standard error over the effect size, where we noticed three outlier data points. We realized that for these three entries (EffectRefs 425, 427, and 428) we had used the wrong formula to calculate the effect size and variance. All of these are studies of multiple groups of *Callithrix jacchus*, each with a small number of females. For these three studies, we had erroneously used the 2-by-2 frequency tables to calculate the standardized mean difference, not the correlation coefficient. We corrected the values for these three entries before performing any of the analyses.

Sampling bias: The funnel plot of the complete dataset showed a strong asymmetry, indicating that our sample is biased towards including many studies with low precision and high positive effect sizes. To better illustrate this sample bias, we used a different way to plot the data (Nakagawa, Lagisz, O’Dea, et al. (2021)) that was suggested after we had written our preregistration. We added further analyses to investigate the potential causes of the bias in our sample, both based on functions in the packages ‘metafor’ (following Nakagawa, Lagisz, Jennions, et al. (2021)) and ‘rethinking’ (following McElreath (2020)), to determine the potential causes of the bias in our sample and the influence on what effects should be expected in new samples.

Multivariate analyses: We constructed the multivariate analyses after completing the univariate analyses. We did not perform the multivariate analyses we had listed in the preregistration where the univariate analyses indicated no influence/interaction (group size + intersexual conflict; diet + population density; harshness + population density). We added a set of multivariate analyses after finding that cooperative breeders have very different effect sizes of dominance rank on female reproductive success than plural/associated breeders to determine how this difference between breeding systems might relate to the influence of some of the additional social variables we included.

Results

We extracted 444 effect sizes of the relationship between dominance rank and reproductive success of female mammals from 187 studies on 86 species during our literature search. More than half of the effect sizes are from primate species (253 effect sizes), with macaques (109) and baboons (76) a particular focus for this research. About two thirds (283) of the reports are from wild populations; rank was predominantly determined on the basis of aggressive interactions (407) rather than on other measures such as age or size (37); and it was about equally frequent that researchers classified rank categorically as dominant versus subordinate (251) than continuously from highest to lowest (193). Most of the reported effects link dominance rank to infant production (198) followed by infant survival (113), with fewer effects reported on interbirth intervals (46), lifetime reproductive success (34), survival (30), or age at first reproduction (23).

1) Does high rank generally lead to higher reproductive success for females in social mammals?

R1.1 Sample bias: A visual inspection of the range of effect sizes at different sample sizes in a funnel plot (Figure 8a) showed that there might be an underrepresentation of studies with small or negative effect sizes and small sample sizes (Egger et al. (1997)). This sample bias is clearer to see in an orchard plot, which shows that extreme effect sizes tend to be of low precision and that there is an overrepresentation of positive effect sizes (Figure 1).

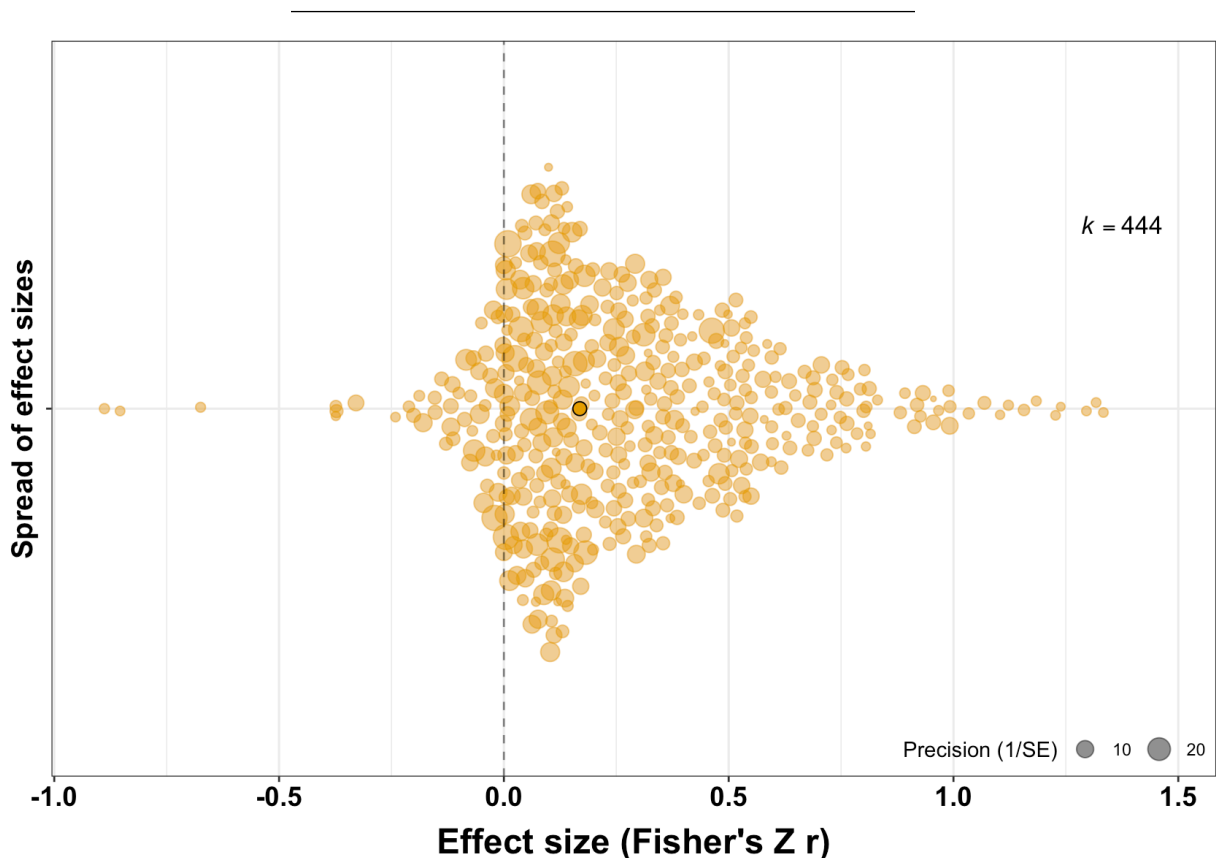


Figure 1. Orchard plot displaying the spread of the 444 effect sizes in our sample (each dot represents a single effect size, the size of the dot indicates the precision). Overall, most studies report a positive

association between dominance rank and reproductive success (darker circle in the center indicates the mean). Our sample does show bias, with effect sizes not distributed symmetrical around the center but showing an overrepresentation of highly positive values.

There are potentially (at least) three sources of sample bias, the first being 'publication bias' with studies with low effect sizes (not reaching traditional levels of significance) not ending up in the published literature, the second being 'study system bias' with research focusing on populations where it is easy to detect effects (e.g. cooperative breeders), and the third being 'study time bias' with studies performed over shorter time frames generally being more imprecise. We added further post-hoc analyses to investigate these patterns individually here, and in combined models after identifying which study systems might show different effect sizes (section R5.1).

Simple tests for 'publication bias' (Preston, Ashby, and Smyth (2004)) suggest that effect sizes with a p-value smaller than 0.05 are about four times more likely to be reported than effect sizes with a p-value larger than 0.50.

As a further indication of 'publication bias,' we find that studies with small sample sizes and small effect sizes (those that presumably did not reach statistical significance) are missing in our dataset such that the average effect sizes at smaller sample sizes are more extreme than those at larger sample sizes (estimate of sample size on effect sizes metafor -0.03 - -0.02, rethinking -0.09 - -0.04) (Figure 2). Nevertheless, the estimated overall effect size in this model remains consistently larger than zero, indicating that even after including any missing studies with small or negative effect sizes there would still be on average a positive relationship between dominance rank and female reproductive success across studies.

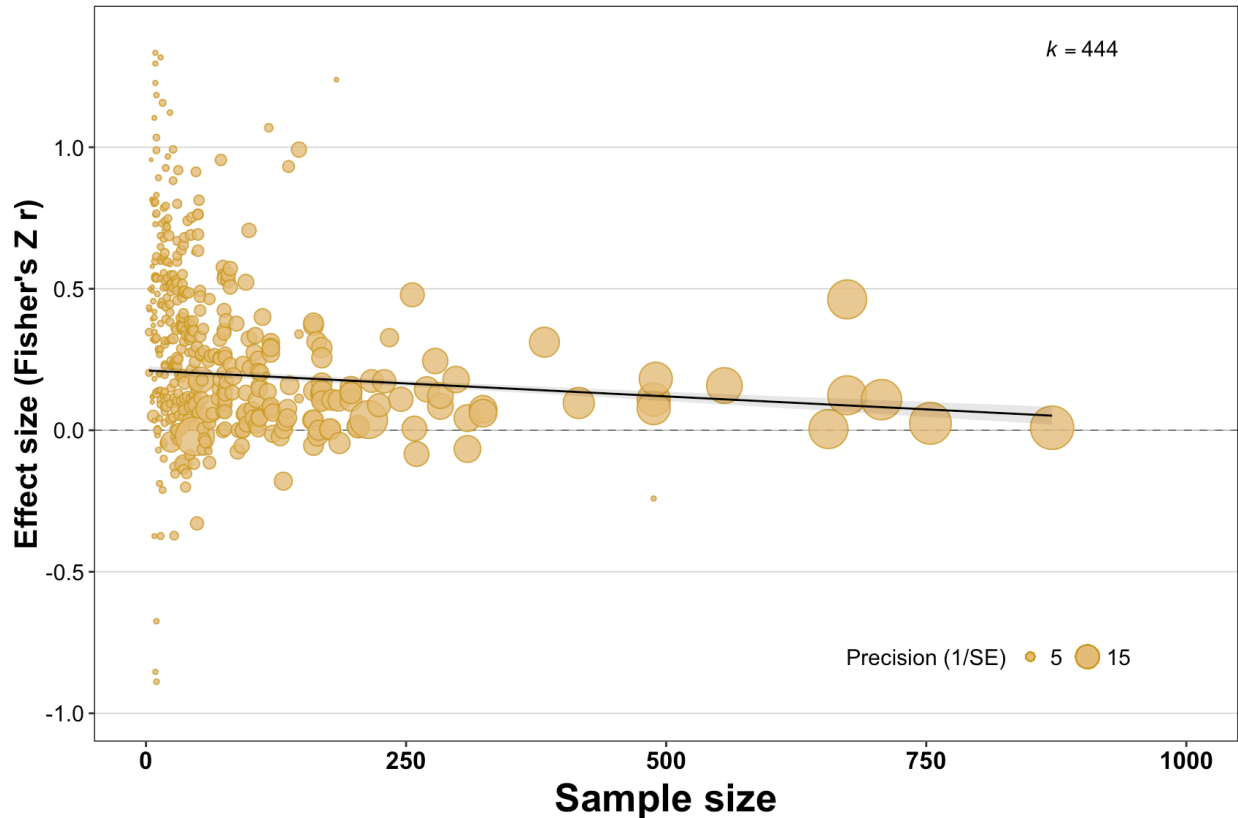


Figure 2. Relationship between the measured size of the effect of dominance rank on female reproductive success and the sample size of the study. Studies with smaller sample sizes show more extreme effect sizes, and also indications of potential publication bias as there are more extremely positive values than what would be expected based on the average effect sizes of studies with larger sample sizes.

Our data also shows indication that the sample bias might result from 'study system bias,' because these base analyses indicate high heterogeneity in our sample (total heterogeneity / total variability: 73.37%). Given the diversity of studies in our sample, we did not expect that the effect sizes represent a sample from a single distribution: for example, studies of offspring mortality tend to have larger sample sizes (because each mother can have multiple offspring) and we predict different effect sizes for these studies. Sections R2 - R4 present the specific analyses for each prediction to assess each of the factors potentially leading to differences between effect size estimates, and we combine them in section R5.1.

Finally, including the number of years a study had been conducted for as a predictor of the effect sizes also indicates that our sample shows 'study time bias.' Effect sizes are lower when studies have been conducted for longer (metafor estimate -0.01 - 0.00, rethinking estimate -0.05 - 0.00), but in particular the variance is reduced once a study has been running for 10 or more years (Figure 3).

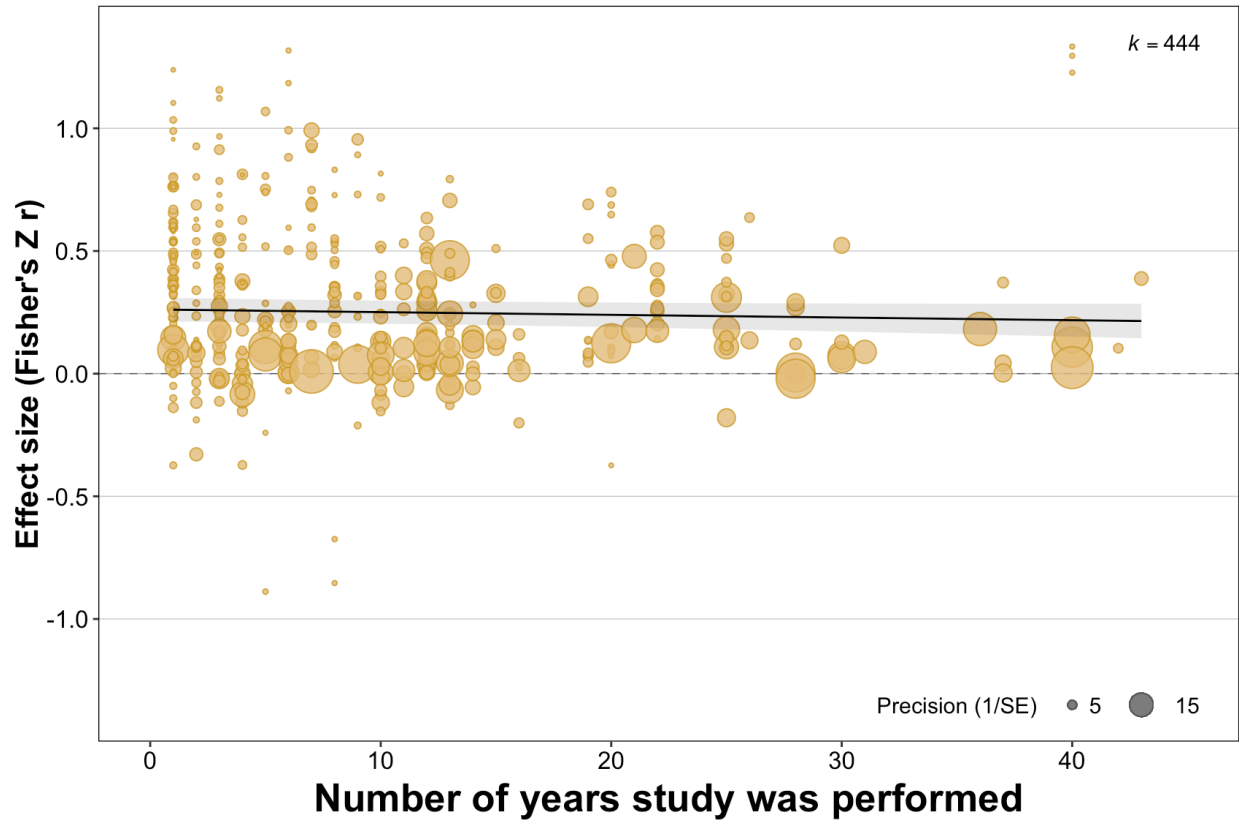


Figure 3. Relationship between the measured size of the effect of dominance rank on female reproductive success and the length a study was conducted for. Studies that have been conducted for 10 or more years tend to have higher precision (larger circle) and tend to be closer to the overall mean.

R1.2 Overall effect: We constructed an intercept-only meta-analytic base model to test for a general effect of dominance rank on reproductive success. Across our sample, there is a strong effect that females with higher dominance rank have higher reproductive success (metafor estimate +0.22 - +0.27, rethinking estimate +0.26 - +0.30; the metafor estimate here and in the additional models is lower than the rethinking estimate because the statistical approach of the former expects the data to be more symmetrical than they are). This overall effect means, for example, that in groups with two individuals dominants would have between 0-6 offspring while subordinates have between 0-4 offspring. There is large variation though in our sample, with effect sizes ranging from -0.89 - +1.33 (Figure 1).

R1.3 Influence of locality/species: To the base model, we added random effects to account for non-independence due to effect sizes originating from within the same study, from studies performed on the same population and on the same species. The estimate of the overall effect size did not change in this model (metafor estimate +0.22 - +0.31, rethinking estimate +0.26 - +0.35). Effect sizes from the same species and the same study, but not the same population, tend to be similar to each other. The absence of a population effect could be because there are only very few observations in our dataset of the same population taken in different studies where there are also observations from multiple additional populations of

the same species. Alternatively, it could be that effects do not vary across populations of the same species, which is also indicated by the absence of differences between wild and captive populations (see below).

R1.4 Influence of phylogeny: To the random effects model, we added a covariance structure to reflect potential similarities in effect sizes arising from closely related species showing similar effects due to their shared phylogenetic history. Both statistical approaches indicate that closely related species tend to have effect sizes that are more similar than those of distantly related species. The metafor approach suggests that about 20% of the variation in effect sizes is associated with covariation among species. The rethinking approach shows high uncertainty in the estimates (Figure 4), reflecting the high heterogeneity in the underlying data with high variation within species and different measures taken among closely related species. It suggests that species of the same genus tend to have similar effect sizes and that shared phylogenetic history might also explain similarities in effect sizes among species in the same Order, but covariance estimates are close to zero for species pairs that are more distantly related (Figure 4; the highest standardized distance between any pair of species in the same Order is 0.40).

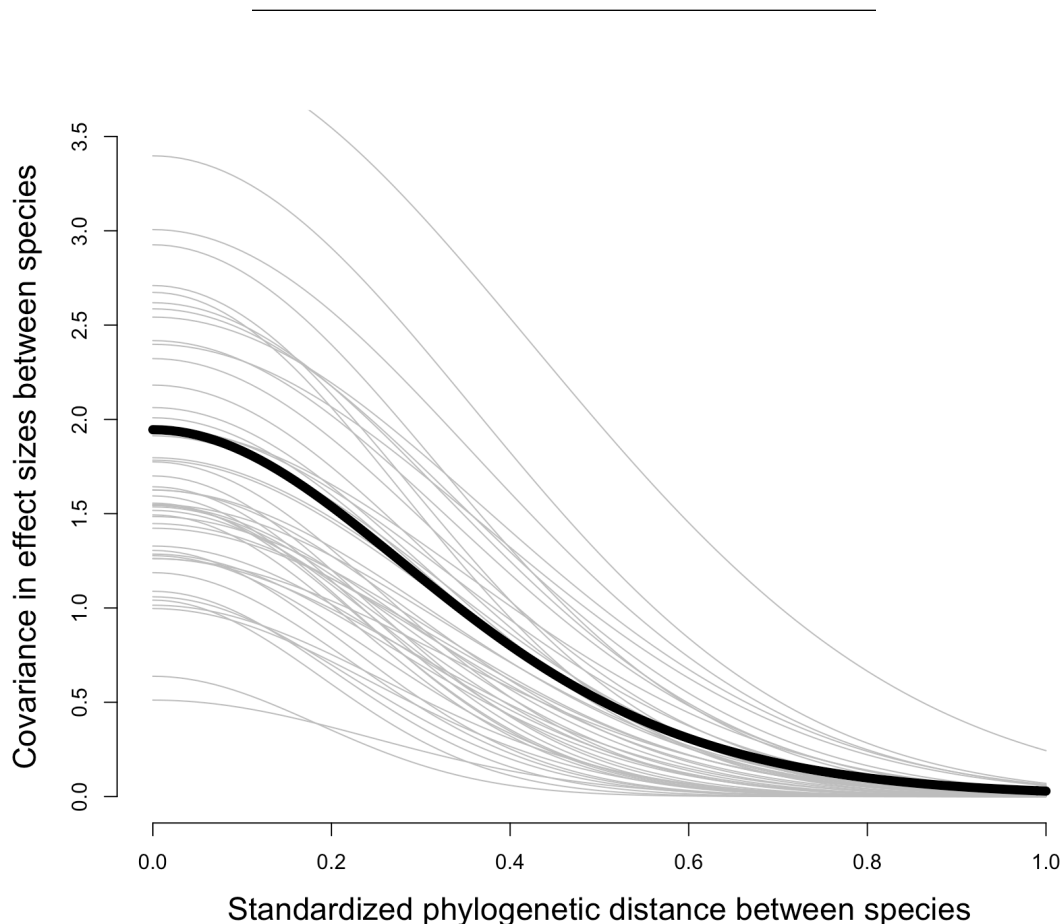


Figure 4. Relationship between the phylogenetic distance between pairs of species and the similarity of their effect sizes (solid black line represents mean estimate of rethinking model, grey lines represent variation in the estimate). Species that are closely related and share most of their phylogenetic history (standardized phylogenetic distance close to zero) show intermediate levels of covariance in their effect sizes of dominance rank on female reproductive success. The covariance drops to low values at a

standardized phylogenetic distance of around 0.4, the level separating species that are part of the same Order.

R1.5 Influence of approach: To the base model, we add random effects reflecting the differences in approaches across studies (dominance ranks classified continuous/categorical; dominance determined through agonism/correlate; population type wild/provisioned/captive; number of years of the study).

Studies which measured dominance rank categorically by classifying individuals as either dominants or subordinates report higher effect sizes (metafor estimate +0.29 - +0.35, rethinking estimate +0.31 - +0.41; n=251 effect sizes) than studies assigning individuals continuous ranks (metafor estimate 0.16-0.22, rethinking estimate +0.17 - +0.28; n=193 effect sizes). In essentially all studies of cooperative breeders (31 of 32 effect sizes), comparisons were between the single dominant female and a class of the remaining subordinate females, which may contribute to higher effect sizes for studies using categorical measures of rank (see section R5.2.1).

Studies which determined the rank of females based on agonistic interactions have lower effect sizes (metafor estimate +0.22 - +0.26, rethinking estimate +0.24 - +0.32; n=407 effect sizes) than studies which used other correlates (body size, age, etc.) to assign dominance ranks (metafor estimate 0.43-0.55, rethinking estimate +0.41 - +0.63; n=37 effect sizes). These 37 effect sizes where rank was assigned based on correlates are from cooperative breeders and/or studies in which groups consisted of mothers and their daughters.

Effect sizes did not vary between studies conducted with captive (metafor estimate +0.24 - +0.30, rethinking estimate +0.27 - +0.37; n=183 effect sizes), provisioned (metafor estimate +0.21 - +0.33, rethinking estimate +0.14 - +0.41; n=23 effect sizes), or wild (metafor estimate +0.22 - +0.34; n=283 effect sizes) individuals, and this does not change when we nest the population type within species (indicating that effect sizes do not differ between captive, provisioned, and wild populations of the same species).

2) What are the life history traits that mediate the benefits of rank on reproductive success?

R2.1 Influence of measure of reproductive success: To the base model, we add a predictor variable reflecting the six classes of measures of reproductive success.

Dominance rank appears to have the highest effect on age at first conception (metafor estimate +0.32 - +0.43, rethinking estimate +0.33 - +0.52; n=23 effect sizes), life time reproductive success (metafor estimate +0.27 - +0.40, rethinking estimate +0.31 - +0.47; n=34 effect sizes), interbirth interval (metafor estimate +0.25 - +0.37, rethinking estimate +0.28 - +0.37; n=46 effect sizes), infant production (metafor estimate +0.21 - +0.33, rethinking estimate +0.23 - +0.38; n=198 effect sizes), adult survival (metafor estimate +0.18 - +0.31, rethinking estimate +0.18 - +0.34; n=30 effect sizes), infant survival (metafor estimate +0.14 - +0.25, rethinking estimate +0.15 - +0.26; n=113 effect sizes). Effects of dominance rank on survival are lower than on other measures of female fitness. In addition, females themselves appear to benefit more than their offspring (adult survival > infant survival). While effect sizes for life time reproductive success are higher than those for the values from which it is usually calculated (adult survival, interbirth interval, infant production),

there does not appear to be a straightforward additive (or multiplicative) combination of the individual effects (Figure 5)

Fig A) Cooperative breeders

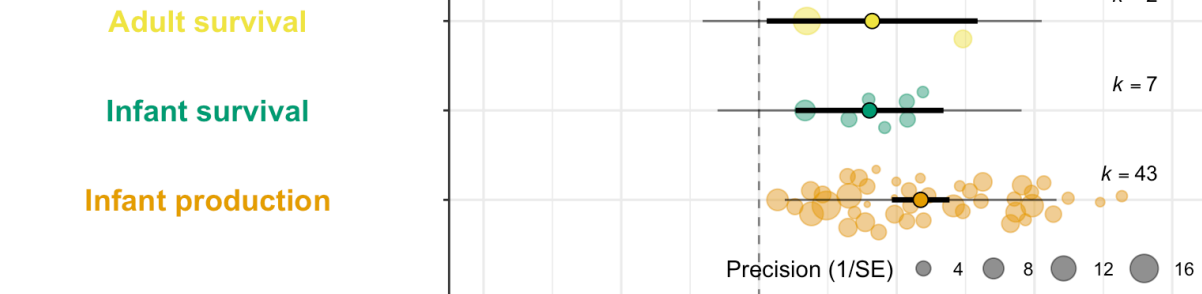


Fig B) Plural breeders

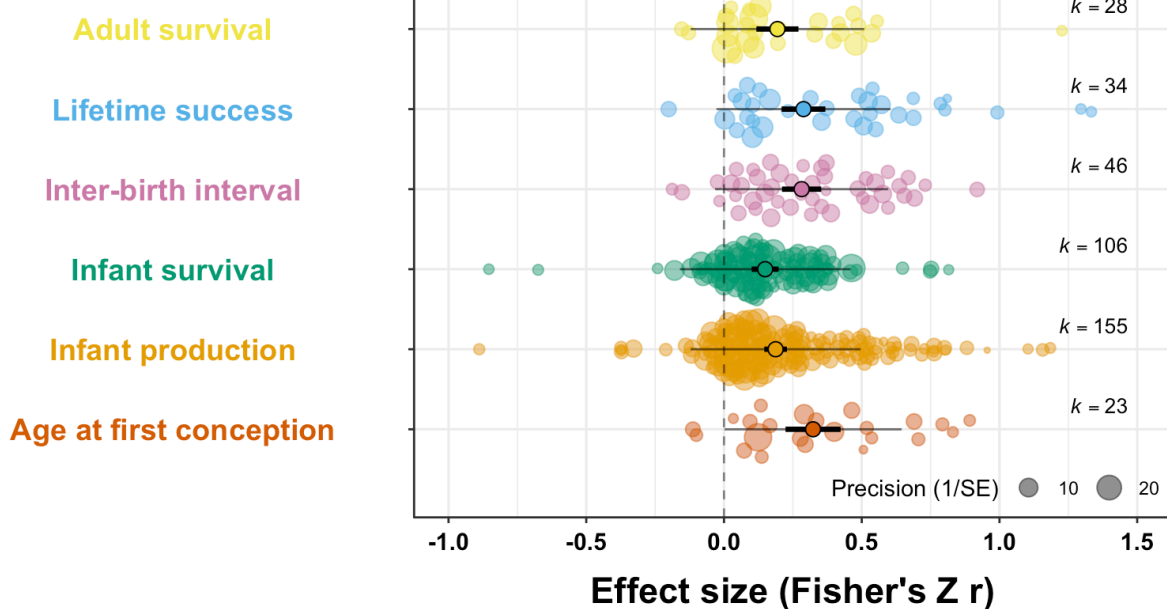


Figure 5. Raw effect sizes of dominance rank on reproductive success are generally higher for cooperative breeders (a) than for plural breeders (b), and differ according to the measure of reproductive success. In general, dominance appears to have stronger effects on reproductive output (lifetime reproductive success, age at first conception, infant production, inter-birth intervals) than on survival (both of the adult females themselves and of their infants). The differences between measures of reproductive success change slightly when accounting for similarity among observations from the same and related species, but the ordering remains the same.

R2.2 Litter Size and Litters Per Year Effects of dominance on reproductive success are higher in species with larger litter sizes (metafor estimate of litter size +0.03 - +0.05, rethinking estimate +0.05 - +0.09; n=444)

effect sizes) and with more litters per year (metafor estimate of litters per year +0.04 - +0.08, rethinking estimate +0.06 - +0.11; n=444 effect sizes). Effect sizes in species where females produce single offspring are on average 0.25 while effect sizes in species where females produce litters are on average 0.34, and effect sizes in species where females produce one or fewer litters per year are on average 0.25 while effect sizes in species where females produce multiple litters each year are on average 0.45. The association of the effect sizes with the number of litters per year remained when accounting for the phylogenetic relatedness among species, but the association with litter size did not, suggesting that it might be influenced by other characteristics that differ among species with variable litter sizes.

3) What are the ecological conditions that mediate the benefits of rank on reproductive success?

R3.1 Diet Category

Effect sizes are larger in carnivores (0.36; n=72 effect sizes) than in omnivores (0.29; n=227 effect sizes), herbivores (0.27; n=117 effect sizes), or frugivores (0.22; n=28 effect sizes) (estimated difference carnivores versus omnivores metafor -0.36 - -0.17 rethinking -0.24 - -0.04, difference carnivores versus herbivores metafor -0.29 - -0.13 rethinking -0.16 - -0.03, difference carnivores versus frugivores metafor -0.27 - -0.11 rethinking -0.14 - -0.02; estimates for all other comparisons cross 0). Carnivores are no longer estimated to have different effect sizes when the phylogenetic relatedness among species is taken into account, potentially due to the higher prevalence of cooperative breeding in carnivores.

R3.2 Environmental Harshness

Our data shows no association between environmental harshness and the effect of dominance rank on reproductive success (metafor estimate -0.3 - +0.4, rethinking -0.6 - +0.1; no change when accounting for shared phylogenetic history; n=259 effect sizes).

R3.3 Population Density

Effect sizes are larger in species with higher population densities (metafor +0.04 - +0.08, rethinking +0.05 - +0.10; n=346 effect sizes), even when including phylogenetic relatedness.

4) What are the social circumstances that mediate the benefits of rank?

R4.1 Breeding system

Effect sizes of cooperative breeders (average 0.58; n=52 effect sizes) are higher than those observed in plural (average 0.25; n=324 effect sizes) or associated breeders (average 0.23; n=68 effect sizes) (estimates for difference cooperative breeder vs plural breeder metafor -0.40 - -0.30, rethinking -0.41 - -0.27; cooperative breeder vs associated breeder metafor -0.47 - -0.35, rethinking -0.45 - -0.26; plural breeder vs associated breeder metafor -0.07 - +0.05, rethinking -0.07 - +0.05). Cooperative breeders are still estimated to have higher effect sizes than species with other breeding systems when accounting for phylogenetic relatedness, but the differences are slightly reduced (Figure 5).

R4.2 Dominance System

Effect sizes are higher in species in which condition plays a major role in determining which females are dominant rather than subordinate (average effect size 0.38; n=94 effect sizes), compared to species in which age (average effect size 0.31; n=100 effect sizes) or nepotism (average effect size 0.24; n=243 effect sizes) influence dominance rank (estimates for difference condition vs age: metafor +0.05 - +0.17, rethinking +0.01 - +0.16; condition vs nepotism: metafor +0.07 - +0.20, rethinking +0.08 - +0.20; age vs nepotism: metafor -0.07 - +0.03, rethinking -0.01 - +0.12). Species with different dominance system are no longer estimated to be different when including the phylogenetic similarity.

We had initially planned to assess whether dominance effect appear across different time scales depending on how dominant females acquire their position. However, this turned out to be more difficult. The species in our dataset have vastly varying lifespans, so simply assessing the number of years a study had been conducted for skews the observation towards short-lived species. The values for the relative duration (number of years studied divided by the maximum lifespan of the species) show that 90% of effect sizes are from studies that lasted less than 10% of the lifespan of the species (median 3%). In all of the 19 species in which studies spanned more than 10% of the lifespan, females acquire rank by nepotism. We did not find any consistent pattern of relationship between effect size and study duration dependent on the system of dominance acquisition.

R4.3 Philopatry

The effects of dominance rank on reproductive success are higher in species in which females disperse and join new groups (average effect size 0.46; n=55 effect sizes) compared to species in which most females were born in the same group they breed (average effect size 0.26; n=360 effect sizes) (metafor estimate of difference -0.24 - -0.12, rethinking estimate -0.25 - -0.11), also when accounting for phylogenetic covariance (Figure 6).

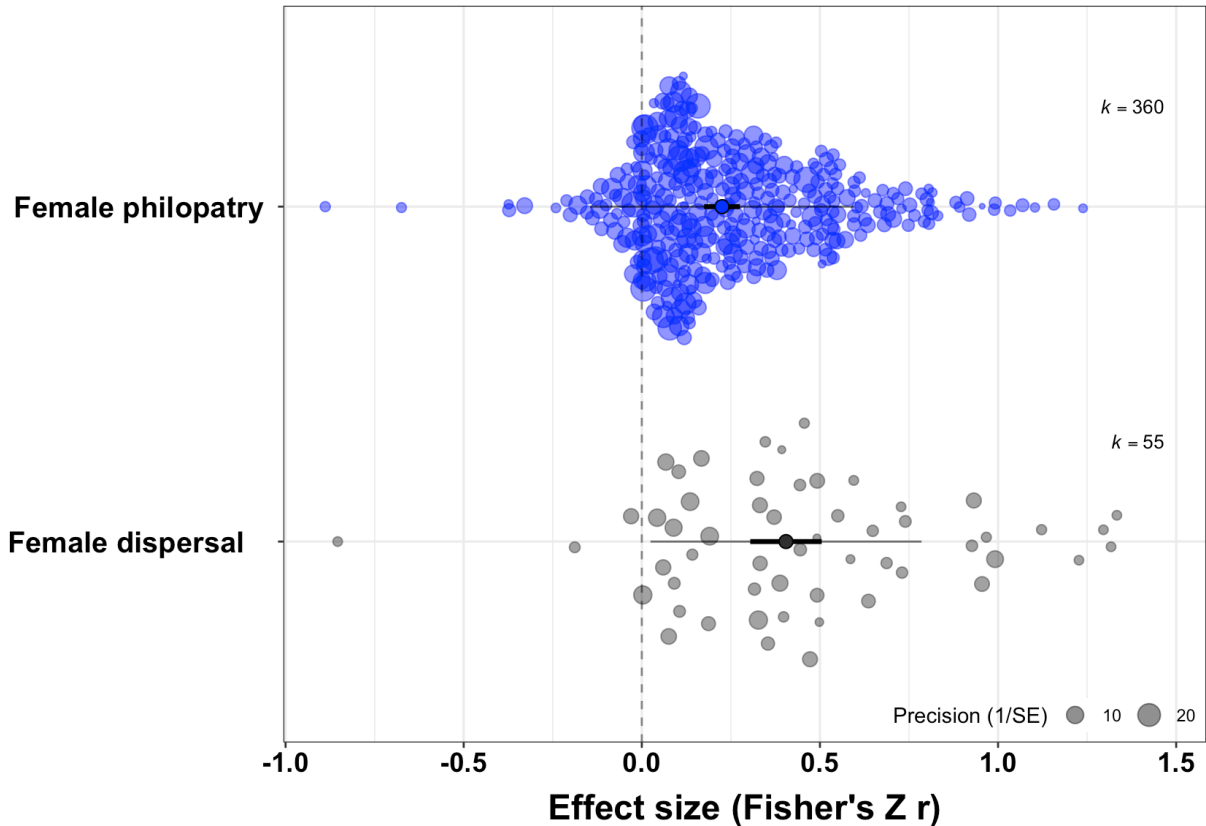


Figure 6. Effect sizes of dominance rank on female reproductive success are lower in species in which females are philopatric and remain in the group/area where they have been born (top, blue dots) than in species in which females disperse to breed (bottom, grey dots).

R4.4 Group size

Both approaches detect a negative association between the effect sizes and group sizes (metafor estimate of log group size $-0.099 - -0.678$, rethinking estimate of standardized group size $-0.10 - -0.05$; $n=444$ effect sizes). Compared to groups of 2 females, groups of 10 females show $\sim 10\%$ lower effect sizes and groups of ~ 50 females show 50% lower effect sizes. The negative association between group size and the effect sizes remains when accounting for similarity among closely related species.

R4.5 Average Relatedness

Effect sizes of dominance rank on reproductive success increase with increasing levels of average relatedness among female group members (metafor estimate $+0.31 - +0.59$, rethinking estimate $+0.31 - +0.71$; $n=288$ effect sizes), though the association is no longer detected when including the shared phylogenetic history among species (metafor estimate $-0.01 - +0.56$; rethinking estimate $-0.02 - +0.65$).

R4.6 Variance in relatedness

We could not assess this prediction because sufficient data was not available.

R4.7 Coalition formation

Species in which females form coalitions show only slightly lower effects of dominance rank on reproductive success (average 0.27; $n=246$ effect sizes) than species in which females do not have support during aggressive interactions (average 0.32; $n=180$ effect sizes) (estimate of difference metafor: $-0.11 - -0.01$, rethinking $-0.09 - +0.01$), with no difference in models accounting for similarity due to phylogenetic relatedness (metafor $-0.10 - +0.07$; rethinking $-0.09 - +0.03$).

R4.8 Intersexual conflict

Effect sizes are larger in species in which sex ratios in social groups are more balanced and lower when there are fewer males per female (metafor estimate $+0.55 - +1.25$, rethinking estimate $+0.07 - +0.11$; $n=328$ effect sizes), and the association remains the same when accounting for shared phylogenetic history.

Effect sizes are lower in species in which males commit infanticide (metafor estimate $-0.20 - 0.00$; rethinking estimate $-0.15 - -0.04$; $n=332$ effect sizes), but the relationship does not hold when accounting for phylogenetic relatedness (metafor $-0.13 - +0.07$, rethinking $-0.07 - +0.06$).

Differences in effect sizes are not associated with the extent of sexual dimorphism in body size across species (metafor estimate $-0.17 - 0.11$; rethinking $-0.05 - +0.01$; similar estimates when accounting for shared phylogenetic history; $n=334$ effect sizes).

R4.9 Macaque dominance styles

Differences in dominance styles among macaques are not associated with the effect of dominance rank on reproductive success (metafor estimates effect sizes of species in Grade 1 to be different from species in Grade 2 $+0.05 - +0.12$ but no differences for the five other pairwise Grade comparisons; rethinking estimates for all comparisons overlap zero; $n = 109$ effect sizes from 9 species). Egalitarian species do not show lower effects of dominance rank on reproductive success than other species and the sample size is too small to determine whether despotic species systematically differ from other species (Figure 7).

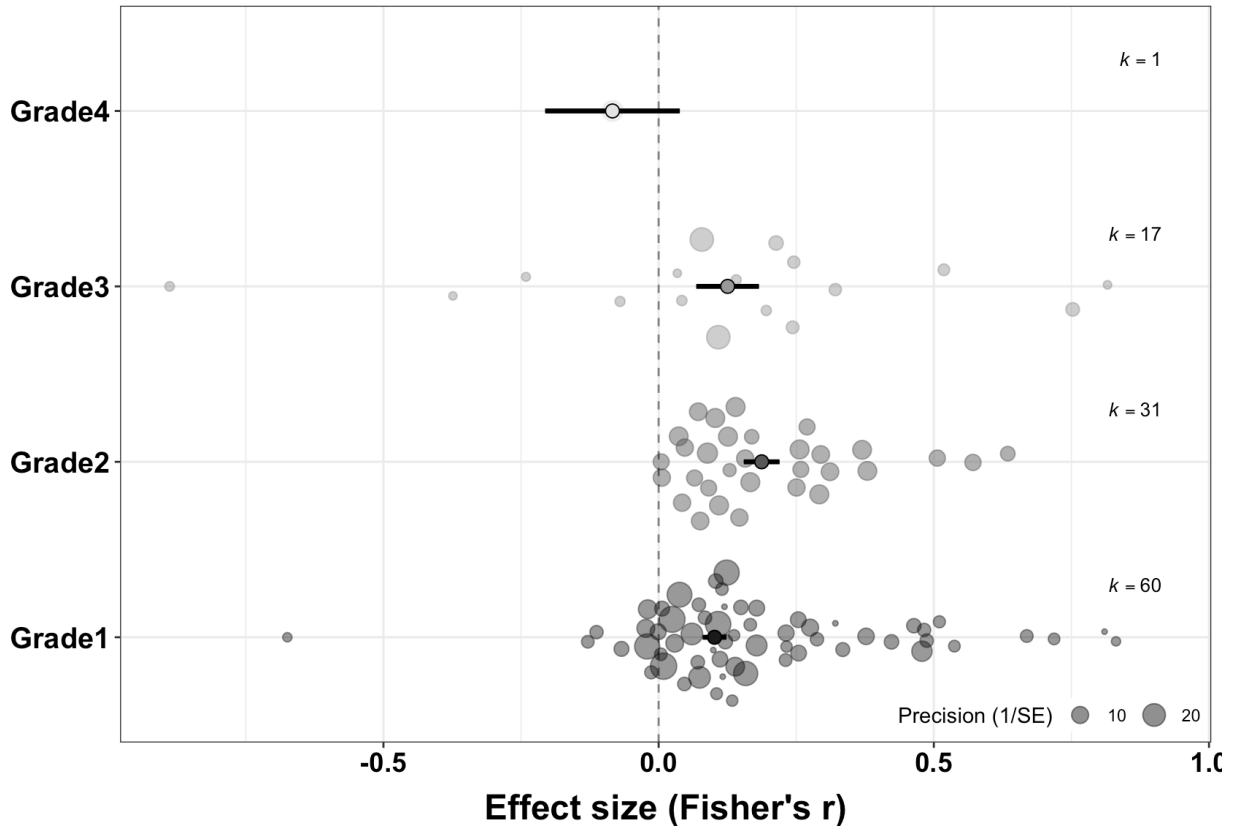


Figure 7. The effect of dominance rank on female reproductive success is similar across macaque species with different dominance styles. Relationships among female group members in species of grade 1 (bottom dark grey) are generally considered egalitarian, while grade 4 (top light grey) is assigned to species in which relationships are deemed highly despotic. Species with different dominance styles are not estimated to be different (all posterior contrasts overlap zero).

Summary of univariate analyses

Overall, our data indicate that females of higher rank generally have higher reproductive success than females of lower rank. In terms of the approach, effect sizes of dominance rank on reproductive success were higher (i) when individuals were assigned a rank category rather than a continuous position, (ii) when rank was determined using indirect measures rather than aggressive interactions, and (iii) in some studies, species, and families of species than in others. We found no differences in effect sizes when studies were conducted in a captive rather than a wild setting. Effect sizes of dominance rank were higher for measures of reproductive output than for measures of survival, and higher for measures of maternal than offspring fitness.

We found that effect sizes of dominance rank on reproductive success are associated with six of our single predictor variables, whereas we did not find an association with another eight of the single predictor variables (Table 1). Five of the six associated predictor variables reflect variation in the social environment, while we

did not find any association with any of the predictor variables reflecting the ecological environment.

Table 1. Overview of **variables associated with variation in effect sizes of dominance rank on female reproductive success** in univariate analyses. The following six variables (of the fourteen we assessed) are estimated to explain variation in the effect sizes with both approaches when accounting for shared phylogenetic history among the species in our sample.

Predictor variable	Metafor compatibility estimate of association	Rethinking compatibility estimate of association
litters per year	+0.03 - +0.05	+0.05 - +0.09
population density	+0.04 - +0.08	+0.05 - +0.10
group size	-0.07 - -0.01	-0.10 - -0.05
cooperative breeding	+0.30 - +0.40	+0.27 - +0.41
philopatry	-0.24 - -0.12	-0.25 - -0.11
sex ratio	+0.44 - +1.25	+0.07 - +0.11

Table 2. Overview of **variables not associated with variation in effect sizes of dominance rank on female reproductive success** in univariate analyses. The following eight variables (of the fourteen we assessed) are estimated to not be linked with variation in the effect sizes when accounting for shared phylogenetic history among the species in our sample.

Predictor variable	Metafor compatibility estimate of association	Rethinking compatibility estimate of association
litter size	-0.01 - +0.03	-0.04 - +0.09
dominance acquisition	-0.07 - +0.03	-0.01 - +0.12
diet	-0.04 - +0.03	-0.10 - +0.06
environmental harshness	-0.30 - +0.40	-0.60 - +0.10
average relatedness	-0.01 - +0.56	-0.01 - +0.12
female coalitions	-0.10 - +0.07	-0.09 - +0.07
male infanticide	-0.13 - +0.07	-0.07 - +0.06
sexual dimorphism	-0.17 - +0.11	-0.05 - +0.01

5) Combined analyses

R5.1 Heterogeneity and sample bias

The sample bias, namely the over-representation of extreme effect sizes, in our data likely results from all three influences of (i) publication bias, (ii) study system bias, and (iii) study time bias. In addition to the direct indications of publication and study system bias in our sample, our univariate analyses identified many factors that could lead to study system bias. For example, while less than 5% of all mammalian species are cooperative breeders, 12% of all effect sizes in our sample come from cooperative breeders which have high positive effect sizes.

To identify the potential interplay between the three biases, we built combined models. If biases occur because study systems with different effect sizes also have particular sample sizes and study duration (e.g. cooperative breeders tend to live in smaller groups), we should no longer detect an association between

sample size and study duration with the effect sizes when controlling for the different study systems. The combined models indicate that the study system factors identified in the uni-variate analyses are directly associated with variation in effect sizes (all their estimates do not overlap zero), as is sample size, but not the number of years a study had been conducted for. This indicates that our sample has both publication and study system bias. The lack of a direct influence of study time bias presumably occurs because sample size is associated with the number of years a study has been conducted for, indicating that large samples both in terms of time period or breadth might reduce noise.

The reduction in publication bias when accounting for the study system bias is visible when comparing the funnel plot of the raw effect sizes in relation to their precision (Figure 8a), which shows a clear asymmetry, to the funnel plot of the effect sizes adjusted for known predictors (Figure 8b), which only indicates some large effect sizes at small precision that are not balanced.

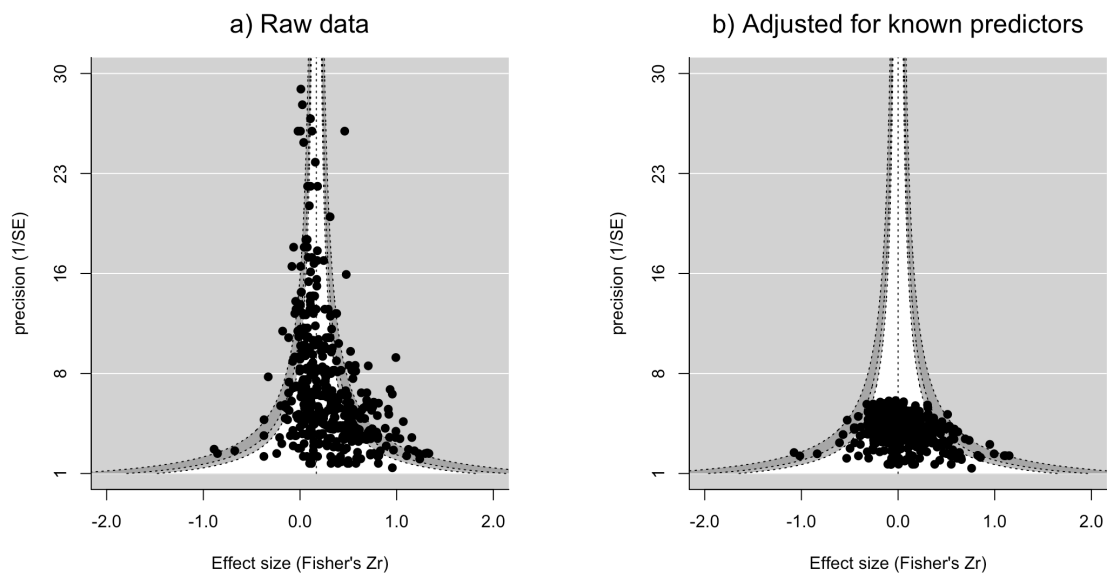


Figure 8. Funnel plots based on raw effect sizes (a) and effect sizes adjusted for known predictors (b). When accounting for the influence of which reproductive trait was measured, whether the species is a cooperative breeder or not, the number of litters per year the species produces, and the phylogenetic covariance among species, the distribution of the 444 effect sizes in our sample appears much less imbalanced (b) than the raw effect sizes (a). The mean effect size (grey dotted line in the center going upwards) is shifted close to zero when adjusting for known predictors because these predictors explain why some studies have positive effect sizes. Precision decreases for most estimates because they no longer represent the measured values but the values inferred from the interaction of the predictors.

R5.2 Differences between cooperative and plural/associated breeders

In our preregistration, we had decided to first construct univariate models as reported above, testing the influence of a single variable at a time to assess support for the specific predictions. One of the main factors that we found to be associated with higher effect sizes is cooperative breeding. Cooperative breeders differ

in many additional aspects, so we first checked whether any of the other associations we detect occur because they covary with cooperative breeding.

R5.2.1 Differences in approach to study cooperative breeders

Approaches of assigning rank depend on the breeding system of the study species, with many studies of cooperative breeders assigning rank into categories (98% categorical, 2% continuous) based on other measures (50% agonism, 50% other) while studies of plural and associated breeders often assign continuous ranks (51% categorical, 49% continuous) based on agonistic interactions (97% agonism, 3% other). Combining the variables representing the different study approaches with the variable representing the classification as cooperative breeder or not into single models indicates that the difference in effect sizes is primarily due to the stronger dominance effects in cooperative breeders (estimate of difference metafor +0.23 - +0.34, rethinking +0.23 - +0.37, n=444 effect sizes) and only very little due to the approaches the authors chose (other measure vs agonisms estimate of difference metafor +0.02 - +0.15, rethinking -0.02 - +0.16; rank categorical vs continuous estimate of difference metafor -0.02 - -0.09, rethinking -0.07 - +0.03, n=444 effect sizes).

R5.2.2 Different life history measures and cooperative breeding

In cooperative breeders, effects of dominance rank were only assessed on three of the six life history traits. We therefore performed separate analyses for cooperative and for plural/associated breeders to identify the life history traits showing specific increases in higher ranking females compared to others.

In cooperative breeders, effect sizes are higher for infant production (metafor estimate +0.49 - +0.72, rethinking estimate +0.55 - +0.69, n=43 effect sizes), and lower for infant survival (metafor +0.13 - +0.54, rethinking +0.20 - +0.61, n=7 effect sizes) and adult survival (metafor estimate +0.02 - +0.59, +0.12 - +0.73, n=2 effect sizes) (Figure 5).

In plural/associated breeders, effect sizes are (depending on the approach) highest for lifetime reproductive success (metafor estimate +0.19 - +0.29, rethinking estimate +0.33 - +0.47, n=34 effect sizes), age at first conception (metafor +0.27 - +0.36, rethinking +0.25 - +0.43, n=23 effect sizes) and interbirth interval (metafor +0.23 - +0.34, rethinking +0.25 - +0.38, n=46 effect sizes), followed by infant production (metafor +0.13 - +0.22, rethinking +0.19 - +0.27, n=155 effect sizes) and adult survival (metafor +0.14 - +0.24, rethinking +0.15 - +0.30, n=28 effect sizes), and are lowest for infant survival (metafor +0.11 - +0.20, rethinking +0.11 - +0.20, n=106 effect sizes) (Figure 5). The two methods give slightly different estimates because there is large variation among the effect sizes within each life history trait. In particular, effect sizes of dominance rank on lifetime reproductive success can be either low or high, often for the same population. For example, an experiment with house mice reported effect sizes ranging from 0.08 to 0.80, depending on the relatedness among the group members (König 1994). For mountain gorillas living in the Virungas, one study reported no effect of dominance rank on lifetime reproductive success (0.00) (Robbins et al. 2007) while another reported the highest effect size in our sample (1.33) after excluding major sources of environmental variability on reproductive success (Robbins et al. 2011).

R5.2.3 Litters per year and cooperative breeding

Cooperative breeders tend to have higher reproductive rates than species with other breeding systems. However, the association between reproductive rate and effect sizes of dominance rank on reproductive success remains across all breeding systems (metafor estimate of cooperative breeding +0.22 - +0.58, litters per year 0.00 - +0.07, interaction -0.10 - +0.04), with larger effect sizes in species producing more litters per year in cooperative (rethinking estimate +0.02 - +0.20; n=52 effect sizes) and plural (rethinking

+0.13 - +0.33; n=324 effect sizes), but not associated breeders (rethinking -0.08 - +0.23; n=68 effect sizes) (estimates take into account phylogenetic relatedness).

R5.2.4 Group size and cooperative breeding

In mammals, groups of cooperative breeders never grow to the same size (in our data, median 2 females per group, n=52) as groups of plural/associated breeders (in our data, median 14 females per group, n=392), potentially introducing an interaction effect. In our data, both group size and cooperative breeding remain independently associated with the effect sizes of dominance rank on reproductive success. The analyses suggest an interaction (metafor estimate for cooperative breeding +0.16 - +0.39, for group size -0.01 - 0.00, interaction 0.00 - +0.03, n=444 effect sizes), with effect sizes increasing with group size in cooperative breeders (rethinking estimate +0.01 - +0.02), where a single dominant continues to monopolize reproduction as groups get larger, and declining with group sizes in other breeding systems (rethinking estimate -0.01 - 0.00), where dominants might be less able to control reproduction of other group members as groups grow larger (Figure 9).

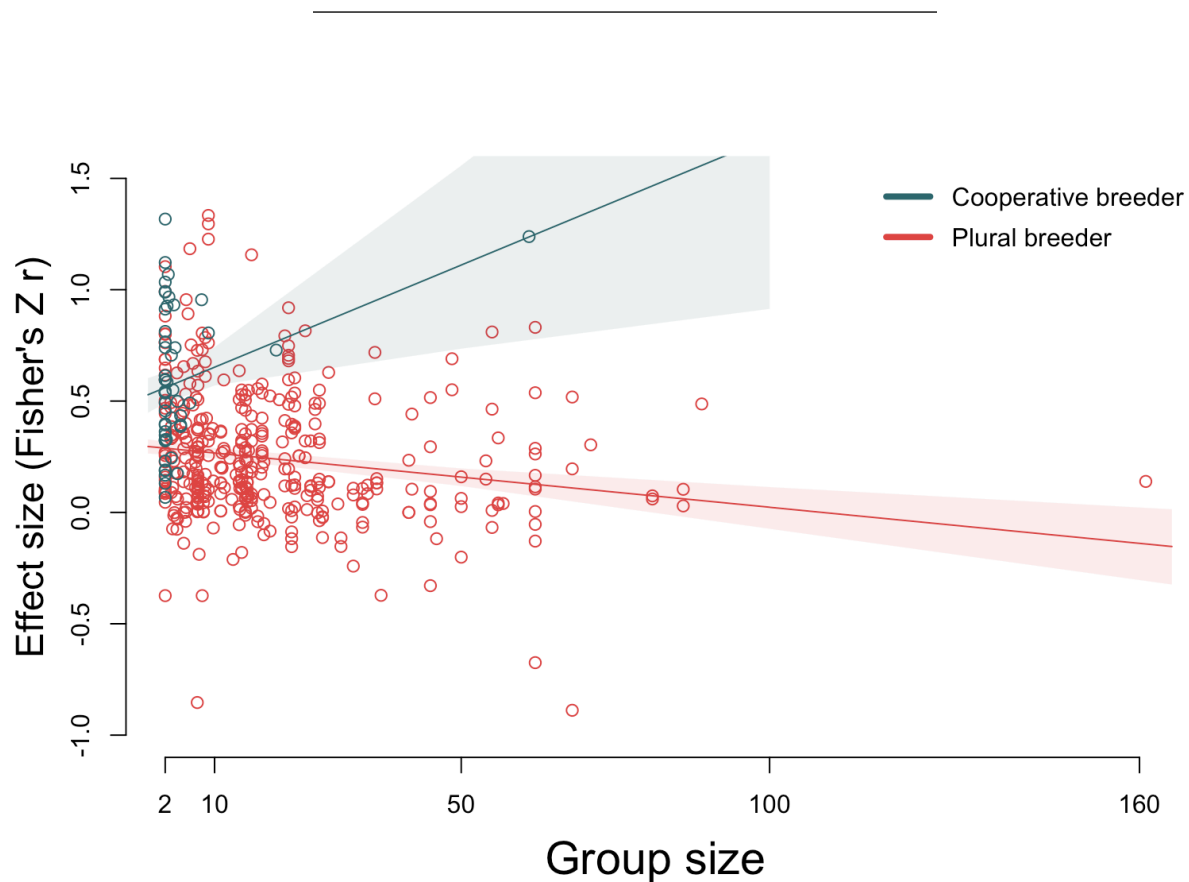


Figure 9. The relationship between the number of females in the group and the effect of dominance on reproductive success depends on whether the species is a cooperative (olive dots show data and olive line with shading shows estimate from rethinking model) or a plural breeder (red dots show data and red line with shading shows estimate from rethinking model). In cooperative breeders, effect sizes increase with increasing group size as a single female continues to monopolize reproduction in the group, whereas effect sizes decrease with increasing group size as dominants can potentially no longer control other females in

the group.

R5.2.5 Average relatedness and cooperative breeding

Similarly, there appears to be an interaction between average relatedness and breeding systems (metafor estimate for cooperative breeding $-0.06 - +0.44$, for average relatedness $-0.75 - +0.03$, for interaction $+0.10 - +1.51$, $n=288$ effect sizes), with effect sizes increasing with higher levels of average relatedness in cooperative breeders (rethinking estimate $0.00 - +0.12$, $n=36$ effect sizes) and decreasing with higher levels of average relatedness in plural/associate breeders (rethinking estimate $-0.06 - 0.00$, $n=252$ effect sizes)

R5.2.6 Philopatry and cooperative breeding

Female dispersal is more common in cooperative breeders (46%) than in plural/associated breeders (9%). However, effect sizes are larger in species with female dispersal also just among the plural/associated breeders (rethinking estimate $-0.19 - -0.02$, $n=363$ effect sizes), though differences between philopatry and dispersal are not associated with effect sizes in cooperative breeders (rethinking estimate $-0.10 - +0.12$, $n=52$ effect sizes) (metafor estimate for cooperative breeding $+0.15 - +0.49$, for philopatry $-0.18 - +0.06$, for interaction $-0.18 - +0.26$).

R5.2.7 Coalition formation and cooperative breeding

Coalition formation does not occur in cooperative breeders, leading to a potential confound. Restricting the analyses to plural/associated breeders, we find that effect sizes are higher in species in which females do form coalitions than in species where they do not (metafor estimate $0.00 - +0.14$, rethinking estimate $+0.01 - +0.11$, $n=374$ effect sizes). This likely reflects the benefits of nepotism in matrilineal groups. For our analysis, we did not differentiate between stabilizing coalitions, which usually occur among kin to maintain matrilineal rank differences, and revolutionary coalitions, which usually occur among unrelated individuals to limit the power of others in the group.

R5.3 Philopatry and group size

Group sizes of species in which females disperse tend to be smaller than group sizes of species in which females are philopatric. Both philopatry and increasing group size appear however to independently lead to lower effect sizes (metafor estimate philopatry $-0.09 - -0.01$ group size $-0.07 - -0.01$, rethinking estimate philopatry $-0.16 - 0.00$ group size $-0.07 - -0.03$, $n=415$ effect sizes).

R5.4 Philopatry and average relatedness

Among plural/associated breeders, average relatedness is lower in species in which females disperse (mean r 0.03 , $n=16$) than in species in which females are philopatric (mean r 0.10 , $n=228$), and among these species, differences in effect sizes are mainly associated with whether females disperse or are philopatric (metafor estimate $-0.11 - -0.03$, rethinking estimate $-0.22 - -0.02$) rather than levels of average relatedness (metafor estimate $+0.03 - +0.10$, rethinking estimate $-0.04 - +0.01$, $n=242$ effect sizes).

R5.5 Population density and group size

Population density and group size have independent influences on effect sizes (population density estimate metafor 0.00 - +0.01, rethinking 0.00 - +0.01; group size estimate metafor -0.03 - 0.01, n=346 effect sizes).

R5.6 Different influences in captive and wild populations

Models in which both the intercept and the slopes can vary according to whether studies were performed in the wild or in captivity also showed that there are no systematic differences of the effects of dominance rank on reproductive success between populations in these settings (for the different life history measurements and for cooperative breeding).

Summary of combined analyses

The analyses of combinations of predictors of the effect size of dominance on rank on reproductive success show that most predictors have a direct influence. However, we find that the approach authors used to measure the effect does not lead to different estimates of the effect size, it is rather that different approaches have been used in different study systems. We also find that average relatedness might not directly mitigate effect sizes, but that it is a co-variate of the breeding system and whether females are philopatric or disperse. In addition, we find some interactions, with group size having divergent influences depending on the breeding system; and coalitions among females reducing effect sizes among plural breeders.

Discussion

Our study finds that, in social mammals, dominant females have higher reproductive success than lower-ranking females. Positive effects of dominance rank are present for all our measures of reproductive success and among plural breeders, where data for all measures of reproductive success exist, are highest for lifetime reproductive success. This suggests that even if dominants might face some trade-offs (e.g. higher stress levels Cavigelli et al. (2003)), obtaining a high ranking position in a social group generally leads to fitness benefits, though how females obtain these benefits (e.g. shorter interbirth intervals versus larger offspring) differs between populations. Our meta-analysis also highlights several factors associated with variation in the strength of the effect of dominance rank on reproductive success, where social factors in particular appear to have a modulating influence while variation in life history and ecological factors appears of less importance. Despite a consistent positive relationship between higher dominance rank and higher reproductive success, the data we were able to bring together for this study show some biases that suggest that further studies might detect lower effects. Our investigation of sample bias indicates a combination of publication bias, study system bias, and study time bias. Unlike often claimed for meta-analyses, the over-representation of positive findings in our case appears not to be primarily due to a file-drawer problem of unpublished negative findings but due to researchers targeting their efforts on feasible systems. Studies into the potential mechanisms of female competition and reproductive suppression have focused on species where there are clear differences in reproductive success between dominants and subordinates. In addition, obtaining reliable reproductive success data in long-lived mammals takes particular effort, again likely limiting the systems that have been studied to investigate the effects of dominance rank. We did find that studies conducted for longer time periods show less variance in their estimates, potentially because they

also have larger sample sizes. Alternatively, or in addition, studies conducted across longer time frames might be less likely to show extreme effect size estimates because natural changes in dominance rank and events that affect all females equally (e.g. infanticide Cheney et al. (2004)) occur relatively regularly across a multi-year study, while estimates derived over short time frames may over-estimate effect sizes.

Overall, we estimated an average effect of 0.28 of rank on reproductive success. What does this mean? First, it is important to highlight that this effect size reflects how well rank predicts reproductive success, but not directly indicates how different the reproductive success of high-ranking females is from that of low-ranking females. While the effect of dominance has to be zero in groups where all females have exactly the same reproductive success, an effect of zero is also found in a group where there are large differences in reproductive success across females which do not align with the females' dominance rank. Just by chance, we would expect differences in reproductive success among females in a social group and we could also expect that these differences are associated with traits that might be used to classify social rank. To assess whether the effects we detect are higher than such random variation, we performed simulations. For this, we simulated artificial groups of females reflecting macaques, the genus most common in our sample. We assumed that each female in each group might have between 0 to 8 offspring, with an average 2 (following a Poisson distribution, so most females have 1 or 2 offspring). We performed 10,000 simulations of six groups of twelve females each (the median group size in our data). When we set no association between rank and reproductive success, less than 0.1% of simulations showed an effect size as high or higher than the 0.28 we observe in the data (Figure 10). Effect sizes for a perfect association between each female's rank and her reproductive success ranged between 0.75-0.95 (mean 0.88). Simulations in which the two highest ranking females always have the highest reproductive success while rank among lower ranking females no longer is associated with success produces effect sizes close to what we observe (mean 0.32), whereas values tend to be slightly lower if only the highest ranking female consistently has the highest success (mean 0.18). These simulations cannot resolve whether high ranking females have higher reproductive success because they obtained this position or whether there are some traits that lead to both higher rank and higher reproductive success - or whether they are simply the lucky ones (Snyder and Ellner (2018)). However, the value of the overall effect size we observe compared to those under random expectations indicates that social rank has a particular association with reproductive success beyond the random variation we expect in social groups.

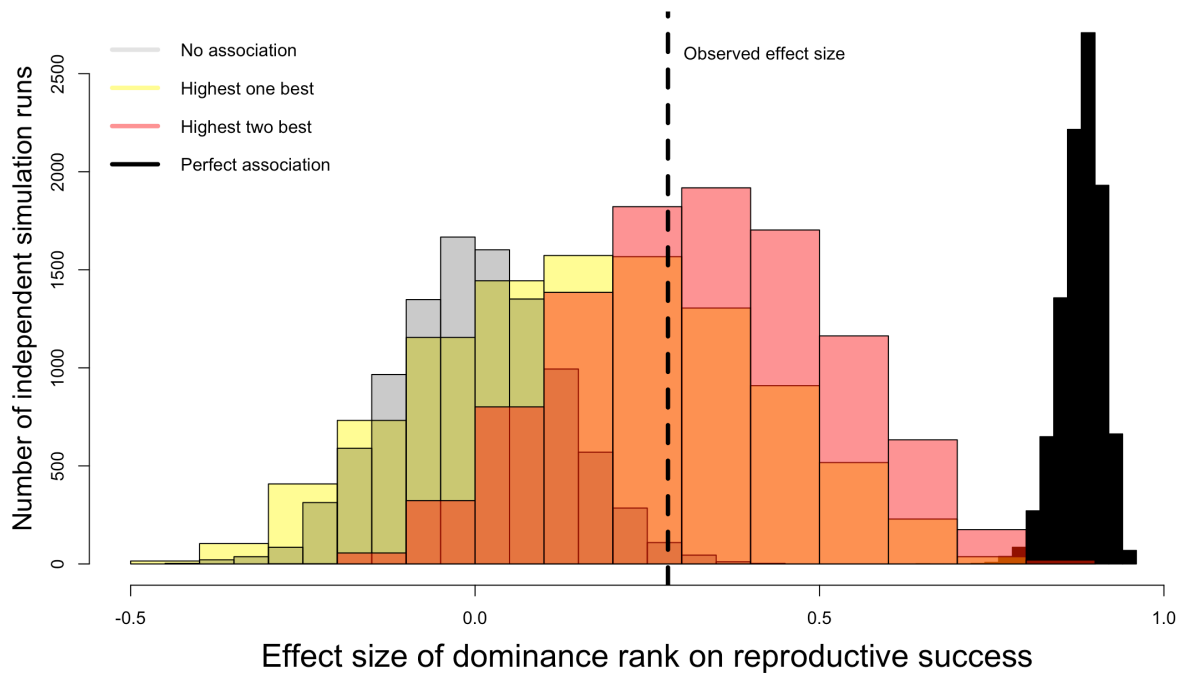


Figure 10. The average effect size of dominance rank on female reproductive success we observe in our sample (0.28; dotted vertical line) is in between the effect sizes expected for social groups in which there is either no (grey histogram) or a perfect association (black histogram) between each rank and the reproductive success of females. The observed value is close to a situation in which the two highest ranking females (red histogram) or only the highest ranking female (yellow histogram) always have the highest success in a group of twelve females.

Among the social traits we investigated, the highest difference in the effect of rank on reproductive success was between cooperative breeders and plural/associated breeders. This result was expected given the higher reproductive skew that has been found among females in cooperative breeders (Lukas and Clutton-Brock (2012)). The contrast between breeding systems appears due to the degree of reproductive control that dominants in cooperative breeders have over their, mostly related, group members. The likely importance of reproductive control of dominant females in cooperative breeders compared to plural/associated breeders are also reflected in the different relationships of the effect sizes with group size in the different breeding systems. While among cooperative breeders there usually is only a single breeding dominant female and large groups occur when her reproductive output is higher, dominant females in plural/associated breeders likely face reduced opportunities to control reproduction in larger groups (Rubenstein, Botero, and Lacey (2016)). In this context, it is again important to note that we only look at the association between rank and the variation in reproductive success within groups. Even though the relative difference between dominant and subordinate females might be lower in larger group sizes, in terms of overall fitness it might still be better to be the dominant in a group of the optimal size rather than a smaller group (e.g. small group where dominant has 3 versus subordinate has 2 offspring (50% higher fitness) compared to large group where dominant has 4 while all other females have 3 offspring (33% higher fitness)). While reproductive control appears important in explaining high reproductive success of dominant females, we did not find that

associations between the effect sizes and how females acquire and maintain rank. Effect sizes were similar when dominant females acquire their position by kin support versus aggression or age, and among macaque species were not associated with dominance styles.

Among plural and associated breeders, effects of dominance rank on female reproductive success are higher when (i) females disperse, (ii) groups are smaller, and (iii) females form coalitions. These observations are somewhat opposite to the processes presumably linked to reproductive suppression in cooperative breeders. In addition, these findings also do not support accounts that focus on nepotism as a primary factor in leading to social groups with large differences among females. It appears that in situations of strong nepotism females in a group might have more similar reproductive success, with patterns such as youngest sister ascendancy potentially reducing differences among kin (Datta (1988), Bergstrom and Fedigan (2010), Lea et al. (2014)). Instead, these findings suggest that competition among females might be highest in social groups in which females form complex relationships and rates of aggression are high (Lukas and Clutton-Brock (2018)). In our sample we for example observe relative strong effects of high dominance rank on reproductive success among equids and among gorillas, who have similar social systems with females benefiting from forming social bonds with unfamiliar/unrelated individuals they encounter when joining new small groups upon reaching maturity (e.g. Cameron, Setsaas, and Linklater (2009)).

Of the ecological variables we investigated, only population density was associated with differences in effect sizes of dominance rank on reproductive success, again supporting the role of social interactions in shaping fitness outcomes of dominance interactions. The observation that other ecological factors do not mitigate the strength of the fitness benefit dominant females receive might suggest that dominants are consistently able to outcompete other females in the group rather than dominance only being important under challenging conditions. While local ecological conditions, rather than the species-level traits we used, might modulate fitness benefits of high dominance rank for females, it seems unlikely that there would be a strong directional influence given that effect sizes from the same species tend to be similar, even in captive conditions. In line with this, previous work has shown that subordinate females may not always be the first to suffer under limiting conditions (Fedigan (1983)). Instead, a number of ecological challenges, such as for example predation (Cheney et al. (2004)), can affect all females independent of their rank and thereby diminishing the relative benefits dominant females acquire (Altmann and Alberts (2003)).

The overall effect size of dominance rank on female reproductive success across the species in our sample is slightly higher than that reported in a previous study, though we find a similar value when we restrict our sample to primate species, the focus of the previous study (the average in our sample is 0.28, for only the primates in our sample 0.23, versus previously reported for primates 0.20 Majolo et al. (2012)). These estimates of the effects of female dominance rank are lower than those previously reported for males. The previous study on primates reports an effect of male dominance rank on fecundity of 0.71 (Majolo et al. (2012)), and estimates in a different study of the effect of dominance rank on males' mating success are ~0.6 (Cowlshaw and Dunbar (1991)). Do these different estimates reflect that males benefit more from high dominance rank than females? We think that we cannot make such an inference at this stage. Measures of mating success might not necessarily translate in equally high skew in reproductive success and studies measuring male reproductive success tend to cover even shorter time periods than the studies that identify female reproductive success. Several of the factors we identified here to modulate the effect of dominance rank on reproductive success may also be linked to differences between females and males. However, it could be expected that males benefit more from rank than females, because female mammals are usually limited in the maximum reproductive success they can have at any given time. The benefits of rank are very different in nature between males and females and only additional symmetrical meta-analyses in males can

answer such a question.

Our findings highlight that social factors can have important influences on demography and genetic evolution by leading to systematic differences in reproductive success. The effect of high dominance rank on reproductive success influence the growth and composition of social groups across generations. In particular when social rank is heritable, strong long-term changes are visible in the few studies which have been able to track reproductive success across multiple generations. For example, among spotted hyenas, the highest ranking female in 1979 is the ancestor of more than half of the females in the clan in 2009 (Holekamp et al. (2012)). This perspective also highlights that even small differences in reproductive success can add up over long time frames. In particular, even if dominant females do not have much higher reproduction under average conditions, if they were the only ones to survive or reproduce under extreme conditions this could have important fitness consequences (Lewontin and Cohen (1969)). For future studies, detailed long-term investigations are not only relevant to understand the long-term consequences of the effect of dominance rank on reproduction, but also to infer the multiple mechanisms that link rank to reproductive output (e.g. Fedigan (1983), Pusey, Williams, and Goodall (1997)).

Ethics

Our study relies on previously published data and did not involve working directly with animals.

Author contributions

Shivani: Hypothesis development, data collection, data analysis and interpretation, revising/editing.

Huchard: Hypothesis development, data analysis and interpretation, write up, revising/editing.

Lukas: Hypothesis development, data collection, data analysis and interpretation, write up, revising/editing, materials/funding.

Funding

Shivani received funding from the INSPIRE programme of the Department of Science & Technology of the Government of India. This research was supported by the Department of Human Behavior, Ecology and Culture at the Max Planck Institute for Evolutionary Anthropology.

Conflict of interest disclosure

We, the authors, declare that we have no financial conflicts of interest with the content of this article. Elise Huchard and Dieter Lukas are Recommenders at PCI Ecology.

Acknowledgements

We thank our PCI Ecology Recommender, Matthieu Paquet, and our reviewers, Bonaventura Majolo and one anonymous reviewer, for their valuable feedback on our preregistration that greatly improved this piece of research. We are grateful to the members of the Department of Human Behavior, Ecology and Culture at the Max Planck Institute for Evolutionary Anthropology for feedback during the early stages of this project.

References

- Alexander, Richard D. 1974. "The Evolution of Social Behavior." *Annual Review of Ecology and Systematics* 5 (1): 325–83.
- Altmann, Jeanne, and Susan C Alberts. 2003. "Variability in Reproductive Success Viewed from a Life-History Perspective in Baboons." *American Journal of Human Biology* 15 (3): 401–9.
- Balasubramaniam, Krishna N, Katharina Dittmar, Carol M Berman, Marina Butovskaya, Mathew A Cooper, Bonaventura Majolo, Hideshi Ogawa, Gabriele Schino, Bernard Thierry, and Frans BM De Waal. 2012. "Hierarchical Steepness, Counter-Aggression, and Macaque Social Style Scale." *American Journal of Primatology* 74 (10): 915–25.
- Barsbai, Toman, Dieter Lukas, and Andreas Ponderfer. 2021. "Local Convergence of Behavior Across Species." *Science* 371 (6526): 292–95.
- Bercovitch, Fred B. 1991. "Social Stratification, Social Strategies, and Reproductive Success in Primates." *Ethology and Sociobiology* 12 (4): 315–33.
- Bergstrom, Mackenzie L, and Linda M Fedigan. 2010. "Dominance Among Female White-Faced Capuchin Monkeys (*Cebus Capucinus*): Hierarchical Linearity, Nepotism, Strength and Stability." *Behaviour*, 899–931.
- Botero, Carlos A, Roi Dor, Christy M McCain, and Rebecca J Safran. 2014. "Environmental Harshness Is Positively Correlated with Intraspecific Divergence in Mammals and Birds." *Molecular Ecology* 23 (2): 259–68.
- Brown, Gillian R, and Joan B Silk. 2002. "Reconsidering the Null Hypothesis: Is Maternal Rank Associated with Birth Sex Ratios in Primate Groups?" *Proceedings of the National Academy of Sciences* 99 (17): 11252–55.
- Burgin, Connor J, Jocelyn P Colella, Philip L Kahn, and Nathan S Upham. 2018. "How Many Species of Mammals Are There?" *Journal of Mammalogy* 99 (1): 1–14.
- Cameron, Elissa Z, Trine H Setsaas, and Wayne L Linklater. 2009. "Social Bonds Between Unrelated Females Increase Reproductive Success in Feral Horses." *Proceedings of the National Academy of Sciences* 106 (33): 13850–53.
- Cavigelli, SA, T Dubovick, W Levash, A Jolly, and A Pitts. 2003. "Female Dominance Status and Fecal Corticoids in a Cooperative Breeder with Low Reproductive Skew: Ring-Tailed Lemurs (*Lemur Catta*)." *Hormones and Behavior* 43 (1): 166–79.
- Chamberlain, Scott A, Stephen M Hovick, Christopher J Dibble, Nick L Rasmussen, Benjamin G Van Allen, Brian S Maitner, Jeffrey R Ahern, et al. 2012. "Does Phylogeny Matter? Assessing the Impact of Phylogenetic Information in Ecological Meta-Analysis." *Ecology Letters* 15 (6): 627–36.
- Cheney, Dorothy L, Robert M Seyfarth, Julia Fischer, J Beehner, T Bergman, SE Johnson, Dawn M Kitchen, RA Palombit, D Rendall, and Joan B Silk. 2004. "Factors Affecting Reproduction and Mortality Among Baboons in the Okavango Delta, Botswana." *International Journal of Primatology* 25 (2): 401–28.
- Clutton-Brock, T, and E Huchard. 2013. "Social Competition and Its Consequences in Female Mammals." *Journal of Zoology* 289 (3): 151–71.

- Clutton-Brock, Tim H, Sarah J Hodge, Tom P Flower, Goran F Spong, and Andrew J Young. 2010. "Adaptive Suppression of Subordinate Reproduction in Cooperative Mammals." *The American Naturalist* 176 (5): 664–73.
- Cowlshaw, Guy, and Robin IM Dunbar. 1991. "Dominance Rank and Mating Success in Male Primates." *Animal Behaviour* 41 (6): 1045–56.
- Datta, Saroj. 1988. "The Acquisition of Dominance Among Free-Ranging Rhesus Monkey Siblings." *Animal Behaviour* 36 (3): 754–72.
- Digby, Leslie J, Stephen F Ferrari, and Wendy Saltzman. 2006. "The Role of Competition in Cooperatively Breeding Species." *Primates in Perspective*. Oxford University Press, New York, 85–106.
- Drummond, Alexei J, Marc A Suchard, Dong Xie, and Andrew Rambaut. 2012. "Bayesian Phylogenetics with BEAUti and the BEAST 1.7." *Molecular Biology and Evolution* 29 (8): 1969–73.
- Egger, Matthias, George Davey Smith, Martin Schneider, and Christoph Minder. 1997. "Bias in Meta-Analysis Detected by a Simple, Graphical Test." *Bmj* 315 (7109): 629–34.
- Ellis, Lee. 1995. "Dominance and Reproductive Success Among Nonhuman Animals: A Cross-Species Comparison." *Ethology and Sociobiology* 16 (4): 257–333.
- Fedigan, Linda Marie. 1983. "Dominance and Reproductive Success in Primates." *American Journal of Physical Anthropology* 26 (S1): 91–129.
- Fedigan, Linda Marie, and Katharine M Jack. 2013. "Sexual Conflict in White-Faced Capuchins." *Evolution's Empress*, Eds Fisher ML, Garcia JR (Oxford Univ Press, New York), 281–303.
- Fortunato, Laura. 2019. "Lineal Kinship Organization in Cross-Specific Perspective." *Philosophical Transactions of the Royal Society B* 374 (1780): 20190005.
- Frank, Laurence G. 1986. "Social Organization of the Spotted Hyaena *Crocuta Crocuta*. II. Dominance and Reproduction." *Animal Behaviour* 34 (5): 1510–27.
- Giles, Sarah L, Christine J Nicol, Patricia A Harris, and Sean A Rands. 2015. "Dominance Rank Is Associated with Body Condition in Outdoor-Living Domestic Horses (*Equus Caballus*)." *Applied Animal Behaviour Science* 166: 71–79.
- Holekamp, Kay E, and Laura Smale. 1991. "Dominance Acquisition During Mammalian Social Development: The 'Inheritance' of Maternal Rank." *American Zoologist* 31 (2): 306–17.
- Holekamp, Kay E, Jennifer E Smith, Christopher C Strelloff, Russell C Van Horn, and Heather E Watts. 2012. "Society, Demography and Genetic Structure in the Spotted Hyena." *Molecular Ecology* 21 (3): 613–32.
- Huchard, Elise, Sinead English, Matt BV Bell, Nathan Thavarajah, and Tim Clutton-Brock. 2016. "Competitive Growth in a Cooperative Mammal." *Nature* 533 (7604): 532–34.
- Isaac, Joanne L. 2005. "Potential Causes and Life-History Consequences of Sexual Size Dimorphism in Mammals." *Mammal Review* 35 (1): 101–15.
- Jarman, Peter. 1983. "Mating System and Sexual Dimorphism in Large Terrestrial, Mammalian Herbivores." *Biological Reviews* 58 (4): 485–520.
- Jones, Kate E, Jon Bielby, Marcel Cardillo, Susanne A Fritz, Justin O'Dell, C David L Orme, Kamran Safi, et al. 2009. "PanTHERIA: A Species-Level Database of Life History, Ecology, and Geography of Extant

- and Recently Extinct Mammals: Ecological Archives E090-184.” *Ecology* 90 (9): 2648–48.
- Kappeler, Peter M, Charles L Nunn, Alexander Q Vining, and Steven M Goodman. 2019. “Evolutionary Dynamics of Sexual Size Dimorphism in Non-Volant Mammals Following Their Independent Colonization of Madagascar.” *Scientific Reports* 9 (1): 1–14.
- Kurz, Solomon. 2019. *Statistical Rethinking with Brms, Ggplot2, and the Tidyverse*. available at: <https://solomonkurz.netlify.com/post/bayesian-meta-analysis/>.
- Lajeunesse, Marc J, J Koricheva, J Gurevitch, and K Mengersen. 2013. “Recovering Missing or Partial Data from Studies: A Survey of Conversions and Imputations for Meta-Analysis.” *Handbook of Meta-Analysis in Ecology and Evolution*, 195–206.
- Lakens, Daniël. 2013. “Calculating and Reporting Effect Sizes to Facilitate Cumulative Science: A Practical Primer for t-Tests and ANOVAs.” *Frontiers in Psychology* 4: 863.
- Lea, Amanda J, Niki H Learn, Marcus J Theus, Jeanne Altmann, and Susan C Alberts. 2014. “Complex Sources of Variance in Female Dominance Rank in a Nepotistic Society.” *Animal Behaviour* 94: 87–99.
- Lewontin, Richard C, and Daniel Cohen. 1969. “On Population Growth in a Randomly Varying Environment.” *Proceedings of the National Academy of Sciences* 62 (4): 1056–60.
- Loison, Anne, Jean-Michel Gaillard, Christophe Pélabon, and Nigel Gilles Yoccoz. 1999. “What Factors Shape Sexual Size Dimorphism in Ungulates?” *Evolutionary Ecology Research* 1 (5): 611–33.
- Lukas, Dieter, and Tim Clutton-Brock. 2012. “Cooperative Breeding and Monogamy in Mammalian Societies.” *Proceedings of the Royal Society B: Biological Sciences* 279 (1736): 2151–56.
- . 2017. “Climate and the Distribution of Cooperative Breeding in Mammals.” *Royal Society Open Science* 4 (1): 160897.
- . 2018. “Social Complexity and Kinship in Animal Societies.” *Ecology Letters* 21 (8): 1129–34.
- Lukas, Dieter, and Elise Huchard. 2014. “The Evolution of Infanticide by Males in Mammalian Societies.” *Science* 346 (6211): 841–44.
- . 2019. “The Evolution of Infanticide by Females in Mammals.” *Philosophical Transactions of the Royal Society B* 374 (1780): 20180075.
- Lukas, Dieter, Vernon Reynolds, Christophe Boesch, and Linda Vigilant. 2005. “To What Extent Does Living in a Group Mean Living with Kin?” *Molecular Ecology* 14 (7): 2181–96.
- Majolo, Bonaventura, Julia Lehmann, Aurora de Bortoli Vizioli, and Gabriele Schino. 2012. “Fitness-Related Benefits of Dominance in Primates.” *American Journal of Physical Anthropology* 147 (4): 652–60.
- McElreath, Richard. 2020. *Statistical Rethinking: A Bayesian Course with Examples in r and Stan*. CRC press.
- Nakagawa, Shinichi, Malgorzata Lagisz, Michael D Jennions, Julia Koricheva, Daniel Noble, Timothy H Parker, Alfredo Sánchez-Tójar, Yefeng Yang, and Rose E O’Dea. 2021. “Methods for Testing Publication Bias in Ecological and Evolutionary Meta-Analyses.”
- Nakagawa, Shinichi, Malgorzata Lagisz, Rose E O’Dea, Joanna Rutkowska, Yefeng Yang, Daniel WA Noble, and Alistair M Senior. 2021. “The Orchard Plot: Cultivating a Forest Plot for Use in Ecology, Evolution, and Beyond.” *Research Synthesis Methods* 12 (1): 4–12.

- Nakagawa, Shinichi, and Eduardo SA Santos. 2012. "Methodological Issues and Advances in Biological Meta-Analysis." *Evolutionary Ecology* 26 (5): 1253–74.
- Pandit, Sagar A, and Carel P van Schaik. 2003. "A Model for Leveling Coalitions Among Primate Males: Toward a Theory of Egalitarianism." *Behavioral Ecology and Sociobiology* 55 (2): 161–68.
- Paradis, Emmanuel, and Klaus Schliep. 2019. "Ape 5.0: An Environment for Modern Phylogenetics and Evolutionary Analyses in r." *Bioinformatics* 35 (3): 526–28.
- Preston, Carrol, Deborah Ashby, and Rosalind Smyth. 2004. "Adjusting for Publication Bias: Modelling the Selection Process." *Journal of Evaluation in Clinical Practice* 10 (2): 313–22.
- Pusey, Anne. 2012. "Magnitude and Sources of Variation in Female Reproductive Performance." *The Evolution of Primate Societies*, 343–66.
- Pusey, Anne, Jennifer Williams, and Jane Goodall. 1997. "The Influence of Dominance Rank on the Reproductive Success of Female Chimpanzees." *Science* 277 (5327): 828–31.
- Rubenstein, Dustin R, Carlos A Botero, and Eileen A Lacey. 2016. "Discrete but Variable Structure of Animal Societies Leads to the False Perception of a Social Continuum." *Royal Society Open Science* 3 (5): 160147.
- Smith, Richard J. 1999. "Statistics of Sexual Size Dimorphism." *Journal of Human Evolution* 36 (4): 423–58.
- Smith, Richard J, and James M Cheverud. 2002. "Scaling of Sexual Dimorphism in Body Mass: A Phylogenetic Analysis of Rensch's Rule in Primates." *International Journal of Primatology* 23 (5): 1095–135.
- Snyder, Robin E, and Stephen P Ellner. 2018. "Pluck or Luck: Does Trait Variation or Chance Drive Variation in Lifetime Reproductive Success?" *The American Naturalist* 191 (4): E90–107.
- Solomon, Nancy G, Jeffrey A French, and others. 1997. *Cooperative Breeding in Mammals*. Cambridge University Press.
- Stockley, Paula, and Jakob Bro-Jørgensen. 2011. "Female Competition and Its Evolutionary Consequences in Mammals." *Biological Reviews* 86 (2): 341–66.
- Swedell, Larissa, Liane Leedom, Julian Saunders, and Mathew Pines. 2014. "Sexual Conflict in a Polygynous Primate: Costs and Benefits of a Male-Imposed Mating System." *Behavioral Ecology and Sociobiology* 68 (2): 263–73.
- Thouless, CR, and FE Guinness. 1986. "Conflict Between Red Deer Hinds: The Winner Always Wins." *Animal Behaviour* 34 (4): 1166–71.
- Upham, Nathan S, Jacob A Esselstyn, and Walter Jetz. 2019. "Inferring the Mammal Tree: Species-Level Sets of Phylogenies for Questions in Ecology, Evolution, and Conservation." *PLoS Biology* 17 (12).
- Van Noordwijk, Maria A, and Carel P Van Schaik. 1988. "Scramble and Contest in Feeding Competition Among Female Long-Tailed Macaques (*Macaca Fascicularis*)." *Behaviour* 105 (1-2): 77–98.
- Vehrencamp, Sandra L. 1983. "A Model for the Evolution of Despotism Versus Egalitarian Societies." *Animal Behaviour* 31 (3): 667–82.
- Viechtbauer, Wolfgang. 2010. "Conducting Meta-Analyses in r with the Metafor Package." *Journal of Statistical Software* 36 (3): 1–48.

- Ward, Ashley, and Mike Webster. 2016. *Sociality: The Behaviour of Group-Living Animals*. Springer.
- West, Mary Jane. 1967. "Foundress Associations in Polistine Wasps: Dominance Hierarchies and the Evolution of Social Behavior." *Science* 157 (3796): 1584–85.
- Williams, George C. 1966. *Adaptation and Natural Selection: A Critique of Some Current Evolutionary Thought*. Vol. 833082108. Princeton science library OCLC.
- Wilman, Hamish, Jonathan Belmaker, Jennifer Simpson, Carolina De La Rosa, Marcelo M Rivadeneira, and Walter Jetz. 2014. "EltonTraits 1.0: Species-Level Foraging Attributes of the World's Birds and Mammals: Ecological Archives E095-178." *Ecology* 95 (7): 2027–27.
- Wilson, D. B. 2019. *Practical Meta-Analysis Effect Size Calculator [Online Calculator]*. retrieved from: <https://www.campbellcollaboration.org/research-resources/research-for-resources/effect-size-calculator.html>.

Supplement: The effect of dominance rank on female reproductive success in social mammals

Shivani, Elise Huchard, Dieter Lukas

07/10/2021

Supplementary data

Data Table. References for the effect sizes of dominance rank on female reproductive success, for the dominance system in a given population, and for the average relatedness among females in social groups in a given population.

Id	Species	Reference effect size	Reference dominance system	Reference relatedness
1	<i>Cervus elaphus</i>	(Clutton-Brock, et al. 1984)	(HALL, 2010)	(Nussey, et al., 2005)
2	<i>Crocuta crocuta</i>	(Holekamp, et al., 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
3	<i>Macaca arctoides</i>	(Nieuwenhuijsen, et al., 1985)	(HOLEKAMP and SMALE, 1991)	NA
4	<i>Macaca fuscata</i>	(Gouzoules, et al. 1982)	(Koyama et al. 2003)	(Baxter and Fedigan, 1979)
5	<i>Macaca fuscata</i>	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
6	<i>Macaca fuscata</i>	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
7	<i>Macaca fuscata</i>	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
8	<i>Macaca mulatta</i>	(Drickamer, 1974)	(Deutsch and Lee, 1991)	NA
9	<i>Mandrillus sphinx</i>	(Setchell, et al. 2005)	(Setchell et al. 2002)	NA
10	<i>Papio cynocephalus</i>	(, 2021)	(Packer, et al., 1995)	NA
11	<i>Papio cynocephalus</i>	(Wasser, et al., 2004)	(Packer, et al., 1995)	(Wasser and Starling, 1988)
12	<i>Rangifer tarandus</i>	(Holand, et al., 2004)	(Holand, et al., 2004)	(Djakovifa et al., 2011)
13	<i>Callithrix jacchus</i>	(Sousa, et al., 2005)	(Digby, 1995)	(Nievergelt et al. 2009)
14	<i>Chlorocebus aethiops</i>	(Fairbanks and McGuire, 1984)	(HOLEKAMP and SMALE, 1991)	(Fairbanks, et al., 2011)
15	<i>Chlorocebus aethiops</i>	(Fairbanks and McGuire, 1984)	(HOLEKAMP and SMALE, 1991)	(Fairbanks, et al., 2011)
16	<i>Crocuta crocuta</i>	(Holekamp, et al., 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
17	<i>Crocuta crocuta</i>	(Holekamp, et al., 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
18	<i>Lemur catta</i>	(Takahata, et al., 2007)	(Taylor and Sussman, 1985)	(Parga, et al., 2015)
19	<i>Macaca fuscata</i>	(Gouzoules, et al. 1982)	(Koyama et al. 2003)	(Baxter and Fedigan, 1979)
20	<i>Macaca fuscata</i>	(Gouzoules, et al. 1982)	(Koyama et al. 2003)	(Baxter and Fedigan, 1979)
21	<i>Macaca fuscata</i>	(Wolfe, 1984)	(Koyama et al. 2003)	(Koyama et al. 2003)
22	<i>Macaca sylvanus</i>	(Kümmerli and Martin, 2005)	(Paul and Kuester, 1987)	(Kuemmerli and Martin, 2008)
23	<i>Macaca sylvanus</i>	(Kümmerli and Martin, 2005)	(Paul and Kuester, 1987)	(Kuemmerli and Martin, 2008)
24	<i>Mesocricetus auratus</i>	(Huck, Lisk, and McKay, 1988)	(Huck, Lisk, and McKay, 1988)	(Huck, Lisk, and McKay, 1988)
25	<i>Mesocricetus auratus</i>	(Huck, Lisk, and McKay, 1988)	(Huck, Lisk, and McKay, 1988)	(Huck, Lisk, and McKay, 1988)

26	<i>Mesocricetus auratus</i>	(Huck, Lisk, and McKay, 1988)	(Huck, Lisk, and McKay, 1988)	(Huck, Lisk, and McKay, 1988)
27	<i>Oreamnos americanus</i>	(Cote and Festa-Bianchet, 2001)	(Cote, 2000)	(Shafer, et al., 2012)
28	<i>Oryctolagus cuniculus</i>	(von Holst, et al., 2002)	(von Holst, et al., 2002)	(Surrridge, et al., 1999)
29	<i>Oryctolagus cuniculus</i>	(von Holst, et al., 2002)	(von Holst, et al., 2002)	(Surrridge, et al., 1999)
30	<i>Papio cynocephalus</i>	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
31	<i>Semnopithecus entellus</i>	(Borries, et al. 1991)	(Borries, Sommer, and Srivastava, 1991)	NA
32	<i>Rangifer tarandus</i>	(Holand, et al., 2004)	(Holand, Gjonstein, Losvar, et al., 2004)	(Djakovifa et al., 2011)
33	<i>Sciurus vulgaris</i>	(Wauters and Dhondt, 1989)	(Wauters and Dhondt, 1989)	NA
34	<i>Sciurus vulgaris</i>	(Wauters and Dhondt, 1989)	(Wauters and Dhondt, 1989)	NA
35	<i>Theropithecus gelada</i>	(DUNBAR and DUNBAR, 1977)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)
36	<i>Papio ursinus</i>	(Cheney et al. 2006)	(HOLEKAMP and SMALE, 1991)	(Silk, Cheney, and Seyfarth, 1999)
37	<i>Papio ursinus</i>	(Bulger and Hamilton, 1987)	(HOLEKAMP and SMALE, 1991)	(Silk, Cheney, and Seyfarth, 1999)
38	<i>Papio ursinus</i>	(Bulger and Hamilton, 1987)	(HOLEKAMP and SMALE, 1991)	(Silk, Cheney, and Seyfarth, 1999)
39	<i>Cervus elaphus</i>	(Clutton-Brock, et al., 1984)	(HALL, 2010)	(Nussey, et al., 2005)
40	<i>Crocuta crocuta</i>	(Holekamp, et al. 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
41	<i>Gorilla beringei</i>	(Robbins, et al., 2007)	(Robbins, et al., 2007)	(Watts, 1994)
42	<i>Lemur catta</i>	(Takahata, et al., 2007)	(Taylor and Sussman, 1985)	(Parga, et al., 2015)
43	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
44	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
45	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
46	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
47	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
48	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
49	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
50	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
51	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
52	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
53	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
54	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
55	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
56	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
57	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
58	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
59	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
60	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
61	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
62	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
63	<i>Macaca fascicularis</i>	(VanNoordwijk & VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
64	<i>Macaca fuscata</i>	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
65	<i>Macaca mulatta</i>	(Meikle and Vessey, 1988)	(Deutsch and Lee, 1991)	NA
66	<i>Oreamnos americanus</i>	(Cote and Festa-Bianchet, 2001)	(Fa, 2000)	(Shafer, et al., 2012)
67	<i>Oreamnos americanus</i>	(Cote and Festa-Bianchet, 2001)	(Fa, 2000)	(Shafer, et al., 2012)
68	<i>Oryctolagus cuniculus</i>	(von Holst, et al., 2002)	(von Holst, et al., 2002)	(Surrridge, et al., 1999)
69	<i>Pan troglodytes</i>	(Pusey, 1997)	(Wittig et al. 2003)	(Vigilant, et al., 2001)

70	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
71	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
72	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
73	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
74	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
75	Papio_cynocephalus	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
76	Papio_cynocephalus	(Silk, 2003)	(Packer, Collins, Sindimwo, et al., 1995)	(Horn, et al., 2007)
77	Papio_cynocephalus	(Silk, 2003)	(Packer, Collins, Sindimwo, et al., 1995)	(Horn, et al., 2007)
78	Semnopithecus_entellus	(Borries, et al., 1991)	(Borries, Sommer, and Srivastava, 1991)	NA
79	Semnopithecus_entellus	(Borries, et al., 1991)	(Borries, Sommer, and Srivastava, 1991)	NA
80	Crocota_crocota	(Hofer and East, 2003)	(Hofer and East, 2003)	NA
81	Papio_ursinus	(Cheney et al. 2006)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
82	Papio_ursinus	(Cheney et al. 2006)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
83	Papio_ursinus	(Bulger and Hamilton, 1987)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
84	Papio_ursinus	(Bulger and Hamilton, 1987)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
85	Macaca_fuscata	(Gouzoules, et al., 1982)	(Koyama et al. 2003)	(Baxter and Fedigan, 1979)
86	Macaca_fuscata	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
87	Mandrillus_sphinx	(Setchell et al. 2002)	(Setchell et al. 2002)	NA
88	Papio_anubis	(Cheney et al. 2006)	(Johnson, 1987)	NA
89	Papio_ursinus	NA	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
90	Papio_ursinus	(Cheney et al. 2006)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
91	Chlorocebus_aethiops	(Fairbanks and McGuire, 1984)	(HOLEKAMP and SMALE, 1991)	(Fairbanks, et al., 2011)
92	Crocota_crocota	(Holekamp, et al., 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
93	Crocota_crocota	(Holekamp, et al., 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
94	Crocota_crocota	(Holekamp, et al., 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
95	Crocota_crocota	(Holekamp, et al., 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
96	Crocota_crocota	(Holekamp, et al., 1996)	(Hofer and East, 2003)	(Horn, et al., 2004)
97	Gorilla_beringei	(Robbins, et al., 2007)	(Robbins, et al., 2005)	(Watts, 1994)
98	Macaca_arctoides	(Nieuwenhuijsen, et al., 1985)	(HOLEKAMP and SMALE, 1991)	NA
99	Mandrillus_sphinx	(Setchell et al. 2002)	(Setchell et al. 2002)	NA
100	Mandrillus_sphinx	(Setchell et al. 2002)	(Setchell et al. 2002)	NA
101	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	NA
102	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
103	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	NA
104	Papio_anubis	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
105	Papio_anubis	(Garcia, Lee, and Rosetta, 2006)	(Johnson, 1987)	NA
106	Papio_anubis	(Garcia, Lee, and Rosetta, 2006)	(Johnson, 1987)	NA
107	Papio_cynocephalus	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
108	Papio_cynocephalus	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
109	Papio_cynocephalus	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
110	Papio_anubis	(Barton and Whiten, 1993)	(Johnson, 1987)	(Lynch 2016)
111	Papio_ursinus	(Bulger and Hamilton, 1987)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
112	Papio_ursinus	(Bulger and Hamilton, 1987)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
113	Gorilla_beringei	(Robbins, et al., 2007)	(Robbins, et al., 2005)	(Watts, 1994)

114	<i>Macaca fascicularis</i>	(VanNoordwijk VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
115	<i>Macaca fascicularis</i>	(VanNoordwijk VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
116	<i>Macaca fascicularis</i>	(VanNoordwijk VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
117	<i>Macaca fascicularis</i>	(VanNoordwijk VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
118	<i>Macaca fascicularis</i>	(VanNoordwijk VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
119	<i>Macaca fascicularis</i>	(VanNoordwijk VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
120	<i>Macaca fascicularis</i>	(VanNoordwijk VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
121	<i>Macaca fascicularis</i>	(VanNoordwijk VanSchaik, 1999)	(van Noordwijk and van Schaik, 1987)	(Ruiter and Geffen, 1998)
122	<i>Macaca fuscata</i>	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
123	<i>Macaca fuscata</i>	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
124	<i>Macaca fuscata</i>	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
125	<i>Macaca fuscata</i>	(Takahata, et al., 1998)	(Koyama et al. 2003)	(Nakagawa, et al., 2015)
126	<i>Mandrillus sphinx</i>	(Setchell, et al., 2005)	(Setchell et al. 2002)	NA
127	<i>Ovis canadensis</i>	(Festa-Bianchet, 1991)	(Festa-Bianchet, 1991)	(Fournier & Festa-Bianchet, 1995)
128	<i>Papio anubis</i>	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
129	<i>Papio anubis</i>	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
130	<i>Papio cynocephalus</i>	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
131	<i>Crocuta crocuta</i>	(Hofer and East, 2003)	(Hofer and East, 2003)	NA
132	<i>Macaca fuscata</i>	(Takahata, 1980)	(Koyama et al. 2003)	(Koyama)2003
133	<i>Oryctolagus cuniculus</i>	(von Holst, Hutzelmeyer, Kaetzke, et al., 2002)	(von Holst, Hutzelmeyer, Kaetzke, et al., 2002)	(Surrridge, et al., 1999)
134	<i>Papio anubis</i>	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
135	<i>Papio anubis</i>	(Packer, et al., 1995)	(Johnson, 1987)	(Kopp 2015)
136	<i>Papio cynocephalus</i>	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
137	<i>Papio cynocephalus</i>	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
138	<i>Papio cynocephalus</i>	(Wasser, et al., 2004)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser and Starling, 1988)
139	<i>Crocuta crocuta</i>	(Hofer and East, 2003)	(Hofer and East, 2003)	NA
140	<i>Papio ursinus</i>	(Cheney et al. 2006)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
141	<i>Papio ursinus</i>	(Cheney et al. 2006)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
142	<i>Cervus elaphus</i>	(Clutton-Brock, et al., 1984)	(HALL, 2010)	(Nussey, et al., 2005)
143	<i>Cervus elaphus</i>	(Clutton-Brock, et al., 1984)	(HALL, 2010)	(Nussey, et al., 2005)
144	<i>Macaca mulatta</i>	(Wilson, et al., 1978)	(Deutsch and Lee, 1991)	(Bernstein and Ehardt, 1986)
145	<i>Macaca mulatta</i>	(Wilson, et al., 1978)	(Deutsch and Lee, 1991)	(Bernstein and Ehardt, 1986)
146	<i>Macaca sinica</i>	(Dittus, 1979)	(Dittus, 1986)	NA
147	<i>Macaca sinica</i>	(Dittus, 1979)	(Dittus, 1986)	NA
148	<i>Lycaon pictus</i>	(Creel, et al., 1997)	(Spiering, et al., 2009)	(Girman, et al., 1997)
149	<i>Fukomys damarensis</i>	(Burland, et al., 2004)	(Gaylard, Harrison, and Bennett, 1998)	(Burland, et al., 2002)
150	<i>Macaca fuscata</i>	(Fedigan, et al., 1986)	(Koyama et al. 2003)	(Baxter and Fedigan, 1979)
151	<i>Macaca fuscata</i>	(Fedigan, et al., 1986)	(Koyama et al. 2003)	(Baxter and Fedigan, 1979)
152	<i>Macaca fuscata</i>	(Fedigan, et al., 1986)	(Koyama et al. 2003)	(Baxter and Fedigan, 1979)
153	<i>Macaca fuscata</i>	(Fedigan, et al., 1986)	(Koyama et al. 2003)	(Baxter and Fedigan, 1979)
154	<i>Helogale parvula</i>	(Keane, et al., 1994)	(Creel, 2005)	(Creel and Waser, 1994)
155	<i>Helogale parvula</i>	(Keane, et al., 1994)	(Creel, 2005)	(Creel and Waser, 1994)
156	<i>Helogale parvula</i>	(Keane, et al., 1994)	(Creel, 2005)	(Creel and Waser, 1994)
157	<i>Marmota caligata</i>	(Wasser and Barash, 1983)	(Patil, Karels, and Hik, 2015)	NA

158	<i>Marmota caligata</i>	(Wasser and Barash, 1983)	(Patil, Karels, and Hik, 2015)	NA
159	<i>Marmota caligata</i>	(Wasser and Barash, 1983)	(Patil, Karels, and Hik, 2015)	NA
160	<i>Marmota caligata</i>	(Wasser and Barash, 1983)	(Patil, Karels, and Hik, 2015)	NA
161	<i>Macaca radiata</i>	(Silk, et al., 1981)	(HOLEKAMP and SMALE, 1991)	NA
162	<i>Macaca radiata</i>	(Silk, et al., 1981)	(HOLEKAMP and SMALE, 1991)	NA
163	<i>Macaca radiata</i>	(Silk, et al., 1981)	(HOLEKAMP and SMALE, 1991)	NA
164	<i>Marmota flaviventris</i>	(Huang, et al., 2011)	(Huang, Wey, and Blumstein, 2011)	(Armitage, et al., 2011)
165	<i>Marmota flaviventris</i>	(Huang, et al., 2011)	(Huang, Wey, and Blumstein, 2011)	(Armitage, et al., 2011)
166	<i>Marmota flaviventris</i>	(Huang, et al., 2011)	(Huang, Wey, and Blumstein, 2011)	(Armitage, et al., 2011)
167	<i>Marmota flaviventris</i>	(Huang, et al., 2011)	(Huang, Wey, and Blumstein, 2011)	(Armitage, et al., 2011)
168	<i>Alouatta palliata</i>	(Glander, 1980)	(Jones, 1980)	NA
169	<i>Alouatta palliata</i>	(Glander, 1980)	(Jones, 1980)	NA
170	<i>Equus quagga</i>	(Pluhacek, and Plausik, 2006)	(Lloyd and Rasa, 1994)	NA
171	<i>Equus quagga</i>	(Pluhacek, and Plausik, 2006)	(Lloyd and Rasa, 1994)	NA
172	<i>Equus zebra</i>	(Lloyd and Rasa, 1989)	(Lloyd and Rasa, 1994)	NA
173	<i>Equus zebra</i>	(Lloyd and Rasa, 1989)	(Lloyd and Rasa, 1994)	NA
174	<i>Equus zebra</i>	(Lloyd and Rasa, 1989)	(Lloyd and Rasa, 1994)	NA
175	<i>Equus zebra</i>	(Lloyd and Rasa, 1989)	(Lloyd and Rasa, 1994)	NA
176	<i>Equus zebra</i>	(Lloyd and Rasa, 1989)	(Lloyd and Rasa, 1994)	NA
177	<i>Equus caballus</i>	(Rubenstein et al. 2009)	(Sinderbrand 2011)	NA
178	<i>Equus caballus</i>	(Rubenstein et al. 2009)	(Sinderbrand 2011)	NA
179	<i>Equus caballus</i>	(Rubenstein et al. 2009)	NA	NA
180	<i>Mirounga angustirostris</i>	(Cheney et al. 1988)	(Christenson and Boeuf, 1978)	NA
181	<i>Ovis canadensis</i>	(Hass, 1991)	(Festa-Bianchet, 1991)	(Fournier & Festa-Bianchet, 1995)
182	<i>Ovis canadensis</i>	(Hass, 1991)	(Festa-Bianchet, 1991)	(Fournier & Festa-Bianchet, 1995)
183	<i>Ovis canadensis</i>	(Hass, 1991)	(Festa-Bianchet, 1991)	(Fournier & Festa-Bianchet, 1995)
184	<i>Hyaena brunnea</i>	(Owens and Owens, 1996)	(OWENS and OWENS, 1996)	(Knowles, et al., 2009)
185	<i>Hyaena brunnea</i>	(Owens and Owens, 1996)	(OWENS and OWENS, 1996)	(Knowles, et al., 2009)
186	<i>Mus musculus</i>	(Rusu and Krackow, 2004)	(Rusu and Krackow, 2004)	(Rusu and Krackow, 2004)
187	<i>Mus musculus</i>	(Koenig, 1994)	(Rusu and Krackow, 2004)	(Koenig, 1994)
188	<i>Mus musculus</i>	(Koenig, 1994)	(Rusu and Krackow, 2004)	(Koenig, 1994)
189	<i>Mus musculus</i>	(Koenig, 1994)	(Rusu and Krackow, 2004)	(Koenig, 1994)
190	<i>Mus musculus</i>	(Koenig, 1994)	(Rusu and Krackow, 2004)	(Koenig, 1994)
191	<i>Rhabdomys pumilio</i>	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)
192	<i>Rhabdomys pumilio</i>	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)
193	<i>Rhabdomys pumilio</i>	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)
194	<i>Rhabdomys pumilio</i>	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)
195	<i>Rhabdomys pumilio</i>	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)
196	<i>Rhabdomys pumilio</i>	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)	(Kinahan and Pillay, 2007)
197	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
198	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
199	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
200	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
201	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)

202	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
203	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
204	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
205	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
206	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
207	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
208	<i>Apodemus sylvaticus</i>	(Gerlach, 2002)	(Gerlach, 2002)	(Gerlach, 2002)
209	<i>Rattus norvegicus</i>	(Schultz and Lore, 1993)	(Ziporyn and McClintock, 1991)	(Schultz and Lore, 1993)
210	<i>Marmota marmota</i>	(Hacklaender, et al., 2003)	(Lardy, and Cohas, 2013)	(Hacklaender, et al. 2003)
211	<i>Heterocephalus glaber</i>	(Faulkes and Bennett, 2001)	(Clarke and Faulkes, 1997)	NA
212	<i>Fukomys damarensis</i>	(Faulkes and Bennett, 2001)	(Gaylard, Harrison, and Bennett, 1998)	(Burland, et al., 2002)
213	<i>Cryptomys hottentotus</i>	(Faulkes and Bennett, 2001)	(Gaylard, Harrison, and Bennett, 1998)	NA
214	<i>Suricata suricatta</i>	(Griffin, 2003)	(Russell, et al., 2004)	(Griffin, 2003)
215	<i>Leontopithecus rosalia</i>	(Henry, et al., 2013)	(Baker et al. 2002)	NA
216	<i>Leontopithecus rosalia</i>	(Henry, et al., 2013)	(Baker et al. 2002)	NA
217	<i>Leontopithecus rosalia</i>	(Henry, et al., 2013)	(Baker et al. 2002)	NA
218	<i>Leontopithecus rosalia</i>	(Dietz and Baker, 1993)	NA	NA
219	<i>Leontopithecus fuscicollis</i>	(Goldizen, et al., 1996)	(Goldizen, et al., 1996)	NA
220	<i>Saguinus mystax</i>	(Garber, et al., 1993)	(Smith 2000)	NA
221	<i>Cebus capucinus</i>	(Fedigan, et al, 2008)	(Fedigan and Bergstrom, 2010)	NA
222	<i>Cebus capucinus</i>	(Fedigan, et al, 2008)	(Fedigan and Bergstrom, 2010)	NA
223	<i>Cercopithecus mitis</i>	(Cords, 2002)	(Klass and Cords, 2015)	NA
224	<i>Chlorocebus aethiops</i>	NA	(HOLEKAMP and SMALE, 1991)	NA
225	<i>Chlorocebus aethiops</i>	(Cheney et al. 1988)	(HOLEKAMP and SMALE, 1991)	NA
226	<i>Chlorocebus aethiops</i>	(Cheney et al. 1988)	(HOLEKAMP and SMALE, 1991)	NA
227	<i>Chlorocebus aethiops</i>	(Whitten et al. 1983)	(HOLEKAMP and SMALE, 1991)	NA
228	<i>Chlorocebus aethiops</i>	(Whitten et al. 1983)	(HOLEKAMP and SMALE, 1991)	NA
229	<i>Chlorocebus aethiops</i>	(Whitten et al. 1983)	(HOLEKAMP and SMALE, 1991)	NA
230	<i>Chlorocebus aethiops</i>	(Whitten et al. 1983)	(HOLEKAMP and SMALE, 1991)	NA
231	<i>Pan troglodytes</i>	(Jones, et al., 2010)	(Wittig et al. 2003)	(Vigilant, et al., 2001)
232	<i>Papio anubis</i>	(Smuts and Nicolson, 1989)	(Johnson, 1987)	NA
233	<i>Papio anubis</i>	(Smuts and Nicolson, 1989)	(Johnson, 1987)	NA
234	<i>Macaca fuscata</i>	(Itoigawa, et al. 1992)	(Koyama et al. 2003)	NA
235	<i>Macaca fuscata</i>	(Itoigawa, et al., 1992)	(Koyama et al. 2003)	NA
236	<i>Macaca fuscata</i>	(Itoigawa, et al., 1992)	(Koyama et al. 2003)	NA
237	<i>Macaca fuscata</i>	(Itoigawa, et al., 1992)	(Koyama et al. 2003)	NA
238	<i>Macaca fuscata</i>	(Itoigawa, et al., 1992)	(Koyama et al. 2003)	NA
239	<i>Macaca fuscata</i>	(Itoigawa, et al., 1992)	(Koyama et al. 2003)	NA
240	<i>Ovis canadensis</i>	(Eccles and Shackleton, 1986)	(Festa-Bianchet, 1991)	(Fournier & Festa-Bianchet, 1995)
241	<i>Ovis canadensis</i>	(Eccles and Shackleton, 1986)	(Festa-Bianchet, 1991)	(Fournier & Festa-Bianchet, 1995)
242	<i>Ammotragus lervia</i>	(Cassinello and Alados, 1996)	(Cassinello, 1995)	NA
243	<i>Ammotragus lervia</i>	(Cassinello and Alados, 1996)	(Cassinello, 1995)	NA
244	<i>Ammotragus lervia</i>	(Cassinello and Alados, 1996)	(Cassinello, 1995)	NA
245	<i>Ammotragus lervia</i>	(Cassinello and Alados, 1996)	(Cassinello, 1995)	NA

246	<i>Antilocapra americana</i>	(Clancey and Byers, 2015)	(Dennehy, 2001)	(Carling, et al., 2003)
247	<i>Antilocapra americana</i>	(Clancey and Byers, 2015)	(Dennehy, 2001)	(Carling, et al., 2003)
248	<i>Antilocapra americana</i>	(Clancey and Byers, 2015)	(Dennehy, 2001)	(Carling, et al., 2003)
249	<i>Nanger dama</i>	(Alados and Escez, 1992)	(Alados and Escvez, 2021)	NA
250	<i>Gazella cuvieri</i>	(Alados and Escez, 1992)	(Alados and Escvez, 2021)	NA
251	<i>Gazella cuvieri</i>	(Alados and Escez, 1992)	(Alados and Escvez, 2021)	NA
252	<i>Gazella cuvieri</i>	(Alados and Escez, 1992)	(Alados and Escvez, 2021)	NA
253	<i>Gazella cuvieri</i>	(Alados and Escez, 1992)	(Alados and Escvez, 2021)	NA
254	<i>Nanger dama</i>	(Alados and Escez, 1992)	(Alados and Escvez, 2021)	NA
255	<i>Nanger dama</i>	(Alados and Escez, 1992)	(Alados and Escvez, 2021)	NA
256	<i>Nanger dama</i>	(Alados and Escez, 1992)	(Alados and Escvez, 2021)	NA
257	<i>Capra nubiana</i>	(Shargal, et al., 2008)	(Greenberg-Cohen, et al., 2010)	NA
258	<i>Ozotoceros bezoarticus</i>	(Morales-Picerva, et al., 2014)	(Morales-Pisterva, et al., 2014)	NA
259	<i>Ozotoceros bezoarticus</i>	(Morales-Picerva, et al., 2014)	(Morales-Pisterva, et al., 2014)	NA
260	<i>Mus musculus</i>	(Drickamer, 1985)	(Rusu and Krackow, 2004)	(Drickamer, 1985)
261	<i>Mus musculus</i>	(Drickamer, 1985)	(Rusu and Krackow, 2004)	(Drickamer, 1985)
262	<i>Mus musculus</i>	(Drickamer, 1985)	(Rusu and Krackow, 2004)	(Drickamer, 1985)
263	<i>Helogale parvula</i>	(Rood, 1980)	(Creel, 2005)	(Creel and Waser, 1994)
264	<i>Macaca mulatta</i>	(Gomendio, et al. 1990)	(Deutsch and Lee, 1991)	NA
265	<i>Macaca mulatta</i>	(Gomendio, et al. 1990)	(Deutsch and Lee, 1991)	NA
266	<i>Cervus elaphus</i>	(Gomendio, et al. 1990)	(HALL, 2010)	(Nussey, et al., 2005)
267	<i>Cervus elaphus</i>	(Gomendio, et al. 1990)	(HALL, 2010)	(Nussey, et al., 2005)
268	<i>Macaca mulatta</i>	(Gomendio, et al. 1990)	(Deutsch and Lee, 1991)	NA
269	<i>Crocota crocuta</i>	(Frank et al. 1995)	(Hofer and East, 2003)	(Horn, et al., 2007)
270	<i>Crocota crocuta</i>	(Frank et al. 1995)	(Hofer and East, 2003)	(Horn, et al., 2007)
271	<i>Crocota crocuta</i>	(Frank et al. 1995)	(Hofer and East, 2003)	(Horn, et al., 2007)
272	<i>Crocota crocuta</i>	(Frank et al. 1995)	(Hofer and East, 2003)	(Horn, et al., 2007)
273	<i>Crocota crocuta</i>	(Frank et al. 1995)	(Hofer and East, 2003)	(Horn, et al., 2007)
274	<i>Ateles paniscus</i>	(Symington, 1987)	(van Roosmalen 1980)	NA
275	<i>Crocota crocuta</i>	(White, 2005)	(Hofer and East, 2003)	(Horn, et al., 2007)
276	<i>Crocota crocuta</i>	(White, 2005)	(Hofer and East, 2003)	(Horn, et al., 2007)
277	<i>Crocota crocuta</i>	(White, 2005)	(Hofer and East, 2003)	(Horn, et al., 2007)
278	<i>Petrogale concinna</i>	(Nelson and Goldstone, 1986)	(Nelson and Goldstone, 1986)	NA
279	<i>Macaca assamensis</i>	(Heesen, et al., 2013)	(Fuertbauerr 2011)	(Moor, et al., 2020)
280	<i>Papio ursinus</i>	(Busse 1982)	(HOLEKAMP and SMALE, 1991)	(Silk, et al. 1999)
281	<i>Macaca fuscata</i>	(Wolfe, 1984)	(Koyama et al. 2003)	(Koyama et al. 2003)
282	<i>Macaca fuscata</i>	(Wolfe, 1984)	(Koyama et al. 2003)	(Koyama et al. 2003)
283	<i>Macaca fuscata</i>	(Wolfe, 1984)	(Koyama et al. 2003)	(Koyama et al. 2003)
284	<i>Theropithecus gelada</i>	(le Roux, et al., 2010)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)
285	<i>Theropithecus gelada</i>	(le Roux, et al., 2010)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)
286	<i>Marmota marmota</i>	(King and Cote, 2002)	(Lardy, and Cohas, 2013)	NA
287	<i>Marmota marmota</i>	(King and Cote, 2002)	(Lardy, and Cohas, 2013)	NA
288	<i>Papio cynocephalus</i>	(Beehner, et al., 2006)	(Packer, et al., 1995)	(Horn, et al., 2007)
289	<i>Papio cynocephalus</i>	(Beehner, et al., 2006)	(Packer, et al., 1995)	(Horn, et al., 2007)

290	Papio_cynocephalus	NA	(Packer, et al., 1995)	(Horn, et al., 2007)
291	Papio_cynocephalus	(Altmann & Alberts 2003)	(Packer, et al., 1995)	(Horn, et al., 2007)
292	Papio_ursinus	(Baniel et al. 2021)	(Holekamp and Smale, 1991)	(Baniel, et al. 2018)
293	Vulpes_vulpes	(Baker, et al., 1998)	(Baker et al., 1998)	(Iossa, et al., 2008)
294	Semnopithecus_entellus	(Dolhinow, et al., 1979)	(Borries, Sommer, and Srivastava, 1991)	NA
295	Sapajus_apella	(DiBitetti et al. 2001)	(Welker, et al., 1990)	NA
296	Miopithecus_talapoin	(Abbott, 1987)	(Abbott, 1987)	NA
297	Mungos_mungo	(Nichols,et al., 2010)	(de Luca and Ginsberg, 2001)	(Nichols, et al., 2012)
298	Mungos_mungo	(Nichols,et al., 2010)	(de Luca and Ginsberg, 2001)	(Nichols, et al., 2012)
299	Mungos_mungo	(Nichols,et al., 2010)	(de Luca and Ginsberg, 2001)	(Nichols, et al., 2012)
300	Mungos_mungo	(Nichols,et al., 2010)	(de Luca and Ginsberg, 2001)	(Nichols, et al., 2012)
301	Mungos_mungo	(de Luca and Ginsberg, 2001)	(de Luca and Ginsberg, 2001)	(Nichols, et al., 2012)
302	Canis_simensis	(Randall, et al., 2007)	(HOLEKAMP and SMALE, 1991)	(Randall, et al., 2007)
303	Procapra_capensis	(Koren and Geffen, 2009)	(Visser, Robinson, and van Vuuren, 2020)	(Visser 2013)
304	Bison_bison	(Vervaecke, Roden, and de Vries, 2005)	(Vervaecke, Roden, and de Vries, 2005)	NA
305	Bison_bison	(Vervaecke, Roden, and de Vries, 2005)	(Vervaecke, Roden, and de Vries, 2005)	NA
306	Capra_pyrenaica	(Santiago-Moreno, et al., 2007)	(Santiago et al. 2013)	NA
307	Sus_scrofa	(Meikle, et al., 2010)	(Gaillard et al. 1993)	(Meikle, et al., 2010)
308	Papio_cynocephalus	(Altmann et al. 1988)	(Packer, Collins, Sindimwo, et al., 1995)	(Horn, et al., 2007)
309	Macaca_sylvanus	(Paul & Kuester 1996)	(Paul and Kuester, 1987)	(Kuemmerli and Martin, 2008)
310	Macaca_sylvanus	(Paul & Kuester 1996)	(Paul and Kuester, 1987)	(Kuemmerli and Martin, 2008)
311	Macaca_sylvanus	NA	(Paul and Kuester, 1987)	(Kuemmerli and Martin, 2008)
312	Papio_ursinus	(Baniel et al. 2021)	(HOLEKAMP and SMALE, 1991)	(Baniel, et al., 2018)
313	Papio_ursinus	(Baniel et al. 2021)	(HOLEKAMP and SMALE, 1991)	(Baniel, et al., 2018)
314	Papio_ursinus	(McFarland, et al., 2017)	(HOLEKAMP and SMALE, 1991)	NA
315	Papio_ursinus	(McFarland, et al., 2017)	(HOLEKAMP and SMALE, 1991)	NA
316	Papio_cynocephalus	(McFarland, et al., 2017)	(Packer, Collins, Sindimwo, et al., 1995)	(Horn, et al., 2007)
317	Lama_guanicoe	(Correa, et al., 2013)	(Correa, et al., 2013)	NA
318	Bos_taurus	(Hohenbrink et al., 2012)	(Spinka et al., 2013)	NA
319	Capra_hircus	(Barroso, et al., 2000)	(Barroso, Alados, and Boza, 2000)	NA
320	Sus_scrofa	(Mendl, et al. 1995)	(Cappa, Lombardini, and Meriggi, 2021)	NA
321	Bison_bison	(Green and Rothstein, 1991)	(Vervaecke, Roden, and de Vries, 2005)	NA
322	Bison_bison	(Green and Rothstein, 1991)	(Vervaecke, Roden, and de Vries, 2005)	NA
323	Antilocapra_americana	(Byers 1997)	(Dennehy, 2001)	(Carling, et al., 2003)
324	Antilocapra_americana	(Byers 1997)	(Dennehy, 2001)	(Carling, et al., 2003)
325	Antilocapra_americana	(Byers 1997)	(Dennehy, 2001)	(Carling, et al., 2003)
326	Antilocapra_americana	(Byers 1997)	(Dennehy, 2001)	(Carling, et al., 2003)
327	Suricata_suricatta	(MacLeod & Clutton-Brock, 2013)	(Russell, Carlson, McIlrath, et al., 2004)	(Griffin, 2003)
328	Suricata_suricatta	(MacLeod & Clutton-Brock, 2013)	(Russell, Carlson, McIlrath, et al., 2004)	(Griffin, 2003)
329	Mesocricetus_auratus	(Pratt and Lisk, 1989)	(Huck, Lisk, and McKay, 1988)	(Huck, et al. 1988)
330	Mesocricetus_auratus	(Pratt and Lisk, 1989)	(Huck, Lisk, and McKay, 1988)	(Huck, et al. 1988)
331	Gorilla_beringei	(Robbins, et al., 2011)	(Robbins, Gerald-Steklis, Robbins, et al., 2005)	(Watts, 1994)
332	Gorilla_beringei	(Robbins, et al., 2011)	(Robbins, Gerald-Steklis, Robbins, et al., 2005)	(Watts, 1994)
333	Gorilla_beringei	(Robbins, et al., 2011)	(Robbins, Gerald-Steklis, Robbins, et al., 2005)	(Watts, 1994)

334	Papio_anubis	(Smuts and Nicolson, 1989)	(Johnson, 1987)	NA
335	Papio_anubis	(Smuts and Nicolson, 1989)	(Johnson, 1987)	NA
336	Papio_anubis	(Smuts and Nicolson, 1989)	(Johnson, 1987)	NA
337	Macaca_mulatta	(Small and Hrdy, 1986)	(Deutsch and Lee, 1991)	NA
338	Cercopithecus_mitis	(Roberts and Cords, 2013)	(Klass and Cords, 2015)	NA
339	Suricata_suricatta	(Macdonald and Doolan, 1997)	(Russell, Carlson, McIlrath, et al., 2004)	NA
340	Microtus_arvalis	(Dobly, 2008)	(Dobly, 2008)	(Dobly, 2008)
341	Microtus_ochrogaster	(Wolff, et al., 2001)	(Wolff, Dunlap, and Ritchhart, 2001)	(Wolff, et al., 2001)
342	Microtus_pinetorum	(Wolff, et al., 2001)	(Wolff, Dunlap, and Ritchhart, 2001)	(Wolff, et al., 2001)
343	Macaca_mulatta	(Meikle, et al. 1984)	(Deutsch and Lee, 1991)	NA
344	Macaca_sylvanus	(Paul and Thommen, 1984)	(Paul and Kuester, 1987)	NA
345	Macaca_sylvanus	(Paul and Thommen, 1984)	(Paul and Kuester, 1987)	NA
346	Macaca_sylvanus	(Paul and Thommen, 1984)	(Paul and Kuester, 1987)	NA
347	Equus_quagga	(Schilder and Boer, 1987)	(Lloyd and Rasa, 1994)	NA
348	Equus_quagga	(Schilder and Boer, 1987)	(Lloyd and Rasa, 1994)	NA
349	Macaca_mulatta	(Berman, 1988)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
350	Macaca_arctoides	(Rhine, 1994)	(HOLEKAMP and SMALE, 1991)	NA
351	Papio_cynocephalus	(Rhine, et al., 1992)	(Packer, Collins, Sindimwo, et al., 1995)	(Wasser & Starling, 1988)
352	Canis_latrans	(Gese 2004)	(Gese 2004)	NA
353	Canis_latrans	(Gese 2004)	(Gese 2004)	NA
354	Macaca_mulatta	(Brent, et al. 2017)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
355	Suricata_suricatta	(Cram,et al., 2018)	(Russell, Carlson, McIlrath, et al., 2004)	(Griffin, 2003)
356	Fukomys_mechowi	(Dammann, et al., 2011)	(Wallace and Bennett, 1998)	(Dammann, et al., 2011)
357	Papio_ursinus	(Silk, et al. 2010)	(HOLEKAMP and SMALE, 1991)	(Silk, et al., 1999)
358	Papio_cynocephalus	(Archie, et al., 2014)	(Packer, Collins, Sindimwo, et al., 1995)	(Horn, et al., 2007)
359	Crocata_crocata	(Watts, et al., 2009)	(Hofer and East, 2003)	(Horn, et al., 2007)
360	Crocata_crocata	(Strauss and Holekamp, 2019)	(Hofer and East, 2003)	(Horn, et al., 2007)
361	Propithecus_verreauxi	(Kubzdela 1998)	(Kubzdela 1998)	(Lawler, et al. 2003)
362	Propithecus_verreauxi	(Kubzdela 1998)	(Kubzdela 1998)	(Lawler, et al. 2003)
363	Propithecus_verreauxi	(Kubzdela 1998)	(Kubzdela 1998)	(Lawler, et al. 2003)
364	Macaca_mulatta	(Blomquist, et al., 2010)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
365	Macaca_mulatta	(Blomquist, et al., 2010)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
366	Macaca_mulatta	(Blomquist, et al., 2010)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
367	Papio_ursinus	(Ron, Henzi, and Motro, 1996)	(HOLEKAMP and SMALE, 1991)	NA
368	Papio_ursinus	(Ron, Henzi, and Motro, 1996)	(HOLEKAMP and SMALE, 1991)	NA
369	Papio_ursinus	(Ron, Henzi, and Motro, 1996)	(HOLEKAMP and SMALE, 1991)	NA
370	Macaca_mulatta	(Simpson and Simpson, 1982)	(Deutsch and Lee, 1991)	NA
371	Macaca_fuscata	(Koyama, et al. 1992)	(Koyama et al. 2003)	(Koyama et al. 2003)
372	Macaca_fuscata	(Koyama, et al. 1992)	(Borries, Sommer, and Srivastava, 1991)	(Koyama et al. 2003)
373	Macaca_mulatta	(Maestripieri, 2001)	(Deutsch and Lee, 1991)	(Bernstein & Ehardt, 1986)
374	Macaca_mulatta	(Maestripieri, 2001)	(Deutsch and Lee, 1991)	(Bernstein & Ehardt, 1986)
375	Semnopithecus_schistaceus	(Vries et al., 2016)	(VRIES, KOENIG, and BORRIES, 2016)	NA
376	Semnopithecus_schistaceus	(Vries et al., 2016)	(VRIES, KOENIG, and BORRIES, 2016)	NA
377	Semnopithecus_schistaceus	(Vries et al., 2016)	(VRIES, KOENIG, and BORRIES, 2016)	NA

378	Mungos_mungo	(Sanderson, et al. 2015)	(de Luca and Ginsberg, 2001)	(Nichols, et al., 2012)
379	Mungos_mungo	(Sanderson, et al. 2015)	(de Luca and Ginsberg, 2001)	(Nichols, et al., 2012)
380	Mesocricetus_auratus	(Chelini, et al., 2011)	(Huck, Lisk, and McKay, 1988)	(Pratt and Lisk, 1989)
381	Mesocricetus_auratus	(Chelini, et al., 2011)	(Huck, Lisk, and McKay, 1988)	(Pratt and Lisk, 1989)
382	Mesocricetus_auratus	(Chelini, et al., 2011)	(Huck, Lisk, and McKay, 1988)	(Pratt and Lisk, 1989)
383	Macaca_mulatta	(Liu, et al. 2018)	(Deutsch and Lee, 1991)	NA
384	Macaca_mulatta	(Liu, et al. 2018)	(Deutsch and Lee, 1991)	NA
385	Macaca_mulatta	(Liu, et al. 2018)	(Deutsch and Lee, 1991)	NA
386	Macaca_mulatta	(Liu, et al. 2018)	(Deutsch and Lee, 1991)	NA
387	Ceratotherium_simum	(Metrione and Harder, 2011)	(Metrione, Penfold, and Waring, 2007)	(Metrione and Harder, 2011)
388	Cebus_capucinus	(Kalbitzer, et al. 2017)	(Fedigan and Bergstrom, 2010)	NA
389	Canis_lupus	(Cafazzo, et al., 2014)	(Cafazzo, Valsecchi, Bonanni, and Natoli, 2010)	NA
390	Macaca_nigra	(Kerhoas, et al., 2014)	(Duboscq, et al., 2017)	NA
391	Equus_caballus	(Cameron, et al., 2009)	(Sinderbrand 2011)	(Cameron, et al., 2009)
392	Equus_caballus	(Cameron, et al., 2009)	(Sinderbrand 2011)	(Cameron, et al., 2009)
393	Odocoileus_virginianus	(Michel, et al., 2015)	(Townsend and Bailey, 1981)	NA
394	Papio_cynocephalus	(Archie, et al., 2014)	(Packer, Collins, Sindimwo, et al., 1995)	(Horn, et al., 2007)
395	Macaca_mulatta	(Ellis, et al., 2019)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
396	Cervus_elaphus	(Ceacero, et al., 2018)	(HALL, 2010)	(Ceacero, et al., 2018)
397	Cervus_elaphus	(Ceacero, et al., 2018)	(HALL, 2010)	(Ceacero, et al., 2007)
398	Cervus_elaphus	(Ceacero, et al., 2018)	(HALL, 2010)	(Ceacero, et al., 2007)
399	Cervus_elaphus	(Ceacero, et al., 2018)	(HALL, 2010)	(Ceacero, et al., 2007)
400	Bos_taurus	(Spinka, and Ceacero, 2017)	(Spinka, et al., 2013)	NA
401	Bos_taurus	(Spinka, and Ceacero, 2017)	(Spinka, et al., 2013)	NA
402	Bos_taurus	(Spinka, and Ceacero, 2017)	(Spinka, et al., 2013)	NA
403	Bos_taurus	(Spinka, and Ceacero, 2017)	(Spinka, et al., 2013)	NA
404	Bos_taurus	(Spinka, and Ceacero, 2017)	(Spinka, et al., 2013)	NA
405	Oryctolagus_cuniculus	(Mykytowycz, 1959)	(von Holst, Hutzelmeyer, Kaetzke, et al., 2002)	NA
406	Oryctolagus_cuniculus	(Mykytowycz, 1959)	(von Holst, Hutzelmeyer, Kaetzke, et al., 2002)	NA
407	Heterocephalus_glaber	(Jarvis, 1981)	(Clarke and Faulkes, 1997)	NA
408	Canis_rufus	(Zimen, 2010)	(Sparkman, et al. 2010)	NA
409	Canis_rufus	(Zimen, 2010)	(Sparkman, et al. 2010)	NA
410	Lycaon_pictus	(Malcolm and Marten, 1982)	(Spiering, Somers, Maldonado, et al., 2009)	(Girman, et al., 1997)
411	Lycaon_pictus	(Malcolm and Marten, 1982)	(Spiering, Somers, Maldonado, et al., 2009)	(Girman, et al., 1997)
412	Macaca_mulatta	(Anderson and Simpson, 1979)	(Deutsch and Lee, 1991)	NA
413	Macaca_fuscata	(Sugiyama and Ohsawa, 1982)	(Koyama et al. 2003)	NA
414	Macaca_fuscata	(Sugiyama and Ohsawa, 1982)	(Koyama et al. 2003)	NA
415	Macaca_fuscata	(Sugiyama and Ohsawa, 1982)	(Koyama et al. 2003)	NA
416	Macaca_fuscata	(Sugiyama and Ohsawa, 1982)	(Koyama et al. 2003)	NA
417	Macaca_mulatta	(Stucki, Dow, and Sade, 1991)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
418	Macaca_mulatta	(Bercovitch and Berard, 1993)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
419	Theropithecus_gelada	(Dunbar, 1980)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)
420	Theropithecus_gelada	(Dunbar, 1980)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)
421	Theropithecus_gelada	(Dunbar, 1980)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)

422	<i>Theropithecus gelada</i>	(Dunbar, 1980)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)
423	<i>Theropithecus gelada</i>	(Dunbar, 1980)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)
424	<i>Theropithecus gelada</i>	(Dunbar, 1985)	(Dunbar, 1980)	(Snyder-Mackler, et al., 2014)
425	<i>Callithrix jacchus</i>	(Rothe, 2010)	(Digby, 1995)	(Rothe, 2010)
426	<i>Callithrix jacchus</i>	(Arruda, et al., 2005)	(Digby, 1995)	(Nievergelt et al. 2000)
427	<i>Callithrix jacchus</i>	(Arruda, et al., 2005)	(Digby, 1995)	(Nievergelt et al. 2000)
428	<i>Callithrix jacchus</i>	(Abbott, et al., 1981)	(Digby, 1995)	(Abbott, et al., 1981)
429	<i>Erythrocebus patas</i>	(Loy, 1981)	(Isbell & Pruettz 1988)	NA
430	<i>Saimiri sciureus</i>	(Coe, et al., 1981)	(Mitchell, Boinski, and van Schaik, 1991)	NA
431	<i>Saimiri sciureus</i>	(Coe, et al., 1981)	(Mitchell, Boinski, and van Schaik, 1991)	NA
432	<i>Saimiri sciureus</i>	(Coe, et al., 1981)	(Mitchell, Boinski, and van Schaik, 1991)	NA
433	<i>Chlorocebus aethiops</i>	(Wrangham, 1981)	(HOLEKAMP and SMALE, 1991)	NA
434	<i>Macaca mulatta</i>	(Blomquist, 2009)	(Deutsch and Lee, 1991)	(Chepko-Sade & Olivier, 1979)
435	<i>Pan troglodytes</i>	(Boesch, 1997)	(Wittig et al. 2003)	(Lukas et al., 2005)
436	<i>Pan troglodytes</i>	(Boesch, 1997)	(Wittig et al. 2003)	(Lukas et al., 2005)
437	<i>Lemur catta</i>	(Nunn and Pereira, 2000)	(Taylor and Sussman, 1985)	(Taylor and Sussman, 1985)
438	<i>Macaca fascicularis</i>	(Schaik, et al., 1989)	(Wittig et al. 2003)	NA
439	<i>Pan troglodytes</i>	(Stanton, et al., 2017)	NA	(Vigilant, et al., 2001)
440	<i>Pan troglodytes</i>	(Stanton, et al., 2017)	(Wittig et al. 2003)	(Vigilant, et al., 2001)
441	<i>Gorilla beringei</i>	(Eckardt, et al., 2016)	(Robbins, Gerald-Steklis, Robbins, et al., 2005)	(Watts, 1994)
442	<i>Macaca sylvanus</i>	(Modolo and Martin, 2007)	(Paul and Kuester, 1987)	(Kuemmerli and Martin, 2008)
443	<i>Lophocebus albigena</i>	(Arlet, et al., 2014)	(Arlet, et al., 2014)	NA
444	<i>Trachypithecus phayrei</i>	(Borries, et al., 2004)	(Koenig, Larney, Lu, and Borries, 2004)	(Larney 2013)

Supplementary references

- [1] Nievergelt, C.M., Digby, L.J., “Ramakrishnan, U. and Woodruff, D.S., 2000. Genetic analysis of group composition and breeding system in a wild common marmoset (*Callithrix jacchus*) population”. In: International Journal of Primatology, 21(1), pp.1-20.
- [2] R. Spinka, et al. “Pay respect to the elders: age, more than body mass, determines dominance in female beef cattle”. In: Animal Behaviour 86.6 (Dec. 2013), pp. 1315-1323. DOI: 10.1016/j.anbehav.2013.10.002. <URL: <https://doi.org/10.1016/j.anbehav.2013.10.002>>.
- [3] R. Spinka, et al., and F. Ceacero. “Higher dominance position does not result in higher reproductive success in female beef cattle_{1,2}”. In: Journal of Animal Science 95.8 (Aug. 2017), pp. 3301-3309. DOI: 10.2527/jas.2017.1415. <URL: <https://doi.org/10.2527/jas.2017.1415>>.
- [4] D. H. Abbott. “Behaviourally mediated suppression of reproduction in female primates”. In: Journal of Zoology 213.3 (Nov. 1987), pp. 455-470. DOI: 10.1111/j.1469-7998.1987.tb03720.x. <URL: <https://doi.org/10.1111/j.1469-7998.1987.tb03720.x>>.
- [5] D. H. Abbott, A. S. McNeilly, S. F. Lunn, et al. “Inhibition of ovarian function in subordinate female marmoset monkeys (*Callithrix jacchus jacchus*)”. In: Reproduction 63.2 (Nov. 1981), pp. 335-345. DOI: 10.1530/jrf.0.0630335. <URL: <https://doi.org/10.1530/jrf.0.0630335>>.
- [6] C. Alados and J. Escós. “The determinants of social status and the effect of female rank on reproductive success in Dama and Cuvier’s gazelles”. In: Ethology Ecology & Evolution 4.2 (2021), pp. 151-164. ISSN: 0394-9370. DOI: 10.1080/08927014.1992.9525336. <URL: <http://dx.doi.org/10.1080/08927014.1992.9525336>>.

- [7] C. Alados and J. Escós. “The determinants of social status and the effect of female rank on reproductive success in Dama and Cuvier’s gazelles”. In: *Ethology Ecology & Evolution* 4.2 (2021), pp. 151-164. ISSN: 0394-9370. DOI: 10.1080/08927014.1992.9525336. <URL: <http://dx.doi.org/10.1080/08927014.1992.9525336>>.
- [8] C. Alados and J. Escós. “The determinants of social status and the effect of female rank on reproductive success in Dama and Cuvier’s gazelles”. In: *Ethology Ecology & Evolution* 4.2 (2021), pp. 151-164. ISSN: 0394-9370. DOI: 10.1080/08927014.1992.9525336. <URL: <http://dx.doi.org/10.1080/08927014.1992.9525336>>.
- [9] C. Alados and J. Escós. “The determinants of social status and the effect of female rank on reproductive success in Dama and Cuvier’s gazelles”. In: *Ethology Ecology & Evolution* 4.2 (Apr. 1992), pp. 151-164. DOI: 10.1080/08927014.1992.9525336. <URL: <https://doi.org/10.1080/08927014.1992.9525336>>.
- [10] D. M. Anderson and M. J. A. Simpson. “Breeding Performance of a Captive Colony of Rhesus Macaques (*Macaca Mulatto*)”. In: *Laboratory Animals* 13.3 (Jul. 1979), pp. 275-282. DOI: 10.1258/00236779780937834. <URL: <https://doi.org/10.1258/00236779780937834>>.
- [11] E. A. Archie, J. Tung, M. Clark, et al. “Social affiliation matters: both same-sex and opposite-sex relationships predict survival in wild female baboons”. In: *Proceedings of the Royal Society B: Biological Sciences* 281.1793 (Oct. 2014), p. 20141261. DOI: 10.1098/rspb.2014.1261. <URL: <https://doi.org/10.1098/rspb.2014.1261>>.
- [12] M. E. Arlet, L. A. Isbell, A. Kaasik, et al. “Determinants of Reproductive Performance Among Female Gray-Cheeked Mangabeys (*Lophocebus albigena*) in Kibale National Park, Uganda”. In: *International Journal of Primatology* 36.1 (Dec. 2014), pp. 55-73. DOI: 10.1007/s10764-014-9810-4. <URL: <https://doi.org/10.1007/s10764-014-9810-4>>.
- [13] K. B. Armitage, D. H. V. Vuren, A. Ozgul, et al. “Proximate causes of natal dispersal in female yellow-bellied marmots, *Marmota flaviventris*”. In: *Ecology* 92.1 (Jan. 2011), pp. 218-227. DOI: 10.1890/10-0109.1. <URL: <https://doi.org/10.1890/10-0109.1>>.
- [14] M. Arruda, A. Araújo, M. Sousa, et al. “Two Breeding Females within Free-Living Groups May Not Always Indicate Polygyny: Alternative Subordinate Female Strategies in Common Marmosets (*Callithrix jacchus*)”. In: *Folia Primatologica* 76.1 (2005), pp. 10-20. DOI: 10.1159/000082451. <URL: <https://doi.org/10.1159/000082451>>.
- [15] P. J. BAKER, C. P. ROBERTSON, S. M. FUNK, et al. “Potential fitness benefits of group living in the red fox, *Vulpes vulpes*”. In: *Animal Behaviour* 56.6 (Dec. 1998), pp. 1411-1424. DOI: 10.1006/anbe.1998.0950. <URL: <https://doi.org/10.1006/anbe.1998.0950>>.
- [16] A. Baniuel, G. Cowlshaw, and E. Huchard. “Context dependence of female reproductive competition in wild chacma baboons”. In: *Animal Behaviour* 139 (May. 2018), pp. 37-49. DOI: 10.1016/j.anbehav.2018.03.001. <URL: <https://doi.org/10.1016/j.anbehav.2018.03.001>>.
- [17] F. Barroso, C. Alados, and J. Boza. “Social hierarchy in the domestic goat: effect on food habits and production”. In: *Applied Animal Behaviour Science* 69.1 (Aug. 2000), pp. 35-53. DOI: 10.1016/S0168-1591(00)00113-1. <URL: [https://doi.org/10.1016/S0168-1591\(00\)00113-1](https://doi.org/10.1016/S0168-1591(00)00113-1)>.
- [18] R. A. Barton and A. Whiten. “Feeding competition among female olive baboons, *Papio anubis*”. In: *Animal Behaviour* 46.4 (Oct. 1993), pp. 777-789. DOI: 10.1006/anbe.1993.1255. <URL: <https://doi.org/10.1006/anbe.1993.1255>>.
- [19] M. J. Baxter and L. M. Fedigan. “Grooming and consort partner selection in a troop of Japanese monkeys (*Macaca fuscata*)”. In: *Archives of Sexual Behavior* 8.5 (Sep. 1979), pp. 445-458. DOI: 10.1007/bf01541200. <URL: <https://doi.org/10.1007/bf01541200>>.

- [20] J. C. Beehner, D. A. Onderdonk, S. C. Alberts, et al. “The ecology of conception and pregnancy failure in wild baboons”. In: *Behavioral Ecology* 17.5 (Jun. 2006), pp. 741-750. DOI: 10.1093/beheco/arl006. <URL: <https://doi.org/10.1093/beheco/arl006>>.
- [21] F. B. Bercovitch and J. D. Berard. “Life history costs and consequences of rapid reproductive maturation in female rhesus macaques”. In: *Behavioral Ecology and Sociobiology* 32.2 (Feb. 1993), pp. 103-109. DOI: 10.1007/bf00164042. <URL: <https://doi.org/10.1007/bf00164042>>.
- [22] C. M. Berman. “Maternal Condition and Offspring Sex Ratio in a Group of Free-Ranging Rhesus Monkeys: An Eleven-Year Study”. In: *The American Naturalist* 131.3 (Mar. 1988), pp. 307-328. DOI: 10.1086/284792. <URL: <https://doi.org/10.1086/284792>>.
- [23] I. S. Bernstein and C. Ehardt. “The influence of kinship and socialization on aggressive behaviour in rhesus monkeys (*Macaca mulatta*)”. In: *Animal Behaviour* 34.3 (Jun. 1986), pp. 739-747. DOI: 10.1016/s0003-3472(86)80057-4. <URL: [https://doi.org/10.1016/s0003-3472\(86\)80057-4](https://doi.org/10.1016/s0003-3472(86)80057-4)>.
- [24] G. E. Blomquist. “Environmental and genetic causes of maturational differences among rhesus macaque matrilineages”. In: *Behavioral Ecology and Sociobiology* 63.9 (May. 2009), pp. 1345-1352. DOI: 10.1007/s00265-009-0792-8. <URL: <https://doi.org/10.1007/s00265-009-0792-8>>.
- [25] G. E. Blomquist, D. S. Sade, and J. D. Berard. “Rank-Related Fitness Differences and Their Demographic Pathways in Semi-Free-Ranging Rhesus Macaques (*Macaca mulatta*)”. In: *International Journal of Primatology* 32.1 (Nov. 2010), pp. 193-208. DOI: 10.1007/s10764-010-9461-z. <URL: <https://doi.org/10.1007/s10764-010-9461-z>>.
- [26] C. BOESCH. “Evidence for dominant wild female chimpanzees investing more in sons”. In: *Animal Behaviour* 54.4 (Oct. 1997), pp. 811-815. DOI: 10.1006/anbe.1996.0510. <URL: <https://doi.org/10.1006/anbe.1996.0510>>.
- [27] C. Borries, E. Larney, A. Derby, et al. “Temporary Absence and Dispersal in Phayre’s Leaf Monkeys (*Trachypithecus phayrei*)”. In: *Folia Primatologica* 75.1 (Jan. 01, 2004), pp. 27-30. ISSN: 0015-5713. DOI: 10.1159/000073428. <URL: <http://dx.doi.org/10.1159/000073428>>.
- [28] C. Borries, V. Sommer, and A. Srivastava. “Dominance, age, and reproductive success in free-ranging female hanuman langurs (*Presbytis entellus*)”. In: *International Journal of Primatology* 12.3 (Jun. 1991), pp. 231-257. DOI: 10.1007/bf02547586. <URL: <https://doi.org/10.1007/bf02547586>>.
- [29] L. J. N. Brent, A. Ruiz-Lambides, and M. L. Platt. “Family network size and survival across the lifespan of female macaques”. In: *Proceedings of the Royal Society B: Biological Sciences* 284.1854 (May. 2017), p. 20170515. DOI: 10.1098/rspb.2017.0515. <URL: <https://doi.org/10.1098/rspb.2017.0515>>.
- [30] J. Bulger and W. J. Hamilton. “Rank and density correlates of inclusive fitness measures in a natural chacma baboon (*Papio ursinus*) troop”. In: *International Journal of Primatology* 8.6 (Dec. 1987), pp. 635-650. DOI: 10.1007/bf02735781. <URL: <https://doi.org/10.1007/bf02735781>>.
- [31] T. M. Burland, N. C. Bennett, J. U. M. Jarvis, et al. “Eusociality in African mole-rats: new insights from patterns of genetic relatedness in the Damaraland mole-rat (*Cryptomys damarensis*)”. In: *Proceedings of the Royal Society of London. Series B: Biological Sciences* 269.1495 (May. 2002), pp. 1025-1030. DOI: 10.1098/rspb.2002.1978. <URL: <https://doi.org/10.1098/rspb.2002.1978>>.
- [32] T. M. BURLAND, N. C. BENNETT, J. U. M. JARVIS, et al. “Colony structure and parentage in wild colonies of co-operatively breeding Damaraland mole-rats suggest incest avoidance alone may not maintain reproductive skew”. In: *Molecular Ecology* 13.8 (Jul. 2004), pp. 2371-2379. DOI: 10.1111/j.1365-294x.2004.02233.x. <URL: <https://doi.org/10.1111/j.1365-294x.2004.02233.x>>.
- [33] S. Cafazzo, R. Bonanni, P. Valsecchi, et al. “Social Variables Affecting Mate Preferences, Copulation and Reproductive Outcome in a Pack of Free-Ranging Dogs”. In: *PLoS ONE* 9.6 (Jun. 2014). Ed. by C. Wicker-Thomas, p. e98594. DOI: 10.1371/journal.pone.0098594. <URL: <https://doi.org/10.1371/journal.pone.0098594>>.

- [34] S. Cafazzo, P. Valsecchi, R. Bonanni, et al. “Dominance in relation to age, sex, and competitive contexts in a group of free-ranging domestic dogs”. In: *Behavioral Ecology* 21.3 (2010), pp. 443-455. DOI: 10.1093/beheco/arq001. <URL: <https://doi.org/10.1093/beheco/arq001>>.
- [35] E. Z. Cameron, T. H. Setsaas, and W. L. Linklater. “Social bonds between unrelated females increase reproductive success in feral horses”. In: *Proceedings of the National Academy of Sciences* 106.33 (Aug. 2009), pp. 13850-13853. DOI: 10.1073/pnas.0900639106. <URL: <https://doi.org/10.1073/pnas.0900639106>>.
- [36] F. Cappa, M. Lombardini, and A. Meriggi. “Influence of seasonality, environmental and anthropic factors on crop damage by wild boar *Sus scrofa*”. In: *Folia Zoologica* 68.4 (2021), p. 261. ISSN: 0139-7893. DOI: 10.25225/fozo.015.2019. <URL: <http://dx.doi.org/10.25225/fozo.015.2019>>.
- [37] M. D. Carling, P. A. Wiseman, and J. A. Byers. “MICROSATELLITE ANALYSIS REVEALS MULTIPLE PATERNITY IN A POPULATION OF WILD PRONGHORN ANTELOPES (*ANTILOCAPRA AMERICANA*)”. In: *Journal of Mammalogy* 84.4 (Nov. 2003), pp. 1237-1243. DOI: 10.1644/brb-116. <URL: <https://doi.org/10.1644/brb-116>>.
- [38] J. Cassinello. “Factors modifying female social ranks in *Ammotragus*”. In: *Applied Animal Behaviour Science* 45.1-2 (Oct. 1995), pp. 175-180. DOI: 10.1016/0168-1591(95)00583-e. <URL: [https://doi.org/10.1016/0168-1591\(95\)00583-e](https://doi.org/10.1016/0168-1591(95)00583-e)>.
- [39] J. Cassinello and C. L. Alados. “Female reproductive success in captive *Ammotragus lervia* (Bovidae, Artiodactyla). Study of its components and effects of hierarchy and inbreeding”. In: *Journal of Zoology* 239.1 (May. 1996), pp. 141-153. DOI: 10.1111/j.1469-7998.1996.tb05442.x. <URL: <https://doi.org/10.1111/j.1469-7998.1996.tb05442.x>>.
- [40] F. Ceacero, M. K. á, A. J. Garc, et al. “Different maternal investment strategies for male and female calves in a polygynous mammal”. In: *Current Zoology* 65.3 (Jun. 2018). Ed. by Z. Jia, pp. 269-277. DOI: 10.1093/cz/zoy049. <URL: <https://doi.org/10.1093/cz/zoy049>>.
- [41] F. Ceacero, T. Landete-Castillejos, A. J. Garc, et al. “Kinship Discrimination and Effects on Social Rank and Aggressiveness Levels in Iberian Red Deer Hinds”. In: *Ethology* 113.12 (Dec. 2007), pp. 1133-1140. DOI: 10.1111/j.1439-0310.2007.01427.x. <URL: <https://doi.org/10.1111/j.1439-0310.2007.01427.x>>.
- [42] M. O. M. Chelini, R. Palme, and E. Otta. “Social stress and reproductive success in the female Syrian hamster: Endocrine and behavioral correlates”. In: *Physiology & Behavior* 104.5 (Oct. 2011), pp. 948-954. DOI: 10.1016/j.physbeh.2011.06.006. <URL: <https://doi.org/10.1016/j.physbeh.2011.06.006>>.
- [43] B. D. Chepko-Sade and T. J. Olivier. “Coefficient of genetic relationship and the probability of intragenealogical fission in *Macaca mulatta*”. In: *Behavioral Ecology and Sociobiology* 5.3 (1979), pp. 263-278. DOI: 10.1007/bf00293675. <URL: <https://doi.org/10.1007/bf00293675>>.
- [44] T. Christenson and B. L. Boeuf. “Aggression in the Female Northern Elephant Seal, *Mirounga Angustirostris*”. In: *Behaviour* 64.1-2 (1978), pp. 158-171. DOI: 10.1163/156853978x00495. <URL: <https://doi.org/10.1163/156853978x00495>>.
- [45] E. Clancey and J. A. Byers. “A comprehensive test of the Trivershypothesis in pronghorn (*Antilocapra americana*)”. In: *Journal of Mammalogy* 97.1 (Oct. 2015), pp. 179-186. DOI: 10.1093/jmammal/gyv168. <URL: <https://doi.org/10.1093/jmammal/gyv168>>.
- [46] F. M. Clarke and C. G. Faulkes. “Dominance and queen succession in captive colonies of the eusocial naked mole, *Heterocephalus glaber*”. In: *Proceedings of the Royal Society of London. Series B: Biological Sciences* 264.1384 (Jul. 1997), pp. 993-1000. DOI: 10.1098/rspb.1997.0137. <URL: <https://doi.org/10.1098/rspb.1997.0137>>.

- [47] T. H. Clutton-Brock, S. D. Albon, and F. E. Guinness. “Maternal dominance, breeding success and birth sex ratios in red deer”. In: *Nature* 308.5957 (Mar. 1984), pp. 358-360. DOI: 10.1038/308358a0. <URL: <https://doi.org/10.1038/308358a0>>.
- [48] C. L. Coe, J. Chen, E. L. Lowe, et al. “Hormonal and behavioral changes at puberty in the squirrel monkey”. In: *Hormones and Behavior* 15.1 (Mar. 1981), pp. 36-53. DOI: 10.1016/0018-506x(81)90033-7. <URL: [https://doi.org/10.1016/0018-506x\(81\)90033-7](https://doi.org/10.1016/0018-506x(81)90033-7)>.
- [49] M. Cords. “Friendship among adult female blue monkeys (*Cercopithecus mitis*)”. In: *Behaviour* 139.2 (2002), pp. 291-314. DOI: 10.1163/156853902760102681. <URL: <https://doi.org/10.1163/156853902760102681>>.
- [50] L. A. Correa, B. Zapata, H. Samaniego, et al. “Social structure in a family group of Guanaco (*Lama guanicoe*, Ungulate): Is female hierarchy based on prior attributes’ or social dynamics’?” In: *Behavioural Processes* 98 (Sep. 2013), pp. 92-97. DOI: 10.1016/j.beproc.2013.05.003. <URL: <https://doi.org/10.1016/j.beproc.2013.05.003>>.
- [51] D. L. Cram, P. Monaghan, R. Gillespie, et al. “Rank-Related Contrasts in Longevity Arise from Extra-Group Excursions Not Delayed Senescence in a Cooperative Mammal”. In: *Current Biology* 28.18 (Sep. 2018), pp. 2934-2939.e4. DOI: 10.1016/j.cub.2018.07.021. <URL: <https://doi.org/10.1016/j.cub.2018.07.021>>.
- [52] S. Creel. “DOMINANCE, AGGRESSION, AND GLUCOCORTICOID LEVELS IN SOCIAL CARNIVORES”. In: *Journal of Mammalogy* 86.2 (Apr. 2005), pp. 255-264. DOI: 10.1644/bhe-002.1. <URL: <https://doi.org/10.1644/bhe-002.1>>.
- [53] S. R. Creel and P. M. Waser. “Inclusive fitness and reproductive strategies in dwarf mongooses”. In: *Behavioral Ecology* 5.3 (1994), pp. 339-348. DOI: 10.1093/beheco/5.3.339. <URL: <https://doi.org/10.1093/beheco/5.3.339>>.
- [54] S. Creel, N. M. Creel, M. G. L. Mills, et al. “Rank and reproduction in cooperatively breeding African wild dogs: behavioral and endocrine correlates”. In: *Behavioral Ecology* 8.3 (1997), pp. 298-306. DOI: 10.1093/beheco/8.3.298. <URL: <https://doi.org/10.1093/beheco/8.3.298>>.
- [55] P. Dammann, R. Šumbera, C. Maßmann, et al. “Extended Longevity of Reproductives Appears to be Common in *Fukomys* Mole-Rats (Rodentia, Bathyergidae)”. In: *PLoS ONE* 6.4 (Apr. 2011). Ed. by G. G. de Polavieja, p. e18757. DOI: 10.1371/journal.pone.0018757. <URL: <https://doi.org/10.1371/journal.pone.0018757>>.
- [56] J. J. Dennehy. “Influence of social dominance rank on diet quality of pronghorn females”. In: *Behavioral Ecology* 12.2 (Mar. 2001), pp. 177-181. DOI: 10.1093/beheco/12.2.177. <URL: <https://doi.org/10.1093/beheco/12.2.177>>.
- [57] J. C. Deutsch and P. C. Lee. “Dominance and feeding competition in captive rhesus monkeys”. In: *International Journal of Primatology* 12.6 (Dec. 1991), pp. 615-628. DOI: 10.1007/bf02547673. <URL: <https://doi.org/10.1007/bf02547673>>.
- [58] J. M. Dietz and A. J. Baker. “Polygyny and female reproductive success in golden lion tamarins, *Leontopithecus rosalia*”. In: *Animal Behaviour* 46.6 (Dec. 1993), pp. 1067-1078. DOI: 10.1006/anbe.1993.1297. <URL: <https://doi.org/10.1006/anbe.1993.1297>>.
- [59] L. J. Digby. “Social organization in a wild population of *Callithrix jacchus*: II. Intragroup social behavior”. In: *Primates* 36.3 (Jul. 1995), pp. 361-375. DOI: 10.1007/bf02382859. <URL: <https://doi.org/10.1007/bf02382859>>.
- [60] W. P. Dittus. “The Evolution of Behaviors Regulating Density and Age-Specific Sex Ratios in a Primate Population”. In: *Behaviour* 69.3-4 (1979), pp. 265-301. DOI: 10.1163/156853979x00511. <URL: <https://doi.org/10.1163/156853979x00511>>.

- [61] W. P. J. Dittus. “Sex differences in fitness following a group take-over among Toque macaques: testing models of social evolution”. In: *Behavioral Ecology and Sociobiology* 19.4 (Sep. 1986), pp. 257-266. DOI: 10.1007/bf00300640. <URL: <https://doi.org/10.1007/bf00300640>>.
- [62] N. Djaković, Ø. Holand, A. L. Hovland, et al. “Association patterns and kinship in female reindeer (*Rangifer tarandus*) during rut”. In: *acta ethologica* 15.2 (Dec. 2011), pp. 165-171. DOI: 10.1007/s10211-011-0121-x. <URL: <https://doi.org/10.1007/s10211-011-0121-x>>.
- [63] A. Doby. “Breeding suppression between two unrelated and initially unfamiliar females occurs with or without social tolerance in common voles (*Microtus arvalis*)”. In: *Journal of Ethology* 27.3 (Oct. 2008), pp. 299-306. DOI: 10.1007/s10164-008-0118-8. <URL: <https://doi.org/10.1007/s10164-008-0118-8>>.
- [64] P. Dolhinow, J. J. McKenna, and J. V. H. Laws. “Rank and reproduction among female langur monkeys: Aging and improvement (They’re not just getting older, they’re getting better)”. In: *Aggressive Behavior* 5.1 (1979), pp. 19-30. DOI: 10.1002/1098-2337(1979)5:1<19::aid-ab2480050104>3.0.co;2-7. <URL: [https://doi.org/10.1002/1098-2337\(1979\)5:1<19::aid-ab2480050104>3.0.co;2-7](https://doi.org/10.1002/1098-2337(1979)5:1<19::aid-ab2480050104>3.0.co;2-7)>.
- [65] L. C. Drickamer. “A Ten-Year Summary of Reproductive Data for Free-Ranging *Macaca mulatta*”. In: *Folia Primatologica* 21.1 (1974), pp. 61-80. DOI: 10.1159/000155596. <URL: <https://doi.org/10.1159/000155596>>.
- [66] L. C. Drickamer. “Social dominance, reproduction, and release of the maturation-delaying chemosignal in the urine of female house mice (*Mus musculus*)”. In: *Journal of Comparative Psychology* 99.4 (Dec. 1985), pp. 411-419. DOI: 10.1037/0735-7036.99.4.411. <URL: <https://doi.org/10.1037/0735-7036.99.4.411>>.
- [67] J. Duboscq, C. Neumann, M. Agil, et al. “Degrees of freedom in social bonds of crested macaque females”. In: *Animal Behaviour* 123 (Jan. 2017), pp. 411-426. DOI: 10.1016/j.anbehav.2016.11.010. <URL: <https://doi.org/10.1016/j.anbehav.2016.11.010>>.
- [68] R. Dunbar. *Reproductive Decisions*. Princeton University Press, Dec. 1985. DOI: 10.1515/9781400853847. <URL: <https://doi.org/10.1515/9781400853847>>.
- [69] R. I. M. Dunbar. “Determinants and evolutionary consequences of dominance among female gelada baboons”. In: *Behavioral Ecology and Sociobiology* 7.4 (Nov. 1980), pp. 253-265. DOI: 10.1007/bf00300665. <URL: <https://doi.org/10.1007/bf00300665>>.
- [70] R. I. M. DUNBAR and E. P. DUNBAR. “Dominance and reproductive success among female gelada baboons”. In: *Nature* 266.5600 (Mar. 1977), pp. 351-352. DOI: 10.1038/266351a0. <URL: <https://doi.org/10.1038/266351a0>>.
- [71] S. Cote. “DOMINANCE HIERARCHIES IN FEMALE MOUNTAIN GOATS: STABILITY, AGGRESSIVENESS AND DETERMINANTS OF RANK”. In: *Behaviour* 137.11 (2000), pp. 1541-1566. DOI: 10.1163/156853900502718. <URL: <https://doi.org/10.1163/156853900502718>>.
- [72] S. Cote and M. Festa-Bianchet. “Reproductive success in female mountain goats: the influence of age and social rank”. In: *Animal Behaviour* 62.1 (Jul. 2001), pp. 173-181. DOI: 10.1006/anbe.2001.1719. <URL: <https://doi.org/10.1006/anbe.2001.1719>>.
- [73] T. Eccles and D. Shackleton. “Correlates and consequences of social status in female bighorn sheep”. In: *Animal Behaviour* 34.5 (Oct. 1986), pp. 1392-1401. DOI: 10.1016/s0003-3472(86)80210-x. <URL: [https://doi.org/10.1016/s0003-3472\(86\)80210-x](https://doi.org/10.1016/s0003-3472(86)80210-x)>.
- [74] W. Eckardt, K. Fawcett, and A. W. Fletcher. “Weaned age variation in the Virunga mountain gorillas (*Gorilla beringei beringei*): influential factors”. In: *Behavioral Ecology and Sociobiology* 70.4 (Feb. 2016), pp. 493-507. DOI: 10.1007/s00265-016-2066-6. <URL: <https://doi.org/10.1007/s00265-016-2066-6>>.

- [75] S. Ellis, N. Snyder-Mackler, A. Ruiz-Lambides, et al. “Deconstructing sociality: the types of social connections that predict longevity in a group-living primate”. In: *Proceedings of the Royal Society B: Biological Sciences* 286.1917 (Dec. 2019), p. 20191991. DOI: 10.1098/rspb.2019.1991. <URL: <https://doi.org/10.1098/rspb.2019.1991>>.
- [76] L. A. Fairbanks, M. J. Jorgensen, J. N. Bailey, et al. “Heritability and genetic correlation of hair cortisol in vervet monkeys in low and higher stress environments”. In: *Psychoneuroendocrinology* 36.8 (Sep. 2011), pp. 1201-1208. DOI: 10.1016/j.psyneuen.2011.02.013. <URL: <https://doi.org/10.1016/j.psyneuen.2011.02.013>>.
- [77] L. A. Fairbanks and M. T. McGuire. “Determinants of fecundity and reproductive success in captive vervet monkeys”. In: *American Journal of Primatology* 7.1 (1984), pp. 27-38. DOI: 10.1002/ajp.1350070106. <URL: <https://doi.org/10.1002/ajp.1350070106>>.
- [78] C. G. Faulkes and N. C. Bennett. “Family values: group dynamics and social control of reproduction in African mole-rats”. In: *Trends in Ecology & Evolution* 16.4 (Apr. 2001), pp. 184-190. DOI: 10.1016/s0169-5347(01)02116-4. <URL: [https://doi.org/10.1016/s0169-5347\(01\)02116-4](https://doi.org/10.1016/s0169-5347(01)02116-4)>.
- [79] L. M. Fedigan, S. D. Carnegie, and K. M. Jack. “Predictors of reproductive success in female white-faced capuchins (*Cebus capucinus*)”. In: *American Journal of Physical Anthropology* 137.1 (Sep. 2008), pp. 82-90. DOI: 10.1002/ajpa.20848. <URL: <https://doi.org/10.1002/ajpa.20848>>.
- [80] L. M. Fedigan, L. Fedigan, S. Gouzoules, et al. “Lifetime Reproductive Success in Female Japanese Macaques”. In: *Folia Primatologica* 47.2-3 (1986), pp. 143-157. DOI: 10.1159/000156271. <URL: <https://doi.org/10.1159/000156271>>.
- [81] L. Fedigan and M. Bergstrom. “Dominance among female white-faced capuchin monkeys (*Cebus capucinus*): hierarchical linearity, nepotism, strength and stability”. In: *Behaviour* 147.7 (2010), pp. 899-931. DOI: 10.1163/000579510x497283. <URL: <https://doi.org/10.1163/000579510x497283>>.
- [82] M. Festa-Bianchet. “The social system of bighorn sheep: grouping patterns, kinship and female dominance rank”. In: *Animal Behaviour* 42.1 (Jul. 1991), pp. 71-82. DOI: 10.1016/s0003-3472(05)80607-4. <URL: [https://doi.org/10.1016/s0003-3472\(05\)80607-4](https://doi.org/10.1016/s0003-3472(05)80607-4)>.
- [83] F. Fournier and M. Festa-Bianchet. “Social dominance in adult female mountain goats”. In: *Animal Behaviour* 49.6 (Jun. 1995), pp. 1449-1459. DOI: 10.1016/0003-3472(95)90066-7. <URL: [https://doi.org/10.1016/0003-3472\(95\)90066-7](https://doi.org/10.1016/0003-3472(95)90066-7)>.
- [84] P. A. Garber, F. E. On, L. Moya, et al. “Demographic and reproductive patterns in moustached tamarin monkeys (*Saguinus mystax*): Implications for reconstructing platyrrhine mating systems”. In: *American Journal of Primatology* 29.4 (1993), pp. 235-254. DOI: 10.1002/ajp.1350290402. <URL: <https://doi.org/10.1002/ajp.1350290402>>.
- [85] C. Garcia, P. Lee, and L. Rosetta. “Dominance and reproductive rates in captive female olive baboons, *Papio anubis*”. In: *American Journal of Physical Anthropology* 131.1 (2006), pp. 64-72. DOI: 10.1002/ajpa.20405. <URL: <https://doi.org/10.1002/ajpa.20405>>.
- [86] A. Gaylard, Y. Harrison, and N. C. Bennett. “Temporal changes in the social structure of a captive colony of the Damaraland mole-rat, *Cryptomys damarensis*: the relationship of sex and age to dominance and burrow-maintenance activity”. In: *Journal of Zoology* 244.3 (Mar. 1998), pp. 313-321. DOI: 10.1111/j.1469-7998.1998.tb00035.x. <URL: <https://doi.org/10.1111/j.1469-7998.1998.tb00035.x>>.
- [87] G. Gerlach. “Reproductive skew, costs, and benefits of cooperative breeding in female wood mice (*Apodemus sylvaticus*)”. In: *Behavioral Ecology* 13.3 (May. 2002), p. 408-418. ISSN: 1465-7279. DOI: 10.1093/beheco/13.3.408. <URL: <http://dx.doi.org/10.1093/beheco/13.3.408>>.
- [88] G. Gerlach. “Reproductive skew, costs, and benefits of cooperative breeding in female wood mice (*Apodemus sylvaticus*)”. In: *Behavioral Ecology* 13.3 (May. 2002), pp. 408-418. DOI: 10.1093/beheco/13.3.408. <URL: <https://doi.org/10.1093/beheco/13.3.408>>.

- [89] L. R. Gesquiere, J. Altmann, E. A. Archie, et al. “Interbirth intervals in wild baboons: Environmental predictors and hormonal correlates”. In: *American Journal of Physical Anthropology* 166.1 (Feb. 2018), pp. 107-126. DOI: 10.1002/ajpa.23407. <URL: <https://doi.org/10.1002/ajpa.23407>>.
- [90] D. J. Girman, M. G. L. Mills, E. Geffen, et al. “A molecular genetic analysis of social structure, dispersal, and interpack relationships of the African wild dog (*Lycaon pictus* 1mu)”. In: *Behavioral Ecology and Sociobiology* 40.3 (Mar. 1997), pp. 187-198. DOI: 10.1007/s002650050332. <URL: <https://doi.org/10.1007/s002650050332>>.
- [91] K. E. Glander. “Reproduction and population growth in free-ranging mantled howling monkeys”. In: *American Journal of Physical Anthropology* 53.1 (Jul. 1980), pp. 25-36. DOI: 10.1002/ajpa.1330530106. <URL: <https://doi.org/10.1002/ajpa.1330530106>>.
- [92] A. W. Goldizen, J. Mendelson, M. van Vlaardingen, et al. “Saddle-back tamarin (*Saguinus fuscicollis*) reproductive strategies: Evidence from a thirteen-year study of a marked population”. In: *American Journal of Primatology* 38.1 (1996), pp. 57-83. DOI: 10.1002/(sici)1098-2345(1996)38:1<57::aid-ajp6>3.0.co;2-s. <URL: [https://doi.org/10.1002/\(sici\)1098-2345\(1996\)38:1<57::aid-ajp6>3.0.co;2-s](https://doi.org/10.1002/(sici)1098-2345(1996)38:1<57::aid-ajp6>3.0.co;2-s)>.
- [93] M. Gomendio. “The influence of maternal rank and infant sex on maternal investment trends in rhesus macaques: birth sex ratios, inter-birth intervals and suckling patterns”. In: *Behavioral Ecology and Sociobiology* 27.5 (1990), pp. 365-375. DOI: 10.1007/bf00164008. <URL: <https://doi.org/10.1007/bf00164008>>.
- [94] M. Gomendio, T. H. Clutton-Brock, S. D. Albon, et al. “Mammalian sex ratios and variation in costs of rearing sons and daughters”. In: *Nature* 343.6255 (Jan. 1990), pp. 261-263. DOI: 10.1038/343261a0. <URL: <https://doi.org/10.1038/343261a0>>.
- [95] H. Gouzoules, S. Gouzoules, and L. Fedigan. “Behavioural dominance and reproductive success in female Japanese monkeys (*Macaca fuscata*)”. In: *Animal Behaviour* 30.4 (Nov. 1982), pp. 1138-1150. DOI: 10.1016/s0003-3472(82)80204-2. <URL: [https://doi.org/10.1016/s0003-3472\(82\)80204-2](https://doi.org/10.1016/s0003-3472(82)80204-2)>.
- [96] W. C. Green and A. Rothstein. “Sex bias or equal opportunity? Patterns of maternal investment in bison”. In: *Behavioral Ecology and Sociobiology* 29.5 (Dec. 1991), pp. 373-384. DOI: 10.1007/bf00165963. <URL: <https://doi.org/10.1007/bf00165963>>.
- [97] D. Greenberg-Cohen, P. U. Alkon, and Y. Yom-Tov. “A Linear Dominance Hierarchy in Female Nubian Ibex”. In: *Ethology* 98.3-4 (Apr. 2010), pp. 210-220. DOI: 10.1111/j.1439-0310.1994.tb01072.x. <URL: <https://doi.org/10.1111/j.1439-0310.1994.tb01072.x>>.
- [98] A. S. Griffin. “A genetic analysis of breeding success in the cooperative meerkat (*Suricata suricatta*)”. In: *Behavioral Ecology* 14.4 (Jul. 2003), pp. 472-480. DOI: 10.1093/beheco/arg040. <URL: <https://doi.org/10.1093/beheco/arg040>>.
- [99] K. Hackländer, E. Möstl, and W. Arnold. “Reproductive suppression in female Alpine marmots, *Marmota marmota*”. In: *Animal Behaviour* 65.6 (Jun. 2003), pp. 1133-1140. DOI: 10.1006/anbe.2003.2159. <URL: <https://doi.org/10.1006/anbe.2003.2159>>.
- [100] M. J. HALL. “Social Organization in an Enclosed Group of Red Deer (*Cervus elaphus* L.) on Rhum. I. The Dominance Hierarchy of Females and their Offspring”. In: *Zeitschrift für Tierpsychologie* 61.3 (Apr. 2010), pp. 250-262. DOI: 10.1111/j.1439-0310.1983.tb01341.x. <URL: <https://doi.org/10.1111/j.1439-0310.1983.tb01341.x>>.
- [101] C. C. Hass. “Social status in female bighorn sheep (*Ovis canadensis*): expression, development and reproductive correlates”. In: *Journal of Zoology* 225.3 (Nov. 1991), pp. 509-523. DOI: 10.1111/j.1469-7998.1991.tb03832.x. <URL: <https://doi.org/10.1111/j.1469-7998.1991.tb03832.x>>.
- [102] M. Heesen, S. Rogahn, J. Ostner, et al. “Food abundance affects energy intake and reproduction in frugivorous female Assamese macaques”. In: *Behavioral Ecology and Sociobiology* 67.7 (Apr. 2013), pp. 1053-1066. DOI: 10.1007/s00265-013-1530-9. <URL: <https://doi.org/10.1007/s00265-013-1530-9>>.

1530-9>.

- [103] M. D. Henry, S. J. Hankerson, J. M. Siani, et al. “High rates of pregnancy loss by subordinates leads to high reproductive skew in wild golden lion tamarins (*Leontopithecus rosalia*)”. In: *Hormones and Behavior* 63.5 (May. 2013), pp. 675-683. DOI: 10.1016/j.yhbeh.2013.02.009. <URL: <https://doi.org/10.1016/j.yhbeh.2013.02.009>>.
- [104] H. Hofer and M. L. East. “Behavioral processes and costs of co-existence in female spotted hyenas: a life history perspective”. In: *Evolutionary Ecology* 17.4 (Jul. 2003), pp. 315-331. DOI: 10.1023/a:1027352517231. <URL: <https://doi.org/10.1023/a:1027352517231>>.
- [105] S. Hohenbrink and S. Meinecke-Tillmann. “Influence of social dominance on the secondary sex ratio and factors affecting hierarchy in Holstein dairy cows”. In: *Journal of Dairy Science* 95.10 (Oct. 2012), pp. 5694-5701. DOI: 10.3168/jds.2011-5281. <URL: <https://doi.org/10.3168/jds.2011-5281>>.
- [106] O. Holand, H. Gjostein, A. Losvar, et al. “Social rank in female reindeer (*Rangifer tarandus*): effects of body mass, antler size and age”. In: *Journal of Zoology* 263.4 (Aug. 2004), pp. 365-372. DOI: 10.1017/s0952836904005382. <URL: <https://doi.org/10.1017/s0952836904005382>>.
- [107] O. Holand, R. B. Weladji, H. Gjostein, et al. “Reproductive effort in relation to maternal social rank in reindeer (*Rangifer tarandus*)”. In: *Behavioral Ecology and Sociobiology* 57.1 (Jul. 2004), pp. 69-76. DOI: 10.1007/s00265-004-0827-0. <URL: <https://doi.org/10.1007/s00265-004-0827-0>>.
- [108] K. E. HOLEKAMP and L. SMALE. “Dominance Acquisition During Mammalian Social Development: The of Maternal Rank”. In: *American Zoologist* 31.2 (Apr. 1991), pp. 306-317. DOI: 10.1093/icb/31.2.306. <URL: <https://doi.org/10.1093/icb/31.2.306>>.
- [109] K. E. Holekamp, L. Smale, and M. Szykman. “Rank and reproduction in the female spotted hyaena”. In: *Reproduction* 108.2 (Nov. 1996), pp. 229-237. DOI: 10.1530/jrf.0.1080229. <URL: <https://doi.org/10.1530/jrf.0.1080229>>.
- [110] D. von Holst, H. Hutzelmeyer, P. Kaetzke, et al. “Social rank, fecundity and lifetime reproductive success in wild European rabbits (*Oryctolagus cuniculus*)”. In: *Behavioral Ecology and Sociobiology* 51.3 (Feb. 2002), pp. 245-254. DOI: 10.1007/s00265-001-0427-1. <URL: <https://doi.org/10.1007/s00265-001-0427-1>>.
- [111] R. C. V. Horn, J. C. Buchan, J. Altmann, et al. “Divided destinies: group choice by female savannah baboons during social group fission”. In: *Behavioral Ecology and Sociobiology* 61.12 (Jun. 2007), pp. 1823-1837. DOI: 10.1007/s00265-007-0415-1. <URL: <https://doi.org/10.1007/s00265-007-0415-1>>.
- [112] R. C. V. Horn, A. L. Engh, K. T. Scribner, et al. “Behavioural structuring of relatedness in the spotted hyena (*Crocuta crocuta*) suggests direct fitness benefits of clan-level cooperation”. In: *Molecular Ecology* 13.2 (Jan. 2004), pp. 449-458. DOI: 10.1046/j.1365-294x.2003.02071.x. <URL: <https://doi.org/10.1046/j.1365-294x.2003.02071.x>>.
- [113] B. Huang, T. W. Wey, and D. T. Blumstein. “Correlates and Consequences of Dominance in a Social Rodent”. In: *Ethology* 117.7 (May. 2011), pp. 573-585. DOI: 10.1111/j.1439-0310.2011.01909.x. <URL: <https://doi.org/10.1111/j.1439-0310.2011.01909.x>>.
- [114] U. Huck, R. D. Lisk, and M. V. McKay. “Social dominance and reproductive success in pregnant and lactating golden hamsters (*Mesocricetus auratus*) under seminatural conditions”. In: *Physiology & Behavior* 44.3 (Jan. 1988), pp. 313-319. DOI: 10.1016/0031-9384(88)90031-5. <URL: [https://doi.org/10.1016/0031-9384\(88\)90031-5](https://doi.org/10.1016/0031-9384(88)90031-5)>.
- [115] G. Iossa, C. D. Soulsbury, P. J. Baker, et al. “Behavioral changes associated with a population density decline in the facultatively social red fox”. In: *Behavioral Ecology* 20.2 (Dec. 2008), pp. 385-395. DOI: 10.1093/beheco/arn149. <URL: <https://doi.org/10.1093/beheco/arn149>>.

- [116] N. Itoigawa, T. Tanaka, N. Ukai, et al. “Demography and reproductive parameters of a free-ranging group of Japanese macaques (*Macaca fuscata*) at Katsuyama”. In: *Primates* 33.1 (Jan. 1992), pp. 49-68. DOI: 10.1007/bf02382762. <URL: <https://doi.org/10.1007/bf02382762>>.
- [117] J. Jarvis. “Eusociality in a mammal: cooperative breeding in naked mole-rat colonies”. In: *Science* 212.4494 (May. 1981), pp. 571-573. DOI: 10.1126/science.7209555. <URL: <https://doi.org/10.1126/science.7209555>>.
- [118] J. A. Johnson. “Dominance rank in juvenile olive baboons, *Papio anubis*: the influence of gender, size, maternal rank and orphaning”. In: *Animal Behaviour* 35.6 (Dec. 1987), pp. 1694-1708. DOI: 10.1016/s0003-3472(87)80062-3. <URL: [https://doi.org/10.1016/s0003-3472\(87\)80062-3](https://doi.org/10.1016/s0003-3472(87)80062-3)>.
- [119] C. B. Jones. “The functions of status in the mantled howler monkey, *Alouatta palliata* Gray: Intraspecific competition for group membership in a folivorous neotropical primate”. In: *Primates* 21.3 (Jul. 1980), pp. 389-405. DOI: 10.1007/bf02390468. <URL: <https://doi.org/10.1007/bf02390468>>.
- [120] J. H. Jones, M. L. Wilson, C. Murray, et al. “Phenotypic quality influences fertility in Gombe chimpanzees”. In: *Journal of Animal Ecology* 79.6 (Apr. 2010), pp. 1262-1269. DOI: 10.1111/j.1365-2656.2010.01687.x. <URL: <https://doi.org/10.1111/j.1365-2656.2010.01687.x>>.
- [121] U. Kalbitzer, M. L. Bergstrom, S. D. Carnegie, et al. “Female sociality and sexual conflict shape offspring survival in a Neotropical primate”. In: *Proceedings of the National Academy of Sciences* 114.8 (Feb. 2017), pp. 1892-1897. DOI: 10.1073/pnas.1608625114. <URL: <https://doi.org/10.1073/pnas.1608625114>>.
- [122] B. Keane, P. Waser, S. Creel, et al. “Subordinate reproduction in dwarf mongooses”. In: *Animal Behaviour* 47.1 (Jan. 1994), pp. 65-75. DOI: 10.1006/anbe.1994.1008. <URL: <https://doi.org/10.1006/anbe.1994.1008>>.
- [123] D. Kerhoas, D. Perwitasari-Farajallah, M. Agil, et al. “Social and ecological factors influencing offspring survival in wild macaques”. In: *Behavioral Ecology* 25.5 (2014), pp. 1164-1172. DOI: 10.1093/beheco/aru099. <URL: <https://doi.org/10.1093/beheco/aru099>>.
- [124] A. A. Kinahan and N. Pillay. “Dominance status influences female reproductive strategy in a territorial African rodent *Rhabdomys pumilio*”. In: *Behavioral Ecology and Sociobiology* 62.4 (Sep. 2007), pp. 579-587. DOI: 10.1007/s00265-007-0482-3. <URL: <https://doi.org/10.1007/s00265-007-0482-3>>.
- [125] W. J. King and D. A. é. “Social, maternal, and environmental influences on reproductive success in female Alpine marmots (*Marmota marmota*)”. In: *Canadian Journal of Zoology* 80.12 (Dec. 2002), pp. 2137-2143. DOI: 10.1139/z02-205. <URL: <https://doi.org/10.1139/z02-205>>.
- [126] K. Klass and M. Cords. “Agonism and dominance in female blue monkeys”. In: *American Journal of Primatology* 77.12 (Sep. 2015), pp. 1299-1315. DOI: 10.1002/ajp.22481. <URL: <https://doi.org/10.1002/ajp.22481>>.
- [127] J. C. Knowles, P. J. V. C. de Groot, I. Wiesel, et al. “Microsatellite Variation in Namibian Brown Hyenas (*Hyaena brunnea*): Population Structure and Mating System Implications”. In: *Journal of Mammalogy* 90.6 (Dec. 2009), pp. 1381-1391. DOI: 10.1644/08-mamm-a-298r1.1. <URL: <https://doi.org/10.1644/08-mamm-a-298r1.1>>.
- [128] A. Koenig, E. Larney, A. Lu, et al. “Agonistic behavior and dominance relationships in female phayre’s leaf monkeys - preliminary results”. In: *American Journal of Primatology* 64.3 (2004), pp. 351-357. DOI: 10.1002/ajp.20084. <URL: <https://doi.org/10.1002/ajp.20084>>.
- [129] B. König. “Fitness effects of communal rearing in house mice: the role of relatedness versus familiarity”. In: *Animal Behaviour* 48.6 (Dec. 1994), pp. 1449-1457. DOI: 10.1006/anbe.1994.1381. <URL: <https://doi.org/10.1006/anbe.1994.1381>>.

- [130] L. Koren and E. Geffen. “Androgens and social status in female rock hyraxes”. In: *Animal Behaviour* 77.1 (Jan. 2009), pp. 233-238. DOI: 10.1016/j.anbehav.2008.09.031. <URL: <https://doi.org/10.1016/j.anbehav.2008.09.031>>.
- [131] N. Koyama, Y. Takahata, M. A. Huffman, et al. “Reproductive parameters of female Japanese macaques: Thirty years data from the arashiyama troops, Japan”. In: *Primates* 33.1 (Jan. 1992), pp. 33-47. DOI: 10.1007/bf02382761. <URL: <https://doi.org/10.1007/bf02382761>>.
- [132] R. Kümmerli and R. D. Martin. “Male and Female Reproductive Success in *Macaca sylvanus* in Gibraltar: No Evidence for Rank Dependence”. In: *International Journal of Primatology* 26.6 (Dec. 2005), pp. 1229-1249. DOI: 10.1007/s10764-005-8851-0. <URL: <https://doi.org/10.1007/s10764-005-8851-0>>.
- [133] R. Kümmerli and R. D. Martin. “Patterns of infant handling and relatedness in Barbary macaques (*Macaca sylvanus*) on Gibraltar”. In: *Primates* 49.4 (Sep. 2008), pp. 271-282. DOI: 10.1007/s10329-008-0100-7. <URL: <https://doi.org/10.1007/s10329-008-0100-7>>.
- [134] S. Lardy, and A. Cohas. “Intrasexual competition and female dominance in a singular breeding mammal, the Alpine marmot”. In: *Animal Behaviour* 86.6 (Dec. 2013), pp. 1155-1163. DOI: 10.1016/j.anbehav.2013.09.017. <URL: <https://doi.org/10.1016/j.anbehav.2013.09.017>>.
- [135] R. R. Lawler, A. F. Richard, and M. A. Riley. “Genetic population structure of the white sifaka (*Propithecus verreauxi verreauxi*) at Beza Mahafaly Special Reserve, southwest Madagascar (1992)”. In: *Molecular Ecology* 12.9 (Jul. 2003), pp. 2307-2317. DOI: 10.1046/j.1365-294x.2003.01909.x. <URL: <https://doi.org/10.1046/j.1365-294x.2003.01909.x>>.
- [136] B. Liu, C. Wu, P. A. Garber, et al. “Effects of group size and rank on mother-infant relationships and reproductive success in rhesus macaques (*Macaca mulatta*)”. In: *American Journal of Primatology* 80.7 (Jun. 2018), p. e22881. DOI: 10.1002/ajp.22881. <URL: <https://doi.org/10.1002/ajp.22881>>.
- [137] P. H. Lloyd and O. A. E. Rasa. “Status, reproductive success and fitness in Cape mountain zebra (*Equus zebra zebra*)”. In: *Behavioral Ecology and Sociobiology* 25.6 (Dec. 1989), pp. 411-420. DOI: 10.1007/bf00300187. <URL: <https://doi.org/10.1007/bf00300187>>.
- [138] P. Lloyd and O. Rasa. “Incest Avoidance and Attainment of Dominance By Females in a Cape Mountain Zebra (*Equus Zebra Zebra*) Population”. In: *Behaviour* 128.3-4 (1994), pp. 169-188. DOI: 10.1163/156853994x00253. <URL: <https://doi.org/10.1163/156853994x00253>>.
- [139] J. Loy. “The reproductive and heterosexual behaviours of adult patas monkeys in captivity”. In: *Animal Behaviour* 29.3 (Aug. 1981), pp. 714-726. DOI: 10.1016/s0003-3472(81)80006-1. <URL: [https://doi.org/10.1016/s0003-3472\(81\)80006-1](https://doi.org/10.1016/s0003-3472(81)80006-1)>.
- [140] D. de Luca and J. Ginsberg. “Dominance, reproduction and survival in banded mongooses: towards an egalitarian social system?” In: *Animal Behaviour* 61.1 (Jan. 2001), pp. 17-30. DOI: 10.1006/anbe.2000.1559. <URL: <https://doi.org/10.1006/anbe.2000.1559>>.
- [141] D. LUKAS, V. REYNOLDS, C. BOESCH, et al. “To what extent does living in a group mean living with kin?” In: *Molecular Ecology* 14.7 (Apr. 2005), pp. 2181-2196. DOI: 10.1111/j.1365-294x.2005.02560.x. <URL: <https://doi.org/10.1111/j.1365-294x.2005.02560.x>>.
- [142] D. W. Macdonald and S. P. Doolan. “Band Structure and Failures of Reproductive Suppression in a Cooperatively Breeding Carnivore, the Slender-Tailed Meerkat (*Suricata suricatta*)”. In: *Behaviour* 134.11-12 (1997), pp. 827-848. DOI: 10.1163/156853997x00179. <URL: <https://doi.org/10.1163/156853997x00179>>.
- [143] K. MacLeod and T. Clutton-Brock. “No evidence for adaptive sex ratio variation in the cooperatively breeding meerkat, *Suricata suricatta*”. In: *Animal Behaviour* 85.3 (Mar. 2013), pp. 645-653. DOI: 10.1016/j.anbehav.2012.12.028. <URL: <https://doi.org/10.1016/j.anbehav.2012.12.028>>.

- [144] D. Maestriperi. “Female-Biased Maternal Investment in Rhesus Macaques”. In: *Folia Primatologica* 72.1 (2001), pp. 44-47. DOI: 10.1159/000049920. <URL: <https://doi.org/10.1159/000049920>>.
- [145] J. R. Malcolm and K. Marten. “Natural selection and the communal rearing of pups in African wild dogs (*Lycaon pictus*)”. In: *Behavioral Ecology and Sociobiology* 10.1 (Feb. 1982), pp. 1-13. DOI: 10.1007/bf00296390. <URL: <https://doi.org/10.1007/bf00296390>>.
- [146] R. McFarland, D. Murphy, D. Lusseau, et al. “The ‘strength of weak ties’ among female baboons: fitness-related benefits of social bonds”. In: *Animal Behaviour* 126 (Apr. 2017), pp. 101-106. DOI: 10.1016/j.anbehav.2017.02.002. <URL: <https://doi.org/10.1016/j.anbehav.2017.02.002>>.
- [147] D. B. Meikle, L. C. Drickamer, S. H. Vessey, et al. “Dominance Rank and Parental Investment in Swine (*Sus scrofa domesticus*)”. In: *Ethology* 102.8 (Apr. 2010), pp. 969-978. DOI: 10.1111/j.1439-0310.1996.tb01174.x. <URL: <https://doi.org/10.1111/j.1439-0310.1996.tb01174.x>>.
- [148] D. B. Meikle, B. L. Tilford, and S. H. Vessey. “Dominance Rank, Secondary Sex Ratio, and Reproduction of Offspring in Polygynous Primates”. In: *The American Naturalist* 124.2 (Aug. 1984), pp. 173-188. DOI: 10.1086/284262. <URL: <https://doi.org/10.1086/284262>>.
- [149] D. B. Meikle and S. H. Vessey. “Maternal dominance rank and lifetime survivorship of male and female rhesus monkeys”. In: *Behavioral Ecology and Sociobiology* 22.6 (Jun. 1988), p. 379–383. ISSN: 1432-0762. DOI: 10.1007/bf00294974. <URL: <http://dx.doi.org/10.1007/BF00294974>>.
- [150] M. Mendl, A. J. Zanella, D. M. Broom, et al. “Maternal social status and birth sex ratio in domestic pigs: an analysis of mechanisms”. In: *Animal Behaviour* 50.5 (1995), pp. 1361-1370. DOI: 10.1016/0003-3472(95)80051-4. <URL: [https://doi.org/10.1016/0003-3472\(95\)80051-4](https://doi.org/10.1016/0003-3472(95)80051-4)>.
- [151] L. C. Metrione and J. D. Harder. “Fecal corticosterone concentrations and reproductive success in captive female southern white rhinoceros”. In: *General and Comparative Endocrinology* 171.3 (May. 2011), pp. 283-292. DOI: 10.1016/j.ygcen.2011.02.010. <URL: <https://doi.org/10.1016/j.ygcen.2011.02.010>>.
- [152] L. C. Metrione, L. M. Penfold, and G. H. Waring. “Social and spatial relationships in captive southern white rhinoceros (*Ceratotherium simum simum*)”. In: *Zoo Biology* 26.6 (Jul. 2007), pp. 487-502. DOI: 10.1002/zoo.20143. <URL: <https://doi.org/10.1002/zoo.20143>>.
- [153] E. S. Michel, S. Demarais, B. K. Strickland, et al. “Contrasting the Effects of Maternal and Behavioral Characteristics on Fawn Birth Mass in White-Tailed Deer”. In: *PLOS ONE* 10.8 (Aug. 2015). Ed. by T. Mappes, p. e0136034. DOI: 10.1371/journal.pone.0136034. <URL: <https://doi.org/10.1371/journal.pone.0136034>>.
- [154] C. L. Mitchell, S. Boinski, and C. P. van Schaik. “Competitive regimes and female bonding in two species of squirrel monkeys (*Saimiri oerstedii* and *S. sciureus*)”. In: *Behavioral Ecology and Sociobiology* 28.1 (Jan. 1991), pp. 55-60. DOI: 10.1007/bf00172139. <URL: <https://doi.org/10.1007/bf00172139>>.
- [155] L. Modolo and R. D. Martin. “Reproductive success in relation to dominance rank in the absence of prime-age males in Barbary macaques”. In: *American Journal of Primatology* 70.1 (2007), pp. 26-34. DOI: 10.1002/ajp.20452. <URL: <https://doi.org/10.1002/ajp.20452>>.
- [156] D. D. Moor, C. Roos, J. Ostner, et al. “Female Assamese macaques bias their affiliation to paternal and maternal kin”. In: *Behavioral Ecology* 31.2 (Jan. 2020). Ed. by L. Barrett, pp. 493-507. DOI: 10.1093/beheco/arz213. <URL: <https://doi.org/10.1093/beheco/arz213>>.
- [157] J. T. Morales-Pineyrua, G. Ciappesoni, and R. Ungerfeld. “Social rank and reproductive performance of pampas deer females (*Ozotoceros bezoarticus*, Linnaeus, 1758)”. In: *Behavioural Processes* 105 (Jun. 2014), pp. 49-52. DOI: 10.1016/j.beproc.2014.03.004. <URL: <https://doi.org/10.1016/j.beproc.2014.03.004>>.

- [158] R. Mykytowycz. “Social behaviour of an experimental colony of wild rabbits, *Oryctolagus cuniculus* (L.) II. First breeding season”. In: *CSIRO Wildlife Research* 4.1 (1959), p. 1. DOI: 10.1071/cwr9590001. <URL: <https://doi.org/10.1071/cwr9590001>>.
- [159] N. Nakagawa, M. Matsubara, Y. Shimooka, et al. “Embracing in a Wild Group of Yakushima Macaques (*Macaca fuscata yakui*) as an Example of Social Customs”. In: *Current Anthropology* 56.1 (Feb. 2015), pp. 104-120. DOI: 10.1086/679448. <URL: <https://doi.org/10.1086/679448>>.
- [160] J. Nelson and A. Goldstone. “Reproduction in *Peradorcas-Concinna* (Marsupialia, Macropodidae)”. In: *Wildlife Research* 13.4 (1986), p. 501. DOI: 10.1071/wr9860501. <URL: <https://doi.org/10.1071/wr9860501>>.
- [161] H. J. Nichols, W. Amos, M. A. Cant, et al. “Top males gain high reproductive success by guarding more successful females in a cooperatively breeding mongoose”. In: *Animal Behaviour* 80.4 (Oct. 2010), pp. 649-657. DOI: 10.1016/j.anbehav.2010.06.025. <URL: <https://doi.org/10.1016/j.anbehav.2010.06.025>>.
- [162] H. J. Nichols, M. B. V. Bell, S. J. Hodge, et al. “Resource limitation moderates the adaptive suppression of subordinate breeding in a cooperatively breeding mongoose”. In: *Behavioral Ecology* 23.3 (Feb. 2012), pp. 635-642. DOI: 10.1093/beheco/ars008. <URL: <https://doi.org/10.1093/beheco/ars008>>.
- [163] H. J. Nichols, N. R. Jordan, G. A. Jamie, et al. “Fine-scale spatiotemporal patterns of genetic variation reflect budding dispersal coupled with strong natal philopatry in a cooperatively breeding mammal”. In: *Molecular Ecology* 21.21 (Sep. 2012), pp. 5348-5362. DOI: 10.1111/mec.12015. <URL: <https://doi.org/10.1111/mec.12015>>.
- [164] K. Nieuwenhuijsen, A. J. J. C. Lammers, K. J. de Neef, et al. “Reproduction and social rank in female stump-tail Macaques (*Macaca arctoides*)”. In: *International Journal of Primatology* 6.1 (Feb. 1985), pp. 77-99. DOI: 10.1007/bf02693697. <URL: <https://doi.org/10.1007/bf02693697>>.
- [165] M. A. van Noordwijk and C. P. van Schaik. “Competition among female long-tailed macaques, *Macaca fascicularis*”. In: *Animal Behaviour* 35.2 (Apr. 1987), pp. 577-589. DOI: 10.1016/s0003-3472(87)80284-1. <URL: [https://doi.org/10.1016/s0003-3472\(87\)80284-1](https://doi.org/10.1016/s0003-3472(87)80284-1)>.
- [166] M. A. van Noordwijk and C. P. van Schaik. “The effects of dominance rank and group size on female lifetime reproductive success in wild long-tailed macaques, *Macaca fascicularis*”. In: *Primates* 40.1 (Jan. 1999), pp. 105-130. DOI: 10.1007/bf02557705. <URL: <https://doi.org/10.1007/bf02557705>>.
- [167] M. A. van Noordwijk and C. P. van Schaik. “The effects of dominance rank and group size on female lifetime reproductive success in wild long-tailed macaques, *Macaca fascicularis*”. In: *Primates* 40.1 (Jan. 1999), p. 105-130. ISSN: 1610-7365. DOI: 10.1007/bf02557705. <URL: <http://dx.doi.org/10.1007/BF02557705>>.
- [168] C. L. Nunn and M. E. Pereira. “Group histories and offspring sex ratios in ring-tailed lemurs (*Lemur catta*)”. In: *Behavioral Ecology and Sociobiology* 48.1 (Jun. 2000), pp. 18-28. DOI: 10.1007/s002650000206. <URL: <https://doi.org/10.1007/s002650000206>>.
- [169] D. H. NUSSEY, D. W. COLTMAN, T. COULSON, et al. “Rapidly declining fine-scale spatial genetic structure in female red deer”. In: *Molecular Ecology* 14.11 (Oct. 2005), pp. 3395-3405. DOI: 10.1111/j.1365-294x.2005.02692.x. <URL: <https://doi.org/10.1111/j.1365-294x.2005.02692.x>>.
- [170] D. OWENS and M. OWENS. “Social dominance and reproductive patterns in brown hyaenas, *Hyaena brunnea*, of the central Kalahari desert”. In: *Animal Behaviour* 51.3 (Mar. 1996), pp. 535-551. DOI: 10.1006/anbe.1996.0058. <URL: <https://doi.org/10.1006/anbe.1996.0058>>.
- [171] C. Packer, D. A. Collins, A. Sindimwo, et al. “Reproductive constraints on aggressive competition in female baboons”. In: *Nature* 373.6509 (Jan. 1995), pp. 60-63. DOI: 10.1038/373060a0. <URL: <https://doi.org/10.1038/373060a0>>.

- [172] J. A. Parga, M. L. Sauther, F. P. Cuzzo, et al. “Genetic Evidence for Male and Female Dispersal in Wild Lemur *catta*”. In: *Folia Primatologica* 86.1-2 (May. 2015), pp. 66-75. DOI: 10.1159/000369386. <URL: <https://doi.org/10.1159/000369386>>.
- [173] V. P. Patil, T. J. Karels, and D. S. Hik. “Ecological, Evolutionary and Social Constraints on Reproductive Effort: Are Hoary Marmots Really Biennial Breeders?” In: *PLOS ONE* 10.3 (Mar. 2015). Ed. by J. M. Waterman, p. e0119081. DOI: 10.1371/journal.pone.0119081. <URL: <https://doi.org/10.1371/journal.pone.0119081>>.
- [174] A. Paul and J. Kuester. “Dominance, kinship and reproductive value in female Barbary macaques (*Macaca sylvanus*) at Affenberg Salem”. In: *Behavioral Ecology and Sociobiology* 21.5 (Nov. 1987), pp. 323-331. DOI: 10.1007/bf00299970. <URL: <https://doi.org/10.1007/bf00299970>>.
- [175] A. Paul and D. Thommen. “Timing of Birth, Female Reproductive Success and Infant Sex Ratio in Semifree-Ranging Barbary Macaques (*Macaca sylvanus*)”. In: *Folia Primatologica* 42.1 (1984), pp. 2-16. DOI: 10.1159/000156140. <URL: <https://doi.org/10.1159/000156140>>.
- [176] J. Pluhacek, L. B. š, and L. Cul'. “High-ranking mares of captive plains zebra *Equus burchelli* have greater reproductive success than low-ranking mares”. In: *Applied Animal Behaviour Science* 99.3-4 (Sep. 2006), pp. 315-329. DOI: 10.1016/j.applanim.2005.11.003. <URL: <https://doi.org/10.1016/j.applanim.2005.11.003>>.
- [177] N. C. Pratt and R. D. Lisk. “Effects of social stress during early pregnancy on litter size and sex ratio in the golden hamster (*Mesocricetus auratus*)”. In: *Reproduction* 87.2 (Nov. 1989), pp. 763-769. DOI: 10.1530/jrf.0.0870763. <URL: <https://doi.org/10.1530/jrf.0.0870763>>.
- [178] A. Pusey. “The Influence of Dominance Rank on the Reproductive Success of Female Chimpanzees”. In: *Science* 277.5327 (Aug. 1997), p. 828-831. ISSN: 1095-9203. DOI: 10.1126/science.277.5327.828. <URL: <http://dx.doi.org/10.1126/science.277.5327.828>>.
- [179] D. A. Randall, J. P. Pollinger, R. K. Wayne, et al. “Inbreeding is reduced by female-biased dispersal and mating behavior in Ethiopian wolves”. In: *Behavioral Ecology* 18.3 (Mar. 2007), pp. 579-589. DOI: 10.1093/beheco/arm010. <URL: <https://doi.org/10.1093/beheco/arm010>>.
- [180] R. J. Rhine. “A twenty-one-year study of maternal dominance and secondary sex ratio in a colony group of stumptailed macaques (*Macaca arctoides*)”. In: *American Journal of Primatology* 32.2 (1994), pp. 145-148. DOI: 10.1002/ajp.1350320207. <URL: <https://doi.org/10.1002/ajp.1350320207>>.
- [181] R. J. Rhine, G. W. Norton, J. Rogers, et al. “Secondary sex ratio and maternal dominance rank among wild yellow baboons (*Papio cynocephalus*) of Mikumi National Park, Tanzania”. In: *American Journal of Primatology* 27.4 (1992), pp. 261-273. DOI: 10.1002/ajp.1350270404. <URL: <https://doi.org/10.1002/ajp.1350270404>>.
- [182] A. M. Robbins, T. Stoinski, K. Fawcett, et al. “Lifetime reproductive success of female mountain gorillas”. In: *American Journal of Physical Anthropology* 146.4 (Oct. 2011), pp. 582-593. DOI: 10.1002/ajpa.21605. <URL: <https://doi.org/10.1002/ajpa.21605>>.
- [183] M. M. Robbins, N. Gerald-Steklis, A. M. Robbins, et al. “Long-term dominance relationships in female mountain gorillas: strength, stability and determinants of rank”. In: *Behaviour* 142.6 (2005), pp. 779-809. DOI: 10.1163/1568539054729123. <URL: <https://doi.org/10.1163/1568539054729123>>.
- [184] M. M. Robbins, A. M. Robbins, N. Gerald-Steklis, et al. “Socioecological influences on the reproductive success of female mountain gorillas (*Gorilla beringei beringei*)”. In: *Behavioral Ecology and Sociobiology* 61.6 (Jan. 2007), pp. 919-931. DOI: 10.1007/s00265-006-0321-y. <URL: <https://doi.org/10.1007/s00265-006-0321-y>>.
- [185] M. M. Robbins, A. M. Robbins, N. Gerald-Steklis, et al. “Socioecological influences on the reproductive success of female mountain gorillas (*Gorilla beringei beringei*)”. In: *Behavioral Ecology and Sociobiology* 61.6 (Jan. 2007), p. 919-931. ISSN: 1432-0762. DOI: 10.1007/s00265-006-0321-y.

<URL: <http://dx.doi.org/10.1007/s00265-006-0321-y>>.

[186] S. Roberts and M. Cords. “Group size but not dominance rank predicts the probability of conception in a frugivorous primate”. In: *Behavioral Ecology and Sociobiology* 67.12 (Jul. 2013), pp. 1995-2009. DOI: 10.1007/s00265-013-1607-5. <URL: <https://doi.org/10.1007/s00265-013-1607-5>>.

[187] T. Ron, S. P. Henzi, and U. Motro. “Do Female Chacma Baboons Compete for a Safe Spatial Position in a Southern Woodland Habitat?” In: *Behaviour* 133.5-6 (1996), pp. 475-490. DOI: 10.1163/156853996x00549. <URL: <https://doi.org/10.1163/156853996x00549>>.

[188] J. P. Rood. “Mating relationships and breeding suppression in the dwarf mongoose”. In: *Animal Behaviour* 28.1 (Feb. 1980), pp. 143-150. DOI: 10.1016/s0003-3472(80)80019-4. <URL: [https://doi.org/10.1016/s0003-3472\(80\)80019-4](https://doi.org/10.1016/s0003-3472(80)80019-4)>.

[189] H. Rothe. “Some Aspects of Sexuality and Reproduction in Groups of Captive Marmosets (*Callithrix jacchus*)”. In: *Zeitschrift für Tierpsychologie* 37.3 (Apr. 2010), pp. 255-273. DOI: 10.1111/j.1439-0310.1975.tb00880.x. <URL: <https://doi.org/10.1111/j.1439-0310.1975.tb00880.x>>.

[190] A. le Roux, J. C. Beehner, and T. J. Bergman. “Female philopatry and dominance patterns in wild geladas”. In: *American Journal of Primatology* 73.5 (Dec. 2010), pp. 422-430. DOI: 10.1002/ajp.20916. <URL: <https://doi.org/10.1002/ajp.20916>>.

[191] J. Ruiter and E. Geffen. “Relatedness of matriline, dispersing males and social groups in longmacaques (*Macaca fascicularis*)”. In: *Proceedings of the Royal Society of London. Series B: Biological Sciences* 265.1391 (Jan. 1998), pp. 79-87. DOI: 10.1098/rspb.1998.0267. <URL: <https://doi.org/10.1098/rspb.1998.0267>>.

[192] A. F. Russell, A. A. Carlson, G. M. McIlrath, et al. “ADAPTIVE SIZE MODIFICATION BY DOMINANT FEMALE MEERKATS”. In: *Evolution* 58.7 (Jul. 2004), pp. 1600-1607. DOI: 10.1111/j.0014-3820.2004.tb01739.x. <URL: <https://doi.org/10.1111/j.0014-3820.2004.tb01739.x>>.

[193] A. Rusu and S. Krackow. “Kin-preferential cooperation, dominance-dependent reproductive skew, and competition for mates in communally nesting female house mice”. In: *Behavioral Ecology and Sociobiology* 56.3 (Apr. 2004). DOI: 10.1007/s00265-004-0787-4. <URL: <https://doi.org/10.1007/s00265-004-0787-4>>.

[194] J. L. Sanderson, H. J. Nichols, H. H. Marshall, et al. “Elevated glucocorticoid concentrations during gestation predict reduced reproductive success in subordinate female banded mongooses”. In: *Biology Letters* 11.10 (Oct. 2015), p. 20150620. DOI: 10.1098/rsbl.2015.0620. <URL: <https://doi.org/10.1098/rsbl.2015.0620>>.

[195] J. Santiago-Moreno, A. Gómez-Brunet, A. Toledano-D', et al. “Social dominance and breeding activity in Spanish ibex (*Capra pyrenaica*) maintained in captivity”. In: *Reproduction, Fertility and Development* 19.3 (2007), p. 436. DOI: 10.1071/rd06122. <URL: <https://doi.org/10.1071/rd06122>>.

[196] C. P. V. Schaik, W. J. Netto, A. J. J. V. Amerongen, et al. “Social rank and sex ratio of captive long-tailed macaque females (*Macaca fascicularis*)”. In: *American Journal of Primatology* 19.3 (1989), pp. 147-161. DOI: 10.1002/ajp.1350190303. <URL: <https://doi.org/10.1002/ajp.1350190303>>.

[197] M. B. Schilder and P. L. Boer. “Ethological investigations on a herd of plains zebra in a safari park: Time-budgets, reproduction and food competition”. In: *Applied Animal Behaviour Science* 18.1 (Jul. 1987), pp. 45-56. DOI: 10.1016/0168-1591(87)90253-x. <URL: [https://doi.org/10.1016/0168-1591\(87\)90253-x](https://doi.org/10.1016/0168-1591(87)90253-x)>.

[198] L. A. Schultz and R. K. Lore. “Communal reproductive success in rats (*Rattus norvegicus*): Effects of group composition and prior social experience.” In: *Journal of Comparative Psychology* 107.2 (1993), pp. 216-222. DOI: 10.1037/0735-7036.107.2.216. <URL: <https://doi.org/10.1037/0735-7036.107.2.216>>.

- [199] J. M. Setchell, M. Charpentier, and E. J. Wickings. “Sexual selection and reproductive careers in mandrills (*Mandrillus sphinx*)”. In: *Behavioral Ecology and Sociobiology* 58.5 (May. 2005), pp. 474-485. DOI: 10.1007/s00265-005-0946-2. <URL: <https://doi.org/10.1007/s00265-005-0946-2>>.
- [200] A. B. A. Shafer, J. M. Northrup, K. S. White, et al. “Habitat selection predicts genetic relatedness in an alpine ungulate”. In: *Ecology* 93.6 (Jun. 2012), pp. 1317-1329. DOI: 10.1890/11-0815.1. <URL: <https://doi.org/10.1890/11-0815.1>>.
- [201] D. Shargal, L. Shore, N. Roteri, et al. “Fecal testosterone is elevated in high ranking female ibexes (*Capra nubiana*) and associated with increased aggression and a preponderance of male offspring”. In: *Theriogenology* 69.6 (Apr. 2008), pp. 673-680. DOI: 10.1016/j.theriogenology.2007.11.017. <URL: <https://doi.org/10.1016/j.theriogenology.2007.11.017>>.
- [202] J. B. Silk. “Social Bonds of Female Baboons Enhance Infant Survival”. In: *Science* 302.5648 (Nov. 2003), pp. 1231-1234. DOI: 10.1126/science.1088580. <URL: <https://doi.org/10.1126/science.1088580>>.
- [203] J. B. Silk, J. C. Beehner, T. J. Bergman, et al. “Strong and Consistent Social Bonds Enhance the Longevity of Female Baboons”. In: *Current Biology* 20.15 (Aug. 2010), pp. 1359-1361. DOI: 10.1016/j.cub.2010.05.067. <URL: <https://doi.org/10.1016/j.cub.2010.05.067>>.
- [204] J. B. Silk, C. B. Clark-Wheatley, P. S. Rodman, et al. “Differential reproductive success and facultative adjustment of sex ratios among captive female bonnet macaques (*Macaca radiata*)”. In: *Animal Behaviour* 29.4 (Nov. 1981), pp. 1106-1120. DOI: 10.1016/s0003-3472(81)80063-2. <URL: [https://doi.org/10.1016/s0003-3472\(81\)80063-2](https://doi.org/10.1016/s0003-3472(81)80063-2)>.
- [205] J. Silk, D. Cheney, and R. Seyfarth. “THE STRUCTURE OF SOCIAL RELATIONSHIPS AMONG FEMALE SAVANNA BABOONS IN MOREMI RESERVE, BOTSWANA”. In: *Behaviour* 136.6 (1999), pp. 679-703. DOI: 10.1163/156853999501522. <URL: <https://doi.org/10.1163/156853999501522>>.
- [206] M. J. A. Simpson and A. E. Simpson. “Birth sex ratios and social rank in rhesus monkey mothers”. In: *Nature* 300.5891 (Dec. 1982), pp. 440-441. DOI: 10.1038/300440a0. <URL: <https://doi.org/10.1038/300440a0>>.
- [207] M. F. Small and S. B. Hrdy. “Secondary sex ratios by maternal rank, parity, and age in captive rhesus macaques (*Macaca mulatta*)”. In: *American Journal of Primatology* 11.4 (1986), pp. 359-365. DOI: 10.1002/ajp.1350110406. <URL: <https://doi.org/10.1002/ajp.1350110406>>.
- [208] B. Smuts and N. Nicolson. “Reproduction in wild female olive baboons”. In: *American Journal of Primatology* 19.4 (1989), pp. 229-246. DOI: 10.1002/ajp.1350190405. <URL: <https://doi.org/10.1002/ajp.1350190405>>.
- [209] N. Snyder-Mackler, S. C. Alberts, and T. J. Bergman. “The socio-genetics of a complex society: female gelada relatedness patterns mirror association patterns in a multilevel society”. In: *Molecular Ecology* 23.24 (Nov. 2014), pp. 6179-6191. DOI: 10.1111/mec.12987. <URL: <https://doi.org/10.1111/mec.12987>>.
- [210] M. B. C. Sousa, A. C. S. da Rocha Albuquerque, F. da Silva Albuquerque, et al. “Behavioral strategies and hormonal profiles of dominant and subordinate common marmoset (*Callithrix jacchus*) females in wild monogamous groups”. In: *American Journal of Primatology* 67.1 (2005), pp. 37-50. DOI: 10.1002/ajp.20168. <URL: <https://doi.org/10.1002/ajp.20168>>.
- [211] A. M. Sparkman, J. R. Adams, T. D. Steury, et al. “Direct fitness benefits of delayed dispersal in the cooperatively breeding red wolf (*Canis rufus*)”. In: *Behavioral Ecology* 22.1 (Dec. 2010), pp. 199-205. DOI: 10.1093/beheco/arq194. <URL: <https://doi.org/10.1093/beheco/arq194>>.

- [212] P. A. Spiering, M. J. Somers, J. E. Maldonado, et al. “Reproductive sharing and proximate factors mediating cooperative breeding in the African wild dog (*Lycaon pictus*)”. In: *Behavioral Ecology and Sociobiology* 64.4 (Nov. 2009), pp. 583-592. DOI: 10.1007/s00265-009-0875-6. <URL: <https://doi.org/10.1007/s00265-009-0875-6>>.
- [213] M. A. Stanton, E. V. Lonsdorf, A. E. Pusey, et al. “Do juveniles help or hinder? Influence of juvenile offspring on maternal behavior and reproductive outcomes in wild chimpanzees (*Pan troglodytes*)”. In: *Journal of Human Evolution* 111 (Oct. 2017), pp. 152-162. DOI: 10.1016/j.jhevol.2017.07.012. <URL: <https://doi.org/10.1016/j.jhevol.2017.07.012>>.
- [214] E. D. Strauss and K. E. Holekamp. “Social alliances improve rank and fitness in convention-based societies”. In: *Proceedings of the National Academy of Sciences* 116.18 (Mar. 2019), pp. 8919-8924. DOI: 10.1073/pnas.1810384116. <URL: <https://doi.org/10.1073/pnas.1810384116>>.
- [215] B. R. Stucki, M. M. Dow, and D. S. Sade. “Variance in intrinsic rates of growth among free-ranging rhesus monkey groups”. In: *American Journal of Physical Anthropology* 84.2 (Feb. 1991), pp. 181-191. DOI: 10.1002/ajpa.1330840208. <URL: <https://doi.org/10.1002/ajpa.1330840208>>.
- [216] Y. Sugiyama and H. Ohsawa. “Population Dynamics of Japanese Monkeys with Special Reference to the Effect of Artificial Feeding”. In: *Folia Primatologica* 39.3-4 (1982), pp. 238-263. DOI: 10.1159/000156080. <URL: <https://doi.org/10.1159/000156080>>.
- [217] A. K. SURRIDGE, K. M. IBRAHIM, D. J. BELL, et al. “Fine-scale genetic structuring in a natural population of European wild rabbits (*Oryctolagus cuniculus*)”. In: *Molecular Ecology* 8.2 (Feb. 1999), pp. 299-307. DOI: 10.1046/j.1365-294x.1999.00570.x. <URL: <https://doi.org/10.1046/j.1365-294x.1999.00570.x>>.
- [218] M. M. Symington. “Sex ratio and maternal rank in wild spider monkeys: when daughters disperse”. In: *Behavioral Ecology and Sociobiology* 20.6 (Jun. 1987), pp. 421-425. DOI: 10.1007/bf00302985. <URL: <https://doi.org/10.1007/bf00302985>>.
- [219] Y. Takahata. “The reproductive biology of a free-ranging troop of Japanese monkeys”. In: *Primates* 21.3 (Jul. 1980), pp. 303-329. DOI: 10.1007/bf02390462. <URL: <https://doi.org/10.1007/bf02390462>>.
- [220] Y. Takahata, N. Koyama, S. Ichino, et al. “The relationship between female rank and reproductive parameters of the ringtailed lemur: a preliminary analysis”. In: *Primates* 49.2 (Dec. 2007), pp. 135-138. DOI: 10.1007/s10329-007-0076-8. <URL: <https://doi.org/10.1007/s10329-007-0076-8>>.
- [221] Y. Takahata, S. Suzuki, N. Agetsuma, et al. “Reproduction of wild Japanese macaque females of Yakushima and Kinkazan Islands: A preliminary report”. In: *Primates* 39.3 (Jul. 1998), pp. 339-349. DOI: 10.1007/bf02573082. <URL: <https://doi.org/10.1007/bf02573082>>.
- [222] L. Taylor and R. W. Sussman. “A preliminary study of kinship and social organization in a semi-free-ranging group of *Lemur catta*”. In: *International Journal of Primatology* 6.6 (Dec. 1985), pp. 601-614. DOI: 10.1007/bf02692291. <URL: <https://doi.org/10.1007/bf02692291>>.
- [223] T. W. Townsend and E. D. Bailey. “Effects of Age, Sex and Weight on Social Rank in Pinned White-tailed Deer”. In: *American Midland Naturalist* 106.1 (Jul. 1981), p. 92. DOI: 10.2307/2425138. <URL: <https://doi.org/10.2307/2425138>>.
- [224] R. C. Van Horn, J. C. Buchan, J. Altmann, et al. “Divided destinies: group choice by female savannah baboons during social group fission”. In: *Behavioral Ecology and Sociobiology* 61.12 (Jun. 2007), p. 1823-1837. ISSN: 1432-0762. DOI: 10.1007/s00265-007-0415-1. <URL: <http://dx.doi.org/10.1007/s00265-007-0415-1>>.
- [225] H. Vervaecke, C. Roden, and H. de Vries. “Dominance, fatness and fitness in female American bison, *Bison bison*”. In: *Animal Behaviour* 70.4 (Oct. 2005), pp. 763-770. DOI: 10.1016/j.anbehav.2004.12.018. <URL: <https://doi.org/10.1016/j.anbehav.2004.12.018>>.

- [226] L. Vigilant, M. Hofreiter, H. Siedel, et al. “Paternity and relatedness in wild chimpanzee communities”. In: *Proceedings of the National Academy of Sciences* 98.23 (Oct. 2001), pp. 12890-12895. DOI: 10.1073/pnas.231320498. <URL: <https://doi.org/10.1073/pnas.231320498>>.
- [227] J. Visser, T. Robinson, and B. J. van Vuuren. “Spatial genetic structure in the rock hyrax (*Procavia capensis*) across the Namaqualand and western Fynbos areas of South Africa — a mitochondrial and microsatellite perspective”. In: *Canadian Journal of Zoology* 98.8 (Aug. 2020), pp. 557-571. DOI: 10.1139/cjz-2019-0154. <URL: <https://doi.org/10.1139/cjz-2019-0154>>.
- [228] D. D. VRIES, A. KOENIG, and C. BORRIES. “Female reproductive success in a species with an age-inversed hierarchy”. In: *Integrative Zoology* 11.6 (Nov. 2016), pp. 433-446. DOI: 10.1111/1749-4877.12201. <URL: <https://doi.org/10.1111/1749-4877.12201>>.
- [229] E. D. Wallace and N. C. Bennett. “The colony structure and social organization of the giant Zambian mole-rat, *Cryptomys mechowii*”. In: *Journal of Zoology* 244.1 (Jan. 1998), pp. 51-61. DOI: 10.1111/j.1469-7998.1998.tb00006.x. <URL: <https://doi.org/10.1111/j.1469-7998.1998.tb00006.x>>.
- [230] S. K. Wasser and D. P. Barash. “Reproductive Suppression Among Female Mammals: Implications for Biomedicine and Sexual Selection Theory”. In: *The Quarterly Review of Biology* 58.4 (Dec. 1983), pp. 513-538. DOI: 10.1086/413545. <URL: <https://doi.org/10.1086/413545>>.
- [231] S. K. Wasser and A. K. Starling. “Proximate and ultimate causes of reproductive suppression among female yellow baboons at Mikumi National Park, Tanzania”. In: *American Journal of Primatology* 16.2 (1988), p. 97–121. ISSN: 1098-2345. DOI: 10.1002/ajp.1350160202. <URL: <http://dx.doi.org/10.1002/ajp.1350160202>>.
- [232] S. K. Wasser and A. K. Starling. “Proximate and ultimate causes of reproductive suppression among female yellow baboons at Mikumi National Park, Tanzania”. In: *American Journal of Primatology* 16.2 (1988), pp. 97-121. DOI: 10.1002/ajp.1350160202. <URL: <https://doi.org/10.1002/ajp.1350160202>>.
- [233] S. Wasser, G. Norton, S. Kleindorfer, et al. “Population trend alters the effects of maternal dominance rank on lifetime reproductive success in yellow baboons (*Papio cynocephalus*)”. In: *Behavioral Ecology and Sociobiology* 56.4 (May. 2004). DOI: 10.1007/s00265-004-0797-2. <URL: <https://doi.org/10.1007/s00265-004-0797-2>>.
- [234] D. P. Watts. “Social relationships of immigrant and resident female mountain gorillas, II: Relatedness, residence, and relationships between females”. In: *American Journal of Primatology* 32.1 (1994), pp. 13-30. DOI: 10.1002/ajp.1350320103. <URL: <https://doi.org/10.1002/ajp.1350320103>>.
- [235] H. E. Watts, J. B. Tanner, B. L. Lundrigan, et al. “Post-weaning maternal effects and the evolution of female dominance in the spotted hyena”. In: *Proceedings of the Royal Society B: Biological Sciences* 276.1665 (Mar. 2009), pp. 2291-2298. DOI: 10.1098/rspb.2009.0268. <URL: <https://doi.org/10.1098/rspb.2009.0268>>.
- [236] L. Wauters and A. A. Dhondt. “Body Weight, Longevity and Reproductive Success in Red Squirrels (*Sciurus vulgaris*)”. In: *The Journal of Animal Ecology* 58.2 (Jun. 1989), p. 637. DOI: 10.2307/4853. <URL: <https://doi.org/10.2307/4853>>.
- [237] C. Welker, H. Höl, and C. Schäl-Witt. “Significance of Kin Relations and Individual Preferences in the Social Behaviour of *Cebus apella*”. In: *Folia Primatologica* 54.3-4 (1990), pp. 166-170. DOI: 10.1159/000156440. <URL: <https://doi.org/10.1159/000156440>>.
- [238] P. A. White. “Maternal rank is not correlated with cub survival in the spotted hyena, *Crocuta crocuta*”. In: *Behavioral Ecology* 16.3 (Feb. 2005), pp. 606-613. DOI: 10.1093/beheco/ari033. <URL: <https://doi.org/10.1093/beheco/ari033>>.

- [239] M. E. Wilson, T. P. Gordon, and I. S. Bernstein. “Timing of Births and Reproductive Success in Rhesus Monkey Social Groups”. In: *Journal of Medical Primatology* 7.4 (1978), pp. 202-212. DOI: 10.1159/000459880. <URL: <https://doi.org/10.1159/000459880>>.
- [240] L. D. Wolfe. “Female rank and reproductive success among arashiyama B Japanese macaques (*Macaca fuscata*)”. In: *International Journal of Primatology* 5.2 (Apr. 1984), pp. 133-143. DOI: 10.1007/bf02735737. <URL: <https://doi.org/10.1007/bf02735737>>.
- [241] J. O. Wolff, A. S. Dunlap, and E. Ritchhart. “Adult female prairie voles and meadow voles do not suppress reproduction in their daughters”. In: *Behavioural Processes* 55.3 (Sep. 2001), pp. 157-162. DOI: 10.1016/s0376-6357(01)00176-0. <URL: [https://doi.org/10.1016/s0376-6357\(01\)00176-0](https://doi.org/10.1016/s0376-6357(01)00176-0)>.
- [242] R. Wrangham. “Drinking competition in vervet monkeys”. In: *Animal Behaviour* 29.3 (Aug. 1981), pp. 904-910. DOI: 10.1016/s0003-3472(81)80027-9. <URL: [https://doi.org/10.1016/s0003-3472\(81\)80027-9](https://doi.org/10.1016/s0003-3472(81)80027-9)>.
- [243] E. Zimen. “On the Regulation of Pack Size in Wolves”. In: *Zeitschrift für Tierpsychologie* 40.3 (Apr. 2010), pp. 300-341. DOI: 10.1111/j.1439-0310.1976.tb00939.x. <URL: <https://doi.org/10.1111/j.1439-0310.1976.tb00939.x>>.
- [244] T. Ziporyn and M. K. McClintock. “Passing as an Indicator of Social Dominance Among Female Wild and Domestic Norway Rats”. In: *Behaviour* 118.1-2 (1991), pp. 26-41. DOI: 10.1163/156853991x00184. <URL: <https://doi.org/10.1163/156853991x00184>>.
- [245] Altmann, Hausfater, J. 1988. “Determinants of Reproductive Success in Savannah Baboons, *Papio Cynocephalus*.” In *Reproductive Success: Studies of Individual Variation in Contrasting Breeding Systems*, edited by T. H. Clutton-Brock, 403–18. Princeton University Press.
- [246] Altmann J, Alberts SC. 2003. “Intraspecific Variability in Fertility and Offspring Survival in a Nonhuman Primate: Behavioral Control of Ecological and Social Sources.” In *Offspring: Human Fertility Behavior in Biodemographic Perspective*, edited by Bulatao RA Wachter KW, 6. ashington (DC): National Academies Press (US).
- [247] Baker AJ, Dietz JM, Bales K. 2002. “Mating System and Group Dynamics in Lion Tamarins.” In *Lion Tamarins: Biology and Conservation*, edited by Rylands AB Kleiman DG, 188–212. Washington, Smithsonian Institution Press.
- [248] Baniel A, Huchard E, Carter AJ. in press. “Exploring Environmental and Social Predictors of Reproductive Pace in a Desert-Dwelling Baboon,” in press. Busse, C. 1982. “Social Dominance and Offspring Mortality Among Female Chacma Baboons.” *Int J Primatol* 3: 267.
- [249] Byers, John A. 1997. *American Pronghorn: Social Adaptations and the Ghosts of Predators Past*. University of Chicago Press.
- [250] Cheney, Dorothy L, Robert M Seyfarth, Julia Fischer, Jacinta C Beehner, Thore J Bergman, Sara E Johnson, Dawn M Kitchen, Ryne A Palombit, Drew Rendall, and Joan B Silk. 2006. “Reproduction, Mortality, and Female Reproductive Success in Chacma Baboons of the Okavango Delta, Botswana.” In *Reproduction and Fitness in Baboons: Behavioral, Ecological, and Life History Perspectives*, 147–76. Springer.
- [251] Cheney, R. M. Seyfarth, D. L., and P. C. Lee. 1988. “Reproductive Success in Vervet Monkeys.” In *Reproductive Success: Studies of Individual Variation in Contrasting Breeding Systems*, edited by T. H. Clutton-Brock, 384–402. Princeton University Press.
- [252] Di Bitetti, Mario S, and Charles H Janson. 2001. “Reproductive Socioecology of Tufted Capuchins (*Cebus Apella Nigrurus*) in Northeastern Argentina.” *International Journal of Primatology* 22 (2): 127–42.
- [253] Frank, Laurence G, Kay E Holekamp, and Laura Smale. 1995. “Dominance, Demography, and Reproductive Success of Female Spotted Hyenas.” *Serengeti II: Dynamics, Management, and Conservation of an Ecosystem*, 364–84.

- [254] Fürtbauer, Ines. 2011. “The Socio-Endocrinology of Female Reproductive Strategies in Wild Assamese Macaques (*Macaca Assamensis*).” PhD thesis, Göttingen University.
- [255] Gaillard, J-M, Serge Brandt, and J-M Jullien. 1993. “Body Weight Effect on Reproduction of Young Wild Boar (*Sus Scrofa*) Females: A Comparative Analysis.” *Folia Zoologica (Brno)* 42 (3): 204–12.
- [256] Gese, Eric M. 2004. “Coyotes in Yellowstone National Park: The Influence of Dominance on Foraging, Territoriality, and Fitness.” *Biology and Conservation of Wild Canids*. Oxford University Press, New York, New York, USA, 271–83.
- [257] Isbell, Lynne A, and Jill D Pruettz. 1998. “Differences Between Vervets (*Cercopithecus Aethiops*) and Patas Monkeys (*Erythrocebus Patas*) in Agonistic Interactions Between Adult Females.” *International Journal of Primatology* 19 (5): 837–55.
- [258] Kopp, Gisela. 2015. “Gene Flow Dynamics in Baboons—the Influence of Social Systems.” PhD thesis, Göttingen University.
- [259] Koyama, Nicola F. 2003. “Matrilineal Cohesion and Social Networks in *Macaca Fuscata*.” *International Journal of Primatology* 24 (4): 797–811.
- [260] Kubzdela, Katarzyna Stefania. 1998. “Sociodemography in Diurnal Primates: The Effects of Group Size and Female Dominance Rank on Intragroup Spatial Distribution, Feeding Competition, Female Reproductive Success, and Female Dispersal Patterns in White Sifaka, *Propithecus Verreauxi Verreauxi*.” PhD thesis, University of Chicago.
- [261] Larney, Eileen. 2013. “The Influence of Genetic and Social Structure on Reproduction in Phayre’s Leaf Monkeys (*Trachypithecus Phayrei Crepusculus*).” PhD thesis, State University of New York at Stony Brook.
- [262] Lynch, Emily Claire. 2016. “Paternal Kinship in a Matrilocal Society of Olive Baboons (*Papio Hamadryas Anubis*) in Laikipia District, Kenya.” PhD thesis, Rutgers The State University of New Jersey-New Brunswick.
- [263] Nievergelt, Caroline M, Leslie J Digby, Uma Ramakrishnan, and David S Woodruff. 2000. “Genetic Analysis of Group Composition and Breeding System in a Wild Common Marmoset (*Callithrix Jacchus*) Population.” *International Journal of Primatology* 21 (1): 1–20.
- [264] Paul, Kuester, A. 1996. “Differential Reproduction in Male and Female Barbary Macaques.” In *Evolution and Ecology of Macaque Societies*, edited by Lindburg D. G. Fa J. E., 293–317. Cambridge University Press.
- [265] Roosmalen, Marc GM van. 1980. “Habitat Preferences, Diet, Feeding Strategy and Social Organization of the Black Spider Monkey (*Ateles Paniscus Paniscus* Linnaeus 1758) in Surinam.” PhD thesis, Roosmalen.
- [266] Rubenstein, Daniel I, and CASSANDRA M Nuñez. 2009. “Sociality and Reproductive Skew in Horses and Zebras.” *Reproductive Skew in Vertebrates: Proximate and Ultimate Causes*, 196–226.
- [267] Santiago-Moreno, Julián, Amelia Gómez-Brunet, Antonio González-Bulnes, Benoit Malpoux, Philippe Chemineau, Antonio Pulido-Pastor, and Antonio López-Sebastián. 2003. “Seasonal Ovulatory Activity and Plasma Prolactin Concentrations in the Spanish Ibex (*Capra Pyrenaica Hispanica*) Maintained in Captivity.” *Reproduction Nutrition Development* 43 (3): 217–24.
- [268] Setchell, Joanna M, Phyllis C Lee, E Jean Wickings, and Alan F Dixon. 2002. “Reproductive Parameters and Maternal Investment in Mandrills (*Mandrillus Sphinx*).” *International Journal of Primatology* 23 (1): 51–68.

- [269] Sinderbrand, Carly Anne. 2011. “The Relationship of Dominance, Reproductive State and Stress in a Non-Cooperative Breeder, the Domestic Horse (*Equus Caballus*).” PhD thesis, Western Kentucky University. Smith, Andrew C, ER Tirado Herrera, Hannah M Buchanan-Smith, and Eckhard W Heymann. 2001. “Multiple Breeding Females and Allo-Nursing in a Wild Group of Moustached Tamarins (*Saguinus Mystax*).” *Neotropical Primates* 9 (2): 67–69.
- [270] Visser, Jacobus Hendrik. 2013. “Gene-Flow in the Rock Hyrax (*Procavia Capensis*) at Different Spatial Scales.” PhD thesis, Stellenbosch: Stellenbosch University.
- [271] Whitten, Patricia L. 1983. “Diet and Dominance Among Female Vervet Monkeys (*Cercopithecus Aethiops*).” *American Journal of Primatology* 5 (2): 139–59.
- [272] Wittig, Roman M, and Christophe Boesch. 2003. “Food Competition and Linear Dominance Hierarchy Among Female Chimpanzees of the Tai National Park.” *International Journal of Primatology* 24 (4): 847–67.