

1 **Sperm-dependent asexual species matter in ecology and evolution**

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16 **Abstract**

17 In a world of metazoans, where sexual reproduction vastly predominates, asexual organisms are
18 nonetheless very important. The aim of this review is to show that asexuals can have general effects
19 on other species, ecosystems and biological networks via mechanisms that deserve more attention.
20 These include 1) impact on the genepool of coexisting sexual species by either restricting their
21 population sizes or by providing bridges for interspecific gene-flow whose type and consequences
22 substantially differ from gene flow mechanisms expected under sexual reproduction; 2) impact on
23 diversification rate, either direct by serving as stepping-stones in speciation or indirect by
24 promoting the formation of pre- and postzygotic reproduction barriers among nascent species; and
25 3) impact on spatial effects, via direct or indirect (apparent) types of competition and Allee effects.
26 We specifically point to these important mechanisms, provide empirical examples how asexuals
27 impact the evolution of sexual species and ecosystems, and, finally, show that these broad effects
28 may last beyond the tenure of the individual asexual lineages causing them. We also propose new
29 research directions to incorporate the aforementioned impacts of asexual organisms which will
30 ultimately enhance the understanding of evolution of genomes and ecosystems in general.

31

32 **Keywords:** speciation, hybridization, meiosis, population dynamics, apparent competition

33

34 **Introduction**

35 Reproduction, or the ability to transmit genomes from one generation to another, is a fundamental
36 property of all living organisms. In the world of metazoans, this is mostly realized by meiotic sex
37 with recombination, which probably has a single origin very early in eukaryotic evolution
38 (Bernstein and Bernstein 2010), but there is considerable variability on this theme as many
39 organisms transmit their genomes partly or entirely clonally, commonly being referred to as
40 asexuals. In fact, many sexual species – including our own – pass on parts of their genome, such
41 as mitochondria and sex chromosomes, essentially in an asexual mode with no or very limited
42 recombination. Thus, while *sex* and *asexuality* (Glossary) are usually presented as a dichotomy,
43 in reality, there is a continuum between full meiosis and *mixis* (Glossary) on the one hand, and
44 vegetative reproduction and ameiotic formation of gametes on the other hand. The term asexuality
45 is thus used very broadly to capture a large diversity of reproductive mechanisms that differ in
46 some aspect from full blown sexual reproduction with meiosis and recombination in every
47 generation (Neiman et al. 2014). Leaving aside cases such as budding, life cycles with sexual and
48 asexual phases, alternating gametic and sporophytic phases (like in some parasites) or
49 polyembryony (the production of multiple clonal embryos from a single fertilized egg (Prodöhl et
50 al. 1996)), forms of asexual reproduction based on production of at least partly clonal gametes
51 have evolved many times independently in plants and most major taxa of animals (Suomalainen
52 et al. 1987). Although obligate asexuality evolved relatively infrequently, a survey of the literature
53 indicates that parthenogenetic taxa with clonal gametes evolved in at least 15 out of 31 metazoan
54 phyla (Supplementary Material, Figure S1 and references there), suggesting this trait has high
55 relevance for evolution.

56 In this review, we do not intend to provide an exhaustive overview of all asexual taxa,
57 which would be a book-length endeavor, nor to summarize all research on asexual organisms,
58 which recently attracted new critical thinking and synthesis (e.g., (Laskowski et al. 2019; Fujita et
59 al. 2020)) and contributed significantly to understanding the disadvantages and advantages of
60 meiotic sex. Indeed, because of their atypical meiosis and non-Mendelian propagation of genomes,
61 asexuals serve as excellent natural models for understanding fundamental questions in evolution,
62 ecology and cell biology (Bengtsson 2009; Meirmans 2009), such as, for example, the *Red Queen*
63 *Hypothesis* (Glossary; (Van Valen 1973; Brockhurst et al. 2014; Lively and Morran 2014)),
64 homology search and recombination, mutational load or sperm-egg interactions during fertilization
65 (recently reviewed by e.g. (Laskowski et al. 2019; Dalziel et al. 2020)). Instead, we present new
66 hypotheses and emphasize specific mechanisms how the very existence of asexuals and the
67 complexity of their reproductive modes directly and indirectly shapes the evolution of interacting

68 sexual taxa and ecosystems they occur in by exerting specific types of evolutionary interactions,
69 which are unparalleled in sexually reproducing lineages.

70 The so-called asexual organisms employ a wide spectrum of independently arisen
71 cytological mechanisms for gamete production, which range from completely ameiotic processes
72 (apomixis) to those involving more or less distorted meiotic divisions (automixis) (Stenberg and
73 Saura 2009, 2013). Consequently, some organisms transmit their genomes in a strictly **clonal** way
74 (Glossary) while others, like hybridogens, transmit clonally only parts of their genomes while the
75 rest undergoes recombination or even mixis (Figure 1). The distinction between sexual and asexual
76 reproductive modes is particularly difficult in plants, where various intermediates exist and several
77 types of apomixis may emerge from one type of sexuality (Hojsgaard and Hörandl 2015).
78 Variability also exists with respect to the process of fertilization since true parthenogens (Figure
79 1A) are completely independent of gamete fusion, while other types of asexuals are referred to as
80 **sperm-dependent parthenogens**, pseudogams or sexual parasites (Glossary) as they rely on sperm,
81 which is usually, but not always, provided by closely related sexual species, e.g. (Choleva et al.
82 2008). The sperm-dependent parthenogens may need the sperm just for egg activation with its
83 genome rejected after fertilization (**gynogenesis**; Glossary and Figure 1B), but the sperm may also
84 contribute genetically to the progeny either by subgenomic components, such as
85 microchromosomes (Figure 1C) (Schartl et al. 1995) or its entire genome may be incorporated into
86 progeny which would either utilize it for ploidy increase (genome addition; Figure 1F) or eliminate
87 it after one generation in the next round of gamete production (**hybridogenesis**; Glossary and
88 Figure 1E). However, the situation often is more complex than such textbook examples, as e.g., in
89 the case of *Ambystoma* salamanders from the northeastern USA, where, over evolutionary time,
90 genomes have been added or replaced in the original hybrid lineages (the mode of reproduction
91 known as kleptogenesis) (Bi et al. 2008; Bogart 2019).

92 To be precise, in the present review, we point out that ongoing research on the
93 aforementioned traditional questions relating to asexuality often lacks careful consideration of the
94 effects – both negative and positive – that asexuals have on the sexually reproducing species they
95 occur with. This is especially the case of sperm-dependent parthenogens, traditionally viewed as
96 combining disadvantages of both reproductive modes, asexuality and sexuality because they are
97 deprived of regular recombination and segregation of their genetic material, while on the other
98 hand they cannot take full advantages of asexual reproduction, being dependent on mating with
99 sexual counterparts (rev. in (Beukeboom and Vrijenhoek 1998)). Therefore, we compile evidence

100 for wide-ranging effects that sperm-dependent asexuals have on interacting sexual species and
101 ecosystems they occur in. In particular, we suggest that even if individual asexual lineages may be
102 short-lived, their impact on ecosystems where they occurred (and eventually perished) is
103 substantial and probably quite higher than currently assumed.

104 Although asexual taxa are not as widespread as sexual taxa, we contend that overlooking
105 or ignoring them may lead to an incomplete understanding of ecological and evolutionary
106 dynamics and can be perilous as historically illustrated by Johann Gregor Mendel's example. Like
107 any good scientist Mendel wanted to "verify" the results he obtained with peas (*Pisum*) in 1866
108 (Bicknell et al. 2016) but he inadvertently made a mistake that delayed the recognition of his
109 findings by many years: He picked a popular model system of the time, *Hieracium* and largely
110 failed to replicate his original findings, as reported in his second paper in 1869. The reason,
111 however, was not that his ideas were incorrect, but rather that *Hieracium* is a facultative asexual
112 plant which Mendel could not know in his time (Krahulcová et al. 1999). Nonetheless, this cast
113 doubt on the generality of Mendel's discovery and delayed the general acceptance of genetics for
114 decades, thus changing history (Bicknell et al. 2016). Had Mendel been able to take the asexual
115 reproduction of his model organism into account, the history of evolutionary biology might have
116 been very different.

117 We argue here that although individual asexual lineages eventually go extinct - just like
118 any other species - the particularities of asexual reproduction likely have characteristic effects on
119 coexisting species extending well beyond the lifespan of such clones. In particular, we focus on
120 sperm-dependent parthenogens and show what important impacts they have on the biological
121 networks they occur in. For example, by playing a role as sexual parasites that "steal" gametes of
122 sexual species for their own reproduction (Hubbs 1964; Avise 2008; Lehtonen et al. 2013), they
123 indirectly affect the effective population size of their sexual counterparts, thereby modifying their
124 gene pool. Based on recent advances we show that the very existence of asexuals in general implies
125 several effects on the ecosystems they are embedded in and very important properties of coexisting
126 sexual species in terms of their 1) genetic architecture, 2) diversification and speciation, and 3)
127 spatial distribution.

128

129 ***1. Impact on the gene pool of sexual species***

130 ***1.1 Direct impact on a sexual species' gene pool via asexual to sexual gene flow***

131 Asexual species are typically assumed to produce clonal progeny, but clonality is seldom
132 perfect and many asexuals can produce gamete types that enable their genes to flow back into
133 sexual populations (Figure 1C, F). In plants, for instance, phylogenetic reconstruction of
134 angiosperm evolution indicated that reversals from apomixis to sexuality occurred (Hörandl and

135 Hojsgaard 2012). Backcrossing to sexual relatives has been directly reported from several
136 apomictic species, e.g., in *Hieracium*, *Ranunculus* and *Taraxacum* (Hörandl and Paun 2007),
137 which supposedly helps to generate new cytotypes (Sailer et al. 2020) and local reversal to sex
138 (Majeský et al. 2012), thereby contradicting Darlington's „dead end of evolution” hypothesis
139 (Darlington 1939). The geneflow from asexual to sexual populations has also been demonstrated
140 in various asexual animals ranging from flatworms to vertebrates (Hotz et al. 1992; Goddard and
141 Schultz 1993; Vorburger 2001; Sousa-Santos et al. 2006; D'Souza and Michiels 2009; Scali 2009;
142 Angers et al. 2018).

143 It may take various forms: For example, let us assume a hybridogenetic female hybrid
144 between two species A and B which pre-meiotically eliminate A-type chromosomes, thus clonally
145 transmitting only the B genome into her eggs. Normally, these eggs are fertilized by A - sperm
146 restoring the hybrid state of AB. However, fertilization with B – sperm would lead to formation
147 of sexually reproducing BB diploids, where now half of their nuclear genome has an asexual
148 history and the other half has not (Figure 1G and other mechanisms of mtDNA replacement in
149 Figure 1H, I) (Mikulíček et al. 2014; Denton et al. 2018; Kwan et al. 2019; Suzuki et al. 2019).
150 Furthermore, some asexual organisms do not pass on their genomes only via females, but also
151 produce males (e.g., *Daphnia* water fleas (Paland et al. 2005), *Pelophylax* frogs (Graf and Polls-
152 Pelaz 1989; Mikulíček et al. 2015)) or sperm in case of hermaphrodites like *Schmidtea* flatworms
153 (D'Souza and Michiels 2010), or *Corbicula* clams (Hedtke et al. 2008; Hedtke and Hillis 2011)
154 which fertilize related sexual females, thereby further facilitating the introgression of asexual
155 genomes back into sexual gene pools.

156 Clearly, such gene flow from an asexual to a sexual genepool fundamentally differs from
157 any classical mechanisms of gene exchange between sexual species because asexuals transmit their
158 genetic material *en bloc*, and in a non-Mendelian fashion (Glémin et al. 2019). Since many
159 asexuals are of hybrid origin and reproductively interact with their two parental species, they thus
160 can serve surprisingly often as a hub between sexual species. However, owing to the specific nature
161 of asexual transmission of genomes, the effect on the recipient genomes would be different from
162 other types of introgression found in 'classical' sexual hybrids.

163 For example, if the aforementioned hybridogenetic hybrid originated from hybridization
164 between a female from species A and a male from the species B, its eggs would possess A-type
165 mtDNA but B-type nucleus (Figure 1G). If fertilized by B – sperm, such gametes would thus create
166 cyto-nuclear mosaics (also known as *cybrids*; Glossary) and hence facilitate the transfer of

167 cytoplasmatic and/or complete nuclear genomes at rates superseding many unidirectional
168 backcrosses via meiotic hybrids. Such massive unidirectional mtDNA flow mediated by sperm-
169 dependent parthenogens without any substantial nuclear admixis has been reported for example in
170 water frogs *Pelophylax ridibundus* (Hotz et al. 1992; Plötner et al. 2008; Mikulíček et al. 2014),
171 several species of the *Bacillus* insect (Scali 2009), asexual *Corbicula* clams (Hedtke et al. 2008;
172 Hedtke and Hillis 2011) as well as various fishes including loaches of the family Cobitidae (Kwan
173 et al. 2019) and cyprinids of the genera *Squalius* (Alves et al. 2001, 2002; Sousa-Santos et al.
174 2006) and *Chrosomus* (Binet and Angers 2005; Angers et al. 2018). Angers et al. (2018)
175 documented that such a type of mtDNA introgression might have had considerable adaptive value
176 during the postglacial range expansion in *Chrosomus*. In an extreme case, the gene flow from
177 asexuals might have even caused complete replacement of the original mtDNA by an allospecific
178 mitochondrial genome as suggested by cytonuclear mosaicism of *Cobitis tanaitica* spined loaches
179 (Choleva et al. 2014). Sperm-dependent asexuals may also mediate interspecific gene flow
180 between nuclear gene pools of related sexual species, as found in the European complex of water
181 frogs where hybridogenetic hybrids *P. esculentus* mediate introgression between parental species
182 *P. ridibundus* and *P. lessonae* (Uzzell et al. 1976; Schmeller et al. 2005; Mikulíček et al. 2014).

183 In addition, when considering the effects of asexual to sexual introgression, we have to
184 emphasize that asexual' genomic elements that introgress into a sexual genepool have evolved
185 many generations in a (quasi)clonal way and passed through very different selection regimes than
186 homologous sequences in a sexual species' genepool. First, asexual genomes are assumed to
187 evolve under relaxed purifying selection due to whole-genome linkage, suggesting that a recipient
188 sexual species may acquire genetic material with high numbers of deleterious mutations,
189 depending on the duration of clonal evolution and the speed of mutation accumulation processes,
190 like *Muller's ratchet* (Glossary). Moreover, asexual species may benefit from complete loss or
191 loss of function of many genes coding traits that are not needed or even maladaptive for asexual
192 reproduction, such as genes related to meiosis or mating behavior (Schartl et al. 1991; Schlupp et
193 al. 1992; Kooi and Schwander 2014; Parker et al. 2019). Introgression of asexual genomes may
194 therefore deteriorate the genepool of the sexual recipient species even more than simple mutation
195 accumulation. It should be noted, though, that predicting the negative impact of gene flow from
196 asexuals into sexual genepools is difficult as models for mutation accumulation in asexuals are far
197 from clear. For example, mutation rates seem to be generally male-biased, which might counteract
198 Muller's ratchet (Redfield 1994) and partly explain why accumulation of deleterious mutations
199 appears to be surprisingly slow in many investigated asexual all-female species (Janko et al. 2011;
200 Pellino et al. 2013; Warren et al. 2018; Kočí et al. 2020). Furthermore, in Amazon mollies
201 (*Poecilia formosa*), a gynogenetic fish species from Southern Texas and Northeastern Mexico,

202 loss of sex-related traits was not found (Warren et al. 2018). By contrast, in fully asexual organisms
203 such as the snail *Potamopyrgus antipodarum*, loss of male function has been reported (Jalinsky et
204 al. 2020).

205 On the other hand, introgression from asexuals may also be beneficial since restriction of
206 recombination may have many positive aspects for genome evolution, because it favors the spread
207 of advantageous combinations of alleles in regions where recombinants are expected to have lower
208 fitness (Neiman and Linksvayer 2006). Selection for restricted recombination has indeed been
209 documented between loci contributing to adaptation (Thompson and Jiggins 2014), speciation
210 (Ortiz-Barrientos et al. 2016), or *de novo* evolution of separate sexes (Charlesworth and
211 Charlesworth 1978). Asexual genepools, for their whole-genome linkage, may therefore represent
212 unprecedented testing fields where selection acts on various allelic combinations with much
213 greater efficiency than could ever be observed in any sexual species, and multiply, due to clonal
214 reproduction, the fittest multilocus genotypes (Barbuti et al. 2012). Therefore, when asexual to
215 sexual gene flow is possible, it is likely to introduce whole chromosomes that evolved for a long
216 time without recombination into particularly suitable combinations of alleles. Positive effects of
217 such a gene exchange with asexual hybrids have already been documented, for example, by
218 Schmeller et al. (2005) who showed that bi-directional introgression between sexual water frog
219 species *P. ridibundus* and *P. perezi* mediated by the hybridogenetic hybrids *P. grafi* increased the
220 probability of local adaptation to hypoxic conditions and range expansion of the sexual species.

221 In summary, when asexual to sexual gene flow is possible, the potential effects on the sexual
222 species' fitness, either positive or negative, may last longer than the existence of individual clones
223 from which the introgression originated.

224

225 **1.2. Indirect impact on a sexual species gene pool via modification of effective population** 226 **size**

227 Yet, even if a sexual species is not directly affected by gene flow from an asexual species,
228 its genepool would still be modified indirectly by interactions with coexisting sperm-dependent
229 parthenogenetic asexuals, because the very presence of sperm-dependent asexuals likely affects
230 selective pressures operating in sexual populations and their demographic parameters. To
231 understand the underlying reasons, it must be kept in mind that although asexual taxa often
232 diverged ecologically from their sexual counterparts (see e.g. (Ross et al. 2012; Van der Kooi et
233 al. 2017), many asexual species, especially the sperm-dependent parthenogens, are ecologically

234 relatively similar to their sexual ancestors, with which they often coexist (Beukeboom and
235 Vrijenhoek 1998; Vrijenhoek and Parker 2009). Consequently, part of the environmental carrying
236 capacity potentially available for sexual population is taken up by asexuals when these are present.

237 In general, sperm-dependent competitors may affect the *effective population size* (N_e)
238 (Glossary) of related sexuals via several mechanisms. These include stochastic effects increasing
239 the strength of genetic drift, such as (a) modifying extinction/recolonization dynamics of sexual
240 metapopulation, and (b) increasing the variance in reproductive success of the sexual species, and
241 (c) modifying selective pressures resulting from biased operational sex ratio (OSR) in mixed
242 sexual-asexual populations. Let us consider these mechanisms in the following paragraphs.

243 (a) Sperm-dependent parthenogenesis depends on access to males of a sexual species and
244 consequently the asexuals may not outcompete their sexual host, lest they lose a vital resource,
245 leading to the collapse of the whole sexual-asexual complex (Schlupp and Riesch 2011). Kokko
246 et al. (2008) published a mathematical model proposing that coexistence with sexuals is possible
247 provided that outcompeting the sexual species by the asexuals occurs locally and asynchronously
248 in discrete populations. Sexuals immigrating from nearby populations may re-colonize the areas
249 of extinction until being invaded by another wave of asexuals in a multi-species metapopulation
250 dynamic. For the sexual species, however, Kokko et al.'s model implies one additional
251 consequence which stems from classical population genetic theory: if the sexual metapopulation
252 is forced to pass through such extinction-recolonization cycles induced by sperm-dependent
253 parthenogens, it follows that its effective population size will be diminished. Additionally, it also
254 follows that at each moment, some parts of range potentially suited for the sexual species will be
255 temporarily unavailable.

256 (b) Coexistence with sperm-dependent parthenogens also likely increases the variance in
257 reproductive success in a host sexual species because sexual individuals may spend a considerable
258 portion of their reproductive potential on mating with sperm-dependent parthenogens rather than
259 conspecific individuals (Schlupp 2010). Consequently, in each generation the sexual gene pool
260 would be re-constituted from fewer fathers than it would normally have in absence of sperm-
261 dependent parthenogens. It follows from classical population genetic theory that such an increased
262 variance in reproductive success would subsequently further reduce the effective population size
263 of the sexuals (Figure 2).

264 (c) Another consequence of the presence of sperm-dependent parthenogenetic females is
265 that male mate choice as well as female competition (Makowicz and Schlupp 2015) can occur
266 because the operational sex ratio is female biased (Schlupp 2009). Under some conditions, the
267 OSR, the ratio of reproductively active males and females, may be altered, which has consequences
268 for which sex is choosier (Amundsen 2018; Schlupp 2021). Simply put, if males become the rare

269 sex, they may switch from competing for females to being choosy. Such a mechanism has, for
270 example, been documented in some sexual fishes such as two-spotted goby, *Gobiusculus*
271 *flavescens* (Forsgren et al. 2004). In this mating system males become rare late in the season, and
272 consequently become choosier than females showing how important ecological conditions can be
273 in modifying sexual selection (Amundsen 2018). The presence of excessive number of asexual
274 females may have drastic effect on sex allocation and ratio in interacting sexual species. This
275 occurs e.g., in sexual brine shrimp *Artemia franciscana*, which adaptively adjust their sex ratio
276 under natural conditions. However, when co-occurring with the related obligate asexual all-female
277 species *A. parthenogenetica* in recently invaded parts of its distribution range *A. franciscana*
278 maladaptively produces extremely male-biased sex ratio (Lievens et al. 2016). The presence of
279 sperm-dependent females in sexual/asexual mating systems may have even a stronger effect and
280 alter the OSR in such a way that female choice is diminished, and male choice prevails (Figure 2).
281 A skewed OSR and increased variance in reproductive success of a sexual species thus further
282 reduces its effective population size, especially in those mixed populations where the proportion
283 of sperm-dependent parthenogenetic females is high. Such high proportions of sperm-dependent
284 parthenogens have been documented in several asexual-sexual complexes, often reaching over
285 70%, for example in frogs (*Pelophylax*, (Graf and Polls-Pelaz 1989; Mikulíček et al. 2015)),
286 salamanders (*Ambystoma*, (Bogart et al. 2009)), and fishes (*Squalius*, (Cunha et al. 2008); *Cobitis*
287 (Janko et al. 2007) and *Poecilia* (Heubel and Schlupp 2008).

288 All those mechanisms reducing effective population size have important implications for
289 the gene-pool of the sexual species, which can be predicted from population genetic theory. For
290 example, populations with diminished effective size have higher likelihood of inbreeding
291 depression and extinction (Byers and Waller 1999). Selection becomes less effective while the
292 effect of genetic drift increases, allowing more frequent fixation of deleterious alleles due to
293 chance. Additionally, the faster fixation of positively selected alleles leads to a greater loss of
294 genetic diversity in small populations (e.g. (Lande 1976; Lacy 1987)). Population size reduction
295 further increases the *Allee effect* (Glossary) in sexual populations and directly impacts
296 metapopulation connectivity by reducing the effective number of migrants (Lowe and Allendorf
297 2010). Consequently, lower efficiency of homogenizing geneflow speeds up local fixation of
298 alternative alleles in small, isolated populations (Cosentino et al. 2012), which may also contribute
299 to faster adaptation to local environments, potentially limiting the plasticity of population-wide
300 responses to stochastic events.

301 Despite clear predictions, the detection of such indirect effects on sexual gene pools is a
302 challenging task. Nevertheless, long-term studies of sperm-dependent asexual complexes may
303 provide suitable model systems to test this hypothesis. For instance, we can assume that in a
304 hybridogenetic complex of water frogs, the parental species *P. lessonae* is more affected by
305 hybridogenetic hybrids *P. esculentus* because in most populations it serves as a gamete donor for
306 them. Hybridogenetic hybrids thus could decrease the effective population size of *P. lessonae*. One
307 of the consequences of reduced population size and the higher rate of genetic drift could be the
308 higher genetic differentiation of *P. lessonae* populations, as we observe in nature, compared to
309 another parental species *P. ridibundus* that does not coexist frequently with hybrids and has lower
310 levels of population differentiation (Pruvost et al. (2015)). Another example of such indirect effects
311 comes from spined loaches, the *Cobitis* hybrid complex, where gametogenic performance of males
312 of the sexual species changed in response to a female-biased sex ratio in a mixed sexual-asexual
313 population (Juchno and Boroń 2006; Jablonska et al. 2020). Specifically, unlike their counterparts
314 from purely sexual populations, sexual males serving as sperm donors in mixed populations had
315 to meet the reproductive challenge of a high proportion of sperm-dependent parthenogenetic
316 females, which lead continual year-round sperm-production, higher production of spermatogonia
317 during and after spawning and lower rates of apoptosis in their testes.

318

319 ***2. Impact on speciation***

320 In this section we discuss how asexual organisms, and sperm-dependent asexuals in
321 particular, contribute to the existing biodiversity either indirectly by facilitating the speciation in
322 coexisting sexual species or directly by forming new species themselves.

323 **2.1. Indirect impact on speciation: promotion of reproductive isolation barriers**

324 We first present several ways, how sperm-dependent parthenogens, by their very presence,
325 may considerably affect population divergence as well as establishment of both prezygotic and
326 postzygotic reproductive isolation barriers (RIB) between populations of related sexuals. Such
327 mechanisms are rather cryptic and were appreciated only recently.

328 ***Postzygotic RIB:*** As already mentioned, the presence of asexuals, especially sperm-
329 dependent parthenogens, has a negative effect on effective population sizes and connectivity
330 among demes of interacting sexual species. This ultimately increases local drift, which in turn
331 increases inter-population differentiation (Figure 2). Moreover, if there is introgression from
332 clones, local sexual populations may diverge from each other even more rapidly because
333 introgression patterns mediated by asexuals substantially differ from those mediated by sexual

334 hybrids (see section 1). Sexual species coexisting with clones may thus become genetically more
335 fragmented than similar species without asexual counterparts.

336 Moreover, asexual hybrids can themselves play a role as primary postzygotic barrier
337 promoting speciation between diverging taxa. Indeed, postzygotic reproductive incompatibilities
338 are known to accumulate with genetic divergence between emerging species (Seehausen et al.
339 2014) and although such ‘speciation clocks’ do not tick at an universal rate for all taxa (Edmands
340 2002; Russell 2003; Bolnick and Near 2005; Matute et al. 2010), the initial stages are generally
341 characterized by decreased fertility or sterility of hybrids, while hybrid viability tends to be
342 compromised only at later stages with substantial genetic divergence. Interestingly, Ernst (1918)
343 noted already a century ago that the likelihood of asexual reproduction in hybrids also correlates
344 with divergence between its parental species, following a continuum from sexually reproducing
345 hybrids between closely related parents to obligate asexual hybrids between distant parental
346 species. Several later publications implied that distortion of hybrid’s gametogenesis towards
347 production of clonal gametes is possible in a particular ‘window of genetic divergence’ before
348 complete hybrid sterility emerges (Wetherington et al. 1987; Moritz et al. 1989; Carman 1997; De
349 Storme and Mason 2014). Some forms of asexual reproduction, both in animals and plants, may
350 even rescue the sterility of interspecific hybrids by allowing them to produce clonal gametes,
351 which further contributes to reproductive isolation of genepools of sexual parents (Carman et al.
352 2019; Dedukh et al. 2020). Finally, as recent analysis of spined loaches demonstrated, formation
353 of hybrid asexuality is not only correlated with divergence of parental species, but it also has
354 common cytogenetic background with hybrid sterility (Janko et al. 2018; Dedukh et al. 2020).

355 Thus, a century after seminal works by Bateson (1909) and Ernst (1918), it is becoming
356 clear that the formation of hybrid asexuality has many analogies with Bateson-Dobzhansky-Muller
357 speciation models. Moreover, since asexually reproducing hybrids restrict the gene flow between
358 parental species to much higher extent than fertile sexual hybrids, hybrid asexuality may be viewed
359 as a special type of postzygotic incompatibility, which arises at earlier stages of speciation than
360 other barriers like complete hybrid sterility or inviability (Janko et al. 2018) (Figure 3). Empirical
361 examples do support the view of a link between the ‘classical speciation continuum’ and hybrid
362 asexuality (Lampert et al. 2007; Janko et al. 2018), which has recently been called ‘*extended*
363 *speciation continuum*’ (Stöck et al. 2021). Among vertebrates, for instance, on one end, there are
364 dynamically hybridizing species pairs that produce diverse assemblages of asexual hybrids such
365 as *Cobitis* (Choleva et al. 2012), *Pelophylax* (Hotz et al. 1985; Hoffmann et al. 2015), and

366 *Poeciliopsis* (Schultz 1973). On the other end there are single clonal hybrid taxa stemming from
367 one or a few ancient events whereas attempts to cross their contemporary sexual relatives fail to
368 produce clones, such as *Poecilia* (Lampert and Schartl 2008; Stöck et al. 2010). Cases like the fish
369 *Chrosomus* (the former *Phoxinus*) may represent an intermediate condition, where historical
370 hybridizations produced a highly diverse asexual assemblage, but new clones can no longer be
371 produced by contemporary diverged parental species (Angers and Schlosser 2007). Similarly,
372 some species pairs appear unable to produce sexual hybrids and their hybridization gives rise to
373 asexuals only (e.g., *Cobitis elongatoides-taenia*) while clonal and sexual hybrid females co-occur,
374 and fertile hybrid males may exist in other hybridizing pairs. This is found in fishes such as
375 *Fundulus* (Hernández Chávez and Turgeon 2007), and *Rutilus rutilus* x *Abramis bramma* (Slyn'ko
376 2000).

377 Such a view of the link between speciation and asexuality (Janko et al. 2018; Stöck et al.
378 2021) has important implications for the perception of the role of asexuality in evolutionary
379 biology. Given that asexual lineages are often assumed to have a short evolutionary life span, e.g.
380 (Butlin et al. 1999), and that the phase when diverging species can produce asexual hybrids is
381 transient, it is possible that many extant “good” sexual species might have historically produced
382 asexual hybrids as part of the speciation process, with the transitory forms gone extinct.

383 **Prezygotic barriers:** The presence of sperm-dependent parthenogens also affects the
384 premating isolation mechanisms, e.g., by exerting selection pressure on mate-recognition systems
385 in co-occurring sexuals. While such asexuals rely on sperm, from the point of view of a sexual
386 host mating with them represents a costly behavior and hence it has been postulated that the
387 stability of sexual- sperm-dependent parthenogen complexes relies on the ability of sexual males
388 to discriminate hybrid and conspecific females (Schlupp and Plath 2005; Mee and Otto 2010;
389 Morgado-Santos et al. 2015); Figure 2. This process is somewhat similar to reinforcement where
390 the evolution of mate choice is often selected for in zones where ranges of hybridizing species
391 overlap (Marshall et al. 2002) and character displacement is predicted (Gabor and Ryan 2001).
392 However, contrary to classical cases, the distribution of sperm-dependent parthenogenetic hybrids
393 represents something like a hybrid zone extended in time and space because such hybrids often
394 expand over large parts of parental ranges, sometimes well beyond their area of origin (Janko et
395 al. 2019). Hence, contrary to a classical reinforcement scenario which takes place in narrow zones
396 of sympatry, sperm-dependent parthenogens exert selective pressures over vast areas deep in
397 allopatry and may therefore considerably speed-up the establishment of prezygotic isolation.

398 While only a few studies examined geographical variation in male mate choice against
399 asexuals, some empirical support for such hypothesis exists. For instance, Gabor and Ryan (Gabor
400 and Ryan 2001) and Gabor et al. (Gabor et al. 2005) found that males of the sexual sailfin molly

401 (*P. latipinna*) living sympatrically with gynogenetic Amazon mollies (*P. formosa*) showed a
 402 significantly stronger mating preference for conspecific females than males from populations that
 403 were allopatric with Amazon mollies. Another study by Gabor et al. (Gabor et al. 2013), however,
 404 showed that male mate choice varied geographically and may be associated with variation among
 405 populations in the length of sympatry with the gynogenetic Amazon molly (*P. formosa*).
 406 Furthermore, metapopulation dynamics may depend also on a conflict between species and mate
 407 quality recognition cues (visual, chemical, and tactile). Asexual – sexual mating systems in mollies
 408 represent complex networks where, moreover, not just male mate choice should evolve, but also
 409 interspecific female competition (Makowicz and Schlupp 2015).

410 In addition, given that at least some sexual and asexual species pairs substantially differ
 411 ecologically (Vrijenhoek 1994; Pantel et al. 2011; Ross et al. 2012; Van der Kooi et al. 2017), it
 412 may also be hypothesized that asexuals could drive character displacement in sexual species also
 413 in its ecological characters. In such a case, the presence of asexuals may ultimately generate
 414 ecological divergence between allopatric sexual populations and those in sympatry with asexuals,
 415 perhaps even providing a first step in ecological speciation.

416

417 **2.2. Direct impact on speciation: Species formation by asexuals**

418 Finally, there is a more direct way how asexuals may contribute to biodiversity; they may
 419 form new species themselves. The question whether speciation can occur without sex is
 420 longstanding (Coyne and Orr 2004; Shcherbakov 2010; Dubois 2011; Hausdorf 2011) but recent
 421 theoretical studies show that species-level taxa may be formed also in asexuals (e.g. (Cohan 2001,
 422 2002; Franklin 2007; Birky and Barraclough 2009)). This has been empirically indicated in some
 423 ancient asexual organisms like bdelloid rotifers, oribatid mites, ostracodes, as well as in plants like
 424 *Boechera*, where genetically, morphologically, and ecologically distinct species-like clusters occur
 425 within asexual clades (Domes et al. 2007; Fontaneto et al. 2007; Schön et al. 2012; Carman et al.
 426 2019). Interestingly, the rates of cladogenesis in asexual taxa can achieve similar or even higher
 427 values than in comparable sexual lineages (Fontaneto et al. 2012). The potential of diversification
 428 in asexuals is thus becoming increasingly appreciated.

429 What may be even more interesting for general biology is that at least in some groups, the
 430 asexuals could play a role as stepping-stones in the evolution of new sexual species. The idea that
 431 asexuals may somehow revert to sex and thereby give rise to new sexual species attracted
 432 considerable attention particularly in the botanical literature and has been used as a possible

433 explanation for the origin of hybrid and polyploid species (Figure 1D). Namely, because the
434 emergence of a novel hybrid/polyploid form is supposedly a rare phenomenon and its
435 establishment is thus threatened by a frequency-dependent disadvantage (i.e., the minority
436 cytotype exclusion principle, e.g. (Husband 2000)), the establishment of novel strains could be
437 facilitated by asexual reproduction, which offers immediate reproductive isolation and clonal
438 multiplication of genotypes (Rieseberg and Willis 2007; Choleva and Janko 2013; Hojsgaard and
439 Hörandl 2015). Asexuality can thus represent the first stages towards hybrid speciation. For
440 example, clonally reproducing triploids were suggested to serve as ‘triploid bridge’ towards
441 tetraploid species with re-assumed sexual reproduction (Cunha et al. 2008; Choleva and Janko
442 2013; Hojsgaard and Hörandl 2015; Dubey et al. 2019). This mechanism is particularly appealing
443 in sperm-dependent asexuals since they rely on sperm source and hence seem particularly prone
444 to fertilization with ploidy increase. However, recent data offer controversial support for this
445 hypothesis since most known tetraploids derived from extant triploids are rather sterile or have a
446 fitness disadvantage (reviewed in (Choleva and Janko 2013)). Still, there exists empirical evidence
447 that established obligatory asexuals may revert to sex, as found for example in fish, *Squalius*
448 (Cunha et al. 2008), Oribatid mites (Domes et al. 2007) and some plant taxa, like *Hieracium*
449 *pilosella* (Fehrer et al. 2005).

450 The apparent paucity of asexual to sexual transitions may result from the fact that established
451 asexuals may be selected for the loss of sexual traits, which are disadvantageous or unnecessary
452 for clonal reproduction (Kooi and Schwander 2014), thereby preventing the re-evolution of sex.
453 However, this should not be the case in sperm-dependent parthenogens, which have to maintain
454 the full genetic machinery allowing them to mate with sexuals (Schlupp et al. 1998; Warren et al.
455 2018). Scarcity of asexual to sexual transitions may thus be only apparent, because the
456 identification of sexually reproducing species which passed through a phase of clonality is
457 extremely difficult, and probably easily overlooked.

458 There is indeed sound evidence for the ability of some sperm-dependent parthenogens to
459 form populations independent of sexual sperm-donors, which might be a first step towards either
460 true parthenogenesis or sexuality. Such cases have been recently documented in gynogenetic
461 lineages of a nematode (Grosmaire et al. 2019) and sexual speciation in *statu nascenti* occurs in
462 hybridogenetic water frogs, where a transition occurred from hemiclinal to sexual hybrids, which
463 are reproductively independent from the parental species and form pure-hybrid populations with a
464 high proportion of triploids. Triploid hybrids form gametes with the genome of each of the parental
465 species and thus substitute parental individuals in pure-hybrid populations (Berger 1983;
466 Christiansen et al. 2005; Christiansen and Reyer 2009) (Figure 1F).

467 What makes such a case extremely interesting is the discovery by Stöck et al. (2002) who
468 described an all-triploid bisexual frog species, *Bufo baturae*. This species combines two
469 genomes (two copies of the so-called NOR⁺ genome and one copy of NOR⁻ genome), whose
470 transmission to gametes sharply differs between males and females. While males eliminate the
471 NOR⁻ genome and recombine and segregate NOR⁺ genomes in order to produce haploid sperm,
472 females produce diploid gametes containing clonally transmitted NOR⁻ genomes and recombined
473 NOR⁺ genomes. Fusion of such gametes restores triploidy in every generation. Given the similarity
474 to the *P. esculentus* system, there is an intriguing possibility that bisexual *B. baturae* evolved
475 through an asexual stage and that other systems, like *P. esculentus* may be on a similar evolutionary
476 pathway just at a different stage.

477 These cases indicate that a transition from asexual to sexual is at least a plausible scenario
478 for how asexuals can directly contribute to formation of the regular sexual species.

479

480 **3. Impact on spatial distribution of sexual species**

481 On a large spatial scale, asexuals are known to occupy more disturbed or weedy habitats,
482 displacing sexual species from these areas. This **geographical parthenogenesis** (Glossary) seems
483 to be widespread and is probably the most prominent pattern of asexuality (Peck et al. 1998;
484 Kearney 2005; Hörandl 2009; Tilquin and Kokko 2016). Due to tight link between asexuality
485 (apomixis) and polyploidy and hybridization, there is an ongoing debate as to what extent the
486 patterns of geographical parthenogenesis relate to asexuality *per se* and to what extent they reflect
487 these confounding effects (Kearney 2005; Lundmark 2006), such as for example the niche shifts
488 related to ploidy (Coughlan et al. 2017; Kirchheimer et al. 2018) or higher competitiveness of
489 polyploids in peripheral areas (Karunarathne et al. 2018). Whatever the underlying reason, the
490 patterns of geographical parthenogenesis imply that parthenogens outcompete related sexual
491 species in marginal, resource-poor environments because sexual populations cannot efficiently
492 adapt to local adverse conditions due to dispersal and admixis with individuals from denser-
493 populated more favorable regions.

494 This means, however, that even after the demise of the asexual (e.g., due to the
495 accumulation of deleterious mutations, or other processes), the sexual species will be less
496 successful in colonizing any open ecological niche because they did not have an opportunity to
497 adapt to the conditions. Instead, they are adapted only to the more favorable habitats to which they
498 were restricted. Hence, the effect of asexuality would persist beyond the extinction of an asexual

499 species. This can turn into a cyclical process, if another asexual clone colonizes a niche before the
500 sexual species can adapt to it.

501 As we have discussed in section 1, the presence of sperm-dependent parthenogens also
502 has considerable demographic consequences for its sexual host whose population sizes and density
503 are reduced by direct competition for males and other resources. For instance, Janko & Eisner
504 (2009) used a mathematical model to demonstrate that a sexual population “infected” with sperm-
505 dependent parthenogens should have a limited potential of spatial expansion and colonization of
506 new habitats compared to a situation where no sperm-dependent parthenogens affect it (Figure 4).
507 Such a reduction in population expansion speed stems from reduced density along the expanding
508 wave and therefore a decreased chance to find a mating partner needed to establish viable
509 populations in invaded areas. Hence, sperm-dependent parthenogens may decisively affect large-
510 scale biogeographic patterns of their sexual hosts that, even after an eventual extinction of
511 asexuals, would not be able to expand into new habitats that might have been occupied by another
512 species in the meantime.

513 The biogeography of European loaches of the genus *Cobitis* potentially offers an
514 empirical example of such a process. While in many European freshwater fishes the postglacial
515 recolonization of Central and Western Europe proceeded from Danubian/Pannonian refugia,
516 *Cobitis* fishes show a contrasting pattern since the Danubian species *C. elongatoides* experienced
517 only a limited postglacial expansion reaching only the upper Odra and Elbe river watersheds, while
518 most of Europe was colonized by *C. taenia* rapidly expanding from Eastern refugia (Janko et al.
519 2005). Interestingly, *C. elongatoides* populations survived the last glacial maximum in Danubian
520 refugia together with sperm-dependent parthenogenetic hybrids and co-expanded with them into
521 northern areas, which might have delayed its colonization rate as compared to *C. taenia*, whose
522 expansion was not burdened by sexual parasites (Janko and Eisner 2009).

523 In addition, sperm-dependent parthenogens may decisively affect the results of
524 interspecific coexistence and competition among interacting sexual species. To appreciate this
525 phenomenon, let us emphasize that many sperm-dependent asexuals originated by hybridization
526 between several sexual species (Neaves and Baumann 2011; Choleva et al. 2012) and they can
527 therefore simultaneously use (or parasitize) two or more sexual species for their own reproduction
528 e.g. (Schlupp 2005; Choleva et al. 2008). In such cases, mathematical models of dispersal with
529 competition showed that the sexual host species with better mate recognition ability or smaller
530 niche overlap with coexisting parthenogens will be less negatively affected by their presence,
531 giving it a potentially decisive advantage in competition with other sexual species, whose
532 demographic performance is harmed to a greater extent (Janko et al. 2019). Gynogens can therefore
533 mediate the so-called *apparent competition* (Glossary) among sexual species and cause an effect

534 analogous to *parasite-mediated competition* (Glossary) (Thomas et al. 2000; Holt and Bonsall
 535 2017). In their presence, even a stronger sexual competitor may be outcompeted by a weaker one
 536 if the latter is less negatively impacted by coexisting sperm-dependent parthenogens (Figure 5). It
 537 follows that the effect on the diversity of sexual species would remain even if the asexuals
 538 eventually go extinct.

539 On the other hand, as discussed above, some asexuals may also have a positive effect on
 540 the distribution of their sexual hosts as they can transmit their genes far away from their own
 541 distribution. The examples are hybridogenetic frogs *P. esculentus* which serve as a "vector"
 542 transmitting clonal genomes of a parental species, *P. ridibundus*, to western Europe, far beyond
 543 its original range (Arano et al. 1994; Pagano et al. 2001), gynogenetic *Cobitis* hybrids that transmit
 544 genes of its Danubian parent, *C. elongatoides*, hundreds of kilometers outside its range to the Rhine
 545 River region and to the southern Ukraine (Choleva et al. 2008), or *Corbicula* clams, whose asexual
 546 hermaphroditic reproduction increases invasive success (Pigneur et al. 2011, 2012). Similarly in
 547 plants, such as in the genus *Rubus*, polyploid apomicts may preserve ancestral alleles lost in their
 548 sexual ancestors during Pleistocene ice-age bottlenecks and spread younger alleles obtained from
 549 diploids via recent gene flow (Sochor et al. 2017).

550

551 **Conclusions**

552 Gregor Mendel was unable to replicate his findings on peas in the *Hieracium* model due to its
 553 asexual reproduction. Of course, he had no way of knowing, but nonetheless, this story illustrates
 554 how perilous it can be to ignore asexuals. Asexuals are worthy objects for studies by themselves,
 555 but their ecological and evolutionary influences on other species and biodiversity in general are
 556 perhaps even more important and should be more appreciated. Here, we listed several mechanisms
 557 how the very existence of asexuals, in particular sperm-dependent parthenogens, can have major
 558 effects on coexisting sexual species and biodiversity overall in terms of their 1) genetic
 559 architecture, 2) diversification and speciation, and 3) spatial distribution.

560 1) The genetic architecture of sexual species might be influenced in systems when a clonal
 561 genome finds its way into a sexual genepool. Such asexual-to-sexual gene flow differs from gene
 562 exchange between sexual species because asexuals transmit their genetic material without
 563 recombination. The recipient sexual genepool might be impacted negatively by introducing
 564 deleterious mutations but also positively by advantageous combinations of alleles that coevolved
 565 in linkage. Asexuals of hybrid origin can further serve as a bridge for introgression of alleles or

566 whole mitochondrial genomes between sexual species. Even without asexual-to-sexual gene flow,
567 extensive mating between sperm-dependent parthenogens and their sexual hosts can reduce
568 effective population size, increase the strength of genetic drift and thus decrease genetic variability
569 and efficiency of natural selection in sexual populations.

570 2) Sperm-dependent parthenogens can contribute to differentiation of populations of sexual
571 species by reducing their effective population size and increasing the strength of genetic drift but
572 can also promote the accomplishment of speciation between hybridizing taxa. Since asexually
573 reproducing hybrids restrict the gene flow between parental species to much higher extent than
574 sexual hybrids, hybrid asexuality may be viewed as a special type of postzygotic incompatibility.
575 It could be also predicted that sperm-dependent parthenogens reinforce the formation of prezygotic
576 barriers by exerting selection pressure on mate-recognition systems in hybridizing sexual species,
577 a process analogous to reinforcement in classical hybrid zones. Finally, asexuals can play an
578 important role in speciation when, through an intermediate stage of polyploid forms, they can
579 evolve into a new sexual species.

580 3) Asexuals may affect large-scale biogeographic patterns of related sexual species. They
581 frequently outcompete their sexual counterparts in disturbed habitats, at higher latitudes and
582 altitudes, or at the edge of distribution ranges (geographical parthenogenesis). Theoretical models
583 also reveal that sperm-dependent asexuals reduce abundance and density of sexual populations and
584 thus may reduce expansion speed of sexual host species. In complexes composed of two sexual
585 species and their sperm-dependent hybrids, sperm-dependent asexuals may significantly affect
586 competition between sexual species by a process analogous to parasite-mediated competition (or
587 apparent competition) well known in classical host-parasite systems. Sperm-dependent
588 parthenogens thus might be important players forming the structure of ecosystems.

589 Recent advances show that most proposed mechanisms do have support from empirical
590 cases of asexual – sexual coexistence in nature. It indicates that even if the existence of individual
591 clonal species may be ephemeral from an evolutionary point of view, their impact on sexual species
592 likely lasts much longer than the existence of individual clonal lineages. These are all important
593 population or species level effects that should be included into our research programs. We hope
594 our review is going to stimulate further research into the questions we raise. In our review we
595 elected to focus mainly on animals with some mentioning of plants. A combined review of the
596 plant and animal literature is desirable but made exceedingly difficult by important differences in
597 reproductive biology, as well as differences in terminology. We also listed several empirical
598 examples supporting their relevance. Many such examples concerned our own work, but this was
599 not out of vanity, but rather because the discussed aspects of asexual – sexual interactions have

600 otherwise received little attention to date. Our review provides testable hypotheses with clear
601 predictions that may be explored by other scientists in other systems, searching for potentially
602 overlooked empirical examples.

603 We also hope that expanded research will create positive feedback for ecology and
604 evolution in general as new tools are developed. As an example, consider testing the hypothesis
605 that some current sexual organisms have passed through an asexual stage in their evolutionary
606 history either because of an asexual bridge, or because they had an asexual phase during speciation.
607 This would require the development of novel analytical tools incorporating not only population
608 genetic approaches but also explicit models of asexual hereditary patterns. Such tools may
609 eventually not just help finding influences of asexuals but will also improve tools for all of biology.

610

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619

620 **References:**

- 621 Alves, M. J., M. M. Coelho, and M. J. Collares-Pereira. 2001. Evolution in action through
622 hybridisation and polyploidy in an Iberian freshwater fish: a genetic review. *Genetica*
623 111:375–385.
- 624 Alves, M. J., M. J. Collares-Pereira, T. E. Dowling, and M. M. Coelho. 2002. The genetics of
625 maintenance of an all-male lineage in the *Squalius alburnoides* complex. *Journal of Fish*
626 *Biology* 60:649–662.
- 627 Amundsen, T. 2018. Sex roles and sexual selection: Lessons from a dynamic model system.
628 *Current Zoology* 64:363–392.
- 629 Angers, B., C. Leung, R. Vétit, L. Deremiens, and R. Vergilino. 2018. The effects of allospecific
630 mitochondrial genome on the fitness of northern redbelly dace (*Chrosomus eos*). *Ecology*
631 *and Evolution* 8:3311–3321.
- 632 Angers, B., and I. J. Schlosser. 2007. The origin of *Phoxinus eos-neogaeus* unisexual hybrids.
633 *Mol. Ecol.* 16:4562–4571.
- 634 Arano, B., G. A. Llorente, P. Herrero, and B. Sanchiz. 1994. Current studies on Iberian water
635 frogs. *Zoologica Poloniae* 39:365–375.
- 636 Avise, I. J. 2008. *Clonality : The Genetics, Ecology, and Evolution of Sexual Abstinence in*
637 *Vertebrate Animals*. Oxford University Press, New York.
- 638 Barbuti, R., S. Mautner, G. Carnevale, P. Milazzo, A. Rama, and C. Sturmbauer. 2012.
639 Population dynamics with a mixed type of sexual and asexual reproduction in a
640 fluctuating environment. *BMC Evolutionary Biology* 12:49.
- 641 Bateson, W. 1909. Heredity and variation in modern lights. Pp. 85–101 *in Darwin and Modern*
642 *Science*. Cambridge University Press: Cambridge.
- 643 Bengtsson, B. O. 2009. Asex and Evolution: A Very Large-Scale Overview. Pp. 1–19 *in I.*
644 *Schön, K. Martens, and P. Dijk, eds. Lost Sex: The Evolutionary Biology of*
645 *Parthenogenesis*. Springer Netherlands, Dordrecht.

- 646 Berger, L. 1983. Western Palearctic water frogs (Amphibia, Ranidae): Systematics, genetics and
647 population compositions. *Experientia* 39:127–130.
- 648 Bernstein, H., and C. Bernstein. 2010. Evolutionary Origin of Recombination during Meiosis.
649 *BioScience* 60:498–505.
- 650 Beukeboom, L. W., and R. C. Vrijenhoek. 1998. Evolutionary genetics and ecology of sperm-
651 dependent parthenogenesis. *Journal of Evolutionary Biology* 11:755–782.
- 652 Bi, K., J. P. Bogart, and J. Fu. 2008. The prevalence of genome replacement in unisexual
653 salamanders of the genus *Ambystoma* (Amphibia, Caudata) revealed by nuclear gene
654 genealogy. *BMC Evolutionary Biology* 8:158.
- 655 Bicknell, R., A. Catanach, M. Hand, and A. Koltunow. 2016. Seeds of doubt: Mendel’s choice of
656 *Hieracium* to study inheritance, a case of right plant, wrong trait. *Theor. Appl. Genet.*
657 129:2253–2266.
- 658 Binet, M.-C., and B. Angers. 2005. Genetic identification of members of the *Phoxinus eos-*
659 *neogaeus* hybrid complex. *Journal of Fish Biology* 67:1169–1177.
- 660 Birky, C. W., and T. G. Barraclough. 2009. Asexual Speciation. Pp. 201–216 in I. Schön, K.
661 Martens, and P. van Dijk, eds. *Lost Sex: The Evolutionary Biology of Parthenogenesis.*
662 Springer, Dordrecht, Heidelberg, London, New York.
- 663 Bogart, J. P. 2019. Unisexual Salamanders in the Genus *Ambystoma*. *Herpetologica* 75:259–267.
- 664 Bogart, J. P., J. Bartoszek, D. W. A. Noble, and K. Bi. 2009. Sex in unisexual salamanders:
665 discovery of a new sperm donor with ancient affinities. *Heredity* 103:483–493.
- 666 Bolnick, D. I., and T. J. Near. 2005. Tempo of Hybrid Inviability in Centrarchid Fishes
667 (Teleostei: Centrarchidae). *Evolution* 59:1754–1767.

668 Brockhurst, M. A., T. Chapman, K. C. King, J. E. Mank, S. Paterson, and G. D. D. Hurst. 2014.
669 Running with the Red Queen: the role of biotic conflicts in evolution. *Proceedings of the*
670 *Royal Society B: Biological Sciences* 281:20141382.

671 Butlin, R. K., I. Schön, and K. Martens. 1999. Origin, age and diversity of clones. *Journal of*
672 *Evolutionary Biology* 12:1020–1022.

673 Byers, D. L., and D. M. Waller. 1999. Do Plant Populations Purge Their Genetic Load? Effects
674 of Population Size and Mating History on Inbreeding Depression. *Annual Review of*
675 *Ecology and Systematics* 30:479–513.

676 Carman, J. G. 1997. Asynchronous expression of duplicate genes in angiosperms may cause
677 apomixis, bispority, tetraspority, and polyembryony. *Biological Journal of the Linnean*
678 *Society* 61:51–94.

679 Carman, J. G., M. Mateo de Arias, L. Gao, X. Zhao, B. M. Kowallis, D. A. Sherwood, M. K.
680 Srivastava, K. K. Dwivedi, B. J. Price, L. Watts, and M. D. Windham. 2019. Apospority
681 and Diplospority in Diploid *Boechera* (Brassicaceae) May Facilitate Speciation by
682 Recombination-Driven Apomixis-to-Sex Reversals. *Front. Plant Sci.* 10, article number
683 724.

684 Charlesworth, B., and D. Charlesworth. 1978. A Model for the Evolution of Dioecy and
685 Gynodioecy. *The American Naturalist* 112:975–997.

686 Choleva, L., A. Apostolou, P. Ráb, and K. Janko. 2008. Making it on their own: sperm-
687 dependent hybrid fishes (*Cobitis*) switch the sexual hosts and expand beyond the ranges
688 of their original sperm donors. *Philosophical Transactions of the Royal Society B:*
689 *Biological Sciences* 363:2911.

690 Choleva, L., and K. Janko. 2013. Rise and Persistence of Animal Polyploidy: Evolutionary
691 Constraints and Potential. *Cytogenetic and Genome Research* 140:151–170.

- 692 Choleva, L., K. Janko, K. De Gelas, J. Bohlen, V. Šlechtová, M. Rábová, and P. Ráb. 2012.
693 Synthesis of Clonality and Polyploidy in Vertebrate Animals by Hybridization Between
694 Two Sexual Species. *Evolution* 66:2191–2203.
- 695 Choleva, L., Z. Musilova, A. Kohoutova-Sediva, J. Paces, P. Rab, and K. Janko. 2014.
696 Distinguishing between Incomplete Lineage Sorting and Genomic Introgressions:
697 Complete Fixation of Allospecific Mitochondrial DNA in a Sexually Reproducing Fish
698 (*Cobitis*; Teleostei), despite Clonal Reproduction of Hybrids. *PLoS ONE* 9:e80641.
- 699 Christiansen, D. G., K. Fog, B. V. Pedersen, and J. J. Boomsma. 2005. Reproduction and hybrid
700 load in all-hybrid populations of *Rana esculenta* water frogs in Denmark. *Evolution*
701 59:1348–1361.
- 702 Christiansen, D. G., and H.-U. Reyer. 2009. From clonal to sexual hybrids: genetic
703 recombination via triploids in all-hybrid populations of water frogs. *Evolution* 63:1754–
704 1768.
- 705 Cohan, F. M. 2001. Bacterial species and speciation. *Syst. Biol.* 50:513–524.
- 706 Cohan, F. M. 2002. Sexual isolation and speciation in bacteria. *Genetica* 116:359–370.
- 707 Cosentino, B. J., C. A. Phillips, R. L. Schooley, W. H. Lowe, and M. R. Douglas. 2012. Linking
708 extinction–colonization dynamics to genetic structure in a salamander metapopulation.
709 *Proc. Biol. Sci.* 279:1575–1582.
- 710 Coughlan, J. M., S. Han, S. Stefanović, and T. A. Dickinson. 2017. Widespread generalist clones
711 are associated with range and niche expansion in allopolyploids of Pacific Northwest
712 Hawthorns (*Crataegus* L.). *Mol. Ecol.* 26:5484–5499.
- 713 Coyne, J. A., and H. A. Orr. 2004. *Speciation*. Illustrated edition. Sinauer Associates.

714 Cunha, C., I. Doadrio, and M. M. Coelho. 2008. Speciation towards tetraploidization after
715 intermediate processes of non-sexual reproduction. *Philos. Trans. R. Soc. Lond. B Biol.*
716 *Sci.* 363:2921–2929.

717 Dalziel, A. C., S. Tirbhowan, H. F. Drapeau, C. Power, L. S. Jonah, Y. A. Gbotsyo, and A.-M.
718 Dion-Côté. 2020. Using asexual vertebrates to study genome evolution and animal
719 physiology: Banded (*Fundulus diaphanus*) x Common Killifish (*F. heteroclitus*) hybrid
720 lineages as a model system. *Evolutionary Applications* 13:1214–1239.

721 Darlington, C. D. 1939. The Evolution of Genetic Systems. *Annals of the Entomological Society*
722 *of America* 32:656–657.

723 De Storme, N., and A. Mason. 2014. Plant speciation through chromosome instability and ploidy
724 change: Cellular mechanisms, molecular factors and evolutionary relevance. *Current*
725 *Plant Biology* 1:10–33.

726 Dedukh, D., Z. Majtánová, A. Marta, M. Pšenička, J. Kotusz, J. Klíma, D. Juchno, A. Boron, and
727 K. Janko. 2020. Parthenogenesis as a Solution to Hybrid Sterility: the Mechanistic Basis
728 of Meiotic Distortions in Clonal and Sterile Hybrids. *Genetics* 215:975–987.

729 Denton, R. D., A. E. Morales, and H. L. Gibbs. 2018. Genome-specific histories of divergence
730 and introgression between an allopolyploid unisexual salamander lineage and two
731 ancestral sexual species. *Evolution* 72:1689–1700.

732 Domes, K., R. A. Norton, M. Maraun, and S. Scheu. 2007. Reevolution of sexuality breaks
733 Dollo’s law. *Proc. Natl. Acad. Sci. U. S. A.* 104:7139–7144.

734 D’Souza, T. G., and N. K. Michiels. 2009. Sex in Parthenogenetic Planarians: Phylogenetic Relic
735 or Evolutionary Resurrection? Pp. 377–397 in I. Schön, K. Martens, and P. Dijk, eds.
736 *Lost Sex: The Evolutionary Biology of Parthenogenesis*. Springer Netherlands,
737 Dordrecht.

738 D’Souza, T. G., and N. K. Michiels. 2010. The Costs and Benefits of Occasional Sex:
739 Theoretical Predictions and a Case Study. *Journal of Heredity* 101:34–41.

- 740 Dubey, S., T. Maddalena, L. Bonny, D. L. Jeffries, and C. Dufresnes. 2019. Population genomics
741 of an exceptional hybridogenetic system of *Pelophylax* water frogs. *BMC Evol. Biol.*
742 19:164.
- 743 Dubois, A. 2011. Species and “strange species” in zoology: Do we need a “unified concept of
744 species”? *Comptes Rendus Palevol.* 10:77–94.
- 745 Edmands, S. 2002. Does parental divergence predict reproductive compatibility? *Trends in*
746 *Ecology & Evolution* 17:520–527.
- 747 Ernst, A. 1918. Bastardierung als Ursache der Apogamie im Pflanzenreich. Eine Hypothese zur
748 experimentellen Vererbungs- und Abstammungslehre. Nabu Press.
- 749 Fehrer, J., R. Šimek, A. Krahulcová, F. Krahulec, J. Chrtek, E. Brautigam, and S. Bräutigam.
750 2005. Evolution, hybridization, and clonal distribution of apo- and amphimictic species
751 of *Hieracium* subgen. *Pilosella* Asteraceae, Lactucaceae in a central European mountain
752 range. Pp. 175–201 in In: Bakker F, Chatrou L, Gravendeel B, Pelsers PB, editors. *Plant*
753 *species-level systematics: new perspectives on pattern and process.* Ruggell,
754 Liechtenstein: Gantner Verlag.
- 755 Fontaneto, D., E. A. Herniou, C. Boschetti, M. Caprioli, G. Melone, C. Ricci, and T. G.
756 Barraclough. 2007. Independently Evolving Species in Asexual Bdelloid Rotifers. *PLoS*
757 *Biol.* 5:e87.
- 758 Fontaneto, D., C. Q. Tang, U. Obertegger, F. Leasi, and T. G. Barraclough. 2012. Different
759 Diversification Rates Between Sexual and Asexual Organisms. *Evol. Biol.* 39:262–270.
- 760 Forsgren, E., T. Amundsen, Å. A. Borg, and J. Bjelvenmark. 2004. Unusually dynamic sex roles
761 in a fish. *Nature* 429:551–554.
- 762 Franklin, L. R. 2007. Bacteria, Sex, and Systematics. *Philosophy of Science* 74:69–95.

763 Fujita, M. K., S. Singhal, T. O. Brunes, and J. A. Maldonado. 2020. Evolutionary Dynamics and
764 Consequences of Parthenogenesis in Vertebrates. *Annual Review of Ecology, Evolution,*
765 *and Systematics* 51:191–214.

766 Gabor, C. R., L. A. da Barbiano, and A. S. Aspbury. 2013. Geographic variation in male mate
767 choice in a gynogenetic species complex: evaluating long-term data across mating
768 contexts. *Evol. Ecol. Res.* 15:653–666.

769 Gabor, C. R., and M. J. Ryan. 2001. Geographical variation in reproductive character
770 displacement in mate choice by male sailfin mollies. *Proceedings of the Royal Society of*
771 *London. Series B: Biological Sciences* 268:1063–1070.

772 Gabor, C. R., M. J. Ryan, and D. C. Morizot. 2005. Character displacement in sailfin mollies,
773 *Poecilia latipinna*: allozymes and behavior. *Environ. Biol. Fish.* 73:75–88.

774 Glémin, S., C. M. François, and N. Galtier. 2019. Genome Evolution in Outcrossing vs. Selfing
775 vs. Asexual Species. Pp. 331–369 in M. Anisimova, ed. *Evolutionary Genomics:*
776 *Statistical and Computational Methods.* Springer, New York, NY.

777 Goddard, K. A., and R. J. Schultz. 1993. Aclonal Reproduction by Polyploid Members of the
778 Clonal Hybrid Species *Phoxinus eos-neogaeus* (Cyprinidae). *Copeia* 1993:650–660.

779 Graf, J.-D., and M. Polls-Pelaz. 1989. Evolutionary genetics of the *Rana esculenta* complex. Pp.
780 289–301 in Dawley RM, Bogart JP, editors. *Evolution and Ecology of Unisexual*
781 *Vertebrates.* New York State Museum, Albany, New York.

782 Grosmaire, M., C. Launay, M. Siegwald, T. Brugière, L. Estrada-Virrueta, D. Berger, C. Burny,
783 L. Modolo, M. Blaxter, P. Meister, M.-A. Félix, P.-H. Gouyon, and M. Delattre. 2019.
784 Males as somatic investment in a parthenogenetic nematode. *Science* 363:1210–1213.

785 Hausdorf, B. 2011. Progress Toward a General Species Concept. *Evolution* 65:923–931.

786 Hedtke, S. M., and D. M. Hillis. 2011. The potential role of androgenesis in cytoplasmic-nuclear
787 phylogenetic discordance. *Syst. Biol.* 60:87–96.

- 788 Hedtke, S. M., K. Stanger-Hall, R. J. Baker, and D. M. Hillis. 2008. All-male asexuality: origin
789 and maintenance of androgenesis in the Asian clam *Corbicula*. *Evolution* 62:1119–1136.
- 790 Hernández Chávez, C., and J. Turgeon. 2007. Asexual and sexual hybrids between *Fundulus*
791 *diaphanus* and *F. heteroclitus* in the Canadian Atlantic region. *Molecular Ecology*
792 16:1467–1480.
- 793 Heubel, K. U., and I. Schlupp. 2008. Seasonal plasticity in male mating preferences in sailfin
794 mollies. *Behav. Ecol.* 19:1080–1086.
- 795 Hoffmann, A., J. Plötner, N. B. M. Pruvost, D. G. Christiansen, S. Röthlisberger, L. Choleva, P.
796 Mikulíček, D. Cogălniceanu, I. Sas-Kovács, D. Shabanov, S. Morozov-Leonov, and H.-
797 U. Reyer. 2015. Genetic diversity and distribution patterns of diploid and polyploid
798 hybrid water frog populations (*Pelophylax esculentus* complex) across Europe. *Molecular*
799 *Ecology* 24:4371–4391.
- 800 Hojsgaard, D., and E. Hörandl. 2015. Apomixis as a Facilitator of Range Expansion and
801 Diversification in Plants. Pp. 305–327 in P. Pontarotti, ed. *Evolutionary Biology:*
802 *Biodiversification from Genotype to Phenotype*. Springer International Publishing,
803 Cham.
- 804 Holt, R. D., and M. B. Bonsall. 2017. Apparent Competition. *Annual Review of Ecology,*
805 *Evolution, and Systematics* 48:447–471.
- 806 Hörandl, E. 2009. Geographical Parthenogenesis: Opportunities for Asexuality. Pp. 161–186 in I.
807 Schön, K. Martens, and P. Dijk, eds. *Lost Sex: The Evolutionary Biology of*
808 *Parthenogenesis*. Springer Netherlands, Dordrecht.
- 809 Hörandl, E., and D. Hojsgaard. 2012. The evolution of apomixis in angiosperms: A reappraisal.
810 *Plant Biosystems* 146:681–693.

811 Hörandl, E., and O. Paun. 2007. Patterns and sources of genetic diversity in apomictic plants:
812 implications for evolutionary potentials. Pp. 169–194 *in* Apomixis: Evolution,
813 Mechanisms, and Perspectives. Koenigstein, Germany.

814 Hotz, H., P. Beerli, and C. Spolsky. 1992. Mitochondrial DNA reveals formation of nonhybrid
815 frogs by natural matings between hemiclinal hybrids. *Mol. Biol. Evol.* 9:610–620.

816 Hotz, H., G. Mancino, S. Bucciinnocenti, M. Ragghianti, L. Berger, and T. Uzzell. 1985. *Rana*
817 *ridibunda* varies geographically in inducing clonal gametogenesis in interspecies hybrids.
818 *Journal of Experimental Zoology* 236:199–210.

819 Hubbs, C. 1964. Interactions between a bisexual fish species and its gynogenetic sexual parasite.
820 *The Bulletin of the Texas Memorial Museum* 8:1–72.

821 Husband, B. C. 2000. Constraints on polyploid evolution: a test of the minority cytotype
822 exclusion principle. *Proc. Biol. Sci.* 267:217–223.

823 Jablonska, O., D. Juchno, A. Leska, K. Kowalewska, and A. Boroń. 2020. The variable presence
824 of apoptosis in the testes of diploid and sterile allotetraploid *Cobitis* (Teleostei,
825 Cobitidae) males during reproductive cycle. *Journal of Experimental Biology*
826 223:jeb212050.

827 Jalinsky, J., J. M. Logsdon, and M. Neiman. 2020. Male phenotypes in a female framework:
828 Evidence for degeneration in sperm produced by male snails from asexual lineages.
829 *Journal of Evolutionary Biology* 33:1050–1059.

830 Janko, K., M. A. Culling, P. Rab, and P. Kotlik. 2005. Ice age cloning-comparison of the
831 Quaternary evolutionary histories of sexual and clonal forms of spiny loaches (*Cobitis*;
832 Teleostei) using the analysis of mitochondrial DNA variation. *Molecular Ecology*
833 14:2991–3004.

834 Janko, K., P. Drozd, and J. Eisner. 2011. Do clones degenerate over time? Explaining the genetic
835 variability of asexuals through population genetic models. *Biology Direct* 6:17.

- 836 Janko, K., and J. Eisner. 2009. Sperm-dependent parthenogens delay the spatial expansion of
837 their sexual hosts. *Journal of Theoretical Biology* 261:431–440.
- 838 Janko, K., J. Eisner, and P. Mikulíček. 2019. Sperm-dependent asexual hybrids determine
839 competition among sexual species. *Scientific Reports* 9:722.
- 840 Janko, K., M. Flajšhans, L. Choleva, J. Bohlen, V. Šlechtová, M. Rábová, Z. Lajbner, V. Šlechta,
841 P. Ivanova, I. Dobrovolov, and others. 2007. Diversity of European spined loaches (genus
842 *Cobitis* L.): an update of the geographic distribution of the *Cobitis taenia* hybrid complex
843 with a description of new molecular tools for species and hybrid determination. *Journal*
844 *of Fish Biology* 71:387–408.
- 845 Janko, K., J. Pačes, H. Wilkinson-Herbots, R. J. Costa, J. Roslein, P. Drozd, N. Iakovenko, J.
846 Rídl, M. Hroudová, J. Kočí, R. Reifová, V. Šlechtová, and L. Choleva. 2018. Hybrid
847 asexuality as a primary postzygotic barrier between nascent species: On the
848 interconnection between asexuality, hybridization and speciation. *Mol. Ecol.* 249–263.
- 849 Juchno, D., and A. Boroń. 2006. Comparative histology of the testes of the spined loach *Cobitis*
850 *taenia* L. and natural allotetraploids of *Cobitis* (Pisces, Cobitidae). *Hydrobiologia*
851 573:45–53.
- 852 Karunarathne, P., M. Schedler, E. J. Martínez, A. I. Honfi, A. Novichkova, and D. Hojsgaard.
853 2018. Intraspecific ecological niche divergence and reproductive shifts foster cytotype
854 displacement and provide ecological opportunity to polyploids. *Ann. Bot.* 121:1183–
855 1196.
- 856 Kearney, M. 2005. Hybridization, glaciation and geographical parthenogenesis. *Trends Ecol.*
857 *Evol.* 20:495–502.
- 858 Kirchheimer, B., J. Wessely, A. Gatringer, K. Hülber, D. Moser, C. C. F. Schinkel, M.
859 Appelhans, S. Klatt, M. Caccianiga, A. Dellinger, A. Guisan, M. Kuttner, J. Lenoir, L.

860 Maiorano, D. Nieto-Lugilde, C. Plutzer, J.-C. Svenning, W. Willner, E. Hörandl, and S.
861 Dullinger. 2018. Reconstructing geographical parthenogenesis: effects of niche
862 differentiation and reproductive mode on Holocene range expansion of an alpine plant.
863 Ecology Letters 21:392–401.

864 Kočí, J., J. Röslein, J. Pačes, J. Kotusz, K. Halačka, J. Koščo, J. Fedorčák, N. Iakovenko, and K.
865 Janko. 2020. No evidence for accumulation of deleterious mutations and fitness
866 degradation in clonal fish hybrids: Abandoning sex without regrets. Molecular Ecology
867 29:3038–3055.

868 Kokko, H., K. U. Heubel, and D. J. Rankin. 2008. How populations persist when asexuality
869 requires sex: the spatial dynamics of coping with sperm parasites. Proc. Biol. Sci.
870 275:817–825.

871 Kooi, C. J. van der, and T. Schwander. 2014. On the fate of sexual traits under asexuality.
872 Biological reviews of the Cambridge Philosophical Society 89:805–819.

873 Krahulcová, A., J. Chrtek, and F. Krahulec. 1999. Autogamy in *Hieracium* Subgen. *Pilosella*.
874 Folia Geobotanica 34:373–376.

875 Kwan, Y.-S., M.-H. Ko, Y.-S. Jeon, H.-J. Kim, and Y.-J. Won. 2019. Bidirectional mitochondrial
876 introgression between Korean cobitid fish mediated by hybridogenetic hybrids. Ecology
877 and Evolution 9:1244–1254.

878 Lacy, R. C. 1987. Loss of Genetic Diversity from Managed Populations: Interacting Effects of
879 Drift, Mutation, Immigration, Selection, and Population Subdivision. Conservation
880 Biology 1:143–158.

881 Lampert, K. P., D. K. Lamatsch, P. Fischer, J. T. Epplen, I. Nanda, M. Schmid, and M. Schartl.
882 2007. Automictic Reproduction in Interspecific Hybrids of Poeciliid Fish. Current
883 Biology 17:1948–1953.

- 884 Lampert, K. p, and M. Schartl. 2008. The origin and evolution of a unisexual hybrid: *Poecilia*
885 *formosa*. Philosophical Transactions of the Royal Society B: Biological Sciences
886 363:2901–2909.
- 887 Lande, R. 1976. Natural Selection and Random Genetic Drift in Phenotypic Evolution. *Evolution*
888 30:314–334.
- 889 Laskowski, K. L., C. Doran, D. Bierbach, J. Krause, and M. Wolf. 2019. Naturally clonal
890 vertebrates are an untapped resource in ecology and evolution research. *Nat. Ecol. Evol.*
891 3:161–169.
- 892 Lehtonen, J., D. J. Schmidt, K. Heubel, and H. Kokko. 2013. Evolutionary and ecological
893 implications of sexual parasitism. *Trends Ecol. Evol.* 28:297–306.
- 894 Lievens, E. J. P., G. J. B. Henriques, Y. Michalakis, and T. Lenormand. 2016. Maladaptive Sex
895 Ratio Adjustment in the Invasive Brine Shrimp *Artemia franciscana*. *Current Biology*
896 26:1463–1467.
- 897 Lively, C. M., and L. T. Morran. 2014. The ecology of sexual reproduction. *J. Evol. Biol.*
898 27:1292–1303.
- 899 Lowe, W. H., and F. W. Allendorf. 2010. What can genetics tell us about population
900 connectivity? *Molecular Ecology* 19:3038–3051.
- 901 Lundmark, M. 2006. Polyploidization, hybridization and geographical parthenogenesis. *Trends*
902 *Ecol. Evol.* 21:9.
- 903 Majeský, L., R. J. Vašut, M. Kitner, and B. Trávníček. 2012. The Pattern of Genetic Variability
904 in Apomictic Clones of *Taraxacum officinale* Indicates the Alternation of Asexual and
905 Sexual Histories of Apomicts. *PLOS ONE* 7:e41868.
- 906 Makowicz, A. M., and I. Schlupp. 2015. Effects of Female-Female Aggression in a
907 Sexual/Unisexual Species Complex. *Ethology* 121:903–914.

908 Marshall, J. L., M. L. Arnold, and D. J. Howard. 2002. Reinforcement: the road not taken.
909 Trends in Ecology & Evolution 17:558–563.

910 Matute, D. R., I. A. Butler, D. A. Turissini, and J. A. Coyne. 2010. A test of the snowball theory
911 for the rate of evolution of hybrid incompatibilities. Science 329:1518–1521.

912 Mee, J. A., and S. P. Otto. 2010. Variation in the Strength of Male Mate Choice Allows Long-
913 Term Coexistence of Sperm-Dependent Asexuals and Their Sexual Hosts. Evolution
914 64:2808–2819.

915 Meirmans, S. 2009. The Evolution of the Problem of Sex. Pp. 21–46 in I. Schön, K. Martens, and
916 P. Dijk, eds. Lost Sex: The Evolutionary Biology of Parthenogenesis. Springer
917 Netherlands, Dordrecht.

918 Mikulíček, P., M. Kautman, B. Demovič, and K. Janko. 2014. When a clonal genome finds its
919 way back to a sexual species: evidence from ongoing but rare introgression in the
920 hybridogenetic water frog complex. J. Evol. Biol. 27:628–642.

921 Mikulíček, P., M. Kautman, J. Kautman, and N. B. M. Pruvost. 2015. Mode of hybridogenesis
922 and habitat preferences influence population composition of water frogs (*Pelophylax*
923 *esculentus* complex, Anura: Ranidae) in a region of sympatric occurrence (western
924 Slovakia). Journal of Zoological Systematics and Evolutionary Research 53:124–132.

925 Morgado-Santos, M., H. M. Pereira, L. Vicente, and M. J. Collares-Pereira. 2015. Mate Choice
926 Drives Evolutionary Stability in a Hybrid Complex. PLoS ONE 10:e0132760.

927 Moritz, C., W. M. Brown, L. D. Densmore, J. W. Wright, D. Vyas, S. Donnellan, M. Adams, and
928 P. Baverstock. 1989. Genetic diversity and the dynamics of hybrid parthenogenesis in
929 *Cnemidophorus* (Teiidae) and *Heteronotia* (Gekkonidae). Pp. 268–280 in Dawley RM,
930 Bogart JP, editors. Evolution and Ecology of Unisexual Vertebrates. New York State
931 Museum, Albany, New York.

932 Neaves, W. B., and P. Baumann. 2011. Unisexual reproduction among vertebrates. Trends in
933 Genetics 27:81–88.

- 934 Neiman, M., and T. A. Linksvayer. 2006. The conversion of variance and the evolutionary
935 potential of restricted recombination. *Heredity* 96:111–121.
- 936 Neiman, M., T. F. Sharbel, and T. Schwander. 2014. Genetic causes of transitions from sexual
937 reproduction to asexuality in plants and animals. *J. Evol. Biol.* 27:1346–1359.
- 938 Ortiz-Barrientos, D., J. Engelstädter, and L. H. Rieseberg. 2016. Recombination Rate Evolution
939 and the Origin of Species. *Trends Ecol. Evol.* 31:226–236.
- 940 Pagano, A., T. Lodé, and P.-A. Crochet. 2001. New contact zone and assemblages among water
941 frog of Southern France. *Journal of Zoological Systematics and Evolutionary Research*
942 39:63–67.
- 943 Paland, S., J. K. Colbourne, and M. Lynch. 2005. Evolutionary history of contagious asexuality
944 in *Daphnia pulex*. *Evolution* 59:800–813.
- 945 Pantel, J. H., T. E. Juenger, and M. A. Leibold. 2011. Environmental gradients structure *Daphnia*
946 *pulex* x *pulicaria* clonal distribution. *Journal of Evolutionary Biology* 24:723–732.
- 947 Parker, D. J., J. Bast, K. Jalvingh, Z. Dumas, M. Robinson-Rechavi, and T. Schwander. 2019.
948 Repeated Evolution of Asexuality Involves Convergent Gene Expression Changes. *Mol.*
949 *Biol. Evol.* 36:350–364.
- 950 Peck, J. R., J. M. Yearsley, and D. Waxman. 1998. Explaining the geographic distributions of
951 sexual and asexual populations. *Nature* 391:889–892.
- 952 Pellino, M., D. Hojsgaard, T. Schmutzer, U. Scholz, E. Hörandl, H. Vogel, and T. F. Sharbel.
953 2013. Asexual genome evolution in the apomictic *Ranunculus auricomus* complex:
954 examining the effects of hybridization and mutation accumulation. *Mol. Ecol.* 22:5908–
955 5921.
- 956 Pigneur, L.-M., S. M. Hedtke, E. Etoundi, and K. Van Doninck. 2012. Androgenesis: a review
957 through the study of the selfish shellfish *Corbicula* spp. *Heredity* 108:581–591.

958 Pigneur, L.-M., J. Marescaux, K. Roland, E. Etoundi, J.-P. Descy, and K. Van Doninck. 2011.
959 Phylogeny and androgenesis in the invasive *Corbicula* clams (Bivalvia, Corbiculidae) in
960 Western Europe. *BMC Evolutionary Biology* 11:147.

961 Plötner, J., T. Uzzell, P. Beerli, C. Spolsky, T. Ohst, S. N. Litvinchuk, G.-D. Guex, H.-U. Reyer,
962 and H. Hotz. 2008. Widespread unidirectional transfer of mitochondrial DNA: a case in
963 western Palaearctic water frogs. *Journal of Evolutionary Biology* 21:668–681.

964 Prodöhl, P. A., W. J. Loughry, C. M. McDonough, W. S. Nelson, and J. C. Avise. 1996.
965 Molecular documentation of polyembryony and the micro-spatial dispersion of clonal
966 sibships in the nine-banded armadillo, *Dasypus novemcinctus*. *Proc. Biol. Sci.* 263:1643–
967 1649.

968 Pruvost, N. B. M., P. Mikulíček, L. Choleva, and H.-U. Reyer. 2015. Contrasting reproductive
969 strategies of triploid hybrid males in vertebrate mating systems. *J. Evol. Biol.* 28:189–
970 204.

971 Redfield, R. J. 1994. Male mutation rates and the cost of sex for females. *Nature* 369:145–147.

972 Rieseberg, L. H., and J. H. Willis. 2007. Plant speciation. *Science* 317:910–914.

973 Ross, L., N. B. Hardy, A. Okusu, and B. B. Normark. 2012. Large population size predicts the
974 distribution of asexuality in scale insects. *Evolution* 67:196–206.

975 Russell, S. 2003. Evolution of intrinsic post-zygotic reproductive isolation in fish. *Ann. Zool.*
976 *Fennici* 321–329.

977 Sailer, C., J. Stöcklin, and U. Grossniklaus. 2020. Dynamics of apomictic and sexual
978 reproduction during primary succession on a glacier forefield in the Swiss Alps.
979 *Scientific Reports* 10:8269.

980 Scali, V. 2009. Metasexual Stick Insects: Model Pathways to Losing Sex and Bringing It Back.
981 Pp. 317–345 in I. Schön, K. Martens, and P. Dijk, eds. *Lost Sex: The Evolutionary*
982 *Biology of Parthenogenesis*. Springer Netherlands, Dordrecht.

- 983 Schartl, M., I. Nanda, I. Schlupp, B. Wilde, J. T. Epplen, M. Schmid, and J. Parzefall. 1995.
984 Incorporation of subgenomic amounts of DNA as compensation for mutational load in a
985 gynogenetic fish. *Nature* 373:68–71.
- 986 Schartl, M., I. Schlupp, A. Schartl, M. K. Meyer, I. Nanda, M. Schmid, J. T. Epplen, and J.
987 Parzefall. 1991. On the stability of dispensable constituents of the eukaryotic genome:
988 stability of coding sequences versus truly hypervariable sequences in a clonal vertebrate,
989 the amazon molly, *Poecilia formosa*. *PNAS* 88:8759–8763.
- 990 Schlupp, I. 2009. Chapter 5 Behavior of Fishes in the Sexual/Unisexual Mating System of the
991 Amazon Molly (*Poecilia formosa*). Pp. 153–183 in *Advances in the Study of Behavior*.
992 Academic Press.
- 993 Schlupp, I. 2021. *Male Choice, Female Competition, and Female Ornamentation in Sexual*
994 *Selection*. Oxford University Press.
- 995 Schlupp, I. 2010. Mate Choice and the Amazon Molly: How Sexuality and Unisexuality Can
996 Coexist. *Journal of Heredity* 101:55–61.
- 997 Schlupp, I. 2005. The evolutionary ecology of gynogenesis. *Annual Review of Ecology,*
998 *Evolution, and Systematics* 36:399–417.
- 999 Schlupp, I., I. Nanda, M. Döbler, D. K. Lamatsch, J. T. Epplen, J. Parzefall, M. Schmid, and M.
1000 Schartl. 1998. Dispensable and indispensable genes in an ameiotic fish, the Amazon
1001 molly *Poecilia formosa*. *Cytogenet. Cell Genet.* 80:193–198.
- 1002 Schlupp, I., J. Parzefall, J. T. Epplen, I. Nanda, M. Schmid, and M. Schartl. 1992. Pseudomale
1003 Behaviour and Spontaneous Masculinization in the All-Female Teleost *Poecilia formosa*
1004 (Teleostei: Poeciliidae). *Behaviour* 122:88–104.
- 1005 Schlupp, I., and M. Plath. 2005. Male Mate Choice and Sperm Allocation in a Sexual/Asexual
1006 Mating Complex of *Poecilia* (Poeciliidae, Teleostei). *Biol. Lett.* 1:169–171.

- 1007 Schlupp, I., and R. Riesch. 2011. Evolution of unisexual reproduction. pp. 50–58 in Evans J,
1008 Pilastro A, Schlupp I, editors. Ecology and Evolution of Poeciliid Fishes. University of
1009 Chicago Press, Chicago.
- 1010 Schmeller, D. S., A. Seitz, A. Crivelli, and M. Veith. 2005. Crossing species' range borders:
1011 interspecies gene exchange mediated by hybridogenesis. *Proc. Biol. Sci.* 272:1625–1631.
- 1012 Schön, I., R. L. Pinto, S. Halse, A. J. Smith, K. Martens, and C. W. Birky. 2012. Cryptic species
1013 in putative ancient asexual darwinulids (Crustacea, Ostracoda). *PLoS ONE* 7:e39844.
- 1014 Schultz, R. J. 1973. Unisexual fish: laboratory synthesis of a “species.” *Science* 179:180–181.
- 1015 Seehausen, O., R. K. Butlin, I. Keller, C. E. Wagner, J. W. Boughman, P. A. Hohenlohe, C. L.
1016 Peichel, G.-P. Saetre, C. Bank, Å. Brännström, A. Brelsford, C. S. Clarkson, F.
1017 Eroukhanoff, J. L. Feder, M. C. Fischer, A. D. Foote, P. Franchini, C. D. Jiggins, F. C.
1018 Jones, A. K. Lindholm, K. Lucek, M. E. Maan, D. A. Marques, S. H. Martin, B.
1019 Matthews, J. I. Meier, M. Möst, M. W. Nachman, E. Nonaka, D. J. Rennison, J.
1020 Schwarzer, E. T. Watson, A. M. Westram, and A. Widmer. 2014. Genomics and the
1021 origin of species. *Nat. Rev. Genet.* 15:176–192.
- 1022 Shcherbakov, V. P. 2010. Biological species is the only possible form of existence for higher
1023 organisms: the evolutionary meaning of sexual reproduction. *Biol. Direct* 5:14.
- 1024 Slyn'ko, Y. V. 2000. Sistema razmnozheniya mezhrodovyykh gibridov plotvy (*Rutilus rutilus* L.),
1025 leshcha (*Abramis brama* L.) i sinca (*Abramis ballerus* L.) (Leuciscinae: Cyprinidae). PhD
1026 Thesis, Available from: [http://www.dissercat.com/content/sistema-razmnozheniya-
1027 mezhrodovyykh-gibridov-plotvy-rutilus-rutilus-l-leshcha-abramis-brama-l](http://www.dissercat.com/content/sistema-razmnozheniya-mezhrodovyykh-gibridov-plotvy-rutilus-rutilus-l-leshcha-abramis-brama-l).
- 1028 Sochor, M., P. Šarhanová, S. Pfanzelt, and B. Trávníček. 2017. Is evolution of apomicts driven
1029 by the phylogeography of the sexual ancestor? Insights from European and Caucasian
1030 brambles (*Rubus*, Rosaceae). *Journal of Biogeography* 44:2717–2728.
- 1031 Sousa-Santos, C., M. J. Collares-Pereira, and V. C. Almada. 2006. Evidence of extensive
1032 mitochondrial introgression with nearly complete substitution of the typical *Squalius*

- 1033 *pyrenaicus*-like mtDNA of the *Squalius alburnoides* complex (Cyprinidae) in an
1034 independent Iberian drainage. *Journal of Fish Biology* 68:292–301.
- 1035 Stenberg, P., and A. Saura. 2009. Cytology of Asexual Animals. Pp. 63–74 in I. Schön, K.
1036 Martens, and P. Dijk, eds. *Lost Sex: The Evolutionary Biology of Parthenogenesis*.
1037 Springer Netherlands, Dordrecht.
- 1038 Stenberg, P., and A. Saura. 2013. Meiosis and Its Deviations in Polyploid Animals. *Cytogenetic*
1039 *and genome research* 140.
- 1040 Stöck, M., D. Dedukh, R. Reifová, D. K. Lamatsch, Z. Starostová, and K. Janko. 2021. Sex
1041 chromosomes in meiotic, hemiclonal, clonal and polyploid hybrid vertebrates: along the
1042 ‘extended speciation continuum.’ *Philosophical Transactions of the Royal Society B:*
1043 *Biological Sciences* 376:20200103.
- 1044 Stöck, M., D. K. Lamatsch, C. Steinlein, J. T. Epplen, W.-R. Grosse, R. Hock, T. Klapperstück,
1045 K. P. Lampert, U. Scheer, M. Schmid, and M. Schartl. 2002. A bisexually reproducing
1046 all-triploid vertebrate. *Nat. Genet.* 30:325–328.
- 1047 Stöck, M., K. P. Lampert, D. Möller, I. Schlupp, and M. Schartl. 2010. Monophyletic origin of
1048 multiple clonal lineages in an asexual fish (*Poecilia formosa*). *Molecular Ecology*
1049 19:5204–5215.
- 1050 Suomalainen, E., A. Saura, and J. Lokki. 1987. *Cytology and evolution in parthenogenesis*. Boca
1051 Raton, Florida, CRC Press.
- 1052 Suzuki, S., S. Miyake, K. Arai, and H. Munehara. 2019. Unisexual hybrids break through an
1053 evolutionary dead end by two-way backcrossing. *Evolution* 74:392–403.
- 1054 Thomas, F., J. Guégan, Y. Michalakis, and F. Renaud. 2000. Parasites and host life-history traits:
1055 implications for community ecology and species co-existence. *Int. J. Parasitol.* 30:669–
1056 674.

1057 Thompson, M. J., and C. D. Jiggins. 2014. Supergenes and their role in evolution. *Heredity*
1058 113:1–8.

1059 Tilquin, A., and H. Kokko. 2016. What does the geography of parthenogenesis teach us about
1060 sex? *Philosophical Transactions of the Royal Society B: Biological Sciences*
1061 371:20150538.

1062 Uzzell, T., R. Günther, and L. Berger. 1976. *Rana ridibunda* and *Rana esculenta*: A Leaky
1063 Hybridogenetic System (Amphibia Salientia). *Proceedings of the Academy of Natural*
1064 *Sciences of Philadelphia* 128:147–171.

1065 Van der Kooi, C. J., C. Matthey-Doret, and T. Schwander. 2017. Evolution and comparative
1066 ecology of parthenogenesis in haplodiploid arthropods. *Evolution Letters* 1:304–316.

1067 Van Valen, L. 1973. A new evolutionary law. *Evolutionary Theory* 1973:1–30.

1068 Vorburger, C. 2001. Non-hybrid offspring from matings between hemiclonal hybrid waterfrogs
1069 suggest occasional recombination between clonal genomes. *Ecology Letters* 4:628–636.

1070 Vrijenhoek, R. C. 1994. Unisexual Fish: Model Systems For Studying Ecology And Evolution.
1071 *Annual Review of Ecology and Systematics* 25:71–96.

1072 Vrijenhoek, R. C., and E. D. Parker. 2009. Geographical Parthenogenesis: General Purpose
1073 Genotypes and Frozen Niche Variation. Pp. 99–131 *in* I. Schön, K. Martens, and P. Dijk,
1074 eds. *Lost Sex: The Evolutionary Biology of Parthenogenesis*. Springer Netherlands,
1075 Dordrecht.

1076 Warren, W. C., R. García-Pérez, S. Xu, K. P. Lampert, D. Chalopin, M. Stöck, L. Loewe, Y. Lu,
1077 L. Kuderna, P. Minx, M. J. Montague, C. Tomlinson, L. W. Hillier, D. N. Murphy, J.
1078 Wang, Z. Wang, C. M. Garcia, G. C. W. Thomas, J.-N. Volff, F. Farias, B. Aken, R. B.
1079 Walter, K. D. Pruitt, T. Marques-Bonet, M. W. Hahn, S. Kneitz, M. Lynch, and M.
1080 Schartl. 2018. Clonal polymorphism and high heterozygosity in the celibate genome of
1081 the Amazon molly. *Nat. Ecol. Evol.* 2:669–679.

- 1082 Wetherington, J. D., K. E. Kitora, and R. C. Vrijenhoek. 1987. A test of the spontaneous
1083 heterosis hypothesis for unisexual vertebrates. *Evolution* 41:721–731.
1084

1085 **Glossary**

1086 **Allee effect:** a phenomenon describing a positive correlation between population size or density
1087 and the mean individual fitness of a population. A higher mean population fitness in more abundant
1088 or dense populations may be associated with better mate finding, better cooperation among
1089 individuals, greater ability to change the environment in favor of the species or with a lower rate
1090 of inbreeding and higher genetic variability.

1091 **Apparent competition:** a form of mostly negative indirect interactions between species that arise
1092 because they share a natural enemy (predators, parasites, pathogens or herbivores). When such an
1093 indirect competition is driven by a shared parasite, it is called **parasite-mediated competition**.

1094 **Asexual reproduction:** reproductive mode in which an organism passes on its genome (or parts
1095 of it) clonally as a result of vegetative reproduction, polyembryony or by circumventing
1096 recombination during gametogenesis. The latter is gametic asexuality.

1097 **Clonal:** this makes reference to a mode of inheritance where all or most of the genome is passed
1098 on unaltered.

1099 **Cybrids:** hybrids containing a nucleus of one species and cytoplasm of another species.

1100 **Effective population size (N_e):** reflects the rate at which genetic diversity will be lost following
1101 genetic drift, and this rate is inversely proportional to a population's N_e . N_e is reduced by unequal
1102 sex ratio, variation in reproductive success, and by the fluctuation of the population size in time.

1103 **Extended speciation continuum:** a conceptual frame work linking the formation of asexual
1104 reproduction in hybrids with the classical speciation continuum assuming gradual formation of
1105 postzygotic reproductive barriers among diverging taxa. It posits that before accumulating genetic
1106 incompatibilities between hybridizing species cause complete sterility or even inviability, they may
1107 occasionally distort hybrid's gametogenesis towards production of unreduced gametes, thereby
1108 sometimes alleviating problems in chromosomal pairing and rescuing hybrid's fertility,
1109 simultaneously triggering its (hemi-) clonal reproduction.

1110 **Geographical parthenogenesis:** a phenomenon describing that asexual organism occupy larger
1111 ranges, occur at higher elevations or at higher latitudes, and colonize more frequently previously
1112 glaciated or otherwise disturbed areas than their sexual relatives.

1113 **Gynogenesis:** females produce typically diploid eggs, which are pseudo-fertilized by sperm of
1114 males from a different species. The sperm genome is typically not incorporated, and inheritance is
1115 maternal.

1116 **Hybridogenesis:** females produce haploid or diploid eggs, which are usually fertilized by males
1117 from a different species. The sperm genome is incorporated and expressed but excluded from the
1118 germ line during gametogenesis. The female genome is thus inherited clonally.

1119 **Mixis:** well defined haploid and diploid phases that alternate.

1120 **Muller's ratchet:** a process of the irreversible accumulation of deleterious mutations in a clonal
1121 genome because of the absence of recombination.

1122 **Red Queen Hypothesis:** making reference to the red queen character in Lewis Carrolls' 1871
1123 novel "Through the Looking Glass", the hypothesis posits that constant genetic change is needed
1124 to stay evolutionary alive under the continuous pressure from parasites.

1125 **Sexual reproduction:** prevailing mode of reproduction in metazoans, characterized by production
1126 of offspring via syngamy of meiotically produced gametes. Recombination and segregation of
1127 chromosomes (alleles) during meiosis result in genetically variable offspring.

1128 **Sperm-dependent parthenogenesis:** a form of clonal inheritance where eggs need to interact with
1129 sperm, either in gynogenesis or hybridogenesis. This is also called pseudogamy or sexual
1130 parasitism.

1131 **Figure 1**

1132 Modes of asexual reproduction, A, B, E) mechanisms of nuclear (C, F) and mitochondrial (G, H,
1133 I) introgression and polyploid speciation (D). Since asexual reproduction might be linked with
1134 interspecific hybridization, asexual forms in this scheme are hybrids (red and blue genomes
1135 originate from the parental species). **(A)** A parthenogenetic hybrid female (e.g. in lizards of the
1136 genus *Darevskia*) forms unreduced eggs from which a new generation of clonal daughters originate
1137 without any contribution of males. **(B)** Gynogenesis (e.g. in the genus *Cobitis*) is a similar mode
1138 of reproduction during which unreduced eggs must be activated by sperm of a sexual male. The
1139 sperm, however, don't fertilize the eggs and a new generation of daughters is clonal. **(C)** In
1140 gynogenetic systems (e.g., in the fish *Poecilia formosa*), microchromosomes (B chromosomes)
1141 might be incorporated to the clonal egg from the paternal sperm. **(D)** Incorporation of sperm to
1142 clonal triploid eggs (observed in the *Squalius alburnoides* complex) can lead to sexual tetraploid
1143 progeny, which is reproductively isolated from other ploidy forms. **(E)** A hybridogenetic hybrid
1144 female (e.g. in the *Pelophylax esculentus* complex) eliminates a genome of one parental species
1145 during gametogenesis and forms clonal eggs which are fertilized by sperm of a sexual male. **(F)**
1146 In triploid forms (e.g. in hybridogenetic *P. esculentus*) two different genomes belonging to one
1147 parental species (red ones in the scheme) can enter meiosis and recombine after premeiotic
1148 elimination of the genome of another parental species (blue genome). This process is called meiotic
1149 hybridogenesis. Here mating between a diploid hybrid female producing both diploid and haploid
1150 eggs and a triploid male is shown. This mating leads to diploid and triploid progeny of both sexes
1151 (in the scheme a male is triploid and a female is diploid just for simplicity) and enables the
1152 perpetuation of a population without the contribution of the parental species. **(G)** Mating between
1153 hybridogenetic hybrids *P. esculentus* and parental species leading to the origin of the parental
1154 species progeny with introgressed mtDNA. **(H)** In androgenetic *Corbicula* clams, the egg is
1155 fertilized by an unreduced biflagellate sperm. The entire maternal nuclear genome is then extruded
1156 from the oocyte, whereas mitochondria and other organelles from the egg are retained. Thus, the
1157 offspring inherit paternal nuclear genome and maternal mtDNA. **(I)** In the *Bacillus* stick insect,
1158 the whole hybrid genome is lost, two male pronuclei may fuse (since phasmid eggs are
1159 physiologically polyspermic) and produce a sexually reproducing progeny of one parental species
1160 with introgressed mitogenome.

1161 Please note that we deliberately avoided reproductive schemes for plants in Fig. 1 since in
1162 apomictic plants (e.g. in *Hieracium*), asexual reproduction is a more complex process due to a
1163 switch from double fertilization (need for endosperm formation) in sexual species to diplospory,
1164 apospory or haploid parthenogenesis in asexuals.

1165

1166 Figure 2

1167 The effect of coexisting sperm-dependent parthenogen on a sexual hosts effective population size
1168 and related phenomena: Compared to a purely sexual population (A), when a sexual population of
1169 the same census size coexists with sperm-dependent parthenogens (B), it suffers from increased
1170 variance in reproductive success since relatively fewer males have access to conspecific sexual
1171 females and ‘waste’ their reproductive effort on asexual females. This negatively affects the long-
1172 term effective size of sexual population. Such intensification of genetic drift also leads to faster
1173 population differentiation.

1174 Effect on operational sex ratio (OSR) and mate-choice: Because males are in a minority when
1175 coexisting with a sperm-dependent all-female population, it changes the OSR. Simultaneously,
1176 given that mating with sperm-dependent females effectively ‘wastes’ males’ reproductive effort,
1177 an intensified selection for mating preferences with conspecific is expected.

1178

1179 Figure 3

1180 The classical scenario of postzygotic reproductive isolation assumes that as nascent species
1181 diverge (stage 1 in the scheme), their genomes (denoted as bicolored chromosomes) become
1182 progressively less compatible and interspecies barriers become more pronounced (as denoted by
1183 dotted to solid lines) until a stage is reached when species cannot produce fertile or viable hybrids
1184 and speciation is complete (stage 3 in the scheme). However, the ability to produce asexually
1185 reproducing hybrids probably also scales with genetic divergence between hybridizing taxa
1186 (Moritz et al. 1989). Empirical data (Janko et al. 2018) suggest that such a phase (stage 2 in our
1187 scheme) may occur generally at earlier stages of species differentiation before complete hybrid
1188 sterility occurs (stage 3). If so, asexual hybrids, although as fertile as “classical” sexual hybrids
1189 produced at early stages (stage 1 in the scheme), may turn into an effective barrier to interspecific
1190 gene flow due to their general inability to backcross into either parental species.

1191

1192 Figure 4

1193 When sperm-dependent parthenogens coexist with a sexual population (B), they tend to decrease
1194 its effective population size and therefore hamper the population growth rate. Expansions of
1195 sexuals to a new environment thus occur at lower frequency as compared to purely sexual
1196 populations (A) and therefore they have a lower probability to find a proper mate to start a new

1197 generation of colonists. This ultimately decreases the growth rate and expansion rate of entire
1198 populations (Janko and Eisner 2009).

1199

1200 **Figure 5**

1201 Compared to purely sexual competing species (A), when a stronger competitor (blue) outcompetes
1202 the weaker one (red), sperm dependent parthenogens may turn the result of interspecific
1203 competition (B). This occurs when the stronger sexual competitor invests more into mating with
1204 sperm dependent parthenogens (or is otherwise more vulnerable to their presence) than the weaker
1205 competitor, whose population growth is therefore faster and may ultimately outcompete the
1206 stronger one (Janko et al. 2019).

1207 **Supplementary Material**

1208 **Figure S1**

1209 Phylogenetic relationships among metazoans showing lineages (phyla) where parthenogenesis
1210 secondarily evolved (red asterisk). A clade named Nematoda involves also the phylum
1211 Nematomorpha in which parthenogenetic species were also discovered. The tree was taken from
1212 the study of Laumer et al. (2019; Figure 2a). Information about parthenogenetic reproduction of
1213 particular taxa comes from works of Hummon (1984), Roe (1986), Read (1988), Lively and
1214 Johnson (1994), Bertolani (2001), Schön et al. (2009), Hanelt et al. (2012) and Schwarz (2017).

1215

1216 **References in Supplementary Material**

1217 Bertolani R. 2001. Evolution of the Reproductive Mechanisms in Tardigrades — A Review.

1218 *Zoologischer Anzeiger - A Journal of Comparative Zoology* **240**:247–252.

1219 Hanelt B, Bolek MG, Schmidt-Rhaesa A. 2012. Going Solo: Discovery of the First
1220 Parthenogenetic Gordiid (Nematomorpha: Gordiida). *PLoS ONE* **7**:e34472.

1221 Hummon MR. 1984. Reproduction and sexual development in a freshwater gastrotrich. 1.
1222 Oogenesis of parthenogenic eggs (Gastrotricha). *Zoomorphology* **104**:33–41.

1223 Laumer CE, Fernandez R, Lemer S, Combosch D, Kocot KM, Riesgo A, Andrade SCS, Sterrer
1224 W, Sorensen MV, Giribet G. 2019. Revisiting metazoan phylogeny with genomic sampling
1225 of all phyla. *Proc. R. Soc. Lond. B* **286**:20190831.

1226 Lively CM, Johnson SG. 1994. Brooding and the Evolution of Parthenogenesis: Strategy Models
1227 and Evidence from Aquatic Invertebrates. *Proc. R. Soc. Lond. B* **256**:89–95.

1228 Read VMStJ. 1988. The Onychophora of Trinidad, Tobago and the Lesser Antilles. *Zoological*
1229 *Journal of the Linnean Society* **93**: 225–257.

1230 Roe P. 1986. Parthenogenesis in *Carcinonemertes* spp. (Nemertea: Hoplonemertea). *Biological*
1231 *Bulletin* **171**:640–646.

1232 Schwarz EM. 2017. Evolution: A Parthenogenetic Nematode Shows How Animals Become
1233 Sexless. *Current Biology* **27**: R1064–R1066.

1234 Schön I, Martens K, Dijk P, editors. 2009. *Lost Sex: The Evolutionary Biology of Parthenogenesis*.
1235 Dordrecht: Springer Netherlands.

1236

Figure 1

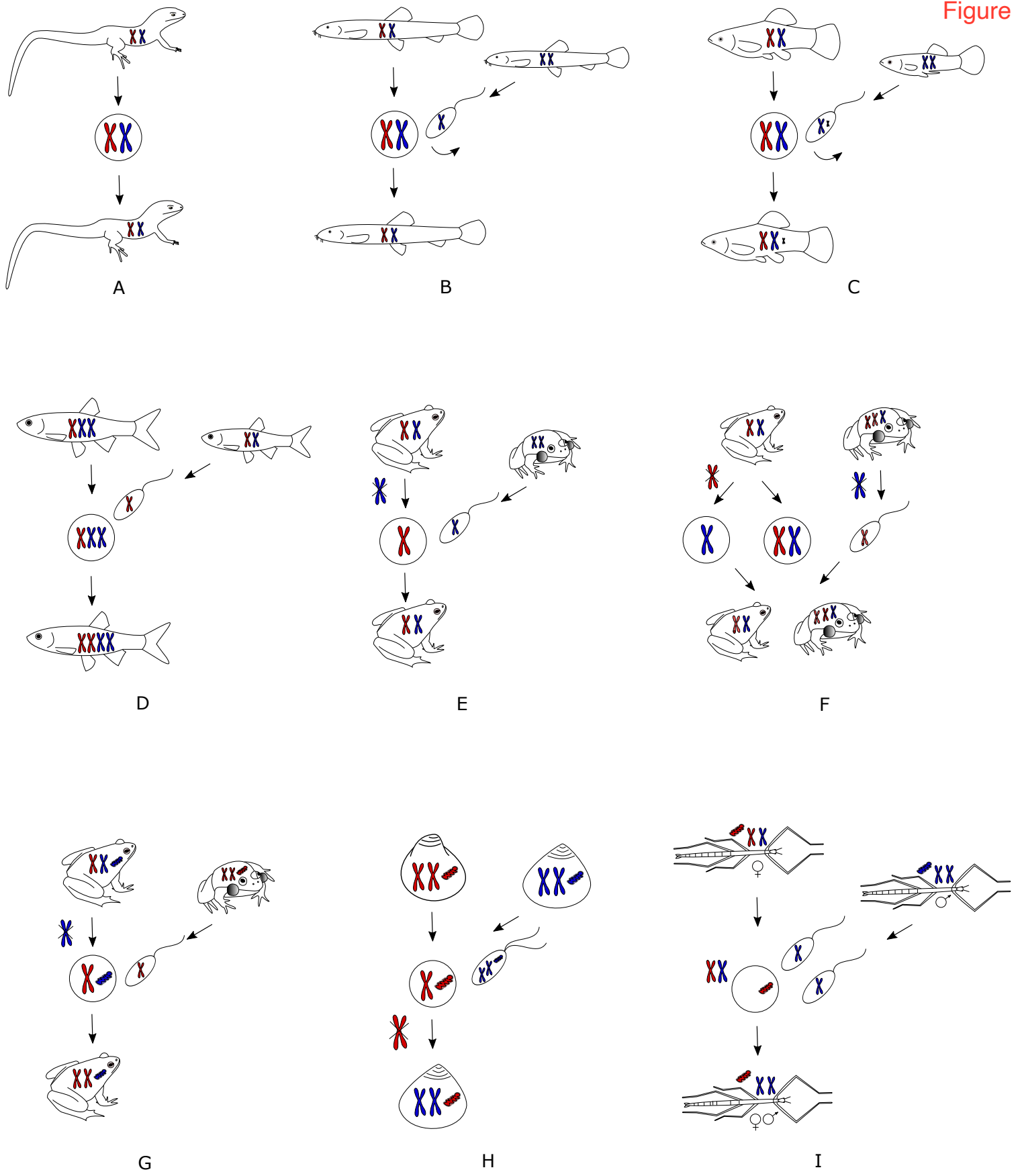
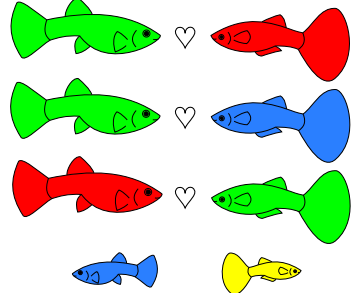
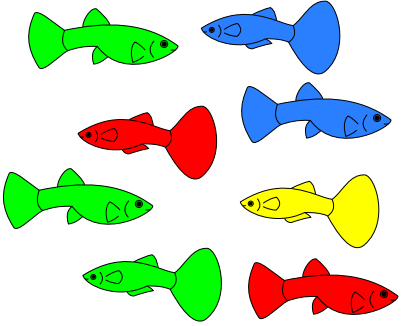
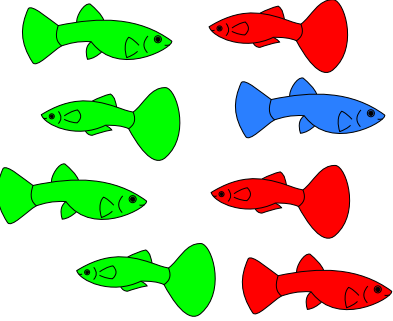


Figure 2

population A without asexuals

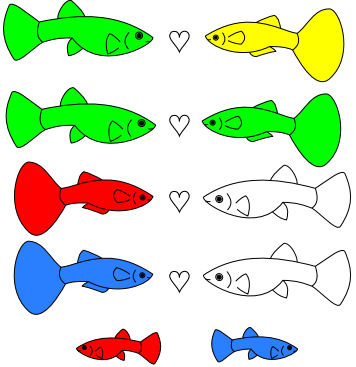
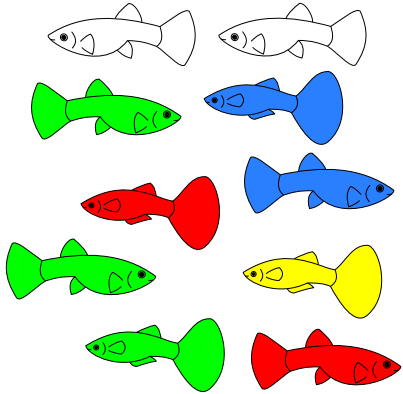


weaker genetic drift

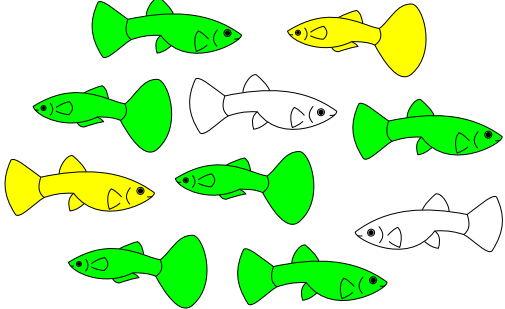


A

population B with asexuals



stronger genetic drift



B

genetic differentiation

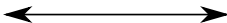


Figure 3

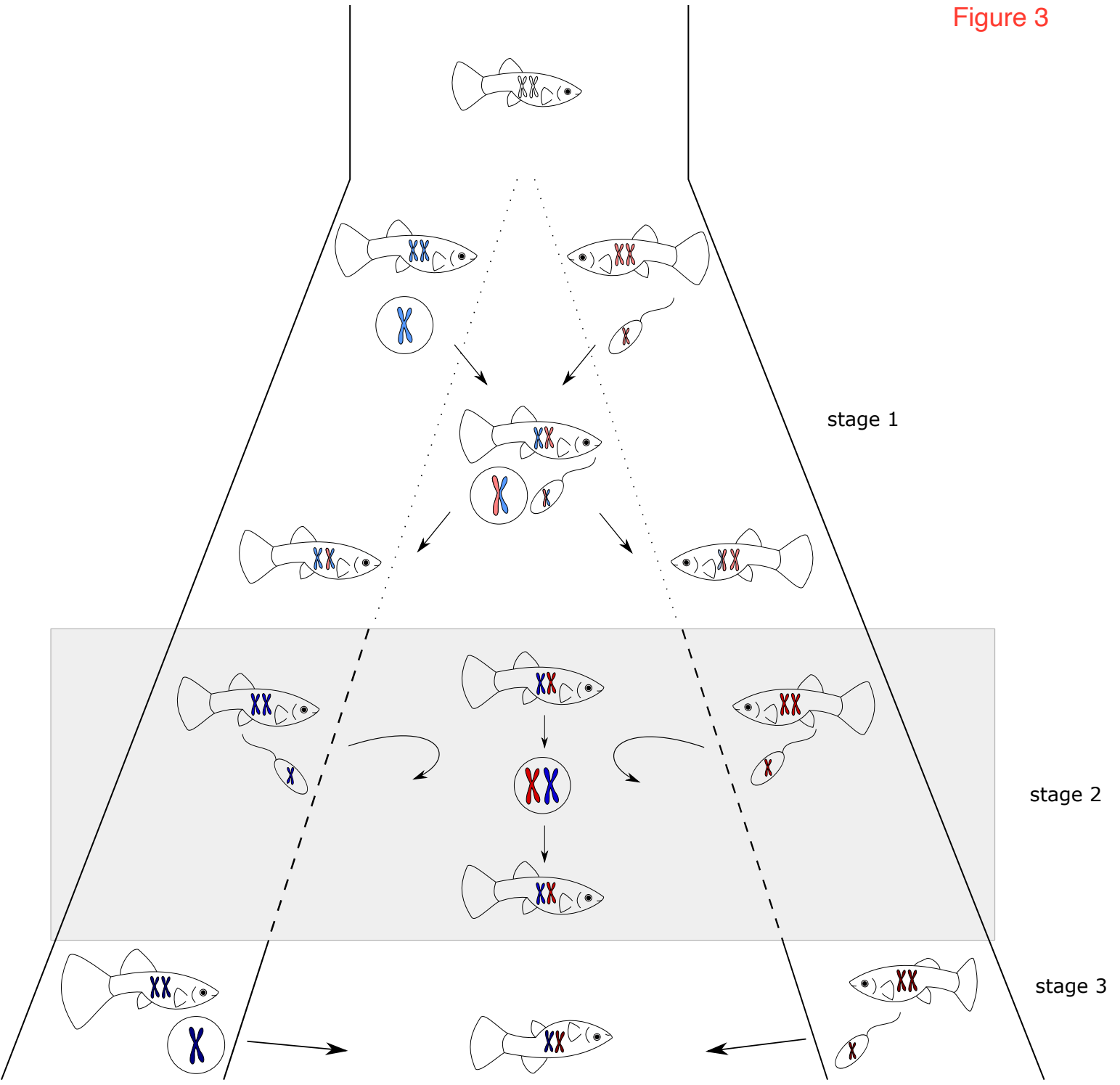
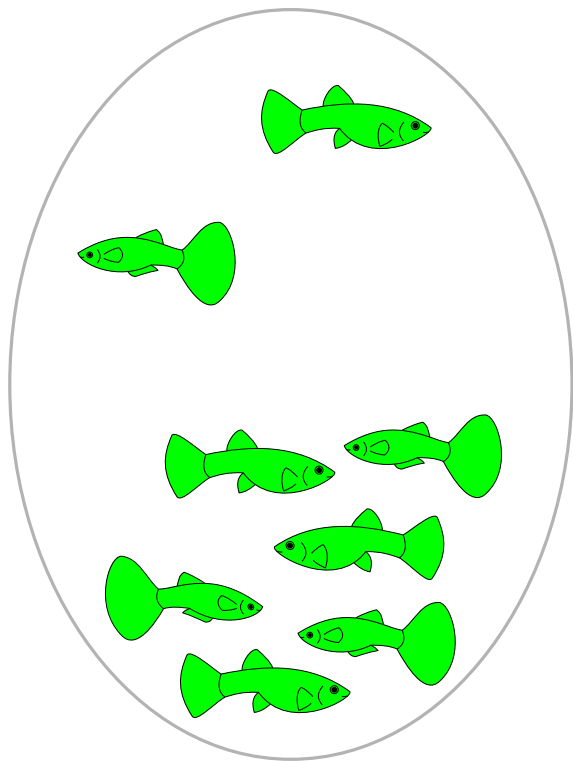
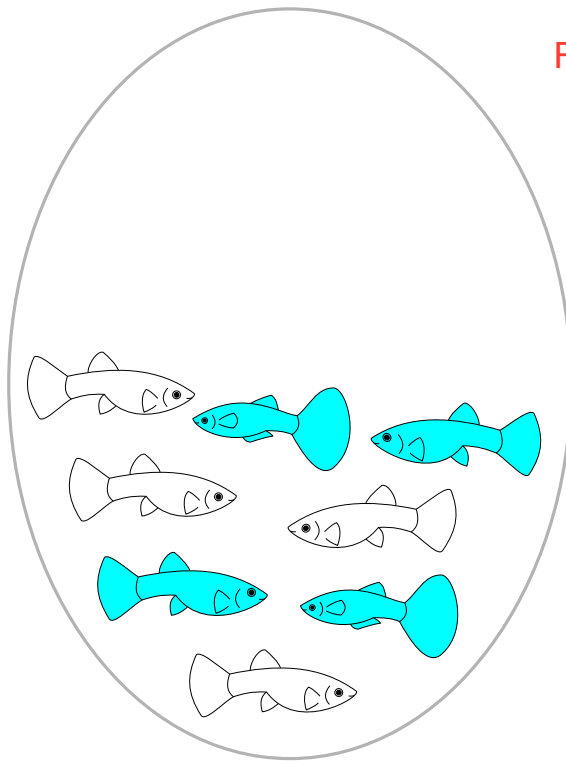


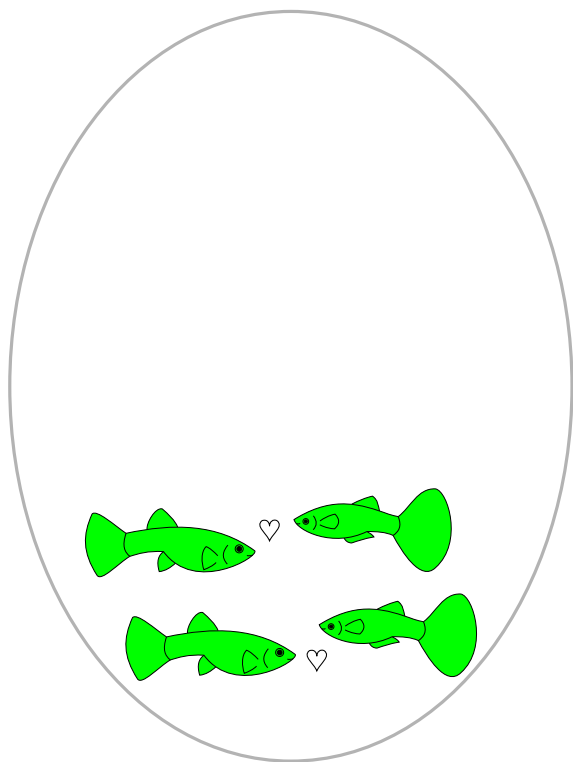
Figure 4



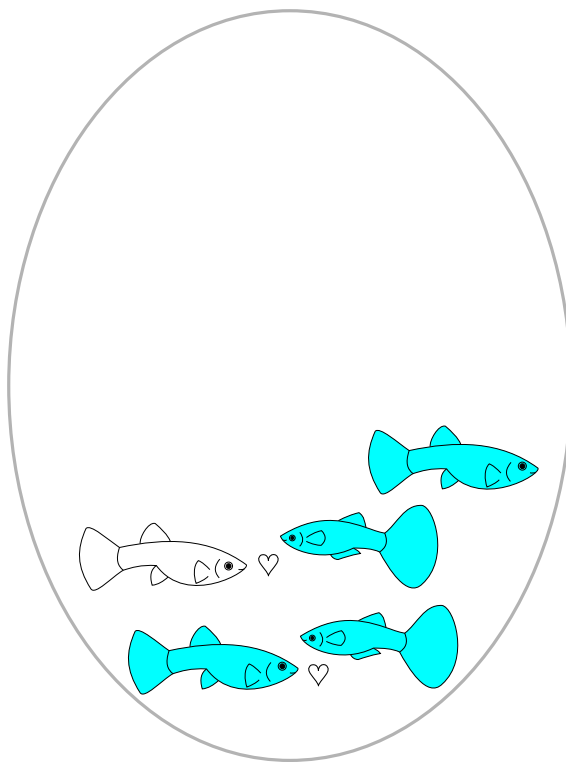
↑ dispersal without
asexuals



↑ dispersal with
asexuals

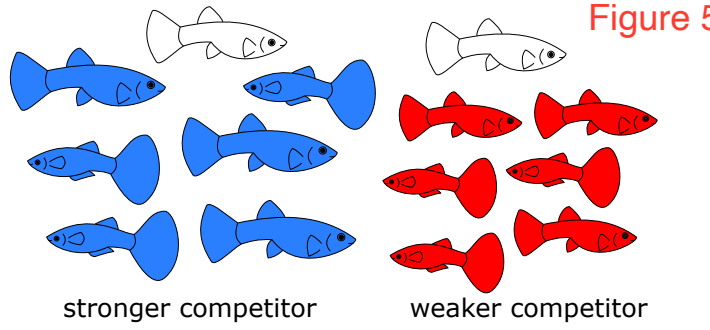
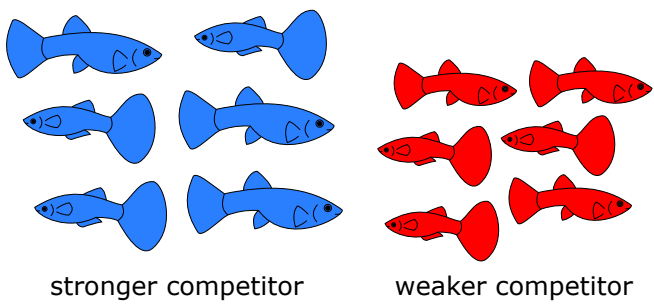


A

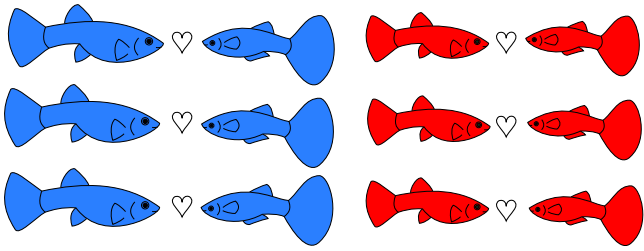


B

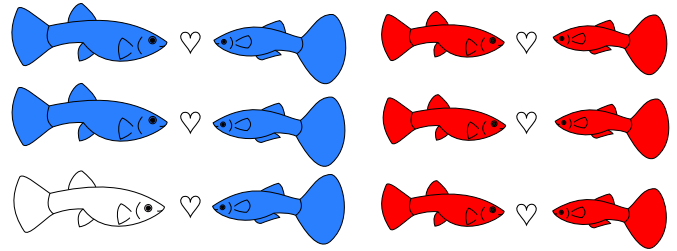
Figure 5



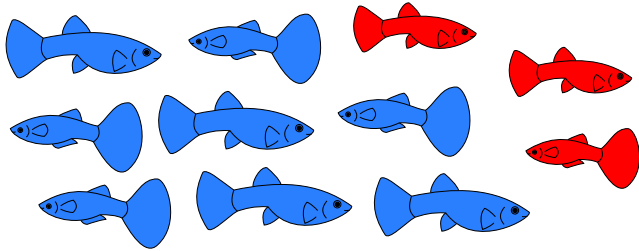
random mating ↓



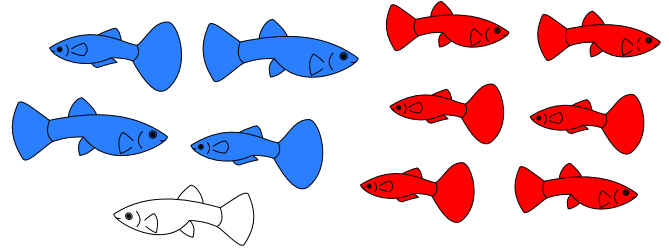
weak assortative mating ↓ strong assortative mating



competition ↓



apparent competition ↓



Supplementary Figure 1

