

1 **Inferring the history of hybridization: A case study in Iochrominae**  
2 **(Solanaceae)**

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8  
9 **ABSTRACT**

10  
11 The role of hybridization in evolution and speciation is one of the most active areas of  
12 evolutionary biology research. One problem that has continually vexed research in hybridization  
13 is accurate and robust detection of hybridization across large samples of species. Advances in  
14 phylogenetic data collection and theory have offered both powerful new datasets and methods  
15 with which to test for hybridization across large phylogenies. In this study, we compare patterns  
16 of hybridization in experimental phylogenetic datasets in order to compare existing methods for  
17 detecting hybridization. These analyses include both natural hybrids and an artificial hybrid as a  
18 phylogenetic ‘positive control’. Although our analyses recover strong indicators of hybridization  
19 in datasets with hybrid taxa, the results frequently disagree across methods or fail to identify  
20 known relationships (e.g., between the artificial hybrid and its parental taxa). These conflicting  
21 outcomes suggest that continued theoretical and empirical studies are needed to identify the  
22 factors that influence statistical power to detect past hybridization events in phylogenomic  
23 analyses.

24

## 25 INTRODUCTION

26

27 The impact of hybridization on evolution and speciation has been well established in  
28 evolutionary biology (Dobzhansky, 1940; Mayr, 1942b, a; Anderson, 1953; Stebbins, 1959;  
29 Grant and Grant, 1971; Rieseberg, 1995; Barton, 2001; Mallet, 2007; Abbott et al., 2013).  
30 Despite the historical notion that hybridization between distant relatives is rare and typically  
31 detrimental (Mayr, 1942a, b), numerous studies have indicated that hybridization occurs  
32 frequently between both closely and distantly related species (Mallet, 2005; Mallet, Besansky,  
33 and Hahn, 2016). Moreover, the introgression of genes via hybridization can be selectively  
34 advantageous (Grant et al., 2004; Consortium, 2012; Kirkpatrick and Barrett, 2015). The  
35 recognition that hybridization is a widespread and important mechanism of adaptation and  
36 speciation has placed a premium on the development of methods to detect signatures of reticulate  
37 evolution and identify the lineages and genomic regions involved (Nakhleh, 2013; Hejase and  
38 Liu, 2016)

39

40 Inferring a history of hybridization, whether among close or distant relatives, has typically relied  
41 upon molecular markers. The earliest approaches were developed and applied in a population  
42 genetic framework, focusing on pairs or ‘swarms’ of putatively hybridizing species (Moran, Bell,  
43 and Matheson, 1980; Avise and Saunders, 1984; Arnold, Hamrick, and Bennett, 1990). Although  
44 population genetic methods are powerful at shallow timescales (Durand et al., 2011), they do not  
45 easily scale to cases involving dozens of taxa. The need to test for hybridization across entire  
46 clades has driven the development of phylogenetic methods that build on population genetic  
47 principles (e.g., Kubatko, 2009; Yu, Degnan, and Nakhleh, 2012). These methods are often  
48 computationally intensive and require large amounts of data to distinguish the signal of  
49 introgression from the background noise generated by the stochasticity of the coalescent process  
50 (Yu et al., 2011; Eaton and Ree, 2013). Nevertheless, advances in sequencing and bioinformatics  
51 have made it increasingly feasible to build genome-wide datasets outside of model taxa (e.g.,  
52 Faircloth et al., 2012; Lemmon, Emme, and Lemmon, 2012; Rubin, Ree, and Moreau, 2012;  
53 Jarvis et al., 2014). Thus, future improvements in estimating evolutionary histories involving  
54 hybridization are likely to come from theoretical and computational advances (e.g., Kubatko and  
55 Chifman, 2015; Solis-Lemus and Ane, 2016).

56

57 Current approaches for studying hybridization in a phylogenetic framework can be divided into  
58 two broad categories, global and local. Global methods aim to detect the signal of hybridization  
59 across the phylogeny without explicitly determining the specific taxa involved (Table 1). These  
60 approaches often combine coalescent simulations with distance metrics or other indices to  
61 exclude the hypothesis that gene tree discordance can be explained by incomplete lineage sorting  
62 alone. In comparison to the global tests, local methods attempt to resolve specific hybridization  
63 relationships (i.e., identify the lineages involved in hybridization events) (Table 1). The more  
64 recent methods in this category estimate the proportion of genes inherited by a hybrid lineage

65 from each parent, in addition to inferring the topology of the species network (Kubatko and  
66 Chifman, 2015; Solis-Lemus and Ane, 2016; Blischak et al., 2018). After finding a global signal  
67 across the tree, local approaches can be used to pinpoint hybridization events. Still, the inference  
68 of the exact hybrid relationships remains challenging, and in some scenarios, it may be  
69 impossible to recover the full evolutionary history, regardless of the amount of available data  
70 (Pardi and Scornavacca, 2015).

71  
72 Here we explore the ability of local and global methods to detect historical hybridization in a  
73 natural and experimental context, using the Iochrominae as a model clade. This group of roughly  
74 40 Andean species in the tomato family has been traditionally divided into six genera, although  
75 many of the species and genera remain crossable and interfertile, even after millions of years of  
76 divergence (Smith and Baum, 2007). Previous phylogenetic studies with nuclear markers have  
77 demonstrated substantial gene tree conflict and suggested at least three reticulation events (Smith  
78 and Baum, 2006; Smith et al., 2008). In this study, we apply phylogenomic approaches to  
79 quantify the signal of hybridization and test specific hybrid relationships based on morphology  
80 and biogeography. Given the interfertility of many Iochrominae, we also created an artificial  
81 hybrid to serve as a ‘positive control’. Our approach is inspired by the pioneering work of  
82 Lucinda McDade (1990, 1992), who used natural hybrids to quantify the impact of hybridization  
83 on phylogenetic inference. Following her lead, we similarly manipulate the number of hybrids in  
84 our dataset to test the impact on the inference of species trees and hybridization events. We  
85 predict that increasing the number of hybrids in the dataset will increase discordance among gene  
86 trees and reduce the confidence in estimated species relationships. We also expect that artificial  
87 hybrids will be easier to detect as they have not had the opportunity to backcross or accumulate  
88 new mutations as in natural hybrids. In addition to improving our understanding of the role of  
89 hybridization in the Iochrominae radiation, these results will shed light on the strengths and  
90 limitations of current approaches and provide direction for future development of computational  
91 methods.

92  
93 **METHODS**

#### 94 95 **Study System**

96  
97 The Iochrominae has been well sampled in previous systematic studies (e.g., Smith and Baum,  
98 2006; Muchhala, Johnsen, and Smith, 2014; Deanna et al., 2018; Gates, Pilson, and Smith,  
99 2018), providing a strong framework for focused analyses of hybridization. This clade is sister to  
100 the large Physalinae (tomatillos and their relatives) (Deanna et al., 2019) and is largely restricted  
101 to the Andes of South America (Smith and Baum, 2006). Species of Iochrominae often grow in  
102 sympatry (Smith et al., 2008) and, in some cases, form hybrid zones (Smith, Kolberg, and Baum,  
103 2008). Phylogenetic studies have sampled several putative hybrids and recovered strongly  
104 conflicting relationships in nuclear gene trees (Smith and Baum, 2006). The strongest support for

105 hybridization from genetic, morphological and biogeographic data corresponds to two taxa (*I.*  
106 *ayabacense* S. Leiva and *I. stenanthum* S. Leiva, Quip. & N. W. Sawyer) (Fig. 1). Native to  
107 northern Peru, *I. ayabacense* grows in a zone of sympatry with several species (*I. cyaneum*  
108 (Lindl.) M. L. Green, *I. confertiflorum* (Miers) Hunz., *I. arborescens* (Schltdl.) J. M. H. Shaw,  
109 and *I. lehmannii* Bitter (syn. *I. squamosum* S. Leiva & Quip.) (Smith, Kolberg, and Baum, 2008).  
110 Alleles from this species fall either in a clade with *I. lehmannii* or with *I. cyaneum* (Smith and  
111 Baum, 2006). Such a biphyletic pattern (with alleles from the same locus in distant clades on the  
112 phylogeny) is a signature consistent with recent hybridization (Barkman and Simpson, 2002).  
113 The chromosome number of *I. ayabacense* is  $2n=24$ , like other *Iochroma* species, suggesting it is  
114 a homoploid hybrid (Smith, Kolberg, and Baum, 2008). Also, its morphology is intermediate,  
115 with sturdy funnel-shaped flowers like *I. lehmannii* but flushes of purple floral pigmentation and  
116 long corolla tube like *I. cyaneum* (Smith, Kolberg, and Baum, 2008).

117  
118 The other putative hybrid taxon, *I. stenanthum*, has similarly intermediate coloration, being  
119 creamy at the base and mauve towards the mouth of the flower. It occurs in another zone of  
120 sympatry in northern Peru and has been proposed to be a hybrid of the lineage comprising the  
121 purple-flowered *I. cornifolium* (Kunth.) Miers and *I. cyaneum* and a white-flowered lineage from  
122 the clade containing *I. confertiflorum* and *I. arborescens* (Smith and Baum, 2006; Smith and  
123 Leiva, 2011). *I. stenanthum* presents an intermediate morphology, being purple at the mouth of  
124 the corolla and fading to cream towards the base. It has long tubular flowers as in *I. cornifolium*  
125 and *I. cyaneum* but more triangular corolla lobes as in *I. confertiflorum* and *I. arborescens*. In  
126 individual gene tree analyses, alleles from *I. stenanthum* show affinity towards both the white-  
127 and purple-flowered lineages (Gates et al., unpublished data). In combined analyses, the  
128 phylogenetic position of *I. stenanthum* is unstable, but it often appears as the sister lineage to the  
129 clade containing the putative parental taxa (Smith and Baum, 2006; De-Silva et al., 2017). The  
130 latter phylogenetic pattern was proposed by McDade (1990) as a common outcome for hybrid  
131 taxa.

132  
133 In addition to these two putative natural hybrids, we included one artificial hybrid, generated in  
134 the greenhouse. This accession is a cross between *I. arborescens* (Bohs 2428 (UT), grown from  
135 seed collected from Las Cruces, Costa Rica) and *I. cyaneum* (Smith 265 (WIS), grown from seed  
136 collected from plants cultivated by W. G. D'Arcy at the Missouri Botanical Garden; likely  
137 provenance, Ecuador). These parental species are thought to be 2 to 4 million years diverged  
138 (De-Silva et al., 2017), and some F<sub>1</sub> seeds produce vigorous offspring. The hybrid flower has an  
139 intermediate phenotype (funnel-shaped flowers, white with purple markings). Its fertility has not  
140 been thoroughly examined, although it seems to be capable of backcrossing to the parental taxa  
141 (S. D. Smith, unpublished data).

## 142 143 **Experimental design** 144

145 In order to assess our ability to detect hybridization with various phylogenomic methods, we  
146 created a series of datasets with different combinations of the two putative natural hybrids (*I.*  
147 *stenanthum* and *I. ayabacense*), their putative parental lineages (*I. arborescens*, *I. lehmannii*, *I.*  
148 *cyaneum*), the artificial F<sub>1</sub> hybrid (*I. arborescens* x *I. cyaneum*), plus an outgroup (*Physalis*  
149 *peruviana* L.). For two of the three putative parents (*I. arborescens* and *I. lehmannii*), we have  
150 two sampled individuals (Table 2). All of the experimental datasets contain the same number of  
151 tips (six) and all have at least one individual of each putative parent plus the outgroup (Fig. 2).  
152 Dataset 1 has no putative hybrids, while dataset 2 has the F<sub>1</sub> hybrid substituting for one of the  
153 two *I. arborescens* individuals. Dataset 3 has the F<sub>1</sub> and *I. ayabacense* replacing one of each of  
154 the *I. arborescens* and *I. lehmannii* individuals. As *I. ayabacense* tends to fall with *I. lehmannii*  
155 in species trees (Gates, unpublished) estimated with STAR (Liu et al., 2009), the topology for  
156 dataset 3 is expected to mirror that for dataset 2 (Fig. 2), albeit with less support due to the  
157 expected hybrid ancestry. The fourth dataset has the two putative natural hybrids (*I. stenanthum*  
158 and *I. ayabacense*) in the place of one of each of the *I. arborescens* and *I. lehmannii* individuals.  
159 The putative parents for *I. stenanthum* are the same as for the F<sub>1</sub> (*I. arborescens* x *I. cyaneum*)  
160 although STAR species tree analyses tend to infer a closer relationship with *I. arborescens*  
161 (Gates, unpublished). Thus, we expect a similar species tree topology to the previous datasets,  
162 but a weaker signal of hybridization since *I. stenanthum* is likely a late generation hybrid  
163 compared to the F<sub>1</sub> (Fig. 2).

164

## 165 **Target capture and assembly**

166

167 We conducted targeted sequence capture to build a molecular dataset for phylogenomic analyses  
168 following Gates et al. (2018). Briefly, we used genomic resources for tomato (Sato et al., 2012)  
169 and *Iochoroma cyaneum* (Gates et al., 2016; Strickler et al., unpublished data) to design probes for  
170 242 single copy genes, most of which were pulled from the conserved orthologous set (Wu et al.,  
171 2006). These loci totaled approximately 1 MB and were used as the template for custom bait  
172 synthesis of 80 nucleotide targets at 2X tiling from MycroArray (MycroArray Inc; Ann Arbor,  
173 MI). Genomic DNA was extracted from silica-dried leaf tissue using a 2XCTAB protocol (Doyle  
174 and Doyle, 1987) and sheared DNA to approximately 500 bases with sonication. For all libraries  
175 except for the F<sub>1</sub> hybrid, we used the TruSeq library preparation kits for sequence adapter  
176 ligation and Mycroarray capture protocol V1. These libraries were sequenced on a single lane of  
177 an Illumina HiSeq2500 on 100bp pair-end rapid mode. For the F<sub>1</sub> hybrid, we used the NEBNext  
178 Ultra II kit (New England Biolabs, Ipswich, MA) for library preparation and Mycroarray capture  
179 protocol V4. This library was sequenced on an Illumina HiSeq 4000, also in 100bp pair-end  
180 mode. We filtered raw reads by using Trimmomatic (Bolger, Lohse, and Usadel, 2014) and  
181 assembled filtered raw reads with the iterative read-mapping program YASRA (Ratan, 2009).  
182 We created a master alignment for all individuals by orienting assemblies to homologous bases  
183 of the reference using custom Blat (Kent, 2002) and samtools (Li et al., 2009) scripts. We then  
184 used this master alignment to subset individuals into each of the four experimental datasets. We

185 conducted gene tree searches on each gene in each of the four datasets using RAxML 8  
186 (Stamatakis, 2014) with 100 bootstrap replicates to assess support. These alignments and gene  
187 tree estimates were used for all downstream analyses (available at  
188 <https://github.com/danjgates/HybrData>).

189

## 190 **Detecting patterns of hybridization from gene tree distributions**

191

192 To quantify the level of gene tree conflict associated with varying numbers of hybrids in our four  
193 datasets, we first applied the internode and tree certainty (IC/TC) measures of Salichos,  
194 Stamatakis, and Rokas (2014) as implemented in RAxML 8 (Stamatakis, 2014). These  
195 information theory-based metrics can be applied to trees generated from any method and  
196 optimality criterion (e.g., parsimony, likelihood) and are computationally tractable with large,  
197 genome-scale data matrices. We computed IC for each of the three internodes in our  
198 experimental datasets (Fig. 2). These values represent the degree of conflict between  
199 relationships inferred with STAR (Liu et al., 2009) and the most common conflicting partition.  
200 When IC approaches 1, there is little conflict at the node, and when it is near zero, there is strong  
201 conflict, with similar numbers of gene trees showing the bipartition in the species tree and the  
202 next most common bipartition. For each of our four datasets, we also computed the tree  
203 credibility (TC) score, which is the sum of the IC scores over all ingroup bipartitions (Salichos,  
204 Stamatakis, and Rokas, 2014). We predicted that the addition of tips with known or inferred  
205 hybrid ancestry would increase gene tree conflict and thus decrease the certainty in relationships  
206 across the phylogeny.

207

208 We further characterized the distribution of the gene trees in each dataset by using “Tree  
209 Incongruence Checking in R” or “TICR” (Stenz et al., 2015; Solis-Lemus, this volume). This  
210 program uses pseudo-loglikelihoods (PLL) calculated from rooted quartet distributions to test the  
211 fit of gene tree distributions to different hypotheses of panmixia or species tree structure. While  
212 rejection of a population tree may be consistent with a history of hybridization and may be used  
213 as evidence of hybridization (Stenz et al., 2015), the program does not directly test a model of  
214 hybridization against the other models of bifurcation or panmixia. We used TICR to compute the  
215 PLL and associated  $\chi^2$  statistic for both the panmixia (star-like ingroup tree) and fully bifurcating  
216 species tree in each of the four datasets. In comparing datasets with hybridization to those  
217 without (e.g. one F<sub>1</sub> dataset vs. the no putative hybrid dataset), we expect that there should be  
218 less support for a bifurcation model as indicated by a greater  $\chi^2$  value and more support for a  
219 panmixia model as indicated by a lowered  $\chi^2$  value in the dataset with greater amounts of  
220 hybridization. The TICR analyses rely on concordance factors (CFs) for each quartet (Ane et al.,  
221 2007), estimated with BUCKY (Larget et al., 2010). We extracted these CFs for each clade as an  
222 additional window into patterns of gene tree discord associated with adding hybrid taxa.

223

## 224 **Testing for hybridization in empirical datasets**

225  
226 In addition to examining uncertainty and discordance across the tree, we also implemented three  
227 specific tests for hybridization. While the two approaches above (IC/TC and TCR) may offer  
228 support for hybridization, these methods do not explicitly test whether a hybridization model is a  
229 more appropriate model than a model without hybridization. In addition to reticulation, conflict  
230 among gene trees and instability in species tree inference can arise due to other factors, such as  
231 incomplete lineage sorting (Degnan and Rosenberg, 2006), recombination (Schierup and Hein,  
232 2000), model misspecification (Buckley, 2002), biased taxon sampling (Hillis, 1998), missing  
233 data (Wiens, 2003), sequencing errors (Kuhner and McGill, 2014), and alignment uncertainty  
234 (Hossain et al., 2015). Thus, without explicit modeling of processes of hybridization, metrics like  
235 IC and CF only offer weak support for hybridization.

236  
237 In order to directly test for hybridization, we first applied HyDe (Hybrid Detection; Blischak et  
238 al., 2018). This program localizes hybridization events (i.e., identifies hybrid taxa and their  
239 putative parental taxa) based on patterns in sequence alignments. Detecting these events relies on  
240 invariants, i.e., phylogenetically informative site patterns in quartet subtrees, such as those used  
241 in the ABBA/BABA patterns used to compute D-statistics (Patterson et al., 2012). Specifically,  
242 HyDe uses the ratio of the differences between specific site patterns, as this approach was found  
243 to provide the highest statistical power (Kubatko and Chifman, 2015). We also implemented a  
244 similar alignment-based method that computes D-statistics across all quartet subtrees to test for  
245 introgression (Eaton and Ree, 2013). While HyDe reports statistics for every combination of two  
246 parental tips and one hybrid tip, the D-statistic approach reports all possible triplet combinations  
247 that are consistent with the species tree. For the purpose of this analysis, we only report the  
248 maximum statistic for each method in each dataset as this should be the strongest proposed  
249 hybrid relationship.

250  
251 Along with these alignment based approaches, we used an additional method, SNaQ (Species  
252 Networks applying Quartets; Solis-Lemus and Ane, 2016; Solis-Lemus, this volume), to fit  
253 reticulation events based upon gene tree distributions. Like the TCR method described above,  
254 SNaQ calculates a quartet-based PLL for models with and without hybridization. Unlike TCR,  
255 however, SNaQ does directly test support for hybridization. For each dataset, we report the  
256 maximum PLL score for the strictly bifurcating species tree as well as the maximum score from  
257 the best network produced by a full search with up to six optimal reticulation events. For SNaQ  
258 as well as HyDe, we expect that inferred reticulation events will follow the predictions based on  
259 morphology and biogeography and that more recent hybridization events (the  $F_1$  hybrid and the  
260 biphyetic *I. ayabacense*) will be more consistently and accurately recovered than the older  
261 events (those that formed *I. stananthum*).

262  
263 RESULTS  
264

265 **Addition of hybrid taxa increases discordance and decreases tree-like signal**

266

267 Comparisons between the no hybrid and one F<sub>1</sub> dataset offer strong evidence that hybridization  
268 has an impact on a number of tree metrics. Values for internode certainty (IC) ranged from 0.19  
269 to 0.60 in the no hybrid tree and dropped to 0 to 0.32 with the substitution of the F<sub>1</sub> hybrid for  
270 one of the putative parental individuals (Fig. 3a,b). Overall tree certainty (TC) fell from 1.04 to  
271 0.32 (Fig. 3a,b). We also saw decreases in concordance factors, from 0.47 to 0.71 without the F<sub>1</sub>  
272 to 0.26 to 0.53 with the F<sub>1</sub>. Given that the F<sub>1</sub> is an artificial hybrid between *I. arborescens* and *I.*  
273 *cyaneum* (Fig. 1), we would expect the branches connecting these taxa to be most strongly  
274 affected by adding the hybrid and, indeed, these were the only internodes with decreases in IC  
275 and CF values. In fact, the unaffected branch, uniting the two *I. lehmannii* individuals, saw a  
276 slight increase in IC and CF (Fig. 3a,b). These increases in gene tree conflict with the addition of  
277 the F<sub>1</sub> hybrid shifted the  $\chi^2$  values from TCR test in line with decreasing tree-like signal.  
278 Although both datasets reject the two extremes (panmixia and a fully bifurcating history), we  
279 observed weakening of the rejection of complete panmixia ( $\chi^2_P$  dropping from 26 to 11) and a  
280 strengthening the rejection of a fully bifurcating tree ( $\chi^2_B$  rising from 58 to 75) with the inclusion  
281 of the F<sub>1</sub> (Fig. 3a,b).

282

283 The increase from one hybrid tip to a second hybrid tip brought additional decreases in certainty  
284 and concordance. In both datasets 3 and 4, all of the internal branches are expected to be  
285 influenced by the hybridization events and indeed, all internodes dropped to IC values less than  
286 0.03 and CFs of less than 0.36 (Fig. 3c,d). Overall tree certainty in both cases was near zero, and  
287 panmixia was strongly favored over a fully-bifurcating history (Fig. 3c,d). The combination of  
288 the F<sub>1</sub> and *I. ayabacense* in dataset 3 had a slightly stronger effect on these values, which may be  
289 due to the recency of their formation. By comparison, *I. stenanthum* appears to be a more ancient  
290 hybrid (with less intermediate morphology, no strongly biphyletic alleles, and a geographic  
291 distribution more distant from its putative parents) (Smith and Baum, 2006). Thus, some of the  
292 conflicting signal originally carried in this lineage may have eroded due to sorting of parental  
293 alleles, the fixation of new mutations, and/or backcrossing to one of the parents.

294

295 **Tests of hybridization support different relationships than expected**

296

297 All three of the tests used to detect hybridization events (HyDe, SNaQ, and D-statistics) provided  
298 strong support for hybrid ancestry but also pointed to some unexpected relationships. First, both  
299 HyDe and the D-statistics supported some degree of reticulation in dataset 1 (the no-hybrid  
300 datasets) and two taxa, *I. arborescens* and *I. lehmannii*, were consistently implicated (Table  
301 3a,b). With correction for multiple tests, these results are marginally significant. SNaQ recovered  
302 the same tips involved, but in this case, the addition of the reticulation did not improve the  
303 pseudo-loglikelihood of the tree (Table 3c). Although the implication of some reticulation  
304 involving *I. arborescens* and *I. lehmannii* was surprising, it is notable that *I. lehmannii* does

305 show instability in traditional phylogenetic analyses and variously appears as more closely  
306 related to *I. arborescens* (Deanna et al., 2018) or *I. cyaneum* (Deanna et al., 2019), always with  
307 weak support (bootstrap < 75%). Species tree analyses favor the former relationship (Gates,  
308 Pilson, and Smith, 2018), but these have not previously included tests for hybridization. Still, the  
309 strength of support for hybridization was weak compared to the other three datasets (Table 3).  
310 Given these mixed results, we consider that *I. lehmannii* does not have a large degree of hybrid  
311 ancestry, and that the patterns may reflect a small rate of gene flow between *I. lehmannii* and *I.*  
312 *arborescens* in southern Ecuador and northern Peru, where the two grow in proximity (S.D.  
313 Smith, pers. obs.).

314

315 The datasets with one or more hybrid taxa consistently supported the presence of hybridization,  
316 and in a few cases, recovered the predicted relationship. We expected the best case scenario to be  
317 the one F<sub>1</sub> dataset, where the parents of the artificial hybrid (*I. arborescens* and *I. cyaneum*) are  
318 known and included in the tree. HyDe identified the F<sub>1</sub> as being involved but placed it as a  
319 parent, with *I. lehmannii* as the other parent and *I. arborescens* as the hybrid (Table 3a). The D-  
320 statistics pointed to the same three taxa as involved in hybridization (Table 3b). By contrast,  
321 SNaQ recovered the correct relationship among the parental taxa and the F<sub>1</sub> and with strong  
322 support for dataset 3 (Table 3c).

323 The results for datasets with two of the six taxa being hybrids were similarly mixed. For dataset  
324 3 (F<sub>1</sub>+*I. ayabacense*), HyDe correctly grouped *I. ayabacense* with *I. arborescens* and *I.*  
325 *cyaneum*, but inferred *I. arborescens* instead of *I. ayabacense* as the hybrid among the three.  
326 Neither D-statistics nor SNaQ estimated a predicted grouping of parents with hybrids for dataset  
327 3. For dataset 4 with two natural putative hybrids (*I. ayabacense* + *I. stenanthum*), only the D-  
328 statistics recovered a predicted grouping of tips involved in hybridization (*I. ayabacense* and its  
329 putative parents, *I. lehmannii* and *I. cyaneum*) (Table 3).

330

331 DISCUSSION

332

### 333 **Effects of hybridization on patterns of gene-tree discordance**

334

335 Our manipulations of the taxon sampling show that adding known or putative hybrids increases  
336 discordance between gene trees and decreases the signal of tree-like relationships. All of the  
337 datasets in which putative or known hybrid taxa were swapped with putative parental taxa  
338 showed marked decreases in concordance factors and internode certainty and a strong preference  
339 for panmixia over tree-like relationships (Fig. 3). The comparison of the one F<sub>1</sub> dataset compared  
340 with the no putative hybrid dataset offers the strongest evidence on this front because the F<sub>1</sub> is an  
341 artificial hybrid generated in the greenhouse. In this dataset, the concordance factors for the  
342 nodes connecting the two putative parental species were reduced by roughly half and the  
343 certainty for those relationships dropped to zero. Doubling the number of hybrid taxa (datasets 3  
344 and 4) further increased gene tree conflict, and with hybrids distributed across the tree, complete

345 panmixia could not be rejected by TICR (Fig. 3). Altogether, we see patterns consistent with  
346 prevalent, detectable hybridization in all trees that we expect to exhibit a signal of reticulate  
347 ancestry.

348  
349 The methods designed to identify hybrid tips and their parental taxa also showed strong evidence  
350 of hybridization in datasets with more putative or known hybrids. For example, SNaQ and HyDe  
351 did not find significant support for hybridization in dataset 1, with no known hybrids (Table 3).  
352 In the remaining three datasets, all of the methods inferred significant reticulation, with the  
353 strength of that inference increasing with the number of hybrid taxa. These results indicate that  
354 the sharp discordance generated by the addition of hybrid tips, as indicated by decreasing  
355 concordance factors and tree certainty (Fig. 3), is generally correctly interpreted by HyDe, D-  
356 statistics and SNaQ as evidence of hybridization during the evolutionary history of *Iochroma*.

357

### 358 **Challenges in determining the exact hybrid relationships**

359

360 Our results also indicate that exact hybrid relationships may be difficult to assign. Looking  
361 across the three methods and the three datasets with augmented hybrid taxa, only two of these  
362 nine combinations estimated a predicted set of three taxa (two hypothesized parental tips along  
363 with the hypothesized hybrid tip). These differed across methods, with D-statistics inferring a  
364 predicted trio of tips in dataset 4, and SNaQ in dataset 2 (shaded in Table 3). Although both  
365 HyDe and SNaQ directly estimate which tip in a trio is the hybrid tip, only SNaQ correctly  
366 inferred a predicted relationship, identifying the  $F_1$  as the product of a cross between *I.*  
367 *arborescens* and *I. cyaneum*. This relationship between the artificial hybrid and its parents was  
368 intended to serve as our ‘positive control’ for detecting hybrid ancestry, and it is worrisome that  
369 only one of the three methods recovered the correct topology.

370

371 The wide range of inferred and seemingly spurious hybrid relationships across methods and  
372 datasets may relate to both our experimental design and our study system. We intentionally built  
373 a small dataset with two putative natural hybrids and their putative parents with the goal of  
374 maximizing the potential for methods to converge upon the correct sets of relationships. While  
375 we are not entirely certain of these relationships, the combination of morphological,  
376 biogeographic and genetic data provides the strongest possible *a priori* predictions, beyond  
377 generating these hybrids ourselves. Still, a larger dataset, with more taxa to break up the  
378 branches between the putative parents, might have provided more power to discriminate among  
379 possible placements for reticulation events. The fact that hybridization appears to be common in  
380 *Iochroma* may also contribute to the challenge of estimating reticulation events. Controlled  
381 greenhouse crosses suggest that all *Iochroma* are able to interbreed to some degree, and they can  
382 also cross with closely related genera (Smith and Baum, 2007). Apparent hybrid zones are well  
383 documented in herbaria as well as in the literature (Smith and Baum, 2006; Smith and Leiva,  
384 2011). Accordingly, all of the *Iochroma* lineages may contain a signal of hybrid ancestry,

385 consistent with rejection of a fully-bifurcating tree by TICR for the no hybrid dataset (Fig. 3a).  
386 This episodic gene flow, combined with incomplete lineage sorting (ILS) along short branches of  
387 the backbone of the species tree (Deanna et al., 2019), may result in relatively little information  
388 for making robust inferences about which tips have a significantly reticulate history.

389  
390 It is important to note that the difficulty of making inferences about hybridization is already well  
391 documented in empirical and theoretical studies. Simulations attempting to reconstruct networks  
392 where there are few tips (<5) and multiple hybridization events have often failed to find the  
393 accurate network (Yu et al., 2011; Solis-Lemus and Ane, 2016). Presumably, this is because with  
394 few taxa and multiple reticulations, the set of gene trees may be equally well explained by  
395 different network topologies (Pardi and Scornavacca, 2015). The fact that multiple  
396 hybridizations can confound inference could explain why SNaQ identified the correct  
397 relationships in the one-F1 dataset but failed in datasets 3 and 4 with two hybrids (Table 3).  
398 Moreover, SNaQ assumes that the edges can only be involved in one reticulation event (Solis-  
399 Lemus and Ane, 2016), and in datasets 3 and 4, *I. cyaneum* likely contributed to two events.  
400 Although making inferences about multiple reticulations from small trees is challenging, a  
401 similar empirical study was able to strongly support up to four reticulations in a dataset with six  
402 species (Wen et al., 2016). Given these conflicting notions in the literature about when  
403 hybridization can be confidently detected, we suggest that future theoretical studies explore a  
404 broad array of scenarios to determine which factors have the strongest influence on the power to  
405 correctly infer reticulate relationships. We expect that population size and divergence time would  
406 have major impacts because these parameters directly govern the amount of signal and noise  
407 (ILS) in multilocus datasets (Degnan and Rosenberg, 2006). Violations of mutation rate  
408 assumptions may also have a large effect upon methods like HyDe and D-statistics because  
409 increases or decreases in mutation rates will make some lineages more similar to distant relatives  
410 without hybridization ever taking place. As empirical studies like ours continue to apply these  
411 relatively new methods, we expect that additional biological factors that potentially influence  
412 statistical power are likely to emerge.

413

#### 414 **Hybridization in *Iochrominae***

415

416 One goal of this study was to assess support for the hybrid origins of *I. ayabacense* and *I.*  
417 *stenanthum* with genome-wide markers. While previous studies had assembled evidence for  
418 these hypotheses from a handful of nuclear markers along with morphological and biogeographic  
419 information (Smith and Baum, 2006), this is the first study to directly test for hybridization with  
420 phylogenomic methods. The analyses of gene tree discordance provide some support for hybrid  
421 ancestry in that the presence of these taxa in the datasets decreased tree-like signal and certainty  
422 in relationships among tips. Also, the analyses with the two natural hybrids (dataset 4) showed  
423 similarly low concordance to those with one natural hybrid and one artificial hybrid (dataset 3),  
424 suggesting that a natural hybrid (in this case, *I. stenanthum*) disrupted tree-like relationships to a

425 similar degree as the F<sub>1</sub> (Fig. 3). Nevertheless, our results are far from conclusive in terms of  
426 which lineages gave rise to these two hybrid species. The local methods each returned different  
427 estimates of relationships among putative parental taxa and putative natural hybrids (Table 3).  
428 Moreover, none of these estimated relationships followed those predicted based on previous  
429 studies (Fig. 1) despite the fact that the ingroup taxa were comprised of putative parental taxa.  
430 Accurate inference of the hybrid ancestry for *I. stenanthum* was expected to be challenging  
431 because its phylogenetic position is more suggestive of an ancient hybrid, e.g. between stem  
432 lineages that gave rise to *I. arborescens* and *I. cyaneum* (Smith and Baum, 2006). By contrast,  
433 we expected the relationships between *I. ayabacense* and its putative parents would be easily  
434 detected given that the strong biphyletic patterns (e.g. one *LFY* allele sister to *I. lehmannii* and  
435 the other sister to *I. cyaneum*) in gene trees (Smith and Baum, 2006). The inability of multiple  
436 approaches to recover the predicted topology for *I. ayabacense*, not to mention the F<sub>1</sub> positive  
437 control, suggests that renewed attempts at testing these hybrid origins will require more  
438 statistical power (more loci, more individuals, and/or more tips). Ultimately, with a larger  
439 dataset, we expect that the majority of local methods should converge on the same set of  
440 relationships, especially since they all build on quartet-based patterns.

441

## 442 CONCLUSIONS

443

444 The growth of phylogenomic datasets along with the well-established prevalence of  
445 hybridization in nature has driven the development of an array of statistical tools for detecting  
446 and localizing reticulation in phylogenies. The application of these methods has led to the  
447 inference of hybridization at multiple phylogenetic scales, from sister species (e.g., Turissini and  
448 Matute 2017) to more distantly related taxa (e.g., Buckley et al. 2006; Eaton et al. 2013). Our  
449 exploratory analyses, which included both known and hypothesized hybrid tips, suggest that,  
450 while the broad signature of reticulate evolution is relatively easily detected (Reid et al. 2012;  
451 Blanco-Pastor et al. 2012; Villiers et al. 2013), the accurate localization of hybridization on the  
452 phylogeny is significantly more difficult. While these inferences may be challenging in  
453 *Ioichroma* because of periodic gene flow across the entire tree, we do not consider this clade to be  
454 an outlier. Hybridization has accompanied the diversification of many taxa (e.g., butterflies,  
455 birds, fish) and is certainly well-documented in plant clades. Groups in which past and on-going  
456 hybridization is suspected are likely to be the primary targets for empirical applications of  
457 methods, such as SNaQ and HyDe. Thus, we encourage further empirical or experimental studies  
458 manipulating the degree of hybrid ancestry in addition to theoretical work to explore the power  
459 to make inferences across a spectrum of biologically realistic scenarios (including recently  
460 formed hybrid species in a backdrop of ILS and periodic gene flow). With more robust  
461 inferences, we may begin to arrive at a broader understanding of how reticulation affects patterns  
462 of genomic variation and how these impacts relate to the timing, duration and scale of  
463 hybridization events.

464

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658

**Table 1.** Various approaches that have been used to detect hybridization. Global analyses are those which test for hybridization involving two or more branches of the phylogeny, but do not attempt to localize the events. Local methods aim to determine the number and location of the reticulation events, and therefore which lineages have been involved.

Pattern	Type	Reference
Tree distance	Global	(Buckley et al., 2006)
Count deep coalescences	Global	(Reid, Demboski, and Sullivan, 2012)
Tree distance	Global	(Blanco-Pastor, Vargas, and Pfeil, 2012)
Network likelihood index	Global	(Konowalik et al., 2015)
Genealogical sorting index	Global	(de Villiers et al., 2013)
Shallow interspecific coalescences	Local	(Joly, McLenachan, and Lockhart, 2009)
Species network likelihood	Local	(Kubatko, 2009)
Minimize deep coalescences	Local	(Yu et al., 2011)
Species network likelihood	Local	(Yu, Degnan, and Nakhleh, 2012)
Four taxa nucleotide distances	Local	(Durand et al., 2011)
Five taxa nucleotide distances	Local	(Pease and Hahn, 2015)
Minimum pairwise sequence distance	Local	(Rosenzweig et al., 2016)
Sequence invariant patterns	Local	(Kubatko and Chifman, 2015; Blischak et al., 2018)
Species network pseudolikelihood	Local	(Solis-Lemus and Ane, 2016)

**Table 2.** Accessions used for this study.

<b>Species</b>	<b>Relationship</b>	<b>Voucher</b>	<b>Locality</b>	<b>DNA ID#</b>
<i>I. arborescens</i>	Putative parent	Smith 312 (MO)	Peru: Contumaza. 7.42409°S 78.90111°W	98
<i>I. arborescens</i>	Putative parent	Smith 209 (WIS)	Ecuador: Alluriquin. 0.32145°S 78.99764°W	250
<i>I. arborescens</i> <i>x I. cyaneum</i>	F1 Hybrid	Smith 687 (COLO)	University of Colorado- Boulder Greenhouses	-
<i>I. ayabacense</i>	Putative hybrid	Smith 337 (MO)	Peru: Ayabaca. 4.61462°W 79.71975°S	126
<i>I. cyaneum</i>	Putative parent	Smith 265 (WIS)	Univ. of Wisconsin- Madison Greenhouses	156
<i>I. lehmannii</i>	Putative parent	Smith 487 (MO)	Ecuador: Cañar 2.37168°W 78.96976°S	228
<i>I. lehmannii</i>	Putative parent	Smith 330 (MO)	Peru: Ayabaca. 4.64422°W 79.71975°S	176
<i>I. stenanthum</i>	Putative hybrid	Smith 313 (MO)	Peru: Contumaza. 7.40116°S 78.89658°W	99
<i>P. peruviana</i>	Outgroup	Smith 217 (MO)	Peru: Quito. 0.16761° S 78.48133° W	91

**Table 3.** Identification of hybrid relationships with HyDe, D-statistics and SNaQ. In (b), columns A, B, and C represent the phylogenetic position in a three tip relationship ((A,B),C) and the p-value indicates hybridization between A and C. For (a) and (b), the asterisks indicate significance after correction for multiple comparisons ( $\alpha = 0.05/30 = 0.0017$  for HyDe and  $\alpha = 0.05/10 = 0.005$  for D-statistics). For (c), the asterisks indicate significance in a likelihood ratio test with one degree of freedom (corresponding to the additional parameter introduced by the reticulation). Instances in which the method inferred a set of relationships consistent with the hypothesized hybrid ancestry are highlighted in gray.

<b>(a) HyDe</b>					
<b>Dataset</b>	<b>Hybrid</b>	<b>Parent 1</b>	<b>Parent 2</b>	<b>Test statistic</b>	<b>P</b>
1. No hybrids	<i>I. lehmannii</i> (176)	<i>I. arborescens</i> (250)	<i>I. lehmannii</i> (228)	3.18	0.002
2. One F <sub>1</sub>	<i>I. arborescens</i> (98)	F <sub>1</sub>	<i>I. lehmannii</i> (176)	5.08	0*
3. F <sub>1</sub> + <i>I. ayabacense</i>	<i>I. arborescens</i> (98)	<i>I. ayabacense</i>	<i>I. cyaneum</i> (156)	8.99	0*
4. <i>I. ayabacense</i> + <i>I. stenanthum</i>	<i>I. cyaneum</i> (156)	<i>I. lehmannii</i> (228)	<i>I. stenanthum</i>	8.64	0*

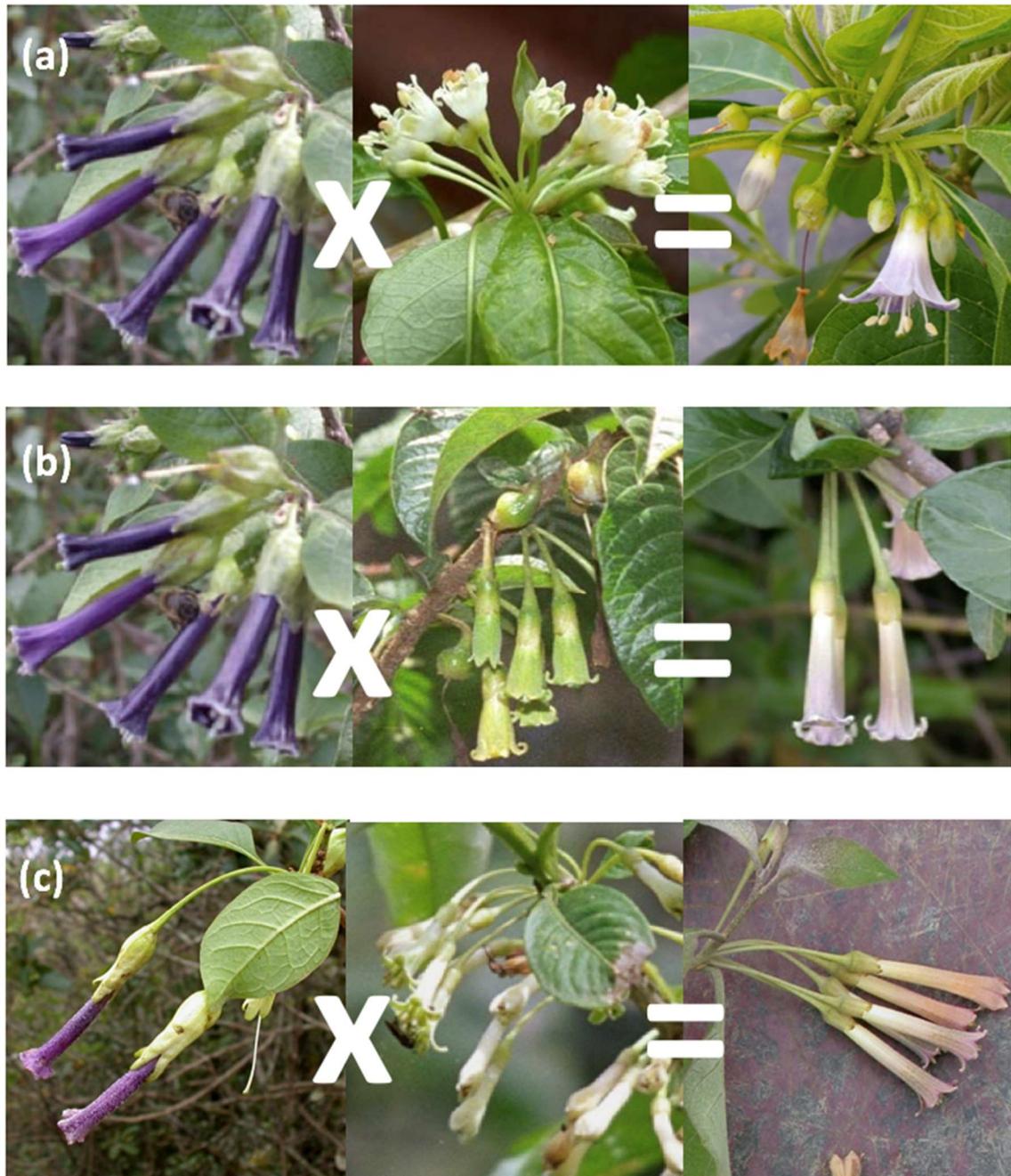
  

<b>(b) D-statistic analysis</b>					
<b>Dataset</b>	<b>A</b>	<b>B</b>	<b>C</b>	<b>Test statistic</b>	<b>P</b>
1. No hybrids	<i>I. lehmannii</i> (176)	<i>I. arborescens</i> (250)	<i>I. lehmannii</i> (228)	0.13	0.002*
2. One F <sub>1</sub>	F <sub>1</sub>	<i>I. arborescens</i> (98)	<i>I. lehmannii</i> (228)	0.42	0*
3. F <sub>1</sub> + <i>I. ayabacense</i>	F <sub>1</sub>	<i>I. ayabacense</i>	<i>I. arborescens</i> (98)	0.25	0*
4. <i>I. ayabacense</i> + <i>I. stenanthum</i>	<i>I. ayabacense</i>	<i>I. lehmannii</i> (228)	<i>I. cyaneum</i> (156)	0.29	0*

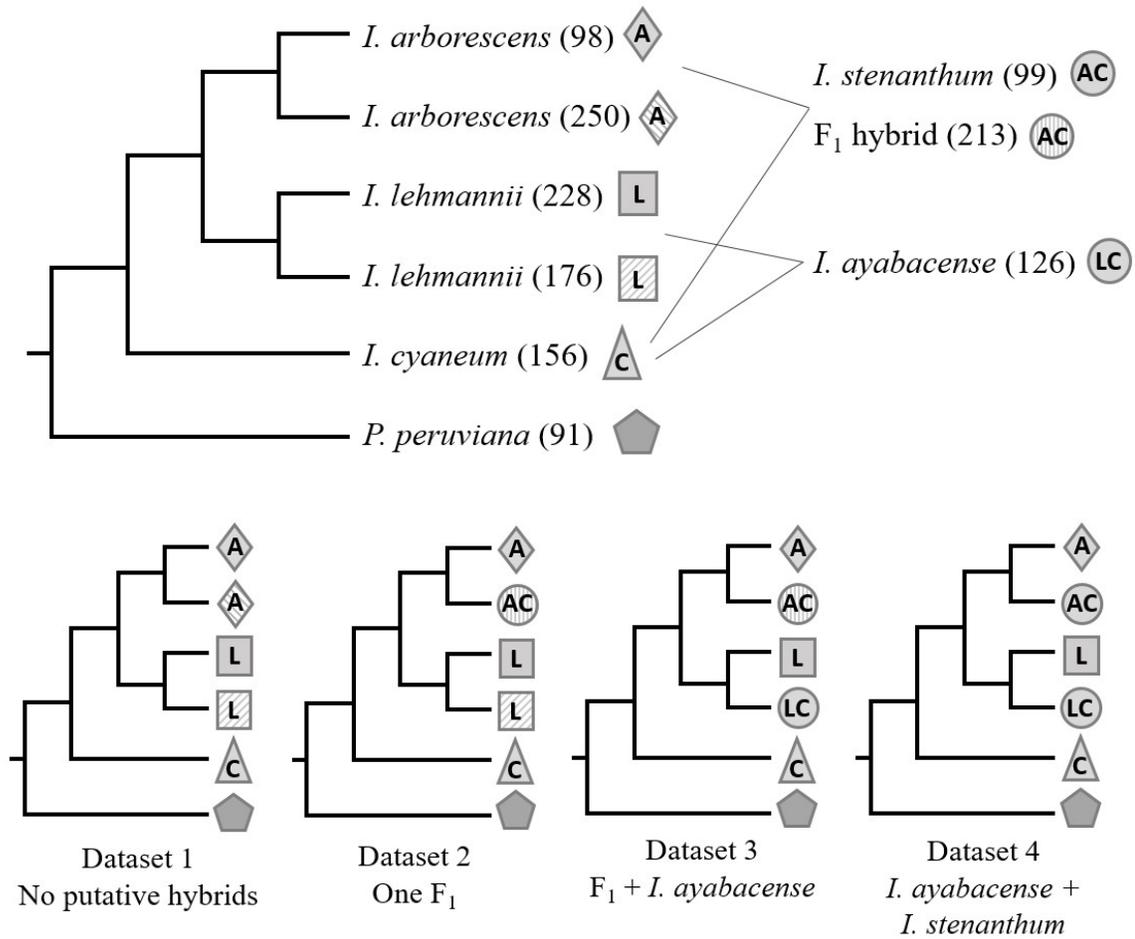
  

<b>(c) SNaQ analysis</b>					
<b>Dataset</b>	<b>Hybrid</b>	<b>Parent 1</b>	<b>Parent 2</b>	<b>Reticulation PLL</b>	<b>Tree PLL</b>
1. No hybrids	<i>I. lehmannii</i> (228)	<i>I. arborescens</i> (98)	<i>I. lehmannii</i> (176)	-1.86	-3.26
2. One F <sub>1</sub>	F <sub>1</sub>	<i>I. arborescens</i> (250)	<i>I. cyaneum</i> (156)	-2.11*	-26.80
3. F <sub>1</sub> + <i>I. ayabacense</i>	<i>I. lehmannii</i> (228)	<i>I. arborescens</i> (250)	<i>I. lehmannii</i> (228)	-31.99*	-91.32
4. <i>I. ayabacense</i> + <i>I. stenanthum</i>	<i>I. lehmannii</i> (228)	<i>I. arborescens</i> (250)	<i>I. lehmannii</i> (228)	-133.28*	-257.97

**Fig. 1. Known and hypothesized hybrid relationships.** (a) *Ichroma cyaneum* and *I. arborescens* were crossed in the greenhouse to create the F<sub>1</sub> hybrid on the right. (b) *I. cyaneum* and *I. lehmannii* are the putative parental taxa of *I. ayabacense*, right. (c) Members of the clade containing *I. cyaneum* and *I. cornifolium* (shown, left) and the clade containing *I. arborescens* and *I. confertiflorum* (middle) are the putative parental taxa of *I. stenanthum*, right.



**Fig. 2. Hypothesized relationships among sampled tips and experimental design.** The top tree shows the relationships among *I. arborescens*, *I. lehmannii*, and *I. cyaneum* based on previous studies (see text). Numbers indicate DNA accession number for reference (Table 2). The lines connect putative or known hybrid taxa (Fig. 1) to their putative parental lineages. The letter scheme indicates the putative parentals (AC denotes an A x C hybrid). Tests for hybridization were carried out on the four six-taxon datasets at the bottom, which range from no putative hybrids to two putative hybrids. See text for complete description.



**Fig. 3. Gene tree conflict and signal across experimental datasets.** Topologies for each dataset follow the species tree inferred by STAR. The symbols for tips follow Fig. 2, with known or putative hybrids indicated with two letters (AC or LC) corresponding to their putative parental lineages. Numbers above the branches are the numbers of gene trees estimated to have this relationship from concordance analysis. Proportions of gene trees (concordance factors) are in parentheses. Numbers below the branches denote internode certainty values, rounded to two decimal places. Tree certainty (TC) are given in the boxes for each tree along with the  $\chi^2$  value for two models (panmixia, P, and fully-bifurcating, B) tested by TICR. Values above 8 are significant at  $P=0.05$  (with three degrees of freedom, Stenz et al. 2015), suggesting that the data reject the model. For example, dataset 4 strongly rejects a bifurcating tree ( $\chi^2_B$  of 94) but does not reject the panmictic model ( $\chi^2_P$  of 1.7).

