

1 **Title**

2 The role of sexual isolation during rapid ecological divergence: evidence for a new dimension of isolation  
3 in *Rhagoletis pomonella*

4

5 **Running title**

6 Sexual isolation in early divergence

7

8 **Authors and affiliations**

9 Alycia C. R. Lackey<sup>1,2,5</sup>, Alyssa C. Murray<sup>2,3</sup>, Nadia A. Mirza<sup>2,4</sup>, Thomas H. Q. Powell<sup>2</sup>

10 <sup>1</sup> University of Louisville, Louisville, KY 40292

11 <sup>2</sup> Binghamton University, Binghamton, NY 13902

12 <sup>3</sup> Present Address: Tufts University, Medford, MA 02153

13 <sup>4</sup> Present Address: Boston University, Boston, MA 02118

14 <sup>5</sup> corresponding author, [alycia.lackey@louisville.edu](mailto:alycia.lackey@louisville.edu)

15

16 **Author contributions**

17 ACRL and THQP conceived of the study and experimental design. ACRL, ACM, and NAM conducted the  
18 experiment. ACRL and ACM analyzed the data. ACRL wrote the manuscript with contributions from  
19 THQP, ACM, and NAM.

20

21 **Acknowledgements**

22 This work was funded by an NSF grant (NSF 1639005) to THQP and D A. Hahn. Thanks to S. Berlocher  
23 and C. Giers for help with access to collection sites in Urbana, Illinois. We greatly appreciate help with

24 lab work and fly husbandry from M. Molina Mera, E. Romeo, D. Fama, I. Pyatetsky, A. Ahmed, A. Dukat,

25 S. Maung, and P. Deneen.

26

27 **Data accessibility statement**

28 Upon acceptance, data will be archived in Dryad.

29 **The role of sexual isolation during rapid ecological divergence: evidence for a new dimension of**  
30 **isolation in *Rhagoletis pomonella***

31

32 **Abstract**

33 The pace of divergence and likelihood of complete speciation may depend how and when different  
34 types of reproductive barriers evolve. After initial reproductive barriers evolve, questions remain about  
35 how subsequently evolving barriers may facilitate additional divergence and potential speciation. We  
36 tested for the presence of sexual isolation (reduced mating between populations due to divergent  
37 mating preferences and traits) in *Rhagoletis pomonella* flies, a model system for incipient ecological  
38 speciation. We measured the strength of sexual isolation between two very recently diverged (~170  
39 generations) sympatric populations, adapted to different host fruits. We found that sexual isolation was  
40 significantly stronger than expectations of random mating. Thus, sexual isolation may play an important  
41 role in reducing gene flow allowed by earlier-acting ecological barriers. We also tested how warmer  
42 temperatures predicted under climate change could alter sexual isolation and found that sexual  
43 isolation was markedly asymmetric between the sexes of each population when flies were reared under  
44 warmer temperatures. Our findings provide a window into the early divergence process and the role of  
45 sexual isolation after initial ecological divergence, in addition to examining how environmental  
46 conditions could shape the likelihood of further divergence.

47

48 **Keywords** reproductive isolation, sexual isolation, speciation, mating, asymmetry

49 **Introduction**

50 During the process of ecological speciation, adaptation to different environments can rapidly drive  
51 divergence (Schluter 2000, Nosil 2012). Yet, while ecological divergence can quickly differentiate  
52 populations, the speciation process frequently remains incomplete (Nosil et al. 2009, Marques et al.  
53 2019) or reversible (Seehausen et al. 1997, Lackey and Boughman 2017, Zhang et al. 2019). How rapidly  
54 or completely divergence proceeds depends on the strengths and types of reproductive barriers that  
55 evolve and when these barriers evolve during divergence (Coyne and Orr 2004, Lowry et al. 2008,  
56 Dopman et al. 2010, Schemske 2010, Lackey and Boughman 2017). Moreover, the coupling of multiple  
57 barrier traits may drive rapid transitions along the speciation continuum, promoting strong reproductive  
58 isolation and widespread genomic differentiation (Barton and De Cara 2009, Flaxman et al. 2014,  
59 Kunerth et al. 2022). Thus, understanding how multifaceted reproductive isolation develops along the  
60 speciation continuum following initial ecological divergence has important implications for the tempo of  
61 diversification.

62  
63 Theoretical and empirical work predicts that speciation is most likely to occur when divergent selection  
64 acts on both mating and non-mating traits (van Doorn et al. 2009, Maan and Seehausen 2011, Weissing  
65 et al. 2011, Wagner et al. 2012). Indeed, sexual isolation, reduced mating between populations due to  
66 divergent mating traits and preferences, can play an essential role during the speciation process. Sexual  
67 isolation often evolves early in divergence and can strongly facilitate speciation (Coyne and Orr 2004,  
68 Mendelson et al. 2007, Lackey and Boughman 2017). Sexual isolation is more likely to facilitate  
69 divergence when it coincides with other barriers (Butlin and Smadja 2018). Sexual isolation often occurs  
70 in conjunction with ecological isolation, and this combination characterizes many cases of rapid  
71 speciation (Boughman 2002, Ritchie 2007, Seehausen et al. 2008, Maan and Seehausen 2011). Ecological  
72 and sexual isolation may evolve rapidly in concert when direct selection acts on ecological and sexual

73 traits (e.g., habitat choice and environmentally-dependent signal production or fitness; McNett and  
74 Coccoft 2008, Boughman and Svanback 2017, Maan and Seehausen 2011, Nosil 2012, Safran et al. 2013,  
75 Scordato et al. 2014, Servedio and Boughman 2017). Additionally, the same trait(s) may shape both  
76 ecological and sexual barriers (Jiggins et al. 2001, Servedio et al. 2011). When sexual isolation occurs  
77 along with ecological isolation, it provides an opportunity to understand the relative roles and  
78 interdependence of these barriers, reveal the mechanisms currently shaping population differentiation,  
79 and potentially understand the origin and evolution of reproductive isolation. This is particularly true  
80 when studying populations in early stages of divergence and comparing them to populations at later  
81 stages along the speciation continuum.

82  
83 Predicting how quickly or completely isolation can evolve also involves evaluating how potential  
84 asymmetries in the strength of isolation between populations shape gene flow. Asymmetric  
85 reproductive isolation can result from differences between populations in the strength of selection on  
86 parental phenotypes or differences in fitness costs for hybrids that are stronger in one direction  
87 (Kaneshiro 1980, Arnold et al. 1996, Tiffin et al. 2001, Turelli and Moyle 2007, Kuwajima et al. 2010,  
88 Ribardiere et al. 2019, Zhang et al. 2022). Strong asymmetries may limit or reverse divergence (Arnold et  
89 al. 1996, Servedio and Kirkpatrick 1997, Chunco et al. 2007). While asymmetries may be common early  
90 in divergence, the extent of asymmetries may diminish as divergence proceeds and selection acts more  
91 symmetrically on each population or as incompatibilities arise (Turelli and Moyle 2007, Lackey and  
92 Boughman 2017). Even if asymmetries persist at later stages of divergence, their effects can be offset by  
93 complementary asymmetries in another barrier (Wade et al. 1995, Kitano et al. 2007, Takami et al.  
94 2007).

95

96 While divergent ecological selection can rapidly generate reproductive isolation, environmental  
97 sensitivity of reproductive barriers has important consequences for gene flow and the potential for  
98 distinct species to evolve and persist. Reproductive isolation that evolves due to divergent ecological  
99 selection may weaken if environmental differences decrease (Seehausen et al. 1997, Grant and Grant  
100 2008, Heath et al. 2010, Vonlanthen et al. 2012, Lackey and Boughman 2017). Sexual isolation may be  
101 particularly sensitive to environmental changes when differences in mating preferences and traits  
102 evolved due to environmental differences (Seehausen et al. 1997, Fisher et al. 2006, Ward and Blum  
103 2012, Lackey and Boughman 2013).

104  
105 Here, we leveraged a well-established study system in ecological speciation, the apple maggot fly,  
106 *Rhagoletis pomonella*, to evaluate how multifaceted reproductive isolation may evolve, particularly  
107 early in divergence. *Rhagoletis pomonella* is a textbook case of ecological speciation-in-action (Dres and  
108 Mallet 2002, Coyne & Orr 2004, Futuyma 2013). A population of these flies shifted from infesting the  
109 fruit of native downy hawthorn (*Crataegus mollis*) to introduced apple (*Malus pumila*) during the mid-  
110 19<sup>th</sup> century (Walsh 1861, Bush 1966) and divergent adaptation to these two host plants in the  
111 subsequent ~170 generations has led to substantial but incomplete reproductive isolation between the  
112 two host-associated populations of *R. pomonella* (Feder et al. 1988, 1994; Michel et al. 2010). The  
113 resulting consistent allele frequency differentiation between sympatric apple and hawthorn-infesting  
114 population pairs support the position of the derived apple fly at the hypothesized “host race” stage of  
115 ecological speciation in phytophagous insects (Berlocher and Feder 2002, Dres and Mallet 2002, Powell  
116 et al. 2013; 2022). The primary axes of divergent host plant adaptation driving reproductive isolation in  
117 this system are chemosensory adaptation to host fruit volatiles, which are the major cues for mating  
118 aggregation (Linn et al. 2003) and diapause-mediated life history timing corresponding to differences in  
119 fruiting phenology of the host plants (Filchak et al. 2000, Feder et al. 2010). These traits act as prezygotic

120 barriers to gene flow by restricting inter-host mating opportunities both spatially and temporally (Feder  
121 et al. 1994; Forbes et al. 2005) and as post-zygotic barriers via maladaptive phenotypes for both traits in  
122 F1 hybrids (Linn et al. 2004, Dambroski & Feder 2007). The divergence in these traits and their role in  
123 this incipient speciation system have been well characterized at the phenotypic, physiological, genetic,  
124 and, in the case of diapause, genomic levels (e.g., Dambroski et al. 2005; Forbes et al. 2005; Olsson et al.  
125 2006; Dambroski and Feder 2007, Tait et al. 2016; 2021, Powell et al. 2020; Dowle et al. 2020; Calvert et  
126 al. 2022).

127  
128 While habitat and temporal isolation strongly limit gene flow, apple and hawthorn flies can still  
129 encounter each other, and mark recapture estimates indicate gross migration of ~6% in sympatry (Feder  
130 et al. 1994). Whether this incomplete state of speciation is a transient phase in a still-progressing  
131 process or a long-term stalemate between divergent selection and migration remains unclear, but  
132 additional reproductive barriers may be necessary for additional divergence to accumulate (Ragland et  
133 al. 2015). Previous research found complete or nearly complete sexual isolation between highly  
134 divergent species pairs in the *Rhagoletis* genus (Hood et al. 2012), indicating that this barrier commonly  
135 contributes to the speciation process in these flies. In very recently diverged populations of *R.*  
136 *pomonella*, however, questions remain as to the presence and strength of sexual isolation as well as the  
137 potential forces that might underlie this barrier.

138  
139 Given the potential potency of sexual isolation acting in concert with known ecological isolation to drive  
140 rapid divergence, we made a novel extension of this classic study system to assess the contribution  
141 sexual isolation to limiting gene flow. First, we measured sexual isolation between recently diverged,  
142 sympatric populations of apple and hawthorn *R. pomonella* flies. Second, we examined potential  
143 asymmetries in sexual isolation by measuring the contribution of each sex from each population to

144 overall sexual isolation. Lastly, we tested whether rearing fly pupae under temperature regimes that  
145 mimic climate change predictions in the next 50-100 years affected mating interactions with  
146 consequences for the strength of sexual isolation as on-going speciation may be altered by  
147 anthropogenic change if reproductive barriers are environmentally sensitive.

148

## 149 **Methods**

### 150 *Insect collection and rearing*

151 We collected fruit infested with *Rhagoletis pomonella* flies from apple (*Malus pumila*) and hawthorn  
152 (*Crataegus mollis*) trees at a sympatric site in Urbana, Illinois in 2017. This sympatric population pair has  
153 been one of the most extensively studied in the *R. pomonella* species complex over the last four decades  
154 and provided one of the first population genetic confirmations of incipient sympatric speciation  
155 (McPheron et al. 1988). Since then, the apple and hawthorn populations in Urbana, IL have contributed  
156 to our understanding of the divergent adaption of chemosensory behavior (e.g., Linn et al. 2003; 2004;  
157 2005, Dambroski et al. 2005, Olsson et al. 2006) and diapause-mediated phenology (e.g., Dambroski &  
158 Feder 2007, Meyers et al. 2016, Powell et al. 2020, Dowle et al. 2020) as well as the population  
159 genomics of differentiation (e.g., Feder et al. 2003; Schwarz et al. 2009; Michel et al. 2010, Ragland et al.  
160 2017, Doellman et al. 2018; 2019, Dowle et al. 2020, Calvert et al. 2022). Thus, the patterns of ecological  
161 divergence and genetic relationship between the apple and hawthorn flies at this site are well-  
162 established, providing a robust foundation for testing for the presence of additional axes of divergence  
163 and reproductive isolation. We collected apples in mid-August and hawthorns in mid-September. We  
164 transported fruit to Binghamton University and maintained fruit at approximately 26°C with 14:10 L:D.  
165 We collected larvae that emerged from fruit daily for three weeks, following the natural emergence  
166 cycle. The flies used in this experiment were derived from a large-scale climate change simulation study  
167 testing for the effect of temperature on pupal developmental timing. Each day, we randomly assigned



168 larvae to two temperature regimes, Control and Warming, described below. We placed larvae into petri  
169 dishes with moist vermiculite in environmental chambers (Percival I41VLC9) with their assigned  
170 temperature regime for 10 days during the transition into the pupal phase. We then transferred viable  
171 pupae into individual 0.2 µl tubes and returned them to their assigned temperature regime until adult  
172 flies eclosed in the spring and summer of 2018.

173  
174 We created temperature regime programs using weekly average minimum, midpoint, and maximum  
175 temperatures calculated from soil temperature data from NOAA's National Climatic Data Center (NCDC)  
176 from 2007 to 2016 (Watseka, Illinois station: 40.79, -87.76). We used soil temperatures at a depth of  
177 10cm, which is the approximate depth of pupal *R. pomonella* during diapause (Feder 1995).  
178 Temperature programs and light:dark cycles replicated natural daily oscillations and weekly changes  
179 throughout the year (see Supplemental methods text for detail). We based the Control temperature  
180 regime on the 10-year weekly averages. Warming temperature regime set points were all 3°C higher  
181 than Control, which falls within the range of expected temperature increases for the Midwest in the  
182 next 50-100 years for multiple emission scenarios (Pryor et al. 2013). We monitored pupae daily for  
183 eclosion after winter programs.

184  
185 We housed newly eclosed flies individually in 50 mL Falcon tubes with food (3:1 sugar to yeast  
186 hydrolysate mixture, Neilson and McAllan 1964) and water for one day to allow for sclerotization of  
187 adult cuticles and wings. Then, flies were assigned to mating trials and painted with randomly assigned  
188 marking codes unique to each of 20 individuals within a trial. We used Testors™(Vernon Hills, Illinois,  
189 USA) enamel paint for marking, and we briefly anesthetized flies on carbon dioxide blocks to apply paint.  
190 Flies were then housed in clear plastic containers with mesh tops (approximately 1L) in same-sex groups  
191 of up to five with food and water *ad libitum* and kept at approximately 26°C and 14:10 L:D cycle.

192

193 *Mating trials*

194 We used multiple choice mating trails with 5 males and 5 females of each population to test whether  
195 copulation is more likely to occur within versus between populations. This design mimics natural  
196 conditions where flies aggregate on host plants to mate (Prokopy 1976, Aluja et al. 2001). Trials with  
197 multiple males and females allow both sexes to engage in mate choice. Thus, we used this design to  
198 measures overall sexual isolation and the contributions of each sex from each population.

199

200 We conducted a mating trial once all flies assigned to a trial had reached reproductive maturity (at least  
201 10 days old; Neilson and McAllan 1965). For each trial we assigned 5 males and 5 females of each  
202 population (Apple and Hawthorn) reared under the same temperature regime (Control or Warming).

203 While we initially assigned 5 flies of each sex from each population to trials, some trials had 4-6 flies of  
204 each sex and population due to early mortality and one case of misassignment. In our analysis, we  
205 accounted for sample size variation in expectations of random mating. We conducted 3-hour mating  
206 trials in tent-shaped enclosures with clear plastic and white mesh sides (BugDorm2™, MegaView  
207 Science Education Services LTD, Taiwan; 61 x 61 x 61cm). Each tent contained two water and two food  
208 stations as well as an apple as a mating stimulus. Both Apple and Hawthorn flies mate readily on and  
209 oviposit into apples in lab trials (Linn et al. 2004, Lyons-Sobaski and Berlocher 2009). In our study,  
210 copulations occurred throughout the mating tent and rarely directly on the fruit. We introduced flies to  
211 the mating arena by allowing them to fly out of their opened housing enclosures. We introduced  
212 females first and allowed them to acclimate for 10 minutes before introducing males. We observed up  
213 to 4 mating trials concurrently during each 3-hour observation using scan sampling. For every attempted  
214 copulation (one fly mounts the other), we recorded copulation duration and identity of the interacting  
215 flies using paint marks. Males typically initiate mating by jumping on the female's back (Smith and

216 Prokopy 1982). Females can resist and dislodge males or accept a mating attempt by extending her  
217 ovipositor. Copulations longer than 5 minutes were categorized as successful (Hood et al. 2012).  
218 Copulations typically last at least 20 minutes (Smith and Prokopy 1982, Schwarz and McPherson 2007).

219

220 *Data analysis*

221 Sexual isolation

222 We calculated sexual isolation using the following equation (Sobel and Chen 2014):

223 
$$SI = 1 - 2 \left( \frac{H}{C+H} \right) \quad (1)$$

224 where  $H$  is the frequency of heterospecific, or between-population, events and  $C$  is the frequency of  
225 conspecific, or within-population events.  $SI$  ranges linearly from -1 (mating only between populations) to  
226 0 (random mating) to 1 (mating only within populations). To account for variation in the number of  
227 males and females of each population in each trial, we calculated expected copulations for each pair  
228 type (Apple female x Apple male, Apple female x Hawthorn male, Hawthorn female x Apple male,  
229 Hawthorn female x Hawthorn male) based on random mating null expectations. For each sex of each  
230 population, we divided the total number of copulations that group had with flies of the opposite sex  
231 from either population with 50:50 mating expectations given the number of Apple males and Hawthorn  
232 males in a trial. For example, if Apple females in a trial had 4 copulations, and there were equal numbers  
233 of Apple (5) and Hawthorn (5) males, then the expected number of copulations given random mating  
234 would be 2 Apple female x Apple male and 2 Apple female x Hawthorn male. If there were unequal  
235 numbers of males (5 Apple, 4 Hawthorn), then the expected number of copulations would be 2.22 Apple  
236 female x Apple male and 1.78 Apple female x Hawthorn male. We used these expected copulations in  
237 the following equation (Sobel and Chen 2014):

238 
$$SI = 1 - 2 \left( \frac{\frac{H_{obs}}{H_{exp}}}{\frac{C_{obs}}{C_{exp}} + \frac{H_{obs}}{H_{exp}}} \right), \quad (2)$$

239 where observed events (*obs*) were divided by expected events (*exp*). We calculated 95% confidence  
240 intervals for total sexual isolation using 127 total copulations as the sample size. To calculate 95%  
241 confidence intervals for the contributions of each sex to sexual isolation, we used the following sample  
242 sizes: 53 copulations with Apple females, 70 copulations with Apple males, 74 copulations with  
243 Hawthorn females, and 57 copulations with Hawthorn males.

244  
245 We also calculated sexual isolation separately by rearing temperature (Control or Warming) to assess  
246 environmental effects on the total strength of SI and the contribution of each sex from each population.  
247 For Control temperatures, the sample sizes were 69 copulations total with 28 copulations with Apple  
248 females, 37 copulations with Apple males, 41 copulations with Hawthorn females, and 32 copulations  
249 with Hawthorn males. For Warming temperatures, the sample sizes were 58 copulations total with 25  
250 copulations with Apple females, 33 copulations with Apple males, 33 copulations with Hawthorn  
251 females, and 25 copulations with Hawthorn males.

252

### 253 Comparing prezygotic isolating barriers

254 To place the strength of sexual isolation in context of other prezygotic barriers linked to divergent  
255 adaptation to different host plants, we measured the strength of temporal and habitat isolation from  
256 existing data. Data for temporal isolation were calculated for Apple and Hawthorn flies reared under  
257 control temperatures (Lackey et al. *in prep*). For habitat isolation, we used data from fruit volatile  
258 preferences in flight tunnels (Linn et al. 2003). After emergence, flies may travel several kilometers to  
259 locate host plants, and fruit volatiles are the major long-range stimulus attracting flies (Maxwell and  
260 Parsons 1968, Linn et al. 2003). We calculated 95% confidence intervals for each barrier. Next, we  
261 calculated the sequential strength of each barrier ordered by their occurrence in the life cycle (i.e.,  
262 temporal, habitat, sexual). The sequential strength of each barrier ( $SS_n$ ) is calculated from its individual

263 strength ( $RI_n$ ) and the amount of gene flow allowed by earlier-acting barriers (Ramsey et al. 2003,  
264 Dopman et al. 2010, Sobel and Chen 2014):

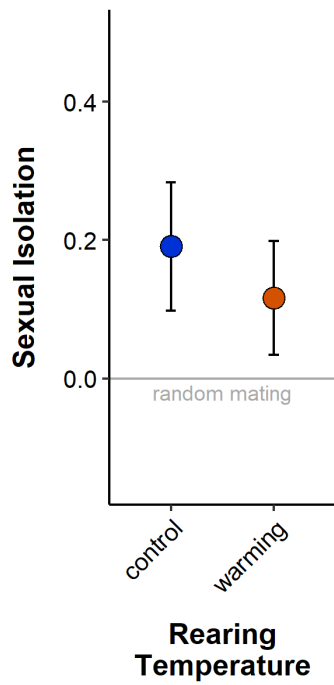
$$265 \quad SS_n = RI_n (1 - \sum_{i=1}^{n-1} SS_i). \quad (3)$$

266

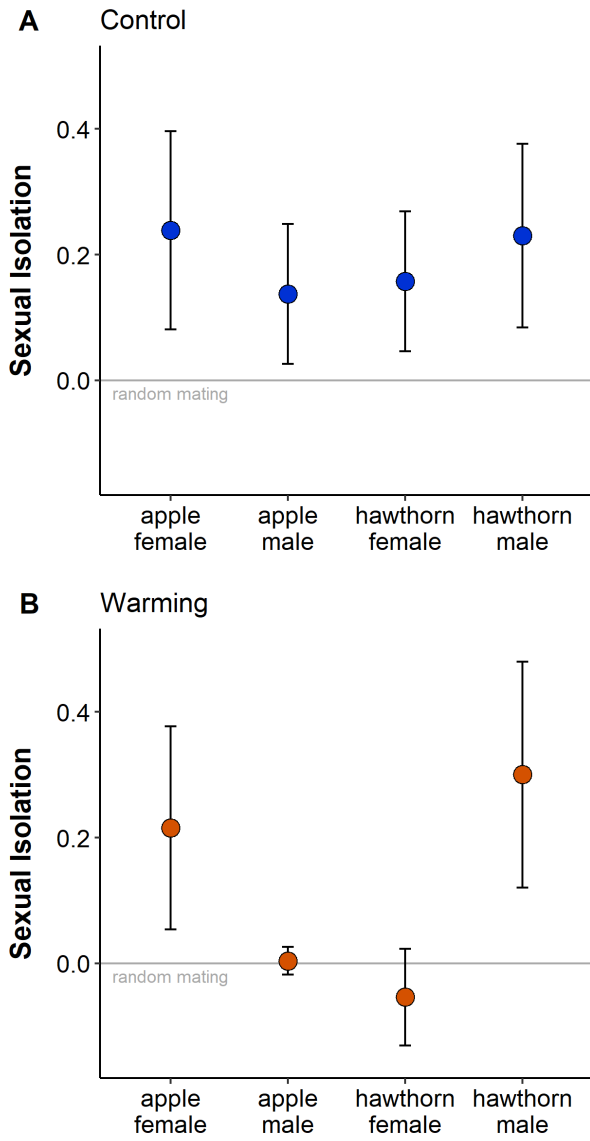
## 267 **Results**

### 268 *Sexual isolation*

269 Sexual isolation between Apple and Hawthorn flies was significantly greater than expectations of  
270 random mating, where isolation is zero, in both Control and Warming rearing treatments, (Control: SI =  
271 0.191 [95%CI: 0.284 - 0.093], Warming: SI = 0.116 [95%CI: 0.199 - 0.034], Figure 1, Supplemental Table  
272 1). Given the overlap of 95% confidence intervals, the strength of sexual isolation did not differ between  
273 temperature treatments (Figure 1, Supplemental Table 1). However, the pattern of the contributions to  
274 total sexual isolation from each sex of each population differed between temperature treatments. In the  
275 Control treatment, all flies mated within population more than between population, and contributions  
276 to sexual isolation from each sex from each population were significantly greater than expectations of  
277 random mating (Figure 2A). In the Warming treatment, in contrast, Apple females and Hawthorn males  
278 mated within population more than between population with measures of sexual isolation significantly  
279 greater than 0, while Apple males and Hawthorn females mated randomly within and between  
280 population (Figure 2B, Supplemental Table 1, Supplemental Figure 1).



281  
 282 *Figure 1. Total sexual isolation for flies reared under Control (blue) and Warming (orange) temperatures.*  
 283 *Circles are point values with 95% CIs. The horizontal grey line at 0 indicates random mating, and positive*  
 284 *values indicate greater mating within populations than between. Values for barrier strengths and 95%*  
 285 *CIs are provided in Supplemental Table 1.*



286  
 287 *Figure 2. Contributions of each sex from each population to sexual isolation for flies reared under Control*  
 288 *(blue) and Warming (orange) temperatures. Circles are point values with 95% CIs. The horizontal grey*

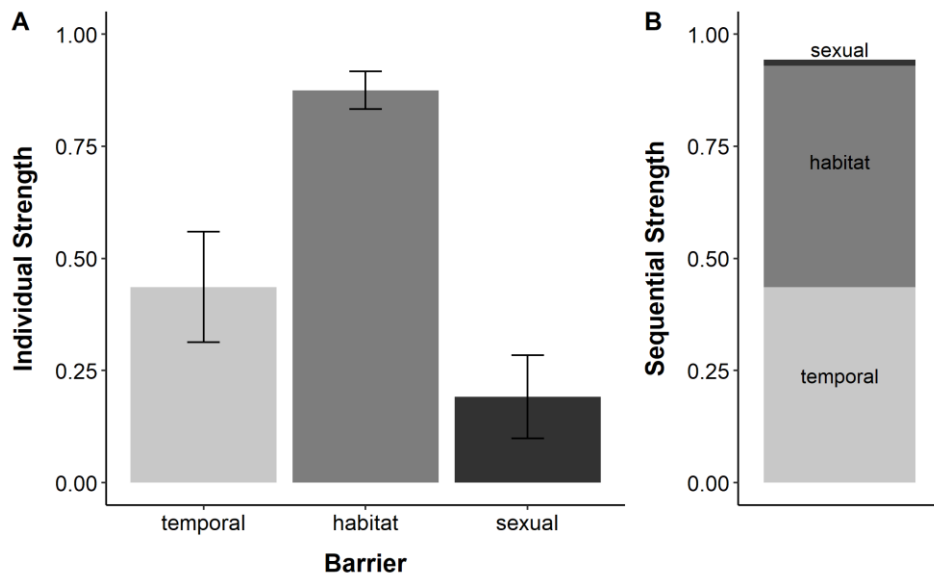
289 *line at 0 indicates random mating, and positive values indicate greater mating within populations than*  
290 *between. Values for barrier strengths and 95% CIs are provided in Supplemental Table 1.*

291

### 292 *Comparing prezygotic isolating barriers*

293 Individual strengths of isolating barriers estimate the proportion of gene flow limited by each barrier if  
294 acting alone. We compared total sexual isolation from Control rearing conditions calculated in this study  
295 to measures of temporal isolation from our unpublished data on eclosion timing and habitat isolation  
296 from previously published work on attraction preference to host fruit volatiles. Temporal isolation was  
297 moderate in strength (RI = 0.44, 95% CI: 0.31 – 0.56). Habitat isolation was the strongest of the three  
298 barriers we estimated (RI = 0.87, 95% CI: 0.83 – 0.92). Sexual isolation was relatively weaker than the  
299 other barriers (RI = 0.15, 95% CI: 0.08 – 0.21), though significantly stronger than expectations of random  
300 mating (RI = 0). The sequential strengths of isolating barriers ordered as each barrier occurs in the life  
301 cycle estimate the proportion of gene flow limited by each barrier given the gene flow allowed by  
302 earlier-acting barriers. Together, temporal and habitat isolation were estimated to limit 93% of potential  
303 gene flow (RI = 0.93). Sexual isolation would strengthen total RI to 0.94, a 14% change in gene flow  
304 allowed.





305  
 306 *Figure 3. (A) Individual and (B) sequential strengths of three prezygotic barriers. Error bars in A are 95%*  
 307 *confidence intervals. Values for barrier strengths and 95% CIs are provided in Supplemental Table 2, and*  
 308 *sexual isolation calculations use mating interactions for flies reared under Control temperatures.*

309  
 310 **Discussion**  
 311 Studying populations early in the process of divergence provides opportunities to measure reproductive  
 312 barriers as they accumulate and detect the evolutionary forces producing isolation (Nosil et al. 2005,  
 313 Merrill et al. 2011, Powell et al. 2014, Hood et al. 2020). In addition to measuring the overall strength of  
 314 reproductive barriers, determining the strength of asymmetries provides insights into the underlying  
 315 evolutionary processes and understand the nature of how reproductive isolation evolves (Arnold et al.  
 316 1996, Servedio and Kirkpatrick 1997, Lackey and Boughman 2017). Moreover, estimating environmental  
 317 sensitivity of reproductive barriers enables predictions of the stability of divergence in the face of

Commented [AL1]: pick up proofreading here

318 environmental change, which is especially important when divergence is primarily driven by  
319 environmental differences.

320

321 Here, we tested for the presence of sexual isolation, a barrier often important in early stages of  
322 divergence, using a well-established case study of rapid divergence with gene flow. Between two very  
323 recently diverged populations of apple and hawthorn flies, we have identified the presence of a new  
324 dimension of reproductive isolation, sexual isolation, that has evolved within ~170 generations. We  
325 found (1) that the strength of sexual isolation was significantly greater than expectations of random  
326 mating, and (2) sexual isolation was symmetric between the sexes of each population for flies reared  
327 under control conditions but asymmetric for flies were reared under warmer temperatures.

328

329 Between apple and hawthorn flies, we provide evidence that sexual isolation could limit approximately  
330 19% of gene flow. While sexual isolation is relatively weaker than habitat and temporal isolation, it may  
331 play an important role in restricting the homogenizing effects of gene flow and, thus, facilitate  
332 divergence. Considering the sequential and combined effects of multiple barriers, temporal and habitat  
333 isolation allow 7% gene flow. Adding sexual isolation reduces potential gene flow to 6%, which is  
334 consistent with estimated gross migration in the field, based on mark-recapture studies (6%, Feder et al.  
335 1994). From the perspective of remaining potential gene flow, the 1% increase in total reproductive  
336 isolation may represent a biologically meaningful reduction. Our observed effect of sexual isolation cuts  
337 the potential remaining gross migration rate by 14% ( $m = 0.07$  to  $0.06$ ). Such incremental reductions in  
338 migration rates may have considerable consequences for migration-selection equilibria (Yeaman and  
339 Whitlock 2011) and may nudge systems closer to “tipping points” after which the pace of divergence  
340 increases rapidly to form reproductively isolated species (Flaxman et al. 2014, Nosil et al. 2017, Schilling  
341 et al. 2018). Moreover, selection on traits that yield sexual isolation may also increase the extent of

342 genome-wide differentiation, strengthening the likelihood of complete and stable speciation (Nosil and  
343 Feder 2012, Kautt et al. 2020).

344

345 The current strength of sexual isolation between apple and hawthorn flies suggests an increase in  
346 isolation compared to an estimate from 30 years ago that found no sexual isolation between different  
347 host-associated populations of *Rhagoletis pomonella* (Smith 1988). Thus, sexual isolation may have  
348 evolved rapidly early in divergence. Across *Rhagoletis* species, sexual isolation increases in strength from  
349 weak to strong as divergence between species increases (Smith 1988, Schwarz and McPherson 2007,  
350 Hood et al. 2012). Notably, geographic isolation alone may be insufficient for the evolution of sexual  
351 isolation; sexual isolation was absent between a pair of populations using the same host plant despite  
352 1.5 million years of geographic isolation (Rull et al. 2010). In the *R. pomonella* species complex,  
353 divergent specialization to different host plants has primarily driven divergence and resulted in  
354 ecological reproductive isolation between populations through divergent life history timing and  
355 olfactory behavioral responses to fruit volatiles (Berlocher 2000, Linn et al. 2005, Dambroski and Feder  
356 2007, Linn et al. 2012, Mattsson et al. 2021). Yet, it is currently unknown whether sexual isolation  
357 evolves in association with host adaptation or independently.

358

359 In ecological speciation, barriers under direct divergent selection evolve first, and subsequent barriers  
360 can evolve as a by-product of divergent adaptation or independently (Schluter 2001, Dieckmann and  
361 Doebeli 2004, Rundle and Nosil 2005). Determining how subsequent barriers evolve is important for  
362 predicting how rapidly divergence can occur (Smadja and Butlin 2011). Sexual isolation can evolve as a  
363 by-product of ecological adaptation to different host fruits if traits under divergent ecological selection  
364 are also mating traits (Servedio et al. 2011). Sexual isolation may also evolve via reinforcement when  
365 selection against costly matings between populations favors the evolution of prezygotic isolation

366 (Servedio and Noor 2003). In *R. pomonella*, F<sub>1</sub> hybrids may suffer an ecological fitness disadvantage due  
367 to reduced responses to host fruit volatiles critical for locating host fruit for reproduction (Linn et al.  
368 2004). Such fitness costs could favor selection for strong mating discrimination via reinforcement. Lastly,  
369 sexual isolation could evolve due to population differences in selection along axes independent of  
370 primary ecological differences (e.g., non-ecologically mediated sexual selection or sexual conflict, Turbek  
371 et al. 2021, Rundle and Rowe 2018) or via non-selective evolutionary processes (e.g., mutation order,  
372 Mendelson et al. 2014). Indeed, species maintenance is more likely when at least some reproductive  
373 barriers evolve independently of environmental differences (Coyne and Orr 2004, Lackey and Boughman  
374 2017). In *Rhagoletis*, future work is needed to determine the extent to which sexual isolation may result  
375 from ecological or non-ecological factors.

376  
377 When isolation is symmetric, gene flow is limited similarly by both populations, and this bi-directional  
378 reduction in gene flow yields more stable isolation. Asymmetric isolation, in contrast, allows gene flow  
379 more in one direction than another between populations and can limit further divergence and halt or  
380 reverse the speciation process, especially if asymmetric isolation persists in later stages of divergence  
381 (Arnold et al. 1996, Servedio and Kirkpatrick 1997, Chunco et al. 2007). In this study, flies reared under  
382 control temperatures, showed no asymmetry in sexual isolation; each sex from each population mated  
383 more within population than between. Thus, sexual isolation can limit gene flow similarly between both  
384 populations. Under warming conditions, however, sexual isolation was asymmetric between the sexes  
385 of each population. Apple males and hawthorn females mated randomly while apple females and  
386 hawthorn males mated more within population than between. Though warmer rearing temperatures  
387 did not change the overall strength of sexual isolation between populations, asymmetric contributions  
388 to total sexual isolation under warming conditions could facilitate asymmetric gene flow. Sexual  
389 isolation may be particularly sensitive to environmental changes when differences in mating preferences

390 and traits evolved due to environmental differences (Seehausen et al. 1997, Fisher et al. 2006, Ward and  
391 Blum 2012, Lackey and Boughman 2013). Potentially, plasticity in mating traits and preferences that  
392 determine the likelihood of mating within or between population could shape the strength and/or  
393 symmetric of sexual isolation (e.g., Jin et al. 2022).

394  
395 In this study, we provide evidence of a new dimension of reproductive isolation between recently  
396 diverged populations of *R. pomonella*. Members of the *R. pomonella* species complex have undergone a  
397 rapid adaptive radiation primarily due to divergent ecological adaptation (Bush 1966, Berlocher 2000,  
398 Powell et al. 2013). However, reproductive isolation is incomplete between recently diverged  
399 populations in this complex (Powell et al. 2013, Arcella et al. 2015, Inskeep et al. 2021). Thus, ecological  
400 divergence alone may be insufficient to complete speciation (e.g., Nosil et al. 2009). Sexual isolation may  
401 play an important role in reducing gene flow to an extent that facilitates further divergence and  
402 potential speciation. This study emphasizes the importance of understanding the strength and evolution  
403 of reproductive barriers that evolve after initial divergence and the role of these barriers in population  
404 divergence.

405 **References**

- 406 Aluja, M., N. Lozada, J. Pinero, A. Birke, V. Hernandez-Ortiz, and F. Diaz-Fleischer. 2001. Basic behavior of  
407 *Rhagoletis turpiniae* (Diptera: Tephritidae) with comparative notes on the sexual behavior of  
408 *Rhagoletis pomonella* and *Rhagoletis zoqui*. *Annals of the Entomological Society of America*  
409 **94**:268-274.
- 410 Arcella, T., G. R. Hood, T. H. Powell, S. B. Sim, W. L. Yee, D. Schwarz, S. P. Egan, R. B. Goughnour, J. J.  
411 Smith, and J. L. Feder. 2015. Hybridization and the spread of the apple maggot fly, *Rhagoletis*  
412 *pomonella* (Diptera: Tephritidae), in the northwestern United States. *Evol Appl* **8**:834-846.
- 413 Arnold, S. J., P. A. Verrell, and S. G. Tilley. 1996. The evolution of asymmetry in sexual isolation: a model  
414 and a test case. *Evolution* **50**:1024-1033.
- 415 Barton, N. H., and M. A. R. De Cara. 2009. The evolution of strong reproductive isolation. *Evolution* **63**:  
416 1171– 1190.
- 417 Benjamini, Y., and Y. Hochberg. 1995. Controlling the false discovery rate: a practical and powerful  
418 approach to multiple testing. *Journal of the Royal Statistical Society B* **57**:289-300.
- 419 Berlocher, S. H. 2000. Radiation and Divergence in the *Rhagoletis Pomonella* Species Group: Inferences  
420 from Allozymes. *Evolution* **54**.
- 421 Berlocher, S. H., and J. L. Feder. 2002. Sympatric speciation phytophagous insects: moving beyond  
422 controversy? *Annual Review of Entomology* **47**:773-815.
- 423 Boughman, J. W. 2002. How sensory drive can promote speciation. *Trends in Ecology & Evolution*  
424 **17**:571-577.
- 425 Boughman, J. W., and R. Svanback. 2017. Synergistic selection between ecological niche and mate  
426 preference primes diversification. *Evolution* **71**:6-22.
- 427 Bush, G. L. 1966. The taxonomy, cytology and evolution of the genus *Rhagoletis* in North America  
428 (Diptera:Tiphritidae). Museum of Comparative Zoology, Cambridge, Massachusetts.

429 Butlin, R. K., and C. M. Smadja. 2018. Coupling, Reinforcement, and Speciation. *Am Nat* **191**:155-172.

430 Calvert, M. B., Doellman, M. M., Feder, J. L., Hood, G. R., Meyers, P., Egan, S. P., Powell, T.H.Q., Glover,  
431 M.M., C. Tait, S.H. Berlocher, P. Nosil, J.J. Smith, Hahn, D.A., Ragland, G. J. 2022. Genomically  
432 correlated trait combinations and antagonistic selection contributing to counterintuitive genetic  
433 patterns of adaptive diapause divergence in *Rhagoletis* flies. *Journal of evolutionary biology* **35**:  
434 146-163.

435 Chunco, A. J., J. S. McKinnon, and M. R. Servedio. 2007. Microhabitat variation and sexual selection can  
436 maintain male color polymorphisms. *Evolution* **61**:2504-2515.

437 Coyne, J. A., and H. A. Orr. 2004. *Speciation*. Sinauer Associates, Inc., Sunderland, Massachusetts.

438 Dambroski, H. R., Linn Jr, C., Berlocher, S. H., Forbes, A. A., Roelofs, W., & Feder, J. L. 2005. The genetic  
439 basis for fruit odor discrimination in *Rhagoletis* flies and its significance for sympatric host shifts.  
440 *Evolution* **59**: 1953-1964.

441 Dambroski, H. R., and J. L. Feder. 2007. Host plant and latitude-related diapause variation in *Rhagoletis*  
442 *pomonella*: a test for multifaceted life history adaptation on different stages of diapause  
443 development. *J Evol Biol* **20**:2101-2112.

444 Davis, J. S., M. J. Pearcy, J. Y. Yew, and L. C. Moyle. 2021. A shift to shorter cuticular hydrocarbons  
445 accompanies sexual isolation among *Drosophila americana* group populations. *Evol Lett* **5**:521-  
446 540.

447 Dieckmann, U., and M. Doebeli. 2004. Adaptive dynamics of speciation: sexual populations. *in* U.  
448 Dieckmann, M. Doebeli, J. A. J. Metz, and D. Tautz, editors. *Adaptive Speciation*. Cambridge  
449 University Press, Cambridge.

450 Dopman, E. B., P. S. Robbins, and A. Seaman. 2010. Components of reproductive isolation between  
451 North American pheromone strains of the European corn borer. *Evolution* **64**:881-902.

452 Drès, M., and J. Mallet. 2002. Host races in plant-feeding insects and their importance in sympatric  
453 speciation. *Philosophical Transactions of the Royal Society London Series B* **357**: 471-492.

454 Dowle, E. J., T. H.Q. Powell, M. M. Doellman, P. J. Meyers, M. B. Calvert, K. K. O. Walden, H. M.  
455 Robertson, S. H. Berlocher, J. L. Feder, D. A. Hahn, and G. J. Ragland. 2020. Genome-wide  
456 variation and transcriptional changes in diverse developmental processes underlie the rapid  
457 evolution of seasonal adaptation. *Proceedings of the National Academy of Sciences* **117**: 23960-  
458 23969.

459 Feder, J. L. 1995. The effects of parasitoids on sympatric host races of *Rhagoletis pomonella*  
460 (Diptera:Tephritidae). *Ecology* **76**:801-813.

461 Feder, J. L., C. A. Chilcote, and G. L. Bush. 1988. Genetic differentiation between sympatric host races of  
462 the apple maggot fly *Rhagoletis pomonella*. *Nature* **336**:61-64.

463 Feder, J. L., S. B. Opp, B. Wlazlo, K. Reynolds, W. Go, and S. Spisak. 1994. Host fidelity is an effective  
464 pre-mating barrier between sympatric races of the apple maggot fly. *Proceedings of the*  
465 *Academy of Natural Sciences of the United States of America* **91**:7990-7994.

466 Feder, J. L., T. H. Q. Powell, K. Filchak, and B. Leung. 2010. The diapause response of *Rhagoletis*  
467 *pomonella* to varying environmental conditions and its significance for geographic and host  
468 plant-related adaptation. *Entomologia Experimentalis Et Applicata* **136**:31-44.

469 Fisher, H. S., B. B. Wong, and G. G. Rosenthal. 2006. Alteration of the chemical environment disrupts  
470 communication in a freshwater fish. *Proc Biol Sci* **273**:1187-1193.

471 Flaxman, S. M., A. C. Wacholder, J. L. Feder, and P. Nosil. 2014. Theoretical models of the influence of  
472 genomic architecture on the dynamics of speciation. *Molecular Ecology* **23**:4074-4088.

473 Futuyma, D. J. 2013. *Evolution*, 3<sup>rd</sup> Edition. Sinauer Associates, Inc., Sunderland, Massachusetts.

474 Grant, B. R., and P. R. Grant. 2008. Fission and fusion of Darwin's finches populations. *Philos Trans R Soc*  
475 *Lond B Biol Sci* **363**:2821-2829.



476 Heath, D., C. M. Bettles, and D. Roff. 2010. Environmental factors associated with reproductive barrier  
477 breakdown in sympatric trout populations on Vancouver Island. *Evol Appl* **3**:77-90.

478 Hood, G. R., S. P. Egan, and J. L. Feder. 2012. Evidence for sexual isolation as a prezygotic barrier to gene  
479 flow between morphologically divergent species of *Rhagoletis* fruit flies. *Ecological Entomology*  
480 **37**:521-528.

481 Hood, G. R., T. H. Q. Powell, M. M. Doellman, S. B. Sim, M. Glover, W. L. Yee, R. B. Goughnour, M.  
482 Mattsson, D. Schwarz, and J. L. Feder. 2020. Rapid and repeatable host plant shifts drive  
483 reproductive isolation following a recent human-mediated introduction of the apple maggot fly,  
484 *Rhagoletis pomonella*. *Evolution* **74**:156-168.

485 Inskip, K. A., M. M. Doellman, T. H. Q. Powell, S. H. Berlocher, N. R. Siefert, G. R. Hood, G. J. Ragland, P.  
486 J. Meyers, and J. L. Feder. 2021. Divergent diapause life history timing drives both allochronic  
487 speciation and reticulate hybridization in an adaptive radiation of *Rhagoletis* flies. *Molecular*  
488 *Ecology*.

489 Jiggins, C. D., R. E. Naisbit, R. L. Coe, and J. Mallet. 2001. Reproductive isolation caused by colour pattern  
490 mimicry. *Nature* **411**:302-305.

491 Jin, B., D. A. Barbash, D. M. Castillo. 2022. Divergent selection on behavioral and chemical traits between  
492 reproductively isolated populations of *Drosophila melanogaster*. *Journal of Evolutionary Biology*.  
493 doi: 10.1111/jeb.14007.

494 Kaneshiro, K. Y. 1980. Sexual isolation, speciation and the direction of evolution. *Evolution* **34**:437-444.

495 Kautt, A. F., C. F. Kratochwil, A. Nater, G. Machado-Schiaffino, M. Olave, F. Henning, J. Torres-Dowdall, A.  
496 Harer, C. D. Hulse, P. Franchini, M. Pippel, E. W. Myers, and A. Meyer. 2020. Contrasting  
497 signatures of genomic divergence during sympatric speciation. *Nature* **588**:106-111.

498 Kitano, J., S. Mori, and C. L. Peichel. 2007. Phenotypic divergence and reproductive isolation between  
499 sympatric forms of Japanese threespine sticklebacks. *Biological Journal of the Linnean Society*  
500 **91**:671-685.

501 Kunerth, H.D., S.M. Bogdanowicz, J.B. Searle, R.G. Harrison, B.S. Coates, G.M. Kozak, and E.B. Dopman.  
502 2022. Consequences of coupled barriers to gene flow for the build-up of genomic differentiation.  
503 *Evolution*. <https://doi.org/10.1111/evo.14466>

504 Kuwajima, M., N. Kobayashi, T. Katoh, and H. Katakura. 2010. Detection of ecological hybrid inviability in  
505 a pair of sympatric phytophagous ladybird beetles (*Henosepilachna* spp.). *Entomologia*  
506 *Experimentalis Et Applicata* **134**:280-286.

507 Lackey, A. C., and J. W. Boughman. 2017. Evolution of reproductive isolation in stickleback fish. *Evolution*  
508 **71**:357-372.

509 Lackey, A. C. R., and J. W. Boughman. 2013. Loss of sexual isolation in a hybridizing stickleback species  
510 pair. *Current Zoology* **59**:591-603.

511 Lande, R., and M. Kirkpatrick. 1988. Ecological speciation by sexual selection. *Journal of Theoretical*  
512 *Biology* **133**:85-98.

513 Linn, C., J. L. Feder, S. Nojima, H. R. Dambroski, S. H. Berlocher, and W. Roelofs. 2003. Fruit odor  
514 discrimination and sympatric host race formation in *Rhagoletis*. *Proceedings of the Academy of*  
515 *Natural Sciences of the United States of America* **100**:11490-11493.

516 Linn, C., S. Nojima, and W. Roelofs. 2005. Antagonist effects of non-host fruit volatiles on discrimination  
517 of host fruit by *Rhagoletis* flies infesting apple (*Malus pumila*), hawthorn (*Crataegus* spp.), and  
518 flowering dogwood (*Cornus florida*). *Entomologia Experimentalis Et Applicata* **114**:97-105.

519 Linn, C. E., H. R. Dambroski, J. L. Feder, S. H. Berlocher, S. Nojima, and W. Roelofs. 2004. Postzygotic  
520 isolating factor in sympatric speciation in *Rhagoletis* flies: Reduced response of hybrids to

521 parental host-fruit odors. Proceedings of the Academy of Natural Sciences of the United States  
522 of America **101**:17753-17758.

523 Linn, C. E., Jr., W. L. Yee, S. B. Sim, D. H. Cha, T. H. Powell, R. B. Goughnour, and J. L. Feder. 2012.  
524 Behavioral evidence for fruit odor discrimination and sympatric host races of *Rhagoletis*  
525 *pomonella* flies in the Western United States. *Evolution* **66**:3632-3641.

526 Lowry, D. B., J. L. Modliszewski, K. M. Wright, C. A. Wu, and J. H. Willis. 2008. The strength and genetic  
527 basis of reproductive isolating barriers in flowering plants. *Philos Trans R Soc Lond B Biol Sci*  
528 **363**:3009-3021.

529 Lyons-Sobaski, S., and S. H. Berlocher. 2009. Life history phenology differences between southern and  
530 northern populations of the apple maggot fly, *Rhagoletis pomonella*. *Entomologia*  
531 *Experimentalis Et Applicata* **130**:149-159.

532 Maan, M. E., and O. Seehausen. 2011. Ecology, sexual selection and speciation. *Ecol Lett* **14**:591-602.

533 Marques, D. A., J. I. Meier, and O. Seehausen. 2019. A combinatorial view on speciation and adaptive  
534 radiation. *Trends in Ecology and Evolution* **34**:531-544.

535 Mattsson, M., G. R. Hood, W. L. Yee, M. M. Doellman, D. J. Bruzese, R. B. Goughnour, A. L. Driscoe, S.  
536 Van Dexter, C. Tait, M. M. Glover, P. Meyers, L. A. Ruedas, and J. L. Feder. 2021. Recursive  
537 adaptation in action: allochronis isolation and divergence of host-associated populations of the  
538 apple maggot fly, *Rhagoletis pomonella*, following its recent introduction to the western USA.  
539 *Entomologia Experimentalis Et Applicata*:1-16.

540 Maxwell, C. W., and E. C. Parsons. 1968. The recapture of marked apple maggot flies in several orchards  
541 from one release point. *Oecologia* **61**:1157-1159.

542 McNett, G. D., and R. B. Cocroft. 2008. Host shifts favor vibrational signal divergence in *Enchenopa*  
543 *binotata* treehoppers. *Behavioral Ecology* **19**:650-656.

544 Mendelson, T. C., V. E. Imhoff, and J. J. Venditti. 2007. The accumulation of reproductive barriers during  
545 speciation: Postmating barriers in two behaviorally isolated species of darters (percidae :  
546 etheostoma). *Evolution* **61**:2596-2606.

547 Mendelson, T. C., M. D. Martin, and S. M. Flaxman. 2014. Mutation-order divergence by sexual  
548 selection: diversification of sexual signals in similar environments as a first step in speciation.  
549 *Ecology Letters* **17**:1053-1066.

550 Merrill, R. M., Z. Gompert, L. M. Dembeck, M. R. Kronforst, W. O. McMillan, and C. D. Jiggins. 2011. Mate  
551 preference across the speciation continuum in a clade of mimetic butterflies. *Evolution* **65**:1489-  
552 1500.

553 Michel, A. P., S. Sim, T. H. Powell, M. S. Taylor, P. Nosil, and J. L. Feder. 2010. Widespread genomic  
554 divergence during sympatric speciation. *Proc Natl Acad Sci U S A* **107**:9724-9729.

555 Neilson, W. T. A., and J. W. McAllan. 1964. Artificial diets for the apple maggot, *Rhagoletis pomonella*. I.  
556 Mass rearing on certain diets. *Journal of Economic Entomology* **57**:333-335.

557 Nosil, P. 2012. *Ecological Speciation*. Oxford University Press, Oxford.

558 Nosil, P., and J. L. Feder. 2012. Genomic divergence during speciation: causes and consequences. *Philos*  
559 *Trans R Soc Lond B Biol Sci* **367**:332-342.

560 Nosil, P., J. L. Feder, S. M. Flaxman, and Z. Gompert. 2017. Tipping points in the dynamics of speciation.  
561 *Nat Ecol Evol* **1**:1.

562 Nosil, P., L. J. Harmon, and O. Seehausen. 2009. Ecological explanations for (incomplete) speciation.  
563 *Trends Ecol Evol* **24**:145-156.

564 Nosil, P., T. H. Vines, and D. J. Funk. 2005. Perspective: reproductive isolation cause by natural selection  
565 against immigrants from divergent habitats. *Evolution* **59**:705-719.

566 Olsson, S. B., Linn Jr, C. E., Michel, A., Dambroski, H. R., Berlocher, S. H., Feder, J. L., & Roelofs, W. L.  
567 2006. Receptor expression and sympatric speciation: unique olfactory receptor neuron  
568 responses in F1 hybrid *Rhagoletis* populations. *J. Exp. Biol.*, **209**, 3729-3741.

569 Opp, S. B., and R. J. Prokopy. 1986. Variation in laboratory oviposition by *Rhagoletis pomonella* (Diptera:  
570 Tephritidae) in relation to mating success. *Annals of the Entomological Society of America*  
571 **79**:705-710.

572 Powell, T. H., A. A. Forbes, G. R. Hood, and J. L. Feder. 2014. Ecological adaptation and reproductive  
573 isolation in sympatry: genetic and phenotypic evidence for native host races of *Rhagoletis*  
574 *pomonella*. *Mol Ecol* **23**:688-704.

575 Powell, T. H., G. R. Hood, M. M. Doellman, P. M. Deneen, J. J. Smith, S. H. Berlocher, and J. L. Feder.  
576 2022. The build-up of population genetic divergence along the speciation continuum during a  
577 recent adaptive radiation of *Rhagoletis* flies. *Gene* **13**:275.

578 Powell, T. H., G. R. Hood, M. O. Murphy, J. S. Heilveil, S. H. Berlocher, P. Nosil, and J. L. Feder. 2013.  
579 Genetic divergence along the speciation continuum: the transition from host race to species in  
580 *rhagoletis* (Diptera: tephritidae). *Evolution* **67**:2561-2576.

581 Powell, T. H. Q., A. D. Nguyen, Q. Xia, J. L. Feder, G. J. Ragland, and D. A. Hahn. 2020. A rapidly evolved  
582 shift in life-history timing during ecological speciation is driven by the transition between  
583 developmental phases. *Journal of Evolutionary Biology* **33**:1371-1386.

584 Prokopy, R. J. 1976. Feeding, mating and oviposition activities of *Rhagoletis fausta* flies in nature. *Annals*  
585 *of the Entomological Society of America* **69**:899-904.

586 Pryor, S. C., R. J. Barthelmie, and J. T. Schoof. 2013. High-resolution projections of climate-related risks  
587 for the Midwestern USA. *Climate Research* **56**:61-79.

588 Ragland, G. J., Almskaar, K., Vertacnik, K. L., Gough, H. M., Feder, J. L., Hahn, D. A., & Schwarz, D. 2015.  
589 Differences in performance and transcriptome-wide gene expression associated with *Rhagoletis*

590 (*Diptera: Tephritidae*) larvae feeding in alternate host fruit environments. *Molecular Ecology* **24**:  
591 2759-2776.

592 Ramsey, J., H. D. Bradshaw, and D. W. Schemske. 2003. Components of reproductive isolation between  
593 the monkeyflowers *Mimulus lewisii* and *M. cardinalis* (Phrymaceae). *Evolution* **57**:1520-1534.

594 Ribardiere, A., E. Pabion, J. Coudret, C. Daguin-Thiebaut, C. Houbin, S. Loisel, S. Henry, and T. Broquet.  
595 2019. Sexual isolation with and without ecological isolation in marine isopods *Jaera albifrons*  
596 and *J. praehirsuta*. *J Evol Biol* **34**:33-48.

597 Rice, W. R., and E. E. Hostert. 1993. Laboratory experiments on speciation: what have we learned in 40  
598 years? *Evolution* **47**:1637-1653.

599 Ritchie, M. G. 2007. Sexual Selection and Speciation. *Annual Review of Ecology, Evolution, and*  
600 *Systematics* **38**:79-102.

601 Rull, J., M. Aluja, and J. L. Feder. 2010. Evolution of intrinsic reproductive isolation among four North  
602 American populations of *Rhagoletis pomonella* (Dipter: Tephritidae). *Biological Journal of the*  
603 *Linnean Society* **100**:213-223.

604 Rundle, H. D., and P. Nosil. 2005. Ecological speciation. *Ecology Letters* **8**:336-352.

605 Rundle, H. D., and L. Rowe. 2018. The contribution of sexual selection to ecological and mutation-order  
606 speciation. *Evolution* **72**:2571-2575.

607 Safran, R. J., E. S. C. Scordato, L. B. Symes, R. L. Rodriguez, and T. C. Mendelson. 2013. Contributions of  
608 natural and sexual selection to the evolution of premating reproductive isolation: a research  
609 agenda. *Trends in Ecology & Evolution* **28**:643-650.

610 Schemske, D. W. 2010. Adaptation and the origin of species. *Am Nat* **176 Suppl 1**:S4-S25.

611 Schilling, M. P., S. P. Mullen, M. Kronforst, R. J. Safran, P. Nosil, J. L. Feder, Z. Gompert, and S. M.  
612 Flaxman. 2018. Transitions from Single- to Multi-Locus Processes during Speciation with Gene  
613 Flow. *Genes (Basel)* **9**.

614 Schluter, D. 2000. The Ecology of Adaptive Radiation. Oxford University Press, Oxford.

615 Schluter, D. 2001. Ecology and the origin of species. Trends in Ecology & Evolution **16**:372-379.

616 Schwarz, D., and B. A. McPherson. 2007. When ecological isolation breaks down: sexual isolation is an  
617 incomplete barriers to hybridization between *Rhagoletis* species. Evolutionary Ecology Research  
618 **9**:829-841.

619 Schwarz, D., Robertson, H. M., Feder, J. L., Varala, K., Hudson, M. E., Ragland, G. J., & Berlocher, S. H.  
620 2009. Sympatric ecological speciation meets pyrosequencing: sampling the transcriptome of the  
621 apple maggot *Rhagoletis pomonella*. BMC genomics, **10**: 1-14.

622 Scordato, E. S., L. B. Symes, T. C. Mendelson, and R. J. Safran. 2014. The role of ecology in speciation by  
623 sexual selection: a systematic empirical review. J Hered **105 Suppl 1**:782-794.

624 Seehausen, O., J. J. M. v. Alphen, and F. Witte. 1997. Cichlid Fish Diversity Threatened by Eutrophication  
625 That Curbs Sexual Selection. Science **277**:1808-1811.

626 Seehausen, O., G. Takimoto, D. Roy, and J. Jokela. 2008. Speciation reversal and biodiversity dynamics  
627 with hybridization in changing environments. Mol Ecol **17**:30-44.

628 Servedio, M. R., and J. W. Boughman. 2017. The Role of Sexual Selection in Local Adaptation and  
629 Speciation. Pages 85-109 in D. J. Futuyma, editor. Annual Review of Ecology, Evolution, and  
630 Systematics, Vol 48.

631 Servedio, M. R., and M. Kirkpatrick. 1997. The effects of gene flow on reinforcement. Evolution **51**:1764-  
632 1772.

633 Servedio, M. R., and M. A. F. Noor. 2003. The Role of Reinforcement in Speciation: Theory and Data.  
634 Annual Review of Ecology, Evolution, and Systematics **34**:339-364.

635 Servedio, M. R., G. S. Van Doorn, M. Kopp, A. M. Frame, and P. Nosil. 2011. Magic traits in speciation:  
636 'magic' but not rare? Trends Ecol Evol **26**:389-397.

637 Smadja, C. M., and R. K. Butlin. 2011. A framework for comparing processes of speciation in the  
638 presence of gene flow. *Mol Ecol* **20**:5123-5140.

639 Smith, D. C. 1988. Genetics and reproductive isolation of *Rhagoletis* flies. University of Illinois at Urbana-  
640 Champaign, Urbana, Illinois.

641 Smith, D. C., and R. J. Prokopy. 1982. Mating behavior of *Rhagoletis mendax* (Diptera: Tephritidae) flies  
642 in nature. *Annals of the Entomological Society of America* **75**:388-392.

643 Sobel, J. M., and G. F. Chen. 2014. Unification of methods for estimating the strength of reproductive  
644 isolation. *Evolution* **68**:1511-1522.

645 Tadeo, E., M. Aluja, and J. Rull. 2018. Precopulatory mating and postzygotic isolation between two  
646 walnut-infesting species of *Rhagoletis* from Mexican highlands. *Entomologia Experimentalis Et*  
647 *Applicata* **166**:713-723.

648 Tait, C., Batra, S., Ramaswamy, S. S., Feder, J. L., & Olsson, S. B. 2016. Sensory specificity and speciation:  
649 a potential neuronal pathway for host fruit odour discrimination in *Rhagoletis pomonella*.  
650 *Proceedings of the Royal Society B: Biological Sciences* **283**: 20162101.

651 Tait, C., Kharva, H., Schubert, M., Kritsch, D., Sombke, A., Rybak, J., Feder, J.L. & Olsson, S. B. 2021. A  
652 reversal in sensory processing accompanies ongoing ecological divergence and speciation in  
653 *Rhagoletis pomonella*. *Proceedings of the Royal Society B* **288**: 20210192.

654 Takami, Y., N. Nagata, M. Sasabe, and T. Sota. 2007. Asymmetry in reproductive isolation and its effect  
655 on directional mitochondrial introgression in the parapatric ground beetles *Carabus yamato* and  
656 *C. albrechti*. *Population Ecology* **49**:337-346.

657 Thibert-Plante, X., and A. P. Hendry. 2011. Factors influencing progress toward sympatric speciation.  
658 *Journal of Evolutionary Biology* **24**:2186-2196.

659 Tiffin, P., M. S. Olson, and L. C. Moyle. 2001. Asymmetrical crossing barriers in angiosperms. *Proc Biol Sci*  
660 **268**:861-867.



661 Turbek, S. P., M. Browne, A. S. Di Giacomo, C. Kopuchian, W. M. Hochachka, C. Estalles, D. A. Lijtmaer, P.  
662 L. Tubaro, L. F. Silveira, I. J. Lovette, R. J. Safran, S. A. Taylor, and L. Campagna. 2021. Rapid  
663 speciation via the evolution of pre-mating isolation in the Ibera Seedeater. *Science* **371**.  
664 Turelli, M., and L. C. Moyle. 2007. Asymmetric postmating isolation: Darwin's corollary to Haldane's rule.  
665 *Genetics* **176**:1059-1088.  
666 van Doorn, G. S., P. Edelaar, and F. J. Weissing. 2009. On the origin of species by natural and sexual  
667 selection. *Science* **326**:1704-1707.  
668 Verhoeven, K. J. F., K. L. Simonsen, and L. M. McIntyre. 2005. Implementing false discovery rate control:  
669 increasing your power. *Oikos* **108**:643-647.  
670 Vonlanthen, P., D. Bittner, A. G. Hudson, K. A. Young, R. Muller, B. Lundsgaard-Hansen, D. Roy, S. Di  
671 Piazza, C. R. Largiader, and O. Seehausen. 2012. Eutrophication causes speciation reversal in  
672 whitefish adaptive radiations. *Nature* **482**:357-362.  
673 Wade, M. J., N. W. Chang, and M. McNaughton. 1995. Incipient speciation in the flour beetle, *Tribolium*  
674 *confusum*: premating isolation between natural populations. *Heredity* **75**:453-459.  
675 Wagner, C. E., L. J. Harmon, and O. Seehausen. 2012. Ecological opportunity and sexual selection  
676 together predict adaptive radiation. *Nature* **487**:366-369.  
677 Walsh, B. D. 1861. On phytophagic varieties and phytophagic species.  
678 Ward, J. L., and M. J. Blum. 2012. Exposure to an environmental estrogen breaks down sexual isolation  
679 between native and invasive species. *Evol Appl* **5**:901-912.  
680 Weissing, F. J., P. Edelaar, and G. S. van Doorn. 2011. Adaptive speciation theory: a conceptual review.  
681 *Behavioral Ecology and Sociobiology* **65**:461-480.  
682 Yeaman, S., and M. C. Whitlock. 2011. The genetic architecture of adaptation under migration-selection  
683 balance. *Evolution* **65**:1897-1911.

684 Yee, W. L., and R. B. Goughnour. 2012. Mating frequencies and production of hybrids by *Rhagoletis*  
685 *pomonella* and *Rhagoletis zephyria* (Diptera: Tephritidae) in the laboratory. *The Canadian*  
686 *Entomologist* **143**:82-90.

687 Zhang, L., X. Thibert-Plante, J. Ripa, R. Svanback, and A. Brannstrom. 2019. Biodiversity loss through  
688 speciation collapse: Mechanisms, warning signals, and possible rescue. *Evolution* **73**:1504-1516.

689 Zhang, L., G.R. Hood, J.R. Ott, and S.P. Egan. 2022. Asymmetric habitat isolation and sexual isolation  
690 predicted by the cost of migration and hybridization generate novel signatures of reinforcing  
691 selection. *BioRxiv*. doi.org/10.1101/2022.01.02.474698

692

693 **Supporting Information**

694

695 Supplemental methods text:

696 In each weekly program, temperatures ramped linearly through four set points: midpoint temperature  
697 at sunrise, maximum temperature at the time halfway between sunrise and sunset, midpoint  
698 temperature at sunset, and minimum temperature at the time halfway between sunset and sunrise. The  
699 timing and length of light:dark cycles were set by sunrise and sunset times for the last day in each week  
700 of 2016 at the Watseka station. When median weekly temperatures would have dropped below 6°C in  
701 each temperature regime, we switched environmental chambers to a winter program with lights off and  
702 2.5°C minimum, 3.0°C midpoint, and 3.5°C maximum set points. When median weekly temperatures  
703 would have risen above 6°C, we switched environmental chambers to resume Control and Warming  
704 regimes based on 10-year weekly temperature averages and light:dark cycles. Given differences in when  
705 Control and Warming median temperatures would drop below and rise above 6°C, winter length  
706 differed between temperature regimes: 20 weeks, November 12 to April 1, for Control; 16 weeks,  
707 November 19 to March 11 for Warming.

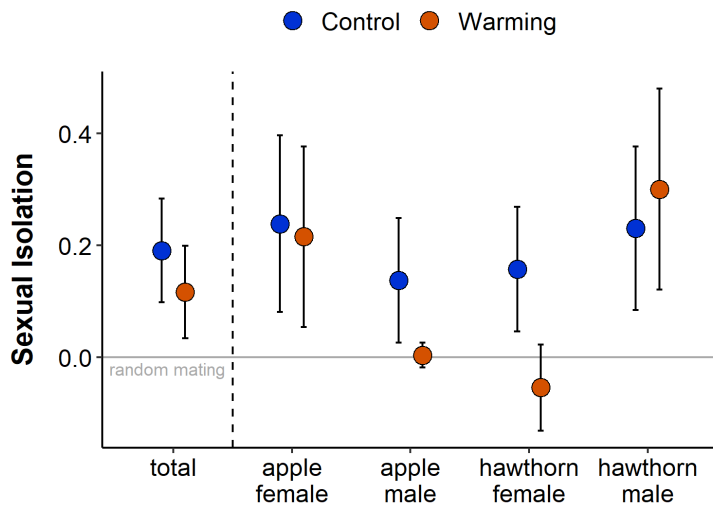
708 Supplemental Table 1: Measures of sexual isolation with 95% confidence interval upper and lower limits  
 709 for total sexual isolation as well as contributions from each sex of each population for flies reared in  
 710 Control and Warming temperature treatments.

	Control treatment			Warming treatment		
	Sexual isolation	95% CI upper limit	95% CI lower limit	Sexual isolation	95% CI upper limit	95% CI lower limit
Total	0.191	0.284	0.098	0.116	0.199	0.034
Apple female	0.239	0.397	0.081	0.216	0.377	0.054
Apple male	0.138	0.249	0.027	0.004	0.026	-0.018
Hawthorn female	0.158	0.269	0.046	-0.054	0.023	-0.131
Hawthorn male	0.231	0.376	0.085	0.301	0.480	0.121

711  
 712 Supplemental Table 2: For each of three prezygotic reproductive barriers, we provide values for the  
 713 individual barrier strength, 95% confidence interval width, upper and lower bounds of the individual  
 714 strength given the confidence interval, and the sequential strength. The sequential strength is calculated  
 715 from its individual strength and the amount of gene flow allowed by earlier-acting barriers. Sexual  
 716 isolation is based on mating interactions in flies from Control rearing temperatures.

barrier	individual strength	95% CI upper limit	95% CI lower limit	sequential strength
temporal	0.4363	0.5597	0.3128	0.4363
habitat	0.8746	0.9167	0.8325	0.4931
sexual	0.1909	0.2837	0.0982	0.0135

717



718

719 Supplemental Figure 1. Total sexual isolation and contributions of each sex from each population for  
720 flies reared under Control (blue) and Warming (orange) temperatures. The dashed vertical line  
721 separates total sexual isolation from contributions of each sex from each population. Circles are point  
722 values with 95% CIs. The horizontal grey line at 0 indicates random mating, and positive values indicate  
723 greater mating within populations than between. Values for barrier strengths and 95% CIs are provided  
724 in Supplemental Table 1.