1	Metamicrobiome-driven homeostasis of nutrient recycling
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21 22 23 24 25 26 27 28 29 30 31 32 33	Abstract Carbon and nutrient recycling by free-living microbial decomposers and fire - two key recycling pathways - are highly sensitive to climatic variation. However, mutualistic associations of microbiomes with plants and animals cause previously underestimated environmental buffering effects. This close cooperation between small and large organisms solves a fundamental allometric trade-off between mass-specific metabolic capacity (decreasing with body size) and homeostatic capacity (environmental buffering; increasing with body size), allowing the combination of the best of both worlds along the body mass spectrum from microbes to elephants. A diverse metamicrobiome, where plant- and animal- associated microbiomes complement the free-living microbiome, consequently increases ecosystem homeostasis of recycling rates in a variable environment. We argue for better integration of this fundamental ecological process in predicting the consequences of current accelerated environmental change.

34 Glossary:

Biome	An area classified according to a characteristic group of species that lives in it, such as the tundra, savanna or tropical rainforest.
Carbon and nutrient recycling	The transformation of carbon and potentially limiting nutrients (e.g. N, P, K, Ca, Mg) from their various organic forms e.g. proteins, fats) to their biologically reactive inorganic forms (ammonium, phosphates, exchangeable bases) that can be used by plants and microbes.
Climate variation	The way aspects of climate (such as temperature and precipitation) differ from an average, due to natural and sometimes periodic changes in the circulation of the air and ocean, volcanic eruptions, and other (including anthropogenic factors at multiple temporal scales (days, seasons, years, decades).
Free-living microbiome	The set of microbes (bacteria, protists, archaea, fungi) without a close and intimate association with larger organisms together responsible for carbon and nutrient recycling in soil.
Ecosystem homeostasis	The degree to which key ecosystem processes, such as primary productivity, secondary productivity, carbon storage and nutrient recycling are kept constant by biotic interactions despite strong variations in environmental conditions imposed from larger spatial scales, such as solar radiation, air temperature and rainfall.
Homeostatic capacity of organisms	The relative ability of an organism to keep the physical and chemical conditions (e.g. temperature, moisture, osmotic potential, pH) in its internal environment (inside the organism) constant despite strong fluctuations in these factors in its external environment.

Metabolic capacity of organisms	The rate of energy turnover (J day-1) of an organism, determining its capacity for nutrient and carbon turnover from organic to inorganic reactive forms.
Metamicrobiome	The interacting set of four main microbiomes responsible for carbon and nutrient recycling in terrestrial ecosystems consisting of i) the free- living microbiome, and the mutualistic microbiomes associated with ii) plants (i.e. mycorrhizae and rhizosphere bacteria), iii) macrodetritivores (i.e. gut microbes or fungal gardens), and iv) with herbivores (gut microbes).
Mutualism/mutualistic	A symbiotic relationship between two species where both partners benefit from their interaction. Mutualistic interactions between animal or plant hosts and microbial habitués span from intracellular (e.g. endophytic fungi) to extracellular (e.g. bacteria and Archaea in gut lumen) to exosymbiontic, in which part of the microbiome is found outside of the host but is actively maintained and tended for by hosts, as in ectomycorrhizal or termite-associated fungi.
Preemption	Interaction between two species or species groups where the first species typically consumes a resource or substrate before the second species can access it.
Saprotroph	An organism that derives nourishment from detritus.
Substrate availability (equivalent term: resource availability)	The rate per unit area or soil volume at which biologically reactive forms of elements that can be directly metabolized by organisms become available. For primary producers, these include nitrate, ammonium, and phosphates while for heterotrophic organisms this is the rate of their food supply, such as sugars (bacteria), grass (herbivores), litter (decomposers).

Stoichiometric	The balance of biologically important elements	
	in organisms and resources	

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36 Carbon and nutrient recycling in a changing climate

37 Carbon and nutrient recycling of plant materials (here-after 'recycling') is central to ecosystem 38 functioning because it maintains primary productivity and regulates the Earth's climate [1]. However, 39 global change is strongly altering the rate and stability of recycling in many terrestrial ecosystems, with 40 consequences for ecosystem dynamics and potential feedback to global biogeochemical cycles and 41 climate [2,3]. Global carbon models generally assume that plant-assimilated carbon and nutrients are 42 predominantly returned into inorganic forms by either the free-living soil microbiome [4] or fire, 43 representing two major terrestrial recycling pathways [5]. Both pathways are highly sensitive to 44 forthcoming environmental change, which can induce strong climatic (e.g. extreme events) and 45 substrate limitations. For example, intensified droughts can halt litter organic matter decomposition for 46 extended periods [6]. However, mutualistic microbiomes associated with plants (mycorrhizal fungi), 47 macroinvertebrates (gut microbes and fungal gardens in mounts and nests), and herbivores (gut 48 microbes) are also important agents of organic matter mineralization and can dominate the recycling 49 process at local to regional scales [7-9]. Recent evidence suggests that these mutualistic microbiomes 50 increase long-term average recycling rates in most terrestrial ecosystems by reducing environmental 51 fluctuations and lifting limitations to microbial breakdown, making them less sensitive to climate 52 variation. Here, we review how the free-living soil microbiome and at least three mutualistic 53 microbiomes interact in a metamicrobiome (Fig. 1). To illustrate how the metamicrobiome concept is 54 key to understanding ecosystem and biome scale recycling (and thus to ecosystem responses to global 55 change), we have to start with the constraints on the functioning of the free-living soil microbiome.

56 Constraints on carbon and nutrient recycling by the free-living microbiome

57 Soil chemo-physical conditions are highly heterogeneous both in time and space. Abiotic factors that 58 determine microbial performance - i.e. soil moisture, temperature, pH and redox status - fluctuate 59 continuously, causing soil microbial activity to be highly restricted to specific microsites and periods 60 where chemo-physical limitations (or climatic limitations) are removed [10]. Because free-living soil 61 bacteria and archaea are so small, they depend primarily on substrates that move towards them 62 passively by diffusion [11]. This makes soil water essential: it prevents desiccation and acts as a solvent 63 for water-soluble organic substrates and a transport medium for microbes and their chemicals and exo-64 enzymes [12]. Their metabolic activity therefore highly depends on temporal variation in water content 65 of the soil. In addition, soil temperature is a key factor for the metabolic activity of ectothermic 66 organisms, including microbes [13]. In regions with a long cold season such as the arctic tundra, this 67 reduces organic matter recycling and promotes carbon sequestration [14].



Free-living microbes associated with soil organic matter

Macrodetritivore-associated Microbiome (DaM)

Mutualistic microbes associated with invertebrate soil macrofauna such as termites or earthworms

Mutualistic microbes associated with vertebrate megafauna, especially large mammalian herbivores

Plant-associated Microbiome (PaM)

Mutualistic microbes associated with plants, as Ectomycorrhiza (EcM), Ericoid Mycorrhiza (ErM), or Arbuscular Mycorrhiza (AM)

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69 Fig. 1: Metamicrobiome ecology: carbon and nutrient recycling in terrestrial ecosystems is generally 70 mediated by a metamicrobiome of four interacting microbiomes, i.e. the Free-living Microbiome (FIM), 71 the Macrodetritivore-associated Microbiome (DaM), the Herbivore-associated Microbiome (HaM), and 72 the Plant-associated Microbiome (PaM). The mutualistic microbiomes DaM, HaM, and PaM profit from 73 the homeostatic conditions offered by their hosts, allowing recycling to continue when environmental 74 conditions for the FIM become unfavourable. Alternatively, carbon and nutrients can be recycled by fire. 75 For other nutrients such as P, K, Mg, and Ca, the same recycling principles hold, except the gaseous 76 components (N2 and NOx) are missing.

77

78 While soil is the largest reservoir of carbon substrates in terrestrial ecosystems, several mechanisms 79 further limit carbon and nutrient recycling by the free-living microbiome. Physical mechanisms such as 80 occlusion within large soil aggregates and adsorption onto mineral surfaces limit substrate accessibility 81 [15,16]. Vegetation composition and structure determine to what extent and when organic substrates 82 remain part of standing biomass (aerial position) or come into contact with the soil surface [17] and thus 83 potentially limit substrate supply to free-living microbes. The mineralization rate of organic matter also 84 depends on the C:N:P ratio of the substrate relative to the decomposers' need [18]. Because microbial

- organisms function best at specific **stoichiometric** C:N:P ratios, nutrients in organic matter that approach that ratio and thereby meet stoichiometric requirements will increase microbial activity and mineralization rates [19]. In summary, substrate availability depends greatly on the microbial landscape; decomposition of organic substrates can only occur when water, oxygen, (high quality) substrates, the
- 89 microbial organism, and their (exo-) enzymes all come together in space and time [12,20].

90 Small organisms: highest metabolic capacity

91 Organisms vary significantly in body mass, ranging by over 20 orders of magnitude from 10⁻¹⁵ g 92 microbes to 2.10⁸ g blue whales. The size of organisms correlates with many ecological and 93 physiological traits in predictable ways [21]. A fundamental relationship underlying many of these 94 patterns is the quarter-power allometric scaling of organismal basal metabolic rate M with body mass 95 B, where B \propto M^{3/4} [22]. It predicts how larger organisms have an increasingly lower rate of energy use 96 per unit of body mass, as B/M \propto M^{-1/4}; and holds across a wide variety of taxonomic groups [23], 97 including plants [24]. This predictably lower metabolic capacity of larger organisms, compared to 98 microbes, has been explained by constraints imposed by the architecture of their internal transport 99 network on the rates at which nutrients and oxygen can be delivered to the cells in metazoans [25,26]. 100 Also, relative body area scales with body mass as $B/M \propto M^{-1/3}$, constraining the capacity for resource 101 uptake through passive diffusion in larger organisms. As a result, bacteria, archaea, protists and fungi 102 have the highest per mass capacity for energy and nutrient turnover (i.e. metabolic capacity, Fig. 2A), 103 but only if environmental conditions such as temperature and moisture are optimal as they have limited 104 capacity to control their internal conditions and direct surroundings.

105 Large organisms: highest homeostatic capacity

106 In contrast, larger, multicellular species have a better capacity to stabilize their internal environment or 107 immediate surroundings through various anatomical, physiological, and behavioral adaptations that are 108 not possible in microbes [27]. This homeostatic capacity also scales with body mass but opposite to 109 metabolic capacity (Fig. 2). Lower surface-to-volume ratios in larger organisms are favourable under 110 environmental conditions where restricting water (droughts) or heat loss (cold periods) are important 111 [28]. For example, larger organisms are less likely to be seasonally disrupted in their daily activities by 112 a need to go into hibernation or torpor under cold conditions [29]. Moreover, the higher capacity (relative 113 to body mass) of larger organisms to store energy and nutrient reserves allows these organisms to 114 remain active during longer periods of resource limitation [30]. Behavioral adaptations of animals (e.g. 115 seeking shade or sun, moving long distances to find suitable food or water, or burrowing to deeper soil 116 layers to prevent climatic stress) also contribute to their homeostatic capacity. This also applies to 117 plants, e.g., through opening and closing of stomata, rolling up of graminoid leaves during drought, and 118 rooting deep to access water by dryland trees. These behavioral adaptations also scale with body size 119 [31], with larger organisms having a higher capacity to move towards essential resources. Also, larger 120 organisms have more options to improve their local environment to their benefit through ecosystem 121 engineering and extended phenotypes (e.g. beaver dams, rodent burrows, bird nests, termite mounds).

122 To summarize, larger organisms have a larger homeostatic capacity when faced with temporarily 123 unfavourable conditions (Fig. 2).



Fig. 2: (A) The trade-off between per mass metabolic capacity (log scale) and homeostatic capacity as a function of body mass (B). An example of the scaling relation of homeostatic capacity with body mass, showing how the capacity to regulate body temperature changes with body mass, where body temperature homeostasis is expressed as the difference between maximum (B_{max}) and minimum (B_{min}) body temperatures. The solid symbols represent actual data and open symbols represent simulated data, while triangles represent data compiled by Stevenson (1985) and the circles represent additional data listed in supplement S1.

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133 The trade-off between metabolic and homeostatic capacity drives mutualisms

134 The observed opposite relationships of body mass with metabolic and homeostatic capacity of 135 organisms suggests a fundamental trade-off: the same organism cannot excel at both (Fig. 2). Microbes 136 are best at achieving high metabolic rates under optimal conditions while large multicellular eukaryotes 137 are best at realizing such optimal conditions in their bodies and immediate surroundings. The long-term 138 outcome of this trade-off is the evolution of mutualisms. Multicellular eukaryotes have repeatedly 139 solved their metabolic constraints over evolutionary time by engaging in mutualisms with microbes that 140 possess complex and efficient metabolic capabilities [32], and microbes have 'accepted this request' 141 because it stabilizes their environment and resource supply. The capacity to acquire microbial partners 142 has been both a key factor in the evolution of first eukaryotes from prokaryotes [33] as well as in the 143 subsequent eukaryote diversification [34], shaping the adaptation to new habitats, lifestyles, and diets. 144 These mutualisms have therefore led to multiple synergies between metabolic and homeostatic 145 capacity (Box 1).

146

147 Box 1: Synergies between metabolic and homeostatic capacity: different mechanisms

148 Stress alleviation benefits

149 In biomes that experience seasonal or multi-annual drought, metazoans often evolve traits that promote 150 their water acquisition and conservation. They can control the water content in their body, or in their 151 direct surroundings, or by selecting constantly moist microhabitats, all of which can be beneficial for 152 their microbial mutualists. For example, fungus-growing termites, which engage in a complex three-way 153 symbiosis with their bacterial gut microbiome and a basidiomycete fungus [92,93], promote the activity 154 of the fungal symbiont by optimizing the humidity inside their mounds. They do this by 'wicking' or 155 actively transporting water to or from their nests, and regulating evaporation and respiration by 156 ventilation [94,95]. Earthworms vertically move up and down in their burrows to stabilize moisture in 157 their direct surroundings [86], supporting the continued activity of their microbiome. Stabilization of 158 chemo-physical conditions also applies to plant hosts. Ectomycorrhizas (EcM) can alleviate drought 159 stress to host trees by enhancing access to soil water [96,97]. However, when water tables fall during 160 summer, trees in Mediterranean forests and woodlands shift their primary water source to deeper soil 161 layers [98] thereby returning the water-provisioning favour to their fungal symbionts [49]. By lifting 162 moisture and temperature constraints, hosts increase the growth, survival, and resource processing 163 activity of microbes, i.e. "metabolic homeostasis" [55]. Mutualistic associations between hosts and 164 microbes, therefore, decouple microbial activity from unfavourable conditions, reducing the overall 165 control of climate on the recycling of organic matter.

166 Complementarity benefits

167 Hosts can also lift substrate quality limitations by facilitating complementary contributions between 168 multiple symbiotic microbial partners. The guts and rumen of mammalian herbivores and 169 macrodetritivores act as biological chemostats where hundreds of microbial species and their 170 metabolites all interact in the digestion process. Substrates that result from the metabolism of some 171 microbial species are often efficiently further metabolized by other species (metabolic cross-feeding), 172 thereby promoting fuller decomposition, from polymer degradation to sugar fermentation [99], in one 173 regulated environment. Moreover, the host continuously removes metabolic end products that are 174 released by mutualistic microbes, which favours further metabolism through reducing feedback 175 inhibition [100]. Similarly, the bacterial community inside the guts of termites, and the basidiomycete 176 fungi that they farm, contribute to the decomposition process in a complementary way [101]. The 177 termites facilitate the process by transferring the substrates and enzymes between the microbial 178 partners [93,102] and thereby speed up the recycling process.

179 *Preemption and shortcutting benefits*

Large hosts also bring mutualistic microbes in continuous contact with substrates through preemption
 (or short-circuiting). Some fungi positioned themselves closer and closer to living plant root cells in
 anticipation of their death, which at some point in evolutionary time likely led to the mycorrhizal habitat

183 in which substrate is colonized before death [103]. The secured and high supply of "easy" carbon 184 through plant hosts allows expansion of mycorrhiza into upper soil layers at the expense of the free-185 living microbiome. Moreover, the supply of host photo-assimilates supports the synthesis of 'expensive' 186 enzymes that can break down very recalcitrant organic substrates, thereby simultaneously lifting 187 substrate quality limitations. Indeed, ecto- and ericoid mycorrhizae are effective scavengers for 188 relatively recalcitrant organic N and P sources and could therefore reinforce a competitive advantage 189 over free-living saprotrophs by limiting the availability of these essential elements [104,105]. Fungus-190 growing termites also preempt organic matter in the soils they occupy [106]. After a first gut passage, 191 collected litter is directly delivered as pseudo-faeces to the termite's fungal symbionts [93]. The 192 continuous directional flow of litter inside mounds explains why soils occupied by termites can be 193 depleted of soil nutrients and organic carbon [107], and why the inter-mound soil matrix may host 194 smaller populations of microbes [108]. Finally, large mammalian herbivores consume resources before 195 green plant parts turn into litter and can, therefore, be considered even better preemptors of energy-196 rich substrates than macrodetritivores [106].

197

198 Metamicrobiome organization along global gradients

199 As the four microbiomes (free-living, the detritivore-associated, plant-associated and large herbivore-200 associated) all interact in a metamicrobiome (Fig. 1), but each with different sets of adaptations, their 201 relative importance is expected to strongly vary between different **biomes**. Because microbial activity 202 increases with temperature and moisture availability, recycling rates are generally faster in tropical 203 regions [35,36]. Higher rainfall promotes closed-canopy systems, which excludes most of the large herbivore-associated microbiome - indirectly increasing the importance of other microbiomes. 204 205 Permanently wet environments also limit abiotic forms of recycling, such as fire [5] and 206 photodegradation [37], making the free-living soil microbiome dominant in recycling organic matter (Fig. 207 3). This dominance under these conditions is also expected from the trade-off between metabolic and 208 homeostatic capacity: the stable environment in tropical forests allows soil microbes to remain active 209 and recycle substrates throughout the year.

The herbivore-associated microbiome (HaM) is expected to dominate intermediate rainfall regions with high soil fertility (Fig. 3) because high plant nutrient concentrations support high levels of herbivory [38,39]. Also under cold or saline conditions, plant quality ingested by herbivores (and thus the HaM) is higher due to restricted productivity, while carbohydrate accumulations can act as osmoregulation or antifreeze [40], and endothermic herbivores provide warm refugia for microbial decomposition. In the harshest and least productive environments (extreme drought or cold), hosts can no longer survive at high densities, and photodegradation becomes increasingly important [41] (Fig. 3).

Nutrient-poor environments generally promote plant species with low-quality tissues, which produce
 poor-quality litter (i.e. substrates) via so-called traits after-life effects [42]. Due to the small size and,

219 therefore, high nutrient requirements of microbes, the activity of the soil microbiome is negatively 220 impacted by recalcitrant compounds and the low concentrations of nutrients in these substrates, which 221 slows recycling and reinforces the low nutrient availability in poor sites. The proportion of mycorrhiza 222 relative to saprotrophs (part of the FIM) increases with decreasing soil fertility in boreal forests [43]. 223 From a metamicrobiome perspective, preemption makes a mycorrhizal association increasingly 224 beneficial compared to living alone in an energy-depleted environment, giving the plant-associated 225 microbiome an outsized role in recycling in poor ecosystems (Fig. 3). Abiotic forms of recycling may 226 also increase in importance when aboveground standing biomass accumulates in poor environments 227 due to low herbivore abundance. Especially in climatically seasonal environments (but with high enough 228 productivity for fuel production), this may lead to increased recycling by fire [44] (Fig. 3) as both the 229 free-living and plant-associated microbiome predominantly act on substrates within soils.

230 Lastly, there are various examples of nest or mound-building macrodetritivores that can lift both climatic 231 and substrate limitations, which may give the macrodetritivore-associated microbiome a dominant role 232 in relatively nutrient-poor environments that are either periodically cold or dry (Fig. 3). Many temperate 233 and boreal zone ant species build above-ground nest-mounds and keep them free from plants to obtain 234 heat from solar radiation. The nests of yellow meadow ants (Lasius flavus and L. umbratus) significantly 235 increase in temperature in the summer, creating favourable conditions for their own gut microbiome and 236 other soil biota and serving as 'hot spots' for recycling [45,46]. Because the burrows of desert isopods 237 provide more stable temperature and higher moisture, the recycling of plant litter is controlled by macro-238 detritivores in desert ecosystems [47].

In summary, the contribution of the different mutualistic microbiomes to recycling should increase with climatic and substrate limitations (Fig. 3). The improved homeostatic and pre-emptive capacity contributed by hosts allows mutualistic microbiomes to maintain recycling even throughout unfavourable periods. However, homeostasis and preemption require investments and only work when costs are balanced by sufficient benefit in exploiting energy-rich substrates, increasing the importance of abiotic forms of recycling in the most limiting environments.



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Fig. 3: The relative importance of the free-living soil microbiome (FIM, grey) and mutualistic Herbivoreassociated Microbiome (HaM, purple) of vertebrate megafauna, invertebrate Macrodetritive-associated
Microbiomemacrofauna (MaM, green), and the Plant-associated Microbiome (PaM, brown) for recycling
of plant organic matter along gradients of climatic and substrate limitation. Large arrows represent the

250 dominant switches in relative importance. See fig. 1 for further explanation of the different microbiomes.

251

252 Homeostasis in variable environments

253 The organization of the metamicrobiome is not only expected to vary with average environmental 254 conditions but also interacts with their temporal variability. Microbial activity is expected to be highest 255 when the environment is not too dry and not too wet (leading to soil anoxia) [48], not too cold and not 256 too hot, leading to a restricted set of conditions for nutrient recycling (green area in Fig. 4A). Soil organic 257 matter thus is subject to temporal variation in both moisture and temperature on multiple time scales 258 (Fig. 4A). In seasonally variable habitats, soil moisture and temperature will frequently move in and out 259 of the optimal range of free-living microbes. Consequently, the rate of recycling by the free-living 260 microbiome is expected to be tightly linked to seasonal changes in rainfall (Fig. 4B) and to vary much 261 more than mutualistic microbiomes, which experience less variation in chemo-physical conditions (Fig. 262 4B).

263 Microbiomes can also alternate over time in their dominance of the recycling process, which stabilizes264 its overall rate. For instance, in temperate woodlands, mycorrhizal fungi are most important during the

- summer, likely due to increased water supply by plants [49], while free-living fungi reach their maximum abundance in autumn [50] associated with peak litter production [51]. Similar alterations are found for
- other microbiomes. Foraging activity by termites (*Macrotermes* sp.) and desert isopods (*Hemilepistus*
- *reaumuril*) peaks during the hot and dry season [52,53], which can lead to a proportional increase in
- 269 macrodetritivore-driven recycling during drier periods relative to the free-living microbiome [54,55] (Fig.
- 4D). Finally, many populations of large herbivores migrate seasonally to overcome periodic limitations
- associated with food quality or quantity [56,57], which can amplify the contribution of grazing to recycling
- 272 during unfavourable climatic conditions. A diverse metamicrobiome where different host-associated
- 273 microbiomes complement the free-living microbiome therefore leads to **ecosystem homeostasis** of
- 274 carbon and nutrient recycling rates in variable environments (Fig. 4C).

275 Global change relevance

276 Climate change, especially extreme events (increase in frequency, duration and amplitude of heat 277 waves and extreme droughts) can bring soil environmental conditions more frequently outside the 278 optimal conditions for the free-living soil microbiome. Drought-induced changes in community 279 composition can cause long-term effects, reducing recycling rates for up to one year, even when water 280 becomes available again [6]. More climatic variation also increasingly exposes the free-living 281 microbiome to repeated dry-rewetting or freeze-thaw cycles, which damage the microbial community 282 [58]. Such cycles may cause nutrient losses through leaching to ground and surface waters [59,60] 283 because soil microbes are important dynamic sinks for carbon and nutrients [61]. Warming furthermore 284 loosens microbial association networks in colder biomes [62], which can lead to a loss of cooperative 285 interactions (e.g. syntrophy) between microbes [63,64]. Also, droughts and intense rainfall events can 286 cause more pulses in microbial recycling [65], leading to more mismatches with plant uptake needs, 287 impairing primary productivity [65,66]. All these factors make recycling by the free-living microbiome 288 highly sensitive to climate change.

289 In contrast, mutualistic microbiomes can provide important buffering effects in a changing climate. Large 290 mammalian herbivores can accelerate recycling through the modification of growing conditions (e.g. 291 sunlight) and the direct return of nutrients to soils through urination and defecation [67-69]. Higher rates 292 of recycling boost plant productivity, even in climatically harsh environments. The arctic region was 293 much colder and drier during the last glacial maximum but was much more productive, a discrepancy 294 that has been termed the "productivity paradox" [70]. The paradox can be explained by the much higher 295 mean body size of the animal hosts that were present at the time [71]. A large body enabled mammoths 296 to efficiently exploit and preempt substrates, giving rise to the highly productive grasslands of the 297 Pleistocene: the mammoth steppe [71]. The loss of megaherbivores during the Pleistocene



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299 Fig. 4. A) Phase space of temperature and moisture content of organic matter. The colour gradient 300 visualizes the optimal conditions (green) for microbial activity. Mutualistic microbiomes experience less 301 variation in chemo-physical conditions, as represented by the smaller circles, compared to free-living 302 microbes in different biomes. B). The rate of recycling by the free-living microbiome should be intimately 303 linked to fluctuations in soil temperature and moisture, while recycling by mutualistic microbiomes is 304 more stable over time. C) Temporal alternation in the relative contribution of the FIM (grey solid line) 305 and mutualistic microbiomes (dotted line) to recycling, leading to D) higher rates of recycling at the 306 ecosystem level even at high climatic and substrate limitations.

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megafaunal extinctions slowed recycling, made recycling more seasonally limited, and consequently
 may have turned the highly productive steppes into tundra ecosystems [72]. Currently, the (sub)arctic

310 is the strongest atmospheric carbon source due to a positive feedback between a warming climate and

311 the thaw of carbon-rich permafrost layers [73]. Reverting the moss and shrub-dominated tundra back 312 to a productive grassy steppe ecosystem by the reintroduction of large herbivores could potentially slow 313 down carbon emission because the increased recycling and productivity would improve carbon capture 314 [74]. Moreover, grazing can prevent fuel accumulation and therefore limit fire frequencies in seasonally 315 dry ecosystems, reducing nutrient and carbon losses [75,76]. This adds evidence to findings that large 316 herbivores are important in mitigating the negative impacts of climate change [77].

317 Plant-associated microbiomes also buffer against the currently observed acceleration of carbon 318 release. Warming promotes higher rates of recycling by the soil microbiome, stimulating a net loss of 319 soil carbon that may further accelerate global warming [78]. Higher atmospheric CO² concentration and 320 temperatures generally result in higher net primary productivity (NPP) which means that (everything 321 else being equal), nutrients can become more limiting [79]. A higher fraction of NPP would therefore 322 have to go to nutrient acquisition, reducing the buffering effect that increased productivity has on rising 323 CO₂ levels. The plant-associated microbiome can delay this so-called 'progressive nutrient limitation' 324 because EcM mines the soil organic matter for nutrients, promoting carbon sequestration also in 325 nutrient-limited areas [80].

326 Concluding remarks

327 The relative contributions of the different mutualistic microbiomes as outlined in Fig. 1 are currently 328 undergoing rapid change in ecosystems worldwide. Among animal hosts, larger herbivores seem to be 329 more sensitive to ongoing land-use change and climate change [81], as was the case during the 330 Pleistocene defaunation. Large animals have vulnerable life-history strategies [82], need more space 331 because of larger home ranges [31,83], and have only a limited capacity to adapt to hotter daytime 332 temperatures [84]. Intensive land use reduces soil insect nest and mound densities [85] and dries out 333 topsoils to unfavourable levels for earthworms [86], and so forms a significant challenge to the 334 macrodetritivore-associated microbiome. The plant-associated microbiome is also vulnerable: 335 ectomycorrhiza-dependent vegetation is declining on all continents and is being replaced by non-336 mycorrhizal vegetation due to forest logging and agricultural intensification [87].

337 Such challenges to mutualistic microbiomes will reduce the rate and stability of recycling, with cascading 338 effects on the biodiversity and productivity of terrestrial ecosystems that will eventually impact the whole 339 system Earth. Better protection of (semi-)natural ecosystems that support a diverse metamicrobiome 340 preserves their resilience to climate change. For example, ecological restoration through rewilding can 341 stop further defaunation by re-introducing missing wildlife and can thereby restore multiple important 342 ecosystem functions [88]. Different studies indeed show that restoration of megaherbivores can lead to 343 a more productive plant community and the moderation of fire regimes [89,90]. Similarly, restoring 344 native (forest) vegetation in abandoned agricultural fields supports the recovery of the plant-associated 345 microbiome [87,91].

346 By mapping the connections between the fields of microbial ecology, community ecology, and 347 ecosystem ecology, we have provided an integrated framework that accounts for the prominent role of 348 host-mediated microbiomes in the recycling of plant matter. Especially for biomes where the activity of 349 the free-living microbiome is chronically or periodically limited, inclusion of mutualistic microbiomes may 350 radically modify predictions on the sensitivity of ecosystems to increased variability and soil carbon 351 sequestration under current and future climate and land-use. To test the general ideas outlined in this 352 paper and advance this line of research, there is now a need for more empirical data on the importance 353 of different microbiomes across biomes and soil types (see outstanding questions). We recommend the 354 simultaneous measurement of the activity of mutualistic microbes in comparison to free-living microbes 355 in controlled field experiments and across main environmental gradients. Finally, we underscore the 356 need for common linguistic and conceptual currencies between microbiologists and ecologists to merge 357 our understanding of the factors that control the activity of the metamicrobiome.

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359 References

- Wardle, D. A. 2013. Communities and ecosystems: linking the aboveground and belowground
 components (MPB-34). Princeton University Press.
- Lashof, D. A., DeAngelo, B. J., Saleska, S. R., & Harte, J. (1997). Terrestrial ecosystem
 feedbacks to global climate change. Annual Review of Energy and the Environment, 22(1),
 75-118.
- 365 3. Heimann, M., & Reichstein, M. (2008). Terrestrial ecosystem carbon dynamics and climate
 366 feedbacks. Nature, 451(7176), 289-292.
- Jones, C., McConnell, C., Coleman, K., Cox, P., Falloon, P., Jenkinson, D., & Powlson, D.
 (2005). Global climate change and soil carbon stocks; predictions from two contrasting
 models for the turnover of organic carbon in soil. Global Change Biology, 11(1), 154-166.
- 370 5. Pausas, J. G., & Bond, W. J. (2020). On the three major recycling pathways in terrestrial
 371 ecosystems. Trends in Ecology & Evolution, 35(9), 767-775.
- Allison, S. D., Lu, Y., Weihe, C., Goulden, M. L., Martiny, A. C., Treseder, K. K., & Martiny, J.
 B. (2013). Microbial abundance and composition influence litter decomposition response to environmental change. Ecology, 94(3), 714-725.
- 375
 7. Veldhuis, M. P., Hulshof, A., Fokkema, W., Berg, M. P., & Olff, H. (2016). Understanding
 376 nutrient dynamics in an African savanna: local biotic interactions outweigh a major regional
 377 rainfall gradient. Journal of Ecology, 104(4), 913-923.
- Ashton, L. A., Griffiths, H. M., Parr, C. L., Evans, T. A., Didham, R. K., Hasan, F., ... &
 Eggleton, P. (2019). Termites mitigate the effects of drought in tropical rainforest. Science,
 363(6423), 174-177.

- Frey, S. D. (2019). Mycorrhizal fungi as mediators of soil organic matter dynamics. Annual
 Review of Ecology, Evolution, and Systematics, 50, 237-259.
- Blagodatskaya, E., & Kuzyakov, Y. (2013). Active microorganisms in soil: critical review of
 estimation criteria and approaches. Soil Biology and Biochemistry, 67, 192-211.
- 385 11. Chapin, F. S., Matson, P. A., Mooney, H. A., & Vitousek, P. M. (2002). Principles of terrestrial
 386 ecosystem ecology.
- 387 12. Dungait, J. A., Hopkins, D. W., Gregory, A. S., & Whitmore, A. P. (2012). Soil organic matter
 388 turnover is governed by accessibility not recalcitrance. Global Change Biology, 18(6), 1781389 1796.
- 390 13. Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M., & Charnov, E. L. (2001). Effects of
 391 size and temperature on metabolic rate. science, 293(5538), 2248-2251.
- 392 14. Hobbie, S. E., Schimel, J. P., Trumbore, S. E., & Randerson, J. R. (2000). Controls over
 393 carbon storage and turnover in high- latitude soils. Global change biology, 6(S1), 196-210.
- 394 15. Kemmitt, S. J., Lanyon, C. V., Waite, I. S., Wen, Q., Addiscott, T. M., Bird, N. R., ... &
 395 Brookes, P. C. (2008). Mineralization of native soil organic matter is not regulated by the size,
 396 activity or composition of the soil microbial biomass—a new perspective. Soil biology and
 397 Biochemistry, 40(1), 61-73.
- Lützow, M. V., Kögel- Knabner, I., Ekschmitt, K., Matzner, E., Guggenberger, G., Marschner,
 B., & Flessa, H. (2006). Stabilization of organic matter in temperate soils: mechanisms and
 their relevance under different soil conditions–a review. European journal of soil science,
 57(4), 426-445.
- 402 17. Rossiter-Rachor, N. A., Setterfield, S. A., Hutley, L. B., McMaster, D., Schmidt, S., & Douglas,
 403 M. M. (2017). Invasive Andropogon gayanus (Gamba grass) alters litter decomposition and
 404 nitrogen fluxes in an Australian tropical savanna. Scientific Reports, 7(1), 1-10.
- 405 18. Sterner, R. W., & Elser, J. J. (2017). Ecological stoichiometry. In Ecological Stoichiometry.
 406 Princeton university press.
- 407 19. Heuck, C., Weig, A., & Spohn, M. (2015). Soil microbial biomass C: N: P stoichiometry and
 408 microbial use of organic phosphorus. Soil Biology and Biochemistry, 85, 119-129.
- 409 20. Kuka, K., Franko, U., & Rühlmann, J. (2007). Modelling the impact of pore space distribution
 410 on carbon turnover. Ecological Modelling, 208(2-4), 295-306.
- 411 21. Peters, R.H. (1983) The Ecological Implications of Body Size, Cambridge University Press.

4	12	22	. Kleiber, M. (1932). Body size and metabolism. Hilgardia, 6(11), 315-353.
4 [.] 4 [.]	13 14	23	. Schmidt-Nielsen, K., & Knut, S. N. (1984). Scaling: why is animal size so important?. Cambridge university press.
4 ⁻ 4 ⁻ 4 ⁻	15 16 17	24	. Kerkhoff, A. J., Enquist, B. J., Elser, J. J., & Fagan, W. F. (2005). Plant allometry, stoichiometry and the temperature- dependence of primary productivity. Global Ecology and Biogeography, 14(6), 585-598.
4 [.] 4 [.]	18 19	25	. West, G. B., Brown, J. H., & Enquist, B. J. (1997). A general model for the origin of allometric scaling laws in biology. Science, 276(5309), 122-126.
42 42	20 21	26	. Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M., & West, G. B. (2004). Toward a metabolic theory of ecology. Ecology, 85(7), 1771-1789.
4: 4:	22 23	27	. Stevenson, R. D. (1985). Body size and limits to the daily range of body temperature in terrestrial ectotherms. The American Naturalist, 125(1), 102-117.
42 42 42 42	24 25 26 27	28	. McCafferty, D. J., Gilbert, C., Paterson, W., Pomeroy, P. P., Thompson, D., Currie, J. I., & Ancel, A. (2011). Estimating metabolic heat loss in birds and mammals by combining infrared thermography with biophysical modelling. Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology, 158(3), 337-345.
4: 4:	28 29	29	. Ruf, T., & Geiser, F. (2015). Daily torpor and hibernation in birds and mammals. Biological Reviews, 90(3), 891-926.
4: 4:	30 31	30	. Kooijman, S.A.L.M. (2000) Dynamic Energy and Mass Budgets in Biological Systems, Cambridge University Press.
4: 4:	32 33	31	. Haskell, J. P., Ritchie, M. E., & Olff, H. (2002). Fractal geometry predicts varying body size scaling relationships for mammal and bird home ranges. Nature, 418(6897), 527-530.
4: 4:	34 35	32	. Douglas, A. E. (2014). Symbiosis as a general principle in eukaryotic evolution. Cold Spring Harbor Perspectives in Biology, 6(2), a016113.
4: 4: 4:	36 37 38	33	. Martin, W. F., Garg, S., & Zimorski, V. (2015). Endosymbiotic theories for eukaryote origin. Philosophical Transactions of the Royal Society B: Biological Sciences, 370(1678), 20140330.
4: 44 44	39 40 41	34	 McFall-Ngai, M., Hadfield, M. G., Bosch, T. C., Carey, H. V., Domazet-Lošo, T., Douglas, A. E., & Wernegreen, J. J. (2013). Animals in a bacterial world, a new imperative for the life sciences. Proceedings of the National Academy of Sciences, 110(9), 3229-3236.

- 442 35. Parton, W., Silver, W. L., Burke, I. C., Grassens, L., Harmon, M. E., Currie, W. S., ... & Fasth,
 443 B. (2007). Global-scale similarities in nitrogen release patterns during long-term
 444 decomposition. science, 315(5810), 361-364.
- 36. Crowther, T. W., Van den Hoogen, J., Wan, J., Mayes, M. A., Keiser, A. D., Mo, L., ... &
 Maynard, D. S. (2019). The global soil community and its influence on biogeochemistry.
 Science, 365(6455), eaav0550.
- 448 37. Austin, A. T. (2011). Has water limited our imagination for aridland biogeochemistry?. Trends
 449 in ecology & evolution, 26(5), 229-235.
- 38. Olff, H., Ritchie, M. E., & Prins, H. H. (2002). Global environmental controls of diversity in
 large herbivores. Nature, 415(6874), 901-904.
- 452 39. Hempson, G. P., Archibald, S., & Bond, W. J. (2015). A continent-wide assessment of the
 453 form and intensity of large mammal herbivory in Africa. Science, 350(6264), 1056-1061.
- 454 40. Fokkema, W., de Boer, W., van der Jeugd, H. P., Dokter, A., Nolet, B. A., De Kok, L. J., ... &
 455 Olff, H. (2016). The nature of plant adaptations to salinity stress has trophic consequences.
 456 Oikos, 125(6), 804-811.
- 457 41. Foereid, B., Rivero, M. J., Primo, O., & Ortiz, I. (2011). Modelling photodegradation in the
 458 global carbon cycle. Soil Biology and Biochemistry, 43(6), 1383-1386.
- 459 42. Cornelissen, J. H. C., Quested, H. M., Gwynn-Jones, D., Van Logtestijn, R. S. P., De Beus,
 460 M. A. H., Kondratchuk, A., ... & Aerts, R. (2004). Leaf digestibility and litter decomposability
 461 are related in a wide range of subarctic plant species and types. Functional Ecology, 779-786.
- 462 43. Clemmensen, K. E., Finlay, R. D., Dahlberg, A., Stenlid, J., Wardle, D. A., & Lindahl, B. D.
 463 (2015). Carbon sequestration is related to mycorrhizal fungal community shifts during long464 term succession in boreal forests. New Phytologist, 205(4), 1525-1536.
- 465 44. Keeley, J. E., Bond, W. J., Bradstock, R. A., Pausas, J. G., & Rundel, P. W. (2012). Fire in
 466 Mediterranean ecosystems: ecology, evolution and management. Cambridge University
 467 Press.
- 468 45. McClain, M. E., Boyer, E. W., Dent, C. L., Gergel, S. E., Grimm, N. B., Groffman, P. M., ... &
 469 Pinay, G. (2003). Biogeochemical hot spots and hot moments at the interface of terrestrial
 470 and aquatic ecosystems. Ecosystems, 301-312.
- 46. Bierbaß, P., Gutknecht, J. L., & Michalzik, B. (2015). Nest-mounds of the yellow meadow ant
 (Lasius flavus) at the "Alter Gleisberg", Central Germany: Hot or cold spots in nutrient
 cycling?. Soil Biology and Biochemistry, 80, 209-217.

474	47.	Sagi, N., Grünzweig, J. M., & Hawlena, D. (2019). Burrowing detritivores regulate nutrient
475		cycling in a desert ecosystem. Proceedings of the Royal Society B, 286(1914), 20191647.
476	48.	Pett-Ridge, J., & Firestone, M. K. (2005). Redox fluctuation structures microbial communities
477		in a wet tropical soil. Applied and Environmental Microbiology, 71(11), 6998-7007.
478	49.	Castaño, C., Lindahl, B. D., Alday, J. G., Hagenbo, A., Martínez de Aragón, J., Parladé, J.,
479 480		Mediterranean pine forest. New Phytologist, 220(4), 1211-1221.
481	50.	Voříšková, J., Brabcová, V., Cajthaml, T., & Baldrian, P. (2014). Seasonal dynamics of fungal
482		communities in a temperate oak forest soil. New Phytologist, 201(1), 269-278.
483	51.	Šnajdr, J., Cajthaml, T., Valášková, V., Merhautová, V., Petránková, M., Spetz, P., &
484		Baldrian, P. (2011). Transformation of Quercus petraea litter: successive changes in litter
485		chemistry are reflected in differential enzyme activity and changes in the microbial community
486		composition. FEMS microbiology ecology, 75(2), 291-303.
487	52.	Ayari, A., Touihri, M., Ghemari, C., & Nasri-Ammar, K. (2018). Hourly and monthly variations
488		in the surface activity patterns of Hemilepistus reaumurii in arid environments of Tunisia.
489		Journal of Arid Land, 10(3), 470-481.
490	53.	Lepage, M. G. (1983). Foraging of Macrotermes spp.(Isoptera: Mcrotermitinae) in the tropics.
491		Social insects in the tropics, 2, 205-218.
492	54.	Freymann, B. P., De Visser, S. N., & Olff, H. (2010). Spatial and temporal hotspots of termite-
493		driven decomposition in the Serengeti. Ecography, 33(3), 443-450.
494	55.	Veldhuis, M. P., Laso, F. J., Olff, H., & Berg, M. P. (2017). Termites promote resistance of
495		decomposition to spatiotemporal variability in rainfall. Ecology, 98(2), 467-477.
496	56.	Fryxell, J. M., & Sinclair, A. R. E. (1988). Causes and consequences of migration by large
497		herbivores. Trends in ecology & evolution, 3(9), 237-241.
498	57.	Hopcraft, J. G. C., Olff, H., & Sinclair, A. R. (2010). Herbivores, resources and risks:
499		alternating regulation along primary environmental gradients in savannas. Trends in ecology
500		& evolution, 25(2), 119-128.
501	58.	Schimel, J. P., & Clein, J. S. (1996). Microbial response to freeze-thaw cycles in tundra and
502		taiga soils. Soil Biology and Biochemistry, 28(8), 1061-1066.
503	59.	Wipf, S., Sommerkorn, M., Stutter, M. I., Wubs, E. J., & Van Der Wal, R. (2015). Snow cover,
504		freeze-thaw, and the retention of nutrients in an oceanic mountain ecosystem. Ecosphere,
505		6(10), 1-16.

- 506 60. Gao, D., Bai, E., Yang, Y., Zong, S., & Hagedorn, F. (2021). A global meta-analysis on
 507 freeze-thaw effects on soil carbon and phosphorus cycling. Soil Biology and Biochemistry,
 508 159, 108283.
- 509 61. Bardgett, R. D., Streeter, T. C., & Bol, R. (2003). Soil microbes compete effectively with plants
 510 for organic-nitrogen inputs to temperate grasslands. Ecology, 84(5), 1277-1287.
- 511 62. Perez-Mon, C., Stierli, B., Plötze, M., & Frey, B. (2022). Fast and persistent responses of
 512 alpine permafrost microbial communities to in situ warming. Science of the Total Environment,
 513 807, 150720.
- 63. Morriën, E., Hannula, S. E., Snoek, L. B., Helmsing, N. R., Zweers, H., De Hollander, M., ... &
 Van Der Putten, W. H. (2017). Soil networks become more connected and take up more
 carbon as nature restoration progresses. Nature communications, 8(1), 1-10.
- 64. Banerjee, S., Walder, F., Büchi, L., Meyer, M., Held, A. Y., Gattinger, A., ... & van der Heijden,
 M. G. (2019). Agricultural intensification reduces microbial network complexity and the
 abundance of keystone taxa in roots. The ISME journal, 13(7), 1722-1736.
- 65. Manzoni, S., Chakrawal, A., Fischer, T., Schimel, J. P., Porporato, A., & Vico, G. (2020).
 Rainfall intensification increases the contribution of rewetting pulses to soil heterotrophic
 respiration. Biogeosciences, 17(15), 4007-4023.
- 66. Augustine, D. J., & McNaughton, S. J. (2004). Temporal asynchrony in soil nutrient dynamics
 and plant production in a semiarid ecosystem. Ecosystems, 7(8), 829-840.
- 525 67. Frank, D. A., Groffman, P. M., Evans, R. D., & Tracy, B. F. (2000). Ungulate stimulation of
 526 nitrogen cycling and retention in Yellowstone Park grasslands. Oecologia, 123(1), 116-121.
- 68. Olofsson, J., Stark, S., & Oksanen, L. (2004). Reindeer influence on ecosystem processes in
 the tundra. Oikos, 105(2), 386-396.
- 69. Bakker, E. S., Olff, H., Boekhoff, M., Gleichman, J. M., & Berendse, F. (2004). Impact of
 herbivores on nitrogen cycling: contrasting effects of small and large species. Oecologia,
 138(1), 91-101.
- 532 70. Yurtsev, B. A. (2001). The Pleistocene "Tundra-Steppe" and the productivity paradox: the
 533 landscape approach. Quaternary Science Reviews, 20(1-3), 165-174.
- 534 71. Zhu, D., Ciais, P., Chang, J., Krinner, G., Peng, S., Viovy, N., ... & Zimov, S. (2018). The large
 535 mean body size of mammalian herbivores explains the productivity paradox during the Last
 536 Glacial Maximum. Nature ecology & evolution, 2(4), 640-649.

- 537 72. Zimov, S. A., Chuprynin, V. I., Oreshko, A. P., Chapin Iii, F. S., Reynolds, J. F., & Chapin, M.
 538 C. (1995). Steppe-tundra transition: a herbivore-driven biome shift at the end of the
 539 Pleistocene. The American Naturalist, 146(5), 765-794.
- 540 73. Koven, C. D., Ringeval, B., Friedlingstein, P., Ciais, P., Cadule, P., Khvorostyanov, D., ... &
 541 Tarnocai, C. (2011). Permafrost carbon-climate feedbacks accelerate global warming.
 542 Proceedings of the National Academy of Sciences, 108(36), 14769-14774.
- 543 74. Macias-Fauria, M., Jepson, P., Zimov, N., & Malhi, Y. (2020). Pleistocene Arctic megafaunal
 544 ecological engineering as a natural climate solution?. Philosophical Transactions of the Royal
 545 Society B, 375(1794), 20190122:
- 546 75. Pellegrini, A. F., Ahlström, A., Hobbie, S. E., Reich, P. B., Nieradzik, L. P., Staver, A. C., ... &
 547 Jackson, R. B. (2018). Fire frequency drives decadal changes in soil carbon and nitrogen and
 548 ecosystem productivity. Nature, 553(7687), 194-198.
- 549 76. Kristensen, J. A., Svenning, J. C., Georgiou, K., & Malhi, Y. (2021). Can large herbivores
 550 enhance ecosystem carbon persistence?. Trends in ecology & evolution.
- 551 77. Malhi, Y., Lander, T., le Roux, E., Stevens, N., Macias-Fauria, M., Wedding, L., ... & Canney,
 552 S. (2022). The role of large wild animals in climate change mitigation and adaptation. Current
 553 Biology, 32(4), R181-R196.
- 554 78. Crowther, T. W., Todd-Brown, K. E., Rowe, C. W., Wieder, W. R., Carey, J. C., Machmuller,
 555 M. B., ... & Bradford, M. A. (2016). Quantifying global soil carbon losses in response to
 556 warming. Nature, 540(7631), 104-108.
- 557 79. Feng, Z., Rütting, T., Pleijel, H., Wallin, G., Reich, P. B., Kammann, C. I., ... & Uddling, J.
 558 (2015). Constraints to nitrogen acquisition of terrestrial plants under elevated CO 2. Global
 559 change biology, 21(8), 3152-3168.
- 560 80. Steidinger, B. S., Crowther, T. W., Liang, J., Van Nuland, M. E., Werner, G. D., Reich, P. B.,
 561 ... & Peay, K. G. (2019). Climatic controls of decomposition drive the global biogeography of
 562 forest-tree symbioses. Nature, 569(7756), 404-408.
- 563 81. Pacifici, M., Visconti, P., Butchart, S. H., Watson, J. E., Cassola, F. M., & Rondinini, C.
 564 (2017). Species' traits influenced their response to recent climate change. Nature Climate
 565 Change, 7(3), 205-208.
- 566 82. Fuller, A., Mitchell, D., Maloney, S. K., & Hetem, R. S. (2016). Towards a mechanistic
 567 understanding of the responses of large terrestrial mammals to heat and aridity associated
 568 with climate change. Climate Change Responses, 3(1), 1-19.

- 569 83. Ofstad, E. G., Herfindal, I., Solberg, E. J., & Sæther, B. E. (2016). Home ranges, habitat and
 570 body mass: simple correlates of home range size in ungulates. Proceedings of the Royal
 571 Society B: Biological Sciences, 283(1845), 20161234.
- 572 84. Veldhuis, M. P., Hofmeester, T. R., Balme, G., Druce, D. J., Pitman, R. T., & Cromsigt, J. P.
 573 (2020). Predation risk constrains herbivores' adaptive capacity to warming. Nature Ecology &
 574 Evolution, 4(8), 1069-1074.
- 575 85. Davies, A. B., Brodrick, P. G., Parr, C. L., & Asner, G. P. (2020). Resistance of mound576 building termites to anthropogenic land-use change. Environmental Research Letters, 15(9),
 577 094038.
- 86. Onrust, J., Wymenga, E., Piersma, T., & Olff, H. (2019). Earthworm activity and availability for
 meadow birds is restricted in intensively managed grasslands. Journal of Applied Ecology,
 56(6), 1333-1342.
- 581 87. Soudzilovskaia, N. A., van Bodegom, P. M., Terrer, C., Zelfde, M. V. T., McCallum, I., Luke
 582 McCormack, M., ... & Tedersoo, L. (2019). Global mycorrhizal plant distribution linked to
 583 terrestrial carbon stocks. Nature communications, 10(1), 1-10.
- 584 88. Bakker, E. S., & Svenning, J. C. (2018). Trophic rewilding: impact on ecosystems under
 585 global change. Philosophical Transactions of the Royal Society B: Biological Sciences,
 586 373(1761), 20170432.
- 587 89. Cromsigt, J. P., & Te Beest, M. (2014). Restoration of a megaherbivore: landscape-level
 588 impacts of white rhinoceros in K ruger N ational P ark, S outh A frica. Journal of Ecology,
 589 102(3), 566-575.
- 590 90. Johnson, C. N., Prior, L. D., Archibald, S., Poulos, H. M., Barton, A. M., Williamson, G. J., &
 591 Bowman, D. M. (2018). Can trophic rewilding reduce the impact of fire in a more flammable
 592 world?. Philosophical Transactions of the Royal Society B: Biological Sciences, 373(1761),
 593 20170443.
- 594 91. Hawkins, B. J., Jones, M. D., & Kranabetter, J. M. (2015). Ectomycorrhizae and tree seedling
 595 nitrogen nutrition in forest restoration. New Forests, 46(5), 747-771.
- 596 92. Aanen, D. K., & Eggleton, P. (2005). Fungus-growing termites originated in African rain forest.
 597 Current biology, 15(9), 851-855.
- 598 93. da Costa, R. R., Hu, H., Li, H., & Poulsen, M. (2019). Symbiotic plant biomass decomposition
 599 in fungus-growing termites. Insects, 10(4), 87.
- 600 94. Mando, A., Stroosnijder, L., & Brussaar, L. (1996). Effects of termites on infiltration into
 601 crusted soil. Geoderma, 74(1-2), 107-113.

- 502 95. Turner, J. S. (2019). Termites as mediators of the water economy of arid savanna
 603 ecosystems. In Dryland ecohydrology (pp. 401-414). Springer, Cham.
- 604 96. Allen, M. F. (2007). Mycorrhizal fungi: highways for water and nutrients in arid soils. Vadose
 605 Zone Journal, 6(2), 291-297.
- 606 97. Mohan, J. E., Cowden, C. C., Baas, P., Dawadi, A., Frankson, P. T., Helmick, K., ... & Witt, C.
 607 A. (2014). Mycorrhizal fungi mediation of terrestrial ecosystem responses to global change:
 608 mini-review. Fungal Ecology, 10, 3-19.
- 80. Voltas, J., Lucabaugh, D., Chambel, M. R., & Ferrio, J. P. (2015). Intraspecific variation in the
 use of water sources by the circum- Mediterranean conifer Pinus halepensis. New
 Phytologist, 208(4), 1031-1041.
- 612 99. Solden, L. M., Naas, A. E., Roux, S., Daly, R. A., Collins, W. B., Nicora, C. D., ... & Wrighton,
 613 K. C. (2018). Interspecies cross-feeding orchestrates carbon degradation in the rumen
 614 ecosystem. Nature microbiology, 3(11), 1274-1284.
- 615 100. Sauro, H. M. (2017). Control and regulation of pathways via negative feedback.
 616 Journal of The Royal Society Interface, 14(127), 20160848.
- 617 101. Poulsen, M., Hu, H., Li, C., Chen, Z., Xu, L., Otani, S., ... & Zhang, G. (2014).
 618 Complementary symbiont contributions to plant decomposition in a fungus-farming termite.
 619 Proceedings of the National Academy of Sciences, 111(40), 14500-14505.
- 620 102. Nobre, T., & Aanen, D. K. (2012). Fungiculture or termite husbandry? The ruminant
 621 hypothesis. *Insects*, *3*(1), 307-323.
- 622 103. Koide, R. T., Sharda, J. N., Herr, J. R., & Malcolm, G. M. (2008). Ectomycorrhizal
 623 fungi and the biotrophy-saprotrophy continuum. New Phytologist, 230-233.
- 624 104. Gadgil, R. L., & Gadgil, P. D. (1971). Mycorrhiza and litter decomposition. Nature,
 625 233(5315), 133-133.
- 626 105. Read, D. J., Leake, J. R., & Perez-Moreno, J. (2004). Mycorrhizal fungi as drivers of
 627 ecosystem processes in heathland and boreal forest biomes. Canadian Journal of Botany,
 628 82(8), 1243-1263.
- 629 106. Veldhuis, M. P., Berg, M. P., Loreau, M., & Olff, H. (2018). Ecological autocatalysis: a
 630 central principle in ecosystem organization?. Ecological Monographs, 88(3), 304-319.
- 631 107. Jones, J. A. (1990). Termites, soil fertility and carbon cycling in dry tropical Africa: a
 632 hypothesis. Journal of Tropical Ecology, 6(3), 291-305.

633 108. Keya, S. O., Mureria, N. K., & Arshad, M. A. (1982). Population dynamics of soil
634 microorganisms in relation to proximity of termite mounds in Kenya. Journal of Arid
635 Environments, 5(4), 353-360.