

1 **When and how does photoinhibition matter for plant fitness?**

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8 Running head: Photoinhibition and plant fitness

9

10 **Abstract**

11 The many biophysical factors that shape how plant species sort across environmental gradients may  
12 include photoinhibition, which I define broadly as oxidative damage that plants and other phototrophs risk  
13 incurring when they absorb excess light energy they cannot safely dissipate. Photoinhibition is seldom  
14 explicitly discussed as a potential driver of plant fitness and distributions. Here, I aim to show that it can  
15 be one by drawing on studies showing that natural gradients or experimental manipulations that increase  
16 the risk of photoinhibition thereby decrease plant fitness, or favor species with stronger photoprotective  
17 adaptations. A corollary is that alleviation of photoinhibition may be a common mechanism of  
18 facilitation. A large share of this research is set in stressful environments like alpine treelines or drylands,  
19 most likely because photoinhibition is most detrimental in the presence of other interacting stress factors.  
20 Nevertheless, knowing the specific role of photoinhibition may have practical value—for example, in  
21 understanding the uses of shading treatments in ecological restoration.

22

23 Much research assumes that photoinhibition alters fitness by reducing carbon assimilation. Nevertheless,  
24 a tension exists in ecological literature on photoinhibition: many kinds of stressful conditions that threaten  
25 oxidative damage to photosynthesis also directly constrain tissue expansion. If carbon sink strength  
26 declines more than source strength—as it often does—a carbon surplus may result and lead to feedback  
27 inhibition of photosynthesis. Under persistent carbon surplus, photoinhibition’s potential influence on  
28 carbon status may not matter much for fitness. This fact might be reconciled with photoinhibition’s  
29 known effects on plant fitness by considering its other consequences. Oxidative damage can have

30 consequences far beyond photosynthesis—altering membranes, blocking phloem loading, and even  
31 triggering cell death. While such effects are harder to measure than photosynthesis, doing so may enrich  
32 our understanding of photoinhibition’s ecological consequences.

33

34 Keywords: fitness; photoinhibition; photoprotection; photosynthesis; reactive oxygen species (ROS); sink  
35 limitation

36

37 *Introduction*

38 The patterns of plant population and community ecology emerge from a few high-level processes, among  
39 which the best studied is ecological selection caused by variation in fitness across environmental  
40 gradients (Vellend 2016). One goal of plant ecophysiology is to explain how such variation emerges from  
41 the vast diversity of plant function at the level of tissues and cells. This knowledge is essential for  
42 predicting how global change will affect plant populations (Cabal et al. 2022).

43  
44 Among the countless biophysical forces that may alter plant fitness, photoinhibition is among the most  
45 enigmatic. Here, I define photoinhibition as damage caused by non-thermal energetic consequences of  
46 light absorption. Kok (1956) provided some of the first evidence of photoinhibition by showing that algal  
47 cultures had lower photosynthesis after exposure to intense light. The study of photoinhibition gained  
48 momentum in the 1980s as researchers learned more about the structure and function of Photosystem II  
49 (PSII), whose D1 protein is particularly susceptible to inactivation by light (Anderson et al. 1998). Also  
50 around this time, the advent of tools like chlorophyll fluorescence made it easy to measure PSII  
51 efficiency. Light-induced damage is usually considered as a result of photosynthetically active radiation  
52 (PAR; 400-700 nm), although much of the ultraviolet (UV) range can also cause it (Takahashi et al.  
53 2010). UV-B can also increase mutation rates by creating pyrimidine dimers in DNA (Schoen & Schultz  
54 2019), which may also alter fitness in a way that is hard to extricate from effects on PSII, except perhaps  
55 by selectively filtering parts of the light spectrum (Lesser 1996; van de Poll et al. 2001).

56  
57 Both PSII and Photosystem I (PSI) are susceptible to photoinhibition. Indeed, PSI damage can be more  
58 severe because it is harder to reverse, but it is also much rarer (Sonoike 2011), so lasting photoinhibition  
59 is most often taken to occur when the rate of PSII damage exceeds the rate of repair through D1  
60 replacement (Long et al. 1994). This situation may occur when photosynthetic pigments absorb light  
61 energy in excess of the plant's ability to dissipate it safely through either photochemical or non-  
62 photochemical quenching (e.g. through the xanthophyll cycle; Demmig-Adams et al. 1996). Because

63 stresses like low temperatures or drought reduce photosynthesis, they increase the share of light in excess  
64 and exacerbate photoinhibition. The predominant mechanism(s) of this effect are still debated. Excitation  
65 energy transfer and electron transport generate reactive oxygen species (ROS) at both PSI and PSII.  
66 Damage to PSII may occur at either its acceptor or its donor side (Pospíšil 2016; Kale et al. 2017). Under  
67 an ‘acceptor-side’ mechanism, ROS are generated and cause D1 inactivation in proportion to the amount  
68 of excess light (Vass 2011). Under a ‘donor-side’ mechanism, D1 inactivation occurs in proportion to the  
69 *total* amount of light absorbed (Tyystjärvi & Aro 1996), but ROS generated by excess light inhibit D1  
70 replacement (Takahashi & Murata 2008). Acceptor- and donor-side mechanisms produce different kinds  
71 of ROS, and perhaps at different levels depending on the quantity and spectrum of light and the degree of  
72 interacting stresses (Hakala et al. 2005; Pospíšil 2016), which may cause them to have different  
73 ecological consequences.

74  
75 In defining photoinhibition as involving ‘damage,’ I distinguish it from biochemical mechanisms of  
76 photoprotection that protect against damage that is longer lasting and more costly to reverse. The line  
77 between sustained biochemical photoprotection and damage is blurry, since it is often ambiguous whether  
78 a given molecular process that indicates reduced electron transport function (like declines in functioning  
79 D1) is, within a certain environmental context, best interpreted as a result of photoinhibitory damage or as  
80 an adaptive mechanism that *averts* more severe consequences (Adams et al. 2013; Tikkanen et al. 2014;  
81 Malnoë 2018). Beyond just the biochemical scale, photoprotective mechanisms are incredibly diverse,  
82 including: positioning of leaves to reduce light absorption, reflective leaf structures, chloroplast  
83 movement, light-screening pigments like flavonoids and anthocyanins, reductions in chlorophyll antenna  
84 size, non-photochemical quenching (NPQ) by pigments, and ROS scavenging by antioxidants (Raven  
85 2011; D’Alessandro 2020). Many of these mechanisms have known trade-offs; for example, all plants  
86 grow in environments where light fluctuates on some time-scale, but declines in light absorption or  
87 induction of NPQ under high light can reduce photosynthesis when light declines (Raven 2011; De Souza

88 et al. 2022). Plants might thus be expected to evolve towards an optimal level of photoprotection for their  
89 growth environment.

90  
91 Although the mechanisms of photoinhibition and photoprotection have become clearer through time, their  
92 relevance for ecology remains obscure. Here, I aim to describe what we know about photoinhibition's  
93 effects on fitness and to re-evaluate this knowledge in the context of our changing understanding of  
94 whole-plant function. I focus on land plants, although at times I draw examples from other photosynthetic  
95 organisms.

96  
97 *Can excess light reduce fitness?*

98 Here, I try to establish that photoinhibition can damage plant fitness. For multiple reasons, I propose to  
99 avoid using biochemical or physiological indicators alone to infer that photoinhibition has ecological  
100 consequences. The first reason is the aforementioned ambiguity about when such indicators are best  
101 interpreted as photoinhibition or photoprotection. The second is that photosynthetic efficiency—as  
102 estimated by fluorescence-based measures like (dark- or light-acclimated)  $F_v/F_m$ —is not the same as  
103 photosynthesis. Given that light is not just a stressor but also an essential resource, an increase in light  
104 very often causes both a decline in photosynthetic efficiency and an increase in photosynthesis, and at  
105 high light photosynthesis is usually limited by RuBisCO kinetics rather than electron transport. The third  
106 reason is that photosynthetic carbon fixation is not always limiting for fitness—a point to which I return  
107 later.

108  
109 So how can we show that photoinhibition does (or doesn't) matter? I focus on evidence about how excess  
110 light affects plant fitness and its consequences, like species distributions, drawing on physiological  
111 evidence only in a supporting role (Fig. 1). Such evidence would imply that fitness declines as incident  
112 light increases, all else held equal. This stipulation that all else be held equal is hard to meet exactly, since  
113 gradients of light (artificial or natural) are often confounded with factors like air temperature and

114 humidity (Niinemets & Valladares 2004). While these other factors should be controlled as well as  
115 possible, experiments need not be done in the complete absence of other abiotic stresses. Indeed, most  
116 abiotic stresses are only dangerous in combination (Mittler 2006; Zandalinas et al. 2021). What *is*  
117 important is that the severity of other stresses stay relatively constant across light environments.

118

119 First, I note that if photoinhibition posed no threat, it would be hard to explain the ubiquity of  
120 photoprotective pathways like the xanthophyll cycle(s) (Esteban et al. 2009; Goss & Lepetit 2015).

121 Likewise, many lineages have putative light avoidance ‘behaviors’ like chloroplast movement in plants  
122 (Howard et al. 2020) and whole organismal movement in phytoplankton (Raven 2011). Species found in  
123 bright environments are often better at avoiding or recovering from photoinhibition (Montgomery et al.  
124 2008; Kothari et al. 2021; Fig. 1A). This pattern holds even when the plants are grown in common  
125 garden-like environments, which implies that it is not just an outcome of plasticity. Indeed, organisms in  
126 extremely bright, hot environments often have extraordinary mechanisms to avoid and dissipate excess  
127 light (Levin et al. 2021). It would require formal evidence synthesis to tell whether such patterns are truly  
128 consistent—but if so, it would imply that photoprotection is important for maintaining fitness under high  
129 light.

130

131 Another form of evidence comes from experiments that grow plants across light environments and show  
132 that high light causes declines in fitness (Fig. 1B). Researchers have long described how high light can  
133 suppress population growth in phytoplankton (Edwards et al. 2015; Croteau et al. 2022), and these  
134 experiments generally raise fewer concerns about confounding between light and other stress factors.

135 Another simplified setting for detecting the influence of photoinhibition is with photoprotection-deficient  
136 mutants of species like *Arabidopsis*. Under natural conditions, mutants deficient in NPQ produce fewer  
137 seeds (Külheim et al. 2002) and mutants deficient in chloroplast movement grow less (Howard et al.  
138 2020).

139

140 Interpretation can get more complicated among land plants in more natural settings. Appendix S1  
141 summarizes many case studies that provide plausible evidence that increases in light can reduce growth or  
142 fitness. Many of these studies monitored the microclimate to test whether their treatments allowed them to  
143 isolate the effect of light. For example, as part of a much broader study of treelines (see also Germino &  
144 Smith 1999), Germino et al. (2002) found that survival in alpine spruce seedlings was halved when  
145 overtopping grasses were removed, even though the removal treatment reduced water and low-  
146 temperature stress. Velasco & Becerra (2020) found that artificial shading had a much more beneficial  
147 effect than irrigation on tree seedling survival in a semiarid grassland, which suggests that the effect of  
148 shading was not due to water status. Bader et al. (2006) used a series of shading and neighbor removal  
149 treatments to show that high light could account for much of the failure of trees to establish beyond a  
150 tropical alpine treeline. Many of the case studies summarized in Appendix S1 took place in typically  
151 ‘stressful’ settings like alpine treelines and drylands, where interacting stresses like low temperature or  
152 drought may make it more plausible that photoinhibition matters. Most studies on trees were also  
153 conducted using seedlings; this may be partly a matter of convenience, but it may also reflect a concern  
154 that early recruitment is an important stage where trees are particularly vulnerable to photoinhibition  
155 (Germino et al. 2002; Slot et al. 2005).

156  
157 A corollary to the apparent importance of photoinhibition is that shading may be a common mechanism of  
158 facilitation. This point is again clear in phytoplankton, among which theoretical (Gerla et al. 2011) and  
159 empirical (Veraart et al. 2012; Faassen et al. 2015) research suggests that intraspecific facilitative  
160 amelioration of photoinhibition cause Allee effects. In trees, Kothari et al. (2021) found that interspecific  
161 facilitation of shade-tolerant species by larger neighbors contributed to whole-community overyielding.  
162 The importance of facilitation between adults and seedlings is also implicit in much of the research on  
163 recruitment at treelines or in canopy gaps (Ball et al. 1991; Egerton et al. 2000; Slot et al. 2005). In the  
164 context of facilitation research guided by the stress-gradient hypothesis (Bertness & Callaway 1994),

165 amelioration of photoinhibition may be one of many mechanisms through which nurse plants aid their  
166 beneficiaries in stressful environments (Cabal et al. 2022).

167

168 Understanding that photoinhibition can reduce fitness can also help us make sense of the efficacy of  
169 certain ecological interventions. For example, reforestation experiments often find that experimental  
170 shading increases survival (Egerton et al. 2000; Velasco & Becerra 2020). While high light is usually  
171 most harmful in combination with other stresses like drought, it may be less expensive to reduce the  
172 intensity of light than to alleviate those other stresses.

173

#### 174 *A whole-plant perspective on photoinhibition*

175 So far, I claim there is evidence that photoinhibition can harm plant fitness. Given that photoinhibition  
176 involves damage to thylakoid membrane complexes, many studies of photoinhibition implicitly assume  
177 that its main cost to fitness is to reduce photosynthesis. Indeed, most attempts at quantifying the costs of  
178 photoinhibition focus on energetic or other resource-related costs, both of foregone photosynthesis and of  
179 protein repair (Long et al. 1994; Raven 2011; Murchie & Niyogi 2011).

180

181 But is this assumption true? The answer depends on how carbon-limited fitness is. In our high-[CO<sub>2</sub>]  
182 world, plant growth under moderate stress is often regulated less by carbon assimilation than by direct  
183 constraints on tissue expansion or carbon export from leaves (Cabon et al. 2022; Fig. 2). In other words,  
184 plants are often sink-limited, particularly under many of the conditions that promote photoinhibition: the  
185 intersection of high light with drought, nutrient limitation, or cold temperatures (Körner 2003). The result  
186 often manifests as a buildup of non-structural carbohydrates (NSCs) in tissues (Körner 2003). Such NSCs  
187 in apparent excess could still be useful as osmolytes that promote drought tolerance, or as insurance  
188 against future disturbance (Sala et al. 2012). Nevertheless, when plant fitness is not carbon-limited, the  
189 apparent importance of photoinhibition and photoprotection becomes a puzzle (Slot et al. 2005).

190

191 These considerations led Adams et al. (2013) to propose that photoinhibition is primarily a *consequence*  
192 of growth limitation, not a cause. Accumulation of NSCs causes plants to reorganize their metabolism to  
193 downregulate photosynthesis and promote sink activity (White et al. 2016; Fig. 2). Experimental  
194 stimulation of source activity or blockage of sink activity can cause many of the physiological hallmarks  
195 of photoinhibition, including declines in dark  $F_v/F_m$  and D1 activity (Adams et al. 2013). Many plants  
196 acclimate by upregulating photoprotective mechanisms (e.g. NPQ) enough to balance the decline in  
197 photochemical quenching, allowing them to avoid damage. Without taking a position on the prevalence of  
198 source- vs. sink-limitation under photoinhibitory conditions, I think this challenge should occasion a  
199 reconsideration of *how* photoinhibition matters for plant fitness.

200  
201 On its own, this account lacks an explanation for how photoinhibition could be harmful for fitness.  
202 Ecophysiologicalists often refer to the easily measurable consequences of excess light-induced ROS  
203 generation in reducing photosynthetic function, but it may help to look toward other aspects of cellular  
204 function. In particular, molecular physiologists have mapped out a much wider range of consequences of  
205 excess light-induced ROS generation (D'Alessandro et al. 2020)—but primarily in a few model  
206 organisms and without having yet demonstrated their importance in nature.

207  
208 It may be time for a synthesis: Could excess light-induced ROS generation have an ecological role in  
209 natural settings beyond its influence on carbon assimilation via electron transport? The species and  
210 quantity of ROS produced are dependent on the mechanism (acceptor-side, donor-side, or other) and  
211 intensity of photoinhibition. ROS species differ in their lifetimes, reactivities, and ability to cross  
212 membranes (Møller et al. 2007; Mittler 2017). In general, though, ROS like singlet oxygen produced  
213 along the electron transport chain can attack the unsaturated lipids in the thylakoid and produce lipid  
214 peroxides (Demmig-Adams et al. 2014; D'Alessandro et al. 2020). These in turn can break into reactive  
215 carbonyl species (RCS), which through their role as signaling molecules and their toxicity can—in  
216 sufficient quantities—damage other components and trigger pathways towards cell death (Møller et al.

217 2007; Chan et al. 2012; D’Alessandro et al. 2020). Even before this point, the signaling functions of ROS  
218 can also trigger other processes beyond the chloroplast, like callose deposition in phloem-loading  
219 complexes, which may serve to block the spread of pathogens (when present) but hinders the export of  
220 sugars and other compounds from leaves (Demmig-Adams et al. 2014).

221  
222 Under certain circumstances, these consequences may be much more relevant than reductions in carbon  
223 assimilation. For example, although most research on PSII inhibition’s influence on mortality in  
224 seagrasses and algae focuses on the potential for carbon starvation, oxidative damage appears to be a  
225 stronger driver (King et al. 2021). Prescott et al. (2020) even suggest that sink-limited plants may dispose  
226 of surplus carbon to avoid feedback inhibition of photosynthesis and the resulting risk of  
227 photoinhibition—a proposal that could only make sense if photoinhibition does more than just reduce  
228 carbon assimilation.

229  
230 Whether any of these adverse consequences occurs depends on the quantity of the potentially harmful  
231 molecules and on plants’ ability to contain them. Within a certain range, ROS and lipid peroxides are  
232 essential as redox signals that coordinate development, signaling, and acclimation to stressful conditions,  
233 including by upregulating photoprotection (Mittler 2006; Foyer 2018). The danger may come when plants  
234 are unable to stop runaway spirals of ROS production. For example, if unrepaired damage to  
235 photosynthetic machinery limits plants’ ability to dissipate light photochemically, the amount of light in  
236 excess may increase, which could increase ROS production and perhaps create a vicious cycle in which  
237 photoinhibition begets more photoinhibition. This possibility is implied by findings that PSII-inhibiting  
238 herbicides can, depending on their mechanism, steeply enhance ROS production (Fufezan et al. 2002;  
239 Chen et al. 2010). ROS and RCS may also attack and inactivate the enzymes that scavenge and detoxify  
240 them (D’Alessandro et al. 2020). Plants adapted to bright or fluctuating light conditions often have a  
241 strong capacity to acclimate and keep ROS within a safe range through protective mechanisms like NPQ  
242 or ROS scavenging, but plants adapted to shade may have more limited capacity (Fig. 2). Interacting

243 stresses may also limit plants' abilities to keep ROS within a safe range. These phenomena may explain  
244 observations of photooxidative bleaching (Powles 1984) or leaf abscission (Egerton et al. 2000) under  
245 high light. Hence, it seems inapt to reach the Panglossian conclusion that ROS generation is always  
246 helpful or benign, or that symptoms like PSII inhibition are always protective rather than an indication of  
247 damage to fitness. Some researchers have suggested that photoinhibition may only be important in  
248 'unrealistic' circumstances, as when plants are outside of their usual environmental niche (Adams et al.  
249 2013). But this is no reason to dismiss its importance: part of the business of community ecology is to  
250 explain why plants don't grow where they don't grow.

251

252 One specific situation in which photoinhibition may reduce fitness other than through energetic costs is  
253 during senescence in winter- (and perhaps drought-) deciduous plants. Efficient nutrient resorption during  
254 senescence is critical for growth in the following year (El Zein et al. 2011) and consequently for fitness  
255 (May & Killingbeck 1992). Senescing leaves are highly vulnerable to light-induced damage (Kar et al.  
256 1993; Merzlyak & Hendry 1994), and the reddish anthocyanins many species produce in the fall may  
257 serve to improve foliar nutrient resorption through photoprotection (Hoch et al. 2003; Renner & Zohner  
258 2019). While failures of nutrient resorption could result from photoinhibitory carbon depletion (Hoch et  
259 al. 2001), they might also result from oxidative damage to proteins involved in breaking down nutrient-  
260 containing molecules, or from callose-induced blockage of phloem loading.

261

262 I close with some ideas for ecologists to explore the potential consequences of photoinhibition more fully.  
263 To establish whether photoinhibition matters at all for fitness, it is important to measure growth or  
264 demography rather than relying solely on physiological indicators. To establish whether photoinhibitory  
265 conditions deplete carbon reserves, it could help to measure NSCs (Slot et al. 2005), although this  
266 approach is best treated just as a heuristic (Weber et al. 2019). In general, photosynthetic and chlorophyll  
267 fluorescence indicators may be useful but should be chosen and interpreted with care. A particular  
268 parameter that may be useful for studying oxidative damage is the fraction of light not dissipated through

269 either photochemistry or NPQ (sometimes denoted  $\Phi_{\text{NO}}$ ; Demmig-Adams et al. 1996; Kramer et al. 2004)  
270 may help indicate the potential for ROS generation (Fig. 2). Studying oxidative damage at the molecular  
271 level is tricky under field conditions, but lipid peroxidation may be a good candidate for measurement  
272 using the relatively simple TBARS assay (Hodges et al. 1999). Given the clear evidence that human  
273 activity is altering global light regimes (Norris et al. 2016; Lu et al. 2017), it is important to understand  
274 when and how photoinhibition matters—not just in the leaf, but across the whole plant.

275

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279

### 280 **Author Contributions**

281 Shan Kothari is the sole author of this paper.

282

### 283 **Data Availability Statement**

284 This is a synthesis article and contains no data.

285 **Literature Cited**

- 286 Adams, W. W., Muller, O., Cohu, C. M., & Demmig-Adams, B. (2013). May photoinhibition be a  
287 consequence, rather than a cause, of limited plant productivity? *Photosynthesis Research*, 117(1),  
288 31–44. <https://doi.org/10.1007/s11120-013-9849-7>
- 289 Anderson, J. M., Park, Y.-I., & Chow, W. S. (1998). Unifying model for the photoinactivation of  
290 Photosystem II in vivo under steady-state photosynthesis. *Photosynthesis Research*, 56(1), 1–13.  
291 <https://doi.org/10.1023/A:1005946808488>
- 292 Bader, M. Y., van Geloof, I., & Rietkerk, M. (2007). High solar radiation hinders tree regeneration above  
293 the alpine treeline in northern Ecuador. *Plant Ecology*, 191(1), 33–45.  
294 <https://doi.org/10.1007/s11258-006-9212-6>
- 295 Ball, M. C., Hodges, V. S., & Laughlin, G. P. (1991). Cold-Induced Photoinhibition Limits Regeneration  
296 of Snow Gum at Tree-Line. *Functional Ecology*, 5(5), 663–668. <https://doi.org/10.2307/2389486>
- 297 Bertness, M. D., & Callaway, R. (1994). Positive interactions in communities. *Trends in Ecology &*  
298 *Evolution*, 9(5), 191–193. [https://doi.org/10.1016/0169-5347\(94\)90088-4](https://doi.org/10.1016/0169-5347(94)90088-4)
- 299 Cabal, C., Valladares, F., & Martinez-Garcia, R. (2022). *The Ecology of Plant Interactions: A Giant with*  
300 *Feet of Clay*. <https://doi.org/10.20944/preprints202009.0520.v3>
- 301 Cabon, A., Kannenberg, S. A., Arain, A., Babst, F., Baldocchi, D., Belmecheri, S., Delpierre, N.,  
302 Guerrieri, R., Maxwell, J. T., McKenzie, S., Meinzer, F. C., Moore, D. J. P., Pappas, C., Rocha,  
303 A. V., Szejner, P., Ueyama, M., Ulrich, D., Vincke, C., Voelker, S. L., ... Anderegg, W. R. L.  
304 (2022). Cross-biome synthesis of source versus sink limits to tree growth. *Science*, 376(6594),  
305 758–761. <https://doi.org/10.1126/science.abm4875>
- 306 Chan, T., Shimizu, Y., Pospíšil, P., Nijo, N., Fujiwara, A., Taninaka, Y., Ishikawa, T., Hori, H., Nanba,  
307 D., Imai, A., Morita, N., Yoshioka-Nishimura, M., Izumi, Y., Yamamoto, Y., Kobayashi, H.,  
308 Mizusawa, N., Wada, H., & Yamamoto, Y. (2012). Quality Control of Photosystem II: Lipid  
309 Peroxidation Accelerates Photoinhibition under Excessive Illumination. *PLOS ONE*, 7(12),  
310 e52100. <https://doi.org/10.1371/journal.pone.0052100>

311 Chen, S., Yin, C., Qiang, S., Zhou, F., & Dai, X. (2010). Chloroplastic oxidative burst induced by  
312 tenuazonic acid, a natural photosynthesis inhibitor, triggers cell necrosis in *Eupatorium*  
313 *adenophorum* Spreng. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, 1797(3), 391–405.  
314 <https://doi.org/10.1016/j.bbabi.2009.12.007>

315 Croteau, D., Lacour, T., Schiffrine, N., Morin, P.-I., Forget, M.-H., Bruyant, F., Ferland, J., Lafond, A.,  
316 Campbell, D. A., Tremblay, J.-É., Babin, M., & Lavaud, J. (2022). Shifts in growth light optima  
317 among diatom species support their succession during the spring bloom in the Arctic. *Journal of*  
318 *Ecology*, 110(6), 1356–1375. <https://doi.org/10.1111/1365-2745.13874>

319 D’Alessandro, S., Beaugelin, I., & Havaux, M. (2020). Tanned or Sunburned: How Excessive Light  
320 Triggers Plant Cell Death. *Molecular Plant*, 13(11), 1545–1555.  
321 <https://doi.org/10.1016/j.molp.2020.09.023>

322 De Souza, A. P., Burgess, S. J., Doran, L., Hansen, J., Manukyan, L., Maryn, N., Gotarkar, D., Leonelli,  
323 L., Niyogi, K. K., & Long, S. P. (2022). Soybean photosynthesis and crop yield are improved by  
324 accelerating recovery from photoprotection. *Science*, 377(6608), 851–854.  
325 <https://doi.org/10.1126/science.adc9831>

326 Demmig-Adams, B., Adams III, W. W., Barker, D. H., Logan, B. A., Bowling, D. R., & Verhoeven, A. S.  
327 (1996). Using chlorophyll fluorescence to assess the fraction of absorbed light allocated to  
328 thermal dissipation of excess excitation. *Physiologia Plantarum*, 98(2), 253–264.  
329 <https://doi.org/10.1034/j.1399-3054.1996.980206.x>

330 Demmig-Adams, B., Stewart, J. J., & Adams, W. W. (2014). Multiple feedbacks between chloroplast and  
331 whole plant in the context of plant adaptation and acclimation to the environment. *Philosophical*  
332 *Transactions of the Royal Society B: Biological Sciences*, 369(1640), 20130244.  
333 <https://doi.org/10.1098/rstb.2013.0244>

334 Edwards, K. F., Thomas, M. K., Klausmeier, C. A., & Litchman, E. (2015). Light and growth in marine  
335 phytoplankton: Allometric, taxonomic, and environmental variation. *Limnology and*  
336 *Oceanography*, 60(2), 540–552. <https://doi.org/10.1002/lno.10033>

337 Egerton, J. J. G., Banks, J. C. G., Gibson, A., Cunningham, R. B., & Ball, M. C. (2000). Facilitation of  
338 Seedling Establishment: Reduction in Irradiance Enhances Winter Growth of Eucalyptus  
339 Pauciflora. *Ecology*, 81(5), 1437–1449. [https://doi.org/10.1890/0012-  
340 9658\(2000\)081\[1437:FOSERI\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2000)081[1437:FOSERI]2.0.CO;2)

341 El Zein, R., Bréda, N., Gérant, D., Zeller, B., & Maillard, P. (2011). Nitrogen sources for current-year  
342 shoot growth in 50-year-old sessile oak trees: An in situ <sup>15</sup>N labeling approach. *Tree Physiology*,  
343 31(12), 1390–1400. <https://doi.org/10.1093/treephys/tpr118>

344 Esteban, R., Olano, J. M., Castresana, J., Fernández-Marín, B., Hernández, A., Becerril, J. M., & García-  
345 Plazaola, J. I. (2009). Distribution and evolutionary trends of photoprotective isoprenoids  
346 (xanthophylls and tocopherols) within the plant kingdom. *Physiologia Plantarum*, 135(4), 379–  
347 389. <https://doi.org/10.1111/j.1399-3054.2008.01196.x>

348 Faassen, E. J., Veraart, A. J., Van Nes, E. H., Dakos, V., Lürling, M., & Scheffer, M. (2015). Hysteresis  
349 in an experimental phytoplankton population. *Oikos*, 124(12), 1617–1623.  
350 <https://doi.org/10.1111/oik.02006>

351 Foyer, C. H. (2018). Reactive oxygen species, oxidative signaling and the regulation of photosynthesis.  
352 *Environmental and Experimental Botany*, 154, 134–142.  
353 <https://doi.org/10.1016/j.envexpbot.2018.05.003>

354 Fufezan, C., Rutherford, A. W., & Krieger-Liszka, A. (2002). Singlet oxygen production in herbicide-  
355 treated photosystem II. *FEBS Letters*, 532(3), 407–410. [https://doi.org/10.1016/S0014-  
356 5793\(02\)03724-9](https://doi.org/10.1016/S0014-5793(02)03724-9)

357 Gerla, D. J., Mooij, W. M., & Huisman, J. (2011). Photoinhibition and the assembly of light-limited  
358 phytoplankton communities. *Oikos*, 120(3), 359–368. [https://doi.org/10.1111/j.1600-  
359 0706.2010.18573.x](https://doi.org/10.1111/j.1600-0706.2010.18573.x)

360 Germino, M. J., & Smith, W. K. (1999). Sky exposure, crown architecture, and low-temperature  
361 photoinhibition in conifer seedlings at alpine treeline. *Plant, Cell & Environment*, 22(4), 407–  
362 415. <https://doi.org/10.1046/j.1365-3040.1999.00426.x>

363 Germino, M. J., Smith, W. K., & Resor, A. C. (2002). Conifer seedling distribution and survival in an  
364 alpine-treeline ecotone. *Plant Ecology*, *162*(2), 157–168.  
365 <https://doi.org/10.1023/A:1020385320738>

366 Goss, R., & Lepetit, B. (2015). Biodiversity of NPQ. *Journal of Plant Physiology*, *172*, 13–32.  
367 <https://doi.org/10.1016/j.jplph.2014.03.004>

368 Hakala, M., Tuominen, I., Keränen, M., Tyystjärvi, T., & Tyystjärvi, E. (2005). Evidence for the role of  
369 the oxygen-evolving manganese complex in photoinhibition of Photosystem II. *Biochimica et*  
370 *Biophysica Acta (BBA) - Bioenergetics*, *1706*(1), 68–80.  
371 <https://doi.org/10.1016/j.bbabi.2004.09.001>

372 Hoch, W. A., Singaas, E. L., & McCown, B. H. (2003). Resorption Protection. Anthocyanins Facilitate  
373 Nutrient Recovery in Autumn by Shielding Leaves from Potentially Damaging Light Levels.  
374 *Plant Physiology*, *133*(3), 1296–1305. <https://doi.org/10.1104/pp.103.027631>

375 Hoch, W. A., Zeldin, E. L., & McCown, B. H. (2001). Physiological significance of anthocyanins during  
376 autumnal leaf senescence. *Tree Physiology*, *21*(1), 1–8. <https://doi.org/10.1093/treephys/21.1.1>

377 Hodges, D. M., DeLong, J. M., Forney, C. F., & Prange, R. K. (1999). Improving the thiobarbituric acid-  
378 reactive-substances assay for estimating lipid peroxidation in plant tissues containing anthocyanin  
379 and other interfering compounds. *Planta*, *207*(4), 604–611.  
380 <https://doi.org/10.1007/s004250050524>

381 Howard, M. M., Bae, A., Pirani, Z., Van, N., & Königer, M. (2020). Impairment of chloroplast movement  
382 reduces growth and delays reproduction of *Arabidopsis thaliana* in natural and controlled  
383 conditions. *American Journal of Botany*, *107*(9), 1309–1318. <https://doi.org/10.1002/ajb2.1537>

384 Kale, R., Hebert, A. E., Frankel, L. K., Sallans, L., Bricker, T. M., & Pospíšil, P. (2017). Amino acid  
385 oxidation of the D1 and D2 proteins by oxygen radicals during photoinhibition of Photosystem II.  
386 *Proceedings of the National Academy of Sciences*, *114*(11), 2988–2993.  
387 <https://doi.org/10.1073/pnas.1618922114>

388 Kar, M., Streb, P., Hertwig, B., & Feierabend, J. (1993). Sensitivity to photodamage increases during  
389 senescence in excised leaves. *Journal of Plant Physiology*, *141*(5), 538–544.  
390 [https://doi.org/10.1016/S0176-1617\(11\)80453-0](https://doi.org/10.1016/S0176-1617(11)80453-0)

391 King, O. C., Smith, R. A., Warne, M. S. J., Merwe, J. P. van de, Connolly, R. M., & Brown, C. J. (2021).  
392 Combined impacts of photosystem II-inhibiting herbicides and light availability on seagrass and  
393 marine microalgae. *Marine Ecology Progress Series*, *668*, 215–230.  
394 <https://doi.org/10.3354/meps13717>

395 Kok, B. (1956). On the inhibition of photosynthesis by intense light. *Biochimica et Biophysica Acta*,  
396 *21*(2), 234–244. [https://doi.org/10.1016/0006-3002\(56\)90003-8](https://doi.org/10.1016/0006-3002(56)90003-8)

397 Körner, C. (2003). Carbon limitation in trees. *Journal of Ecology*, *91*(1), 4–17.  
398 <https://doi.org/10.1046/j.1365-2745.2003.00742.x>

399 Kothari, S., Montgomery, R. A., & Cavender-Bares, J. (2021). Physiological responses to light explain  
400 competition and facilitation in a tree diversity experiment. *Journal of Ecology*, *109*(5), 2000–  
401 2018. <https://doi.org/10.1111/1365-2745.13637>

402 Kramer, D. M., Johnson, G., Kiirats, O., & Edwards, G. E. (2004). New Fluorescence Parameters for the  
403 Determination of QA Redox State and Excitation Energy Fluxes. *Photosynthesis Research*, *79*(2),  
404 209–218. <https://doi.org/10.1023/B:PRES.0000015391.99477.0d>

405 Külheim, C., Ågren, J., & Jansson, S. (2002). Rapid Regulation of Light Harvesting and Plant Fitness in  
406 the Field. *Science*, *297*(5578), 91–93. <https://doi.org/10.1126/science.1072359>

407 Laughlin, D. C., & Messier, J. (2015). Fitness of multidimensional phenotypes in dynamic adaptive  
408 landscapes. *Trends in Ecology & Evolution*, *30*(8), 487–496.  
409 <https://doi.org/10.1016/j.tree.2015.06.003>

410 Lesser, M. (1996). Acclimation of phytoplankton to UV-B radiation: Oxidative stress and photoinhibition  
411 of photosynthesis are not prevented by UV-absorbing compounds in the dinoflagellate  
412 *Prorocentrum micans*. *Marine Ecology Progress Series*, *132*, 287–297.  
413 <https://doi.org/10.3354/meps132287>

414 Levin, G., Kulikovskiy, S., Liveanu, V., Eichenbaum, B., Meir, A., Isaacson, T., Tadmor, Y., Adir, N., &  
415 Schuster, G. (2021). The desert green algae *Chlorella ohadii* thrives at excessively high light  
416 intensities by exceptionally enhancing the mechanisms that protect photosynthesis from  
417 photoinhibition. *The Plant Journal*, 106(5), 1260–1277. <https://doi.org/10.1111/tpj.15232>

418 Long, S. P., Humphries, S., & Falkowski, P. G. (1994). Photoinhibition of Photosynthesis in Nature.  
419 *Annual Review of Plant Physiology and Plant Molecular Biology*, 45(1), 633–662.  
420 <https://doi.org/10.1146/annurev.pp.45.060194.003221>

421 Lu, X., Chen, M., Liu, Y., Miralles, D. G., & Wang, F. (2017). Enhanced water use efficiency in global  
422 terrestrial ecosystems under increasing aerosol loadings. *Agricultural and Forest Meteorology*,  
423 237–238, 39–49. <https://doi.org/10.1016/j.agrformet.2017.02.002>

424 Malnoë, A. (2018). Photoinhibition or photoprotection of photosynthesis? Update on the (newly termed)  
425 sustained quenching component qH. *Environmental and Experimental Botany*, 154, 123–133.  
426 <https://doi.org/10.1016/j.envexpbot.2018.05.005>

427 May, J. D., & Killingbeck, K. T. (1992). Effects of Preventing Nutrient Resorption on Plant Fitness and  
428 Foliar Nutrient Dynamics. *Ecology*, 73(5), 1868–1878. <https://doi.org/10.2307/1940038>

429 Merzlyak, M. N., & Hendry, G. a. F. (1994). Free radical metabolism, pigment degradation and lipid  
430 peroxidation in leaves during senescence. *Proceedings of the Royal Society of Edinburgh, Section*  
431 *B: Biological Sciences*, 102, 459–471. <https://doi.org/10.1017/S0269727000014482>

432 Mittler, R. (2006). Abiotic stress, the field environment and stress combination. *Trends in Plant Science*,  
433 11(1), 15–19. <https://doi.org/10.1016/j.tplants.2005.11.002>

434 Møller, I. M., Jensen, P. E., & Hansson, A. (2007). Oxidative Modifications to Cellular Components in  
435 Plants. *Annual Review of Plant Biology*, 58(1), 459–481.  
436 <https://doi.org/10.1146/annurev.arplant.58.032806.103946>

437 Montgomery, R. A., Goldstein, G., & Givnish, T. J. (2008). Photoprotection of PSII in Hawaiian  
438 lobeliads from diverse light environments. *Functional Plant Biology*, 35(7), 595–605.

439 Murchie, E. H., & Niyogi, K. K. (2011). Manipulation of Photoprotection to Improve Plant  
440 Photosynthesis. *Plant Physiology*, 155(1), 86–92. <https://doi.org/10.1104/pp.110.168831>

441 Niinemets, Ü., & Valladares, F. (2004). Photosynthetic Acclimation to Simultaneous and Interacting  
442 Environmental Stresses Along Natural Light Gradients: Optimality and Constraints. *Plant*  
443 *Biology*, 6(3), 254–268. <https://doi.org/10.1055/s-2004-817881>

444 Norris, J. R., Allen, R. J., Evan, A. T., Zelinka, M. D., O'Dell, C. W., & Klein, S. A. (2016). Evidence for  
445 climate change in the satellite cloud record. *Nature*, 536(7614), 72–75.  
446 <https://doi.org/10.1038/nature18273>

447 Pospíšil, P. (2016). Production of Reactive Oxygen Species by Photosystem II as a Response to Light and  
448 Temperature Stress. *Frontiers in Plant Science*, 7.  
449 <https://www.frontiersin.org/articles/10.3389/fpls.2016.01950>

450 Powles, S. B. (1984). Photoinhibition of Photosynthesis Induced by Visible Light. *Annual Review of*  
451 *Plant Physiology*, 35(1), 15–44. <https://doi.org/10.1146/annurev.pp.35.060184.000311>

452 Prescott, C. E., Grayston, S. J., Helmisaari, H.-S., Kaštovská, E., Körner, C., Lambers, H., Meier, I. C.,  
453 Millard, P., & Ostonen, I. (2020). Surplus Carbon Drives Allocation and Plant–Soil Interactions.  
454 *Trends in Ecology & Evolution*, 35(12), 1110–1118. <https://doi.org/10.1016/j.tree.2020.08.007>

455 Raven, J. A. (2011). The cost of photoinhibition. *Physiologia Plantarum*, 142(1), 87–104.  
456 <https://doi.org/10.1111/j.1399-3054.2011.01465.x>

457 Renner, S. S., & Zohner, C. M. (n.d.). The occurrence of red and yellow autumn leaves explained by  
458 regional differences in insolation and temperature. *New Phytologist*, 0(0).  
459 <https://doi.org/10.1111/nph.15900>

460 Sala, A., Woodruff, D. R., & Meinzer, F. C. (2012). Carbon dynamics in trees: Feast or famine? *Tree*  
461 *Physiology*, 32(6), 764–775. <https://doi.org/10.1093/treephys/tpr143>

462 Schoen, D. J., & Schultz, S. T. (2019). Somatic Mutation and Evolution in Plants. *Annual Review of*  
463 *Ecology, Evolution, and Systematics*, 50(1), 49–73. <https://doi.org/10.1146/annurev-ecolsys-110218-024955>

464

465 Slot, M., Wirth, C., Schumacher, J., Mohren, G. M. J., Shibistova, O., Lloyd, J., & Ensminger, I. (2005).  
466 Regeneration patterns in boreal Scots pine glades linked to cold-induced photoinhibition. *Tree*  
467 *Physiology*, 25(9), 1139–1150. <https://doi.org/10.1093/treephys/25.9.1139>

468 Takahashi, S., Milward, S. E., Yamori, W., Evans, J. R., Hillier, W., & Badger, M. R. (2010). The Solar  
469 Action Spectrum of Photosystem II Damage. *Plant Physiology*, 153(3), 988–993.  
470 <https://doi.org/10.1104/pp.110.155747>

471 Takahashi, S., & Murata, N. (2008). How do environmental stresses accelerate photoinhibition? *Trends in*  
472 *Plant Science*, 13(4), 178–182. <https://doi.org/10.1016/j.tplants.2008.01.005>

473 Tikkanen, M., Mekala, N. R., & Aro, E.-M. (2014). Photosystem II photoinhibition-repair cycle protects  
474 Photosystem I from irreversible damage. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*,  
475 1837(1), 210–215. <https://doi.org/10.1016/j.bbabi.2013.10.001>

476 Tyystjärvi, E., & Aro, E. M. (1996). The rate constant of photoinhibition, measured in lincomycin-treated  
477 leaves, is directly proportional to light intensity. *Proceedings of the National Academy of*  
478 *Sciences*, 93(5), 2213–2218. <https://doi.org/10.1073/pnas.93.5.2213>

479 Van De Poll, W. H., Eggert, A., Buma, A. G. J., & Breeman, A. M. (2001). Effects of UV-B-Induced  
480 DNA Damage and Photoinhibition on Growth of Temperate Marine Red Macrophytes: Habitat-  
481 Related Differences in UV-B Tolerance. *Journal of Phycology*, 37(1), 30–38.  
482 <https://doi.org/10.1046/j.1529-8817.2001.037001030.x>

483 Vass, I. (2011). Role of charge recombination processes in photodamage and photoprotection of the  
484 photosystem II complex. *Physiologia Plantarum*, 142(1), 6–16. <https://doi.org/10.1111/j.1399-3054.2011.01454.x>

485

486 Velasco, N., & Becerra, P. I. (2020). Species-specific effects of the herbaceous layer on recruitment of  
487 woody species under different shading and precipitation conditions. *Forest Ecology and*  
488 *Management*, 460, 117864. <https://doi.org/10.1016/j.foreco.2020.117864>

489 Vellend, M. (2016). *The Theory of Ecological Communities*. Princeton University Press, Princeton, NJ,  
490 United States.

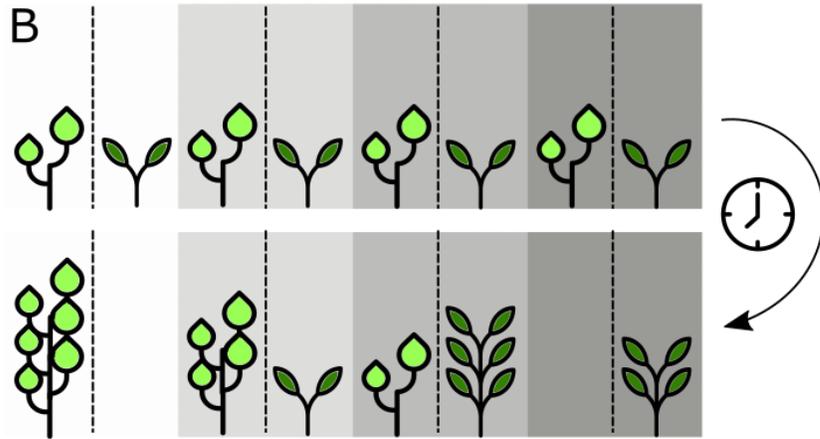
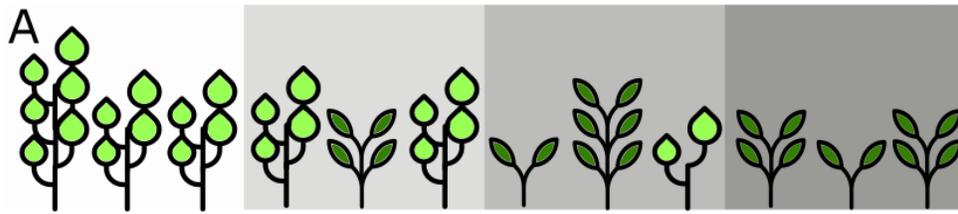
- 491 Veraart, A. J., Faassen, E. J., Dakos, V., van Nes, E. H., Lürling, M., & Scheffer, M. (2012). Recovery  
492 rates reflect distance to a tipping point in a living system. *Nature*, *481*(7381), 357–359.  
493 <https://doi.org/10.1038/nature10723>
- 494 Weber, R., Gessler, A., & Hoch, G. (2019). High carbon storage in carbon-limited trees. *New Phytologist*,  
495 *222*(1), 171–182. <https://doi.org/10.1111/nph.15599>
- 496 White, A. C., Rogers, A., Rees, M., & Osborne, C. P. (2016). How can we make plants grow faster? A  
497 source–sink perspective on growth rate. *Journal of Experimental Botany*, *67*(1), 31–45.  
498 <https://doi.org/10.1093/jxb/erv447>
- 499 Zandalinas, S. I., Sengupta, S., Fritschi, F. B., Azad, R. K., Nechushtai, R., & Mittler, R. (2021). The  
500 impact of multifactorial stress combination on plant growth and survival. *New Phytologist*,  
501 *230*(3), 1034–1048. <https://doi.org/10.1111/nph.17232>

502 **Figures**

503 Figure 1: Two potential approaches to test the importance of photoinhibition and photoprotective  
504 adaptations for fitness. These approaches are respectively analogous to the ‘likelihood’ and ‘vital rates’  
505 approaches described in a much more general context in Laughlin & Messier (2015). Here, we consider  
506 two species (i and ii). I depict species (i) with more yellowish leaves and steeper leaf angles to imply  
507 stronger photoprotective adaptations—high NPQ by carotenoids and light avoidance at midday,  
508 respectively. For simplicity, I assume these traits are not plastic. In reality, photoprotection can include an  
509 immense variety of mechanisms with no obvious common currency to summarize a plant’s total  
510 photoprotective capacity.

511 (A) One way to test the importance of photoinhibition is to observe whether photoprotective traits are  
512 correlated consistently with environmental gradients in the risk of photoprotection. Here, as light  
513 increases, species (i) comprises a greater fraction of community composition. As with all observational  
514 data, causal inference may be hard and interpretation requires special care to rule out competing  
515 explanations. For example, species (i) may be absent under high light due to environmental filtering  
516 caused by photoinhibition or other abiotic stresses, because it is outcompeted there by species (ii), or a  
517 combination. Nevertheless, if sites with higher light consistently have species with greater constitutive  
518 expression of photoprotective adaptations, it would imply that there is ecological selection for those  
519 adaptations. A more rigorous test might involve determining whether including photoprotective traits  
520 improves predictive models of community assembly (Laughlin & Messier 2015).

521 (B) Another way to test the importance of photoinhibition is to monitor vital rates across natural or  
522 (ideally) experimental gradients of light. Here, we see that species (i) has its greatest growth rates at high  
523 light, while species (ii)’s growth is reduced at high light levels. Dashed lines separate the individuals in  
524 each treatment to clarify that they are not directly interacting. This kind of experiment can be done with a  
525 single species/genotype or with multiple; in the latter case, researchers can test whether plants that are  
526 more shade-tolerant (and high light-intolerant) have fitness optima at lower light levels.

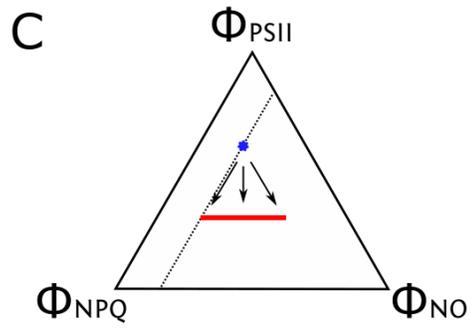
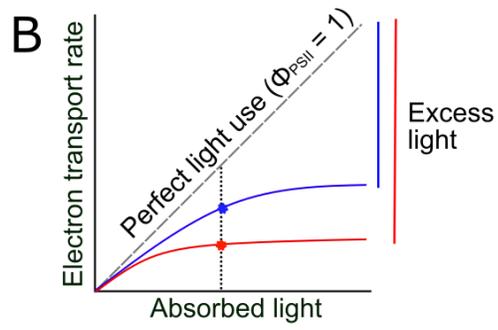
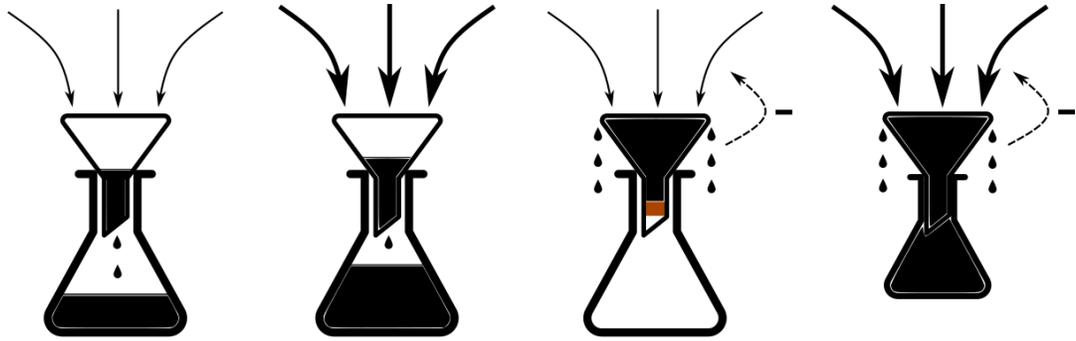


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529 Figure 2: (A) As a rough analogy, the balance between carbon sources and sinks may be described as  
530 liquid being poured (photosynthesis) into a funnel (phloem transport) draining into a flask (carbon sinks).  
531 Here, the carbon that flows into the flask is available for metabolism and growth at the meristems, and in  
532 reality would be steadily consumed. When the funnel's spout is stopped (as when phloem transport is  
533 blocked) or when the flask is small (limited sink capacity), carbon can build up in the source tissues.  
534 Often, this buildup causes feedback inhibition of photosynthesis (dashed arrows), reducing the rate of  
535 inflow. This representation is inspired by Adams et al. (2013). (B) Feedback inhibition or more direct  
536 constraints on light use (e.g. low temperature) can reduce the electron transport rate at a given light level  
537 (red) compared to a healthy plant under conditions conducive to growth (blue). As a result, the fraction of  
538 absorbed light that is used to power electron transport ( $\Phi_{\text{PSII}}$ ) declines and a greater fraction of light is in  
539 excess. (C) Absorbed light energy can be partitioned into three quantum yields— $\Phi_{\text{PSII}}$ ,  $\Phi_{\text{NPQ}}$ , and  $\Phi_{\text{NO}}$ .  
540  $\Phi_{\text{NPQ}}$  is the fraction of light dissipated through NPQ, while  $\Phi_{\text{NO}}$  is the fraction that undergoes non-  
541 regulated dissipation—including through pathways that produce ROS. Under internal or external  
542 constraints to electron transport,  $\Phi_{\text{PSII}}$  declines, but the share of consequent increase in  $\Phi_{\text{NPQ}}$  vs.  $\Phi_{\text{NO}}$   
543 depends on how much the plant can induce NPQ, allowing a range of scenarios (red line). Plants with  
544 limited capacity for NPQ may have elevated ROS production.

**A**  
 carbon  
 assimilation  
  
 phloem  
 transport  
  
 growth and  
 metabolism



545

546 **Appendix S1**

547 **Table S1:** A brief summary of studies that might be taken as evidence that photoinhibition due to excess light can have negative effects on plant fitness. These  
548 studies all report declines in some fitness-related variable (survival, growth, or reproductive output) in conditions with high light. I include some studies that report  
549 only distributions of existing individuals if there is a strong case that absences are due to mortality rather than other factors like dispersal limitation. In general, I  
550 omit studies that only compare fitness in across coarse natural gradients (e.g. understory vs. gaps) because such gradients often include changes in multiple  
551 environmental variables and cannot be used to isolate the importance of photoinhibition. However, I include a few examples where such contrasts are accompanied  
552 by physiological and/or microclimatic measurements that allow a more convincing attribution of fitness gradients to the effects of light. This table should not be  
553 mistaken for a rigorous, formal evidence synthesis, since I did not use a well-defined search procedure and I only summarize studies that appear to show some  
554 positive evidence that photoinhibition can affect fitness. Nevertheless, I hope that they convince the reader of the possibility that excess light can damage fitness.

Citation	Species	Ecosystem or setting	Treatment	Physiological response	Fitness response	Notes
Agyeman et al. 2003	Seedlings of 16 west African tree species	Shadehouses in Ghana	Trees were grown in one of six shadehouses with varying degrees of shade.	None reported	In many (but not all) species, whole-plant relative growth rate was highest at irradiances below the maximum growth irradiance. Shade-tolerant species showed the greatest inhibition of growth at high irradiance.	More severe shading treatments tended to reduce maximum air temperature and slightly increased relative humidity.
Akhalkatsi et al. 2006	<i>Betula litwinowii</i> seedlings	Alpine treelines in the country of Georgia	The number of seedlings was surveyed across a number of plots within sites that varied in slope, aspect, elevation, and overstory cover of <i>B. litwinowii</i> or the shrub <i>Rhododendron caucasicum</i>	Seedlings in sun-exposed microsites had more red anthocyanic leaves.	More sheltered microsites beneath shrub or tree cover, or on the north-facing walls of soil depressions, had greater <i>B. litwinowii</i> seedling abundance.	The sheltered sites with greater seedling abundance had lower air and soil temperatures than more exposed sites.
Bader et al. 2006	Seedlings of six (in the observational component)	Alpine treelines in northern Ecuador	The number of sprouts was surveyed along transects that crossed the treeline from the	None reported	Out of the six species in the observational survey, four were almost completely absent beyond the treeline but the	Microclimatic monitoring revealed that the forest buffered temperature extremes, but the four páramo treatments were

	or eight (in the experimental component) tree species		cloud forest into the páramo. In addition, some seedlings were experimentally transplanted into either the forest or the páramo, and the latter were subjected to shading by shade cloth, clearing of neighbors, or both treatments		other two showed dramatic increases in abundance. In the experimental component, survival was greatest in the forest, then in the páramo under shade cloth, then in páramo controls. Clearing of neighbors also had a negative effect on survival.	similar. However, cleared and unshaded plots did have higher soil temperature and apparently lower soil moisture. The authors note several additional observations that suggest a role for photoinhibition. For example, the species that survive best in the páramo appear to have photoprotective adaptations like reflective leaves or specialized pigments. Also, in many species the light-exposed parts of the leaves were the only ones to show apparent damage.
Ball et al. 1991	<i>Eucalyptus pauciflora</i> seedlings	Alpine treelines in southeastern Australia	Plants were naturally growing on slopes in aspects either more protected or more exposed to sunlight during the winter; for physiological measurements, some trees were also shaded artificially	Trees on exposed sites had lower rates of photosynthesis under light-limited conditions, but not under high light.	Juvenile trees tended to be distributed close to the canopy margin and along an arc to the better-protected south of the canopy. Juvenile trees in exposed areas were often stunted.	
Ballestreri et al. 2021	Four subdominant or understory tree species	Restoration experiment in the Atlantic Forest of southern Brazil	Trees were grown in the varying degrees of shade provided by one of four overstory tree species	Deeper shading caused increases in chlorophyll content and dark-acclimated $F_v/F_m$ , and decreases in stomatal density. All species appeared to show greater photosynthetic capacity under deeper shade.	One species ( <i>Eugenia involucrata</i> ) had a major increase in mortality under the greatest light level. Many species showed greatest growth in height or stem diameter under partial shade.	Because the treatment here involves interactions with neighbors, the outcome may result not just from shading but also from other sorts of mechanisms of interactions among neighboring plants.
Close et al. 2002	<i>Eucalyptus nitens</i> and <i>Eucalyptus globulus</i> seedlings	Alpine treelines in Tasmania, Australia	Plants were either left exposed or surrounded with open-top shade cloth shelters for the first fourteen weeks after planting	Unshaded <i>E. globulus</i> developed high leaf anthocyanin content, as well as severe leaf abscission and severe but transient declines in dark-acclimated $F_v/F_m$ . Unshaded <i>E. nitens</i> had the same responses, but much more moderate. Shaded trees showed slight increases in anthocyanins and declines in dark-acclimated	Unshaded <i>E. globulus</i> suffered severe leaf abscission and had high mortality and reduced height growth compared to shaded <i>E. globulus</i> . However, unshaded <i>E. nitens</i> had much more modest levels of abscission and greater height growth than shaded <i>E. nitens</i> .	The researchers measured air temperature and light quality and found that the shelters had minimal effects on both.

				Fv/Fm after the shadecloth was removed.		
del Valle et al. 2020	<i>Silene littorea</i>	Greenhouse and experimental garden in Mediterranean southern Spain	Plants were grown either under a methacrylate filter that transmitted most UV or a polycarbonate filter that excluded most UV.	Plants under UV exclusion produced less anthocyanins and phenolics. They also had greater midday F <sub>v</sub> /F <sub>m</sub> , but the similar pre-dawn F <sub>v</sub> /F <sub>m</sub> .	Plants under UV exclusion had the same ovule and seed production per flower, but considerably lower pollen production and seed set.	
Egerton et al. 2000	<i>Eucalyptus pauciflora</i> seedlings	Treeless pasture in southeastern Australia	Plants were grown on either the exposed or sheltered sides of vertical screens that intercepted half of incoming light	Trees on the exposed side of the screens had lower dark-acclimated and midday F <sub>v</sub> /F <sub>m</sub> and lowered photosynthetic light-response curves. These trees also lost much of their leaf area to abscission during the winter.	Exposed trees had lower shoot and total biomass after the winter.	The researchers measured air temperature and soil moisture. They found that the shelters had no significant influence on air temperature, but caused a small increase of soil moisture on the exposed side.
Gatti et al. 2014	Saplings of the palm <i>Euterpe edulis</i>	Shadehouse and various sites in subtropical northern Argentina	Plants were grown in either high or low light in shadehouses or in natural canopy understories/gaps	Seedlings grown under higher irradiance had a smaller relative decline in F <sub>v</sub> /F <sub>m</sub> and faster recovery. High growth irradiance did not affect stem water potential but did cause substantial decreases in leaf water potential and increases in plant Huber value.	In the shadehouse, relatively high irradiance treatments are associated with somewhat lower growth and irradiance. In natural gaps, survival was much lower but growth was much higher than in understories.	In this study, the subtle but perhaps important influence of the treatments on water relations makes it hard to uniquely pinpoint an effect of photoinhibition, since high mortality at high irradiance could have been exacerbated by water deficits.
Germino & Smith 1999	<i>Picea engelmannii</i> and <i>Abies lasiocarpa</i> seedlings	Alpine treelines in Wyoming, United States	The position of plants was surveyed along four distinct transects. For physiological measurements, some potted plants were also experimentally subjected to nighttime warming (transparent plastic sheets), daytime	Potted <i>A. lasiocarpa</i> seedlings subjected to daytime shading had higher saturated photosynthetic rates than unshaded seedlings, both comparing shading to control and shading + warming to just warming. There was no major effect in <i>P. engelmannii</i> . Among natural seedlings, those	Both species were almost entirely found in the one transect with high tree cover. <i>A. lasiocarpa</i> in particular was largely missing from highly open microsites.	The authors measured light spectral quality and found that it was unaffected by the treatments.

			shading, or both	in exposed environments had more inclined and clustered needles.		
Germino et al. 2002	<i>Picea engelmannii</i> and <i>Abies lasiocarpa</i> seedlings	Alpine treelines in Wyoming, United States	Naturally occurring seedlings were surveyed repeatedly for survival as a function of factors that influence their exposure to stress. For some naturally occurring seedlings, neighboring grasses were removed. Some seedlings were also artificially sown among several sites with varying exposure to high light and other potential stress factors.	None reported	<i>P. engelmannii</i> seedlings (but not saplings or emergents) had greater survival close to the cover of the tree island. <i>P. engelmannii</i> emergents had greater mortality on more exposed south and east-facing slopes. In general, survival was greater when there were features upwind or overhead, or when trees were surrounded by moss or overtopped by grass.	Removal of neighbors alleviated water and low temperature stress but still increased mortality, which the authors attribute to photoinhibition due to increased light. Seedlings with removed neighbors also experienced greater wind speeds.
Gómez-Aparicio et al. 2006	Two evergreen and two deciduous late-successional tree species	Montane botanical garden in Mediterranean southeastern Spain	Plants were growing in pots either under full sunlight or under mild or severe shading treatments imposed by shade cloth.	Under shade, all species increased chlorophyll content and decreased carotenoid content. Shade also caused increases in $F_v/F_m$ throughout the day, including before dawn. The two deciduous species had lowered photosynthetic light-response curves under high light, while the two evergreen species showed the opposite pattern. The authors report many other changes in plant allocation and physiology that are less relevant here.	One deciduous species ( <i>Acer opalus</i> ) experienced a very high rate of mortality at full sunlight, which was reduced by two-thirds under even a moderate shading treatment. No other species experienced negative survival or growth (total dry mass) consequences from high light.	Soil moisture was held constant across treatments. The moderate shade treatment had no major effect on air temperature, but the more intense shade treatment tended to reduce temperature substantially.

Holly et al. 1994	<i>Eucalyptus polyanthemos</i> seedlings	Treeless pasture in southeastern Australia	Plants were either grown in the open or in a series of shelters that excluded various fractions of incident light	Trees had the highest pre-dawn $F_v/F_m$ under 50% or 30% of natural light, both in midwinter and in spring.	Trees showed the greatest stem elongation under 50% or 30% of natural light.	The researchers measured humidity, air temperature, and wind speed. While they found that while most shelter designs had minimal effects on humidity and temperature, shelters reduced wind speed in proportion to their shading intensity.
Howard et al. 2020	<i>Arabidopsis thaliana</i> —an NPQ-deficient mutant, a chloroplast movement-impaired mutant, three mutants deficient in chloroplast movement as well as other aspects of physiological function, and a wild-type	Growth chambers and an experimental garden in Massachusetts, United States	Plants were grown either outdoors or in a growth chamber with light fluctuating on an hour-long cycle	Under natural conditions, the chloroplast movement-impaired mutant had higher dark-acclimated $F_v/F_m$ than the wild-type.	Under natural conditions, the chloroplast movement-impaired movement had smaller rosettes, smaller and fewer leaves, fewer siliques, fewer secondary inflorescences, and earlier bolting than the wild-type. The NPQ-deficient mutant was largely unaffected.	
Kothari et al. 2021	Eight temperate, winter-deciduous broadleaf tree species in the first eight years of growth	Densely planted tree diversity experiment in Minnesota, United States	Plants were grown either in monocultures, where they were generally unshaded, or in bicultures with faster-growing conifers that provided substantial shade	Plants grown in bicultures had substantially higher pre-dawn $F_v/F_m$ and lower production of photoprotective pigments. The most shade-tolerant species had the greatest increase in pre-dawn $F_v/F_m$ in biculture. At least in one species ( <i>Tilia americana</i> ), they also showed high photosynthetic rates under high light levels and delayed leaf senescence.	Two species ( <i>T. americana</i> and <i>Acer negundo</i> ) grew faster with larger neighbors; most of the others grew slower.	Because the treatment here involves interactions with neighbors, the outcome may result not just from shading but also from other sorts of mechanisms of interactions among neighboring plants. Trees under the shade of larger neighbors generally had more negative pre-dawn leaf water potential, but less negative midday water potential.

Külheim et al. 2002	<i>Arabidopsis thaliana</i> —two NPQ-deficient mutants and a wild-type	Growth chambers and an experimental garden in northern Sweden	Plants were grown in growth chambers with constant and fluctuating light conditions, as well as in the field.	The two mutants had lower midday $F_v/F_m$ under field conditions, especially under high light.	In the field and under fluctuating light, the two mutants had fewer seeds and fruits per plant. Under constant light, there was no effect.	
Murray et al. 1993	Multiple species of <i>Sphagnum</i> mosses	Moist mountain slopes in northern Alaska	At one site, all shade-producing vascular plants were removed in several plots, which were compared to control plots. At another site without high vascular plant cover, shadecloth shelters were set up and compared to control plots. In addition, physiological measurements were taken in the laboratory from various Arctic and temperate mosses.	Mosses under high light had lower dark-acclimated $F_v/F_m$ and lower photosynthetic capacity than those under lower light.	At both sites with shade experiments, mosses under shade (from shadecloth or vascular plants) had much greater growth in length.	Shaded treatments tended to have somewhat lower moss surface temperature.
Ronco 1970	<i>Picea engelmannii</i> seedlings	Nursery in the Central Rocky Mountains	Trees were grown in one of three shade treatments	Unshaded seedlings had much lower photosynthetic light-response curves than fully shaded seedlings. Unshaded and partially shaded seedlings had no major differences in their water deficit.	The paper relies on a 1961 research note by the same author, which is not widely available, to support the claim that light influences seedling survival. It reports that trees growing under full sun suffered from chlorosis and had greater mortality than those under partial shade. It also reports that drought, frost heaving, and herbivory are not major causes of mortality.	
Semchenko et al. 2012	46 herbaceous species from temperate grasslands	Common garden in Estonia	Plants were grown either in full sunlight or in three shading treatments of varying intensity	None reported	Moderate shade increased total plant biomass relative to full sunlight	The shading treatments moderately buffered air temperature and increased relative humidity, particularly the more intense ones. The authors attribute the positive effect of shade on growth to dynamic regulation of growth

						rather than amelioration of abiotic stress. However, it is unclear how this is possible, considering that plants cannot build tissue with material they have not acquired.
Slot et al. 2005	<i>Pinus sylvestris</i> juveniles	Glades with low tree density within forests in western Siberia	Plants were naturally growing on slopes in aspects either more protected or more exposed to sunlight during the winter. Some seedlings were also either left exposed or artificially shaded following removal of adult trees.	In the smaller size class across the growing season, more naturally exposed trees had lower pre-dawn and midday $F_v/F_m$ and lower photosynthetic rates at equivalent light availability. These exposed small trees had greater non-photochemical quenching, xanthophyll pigment to chlorophyll ratios, and xanthophyll de-epoxidation state. Similar results held when comparing artificially exposed and shaded trees. The researchers found that more exposed and photoinhibited trees were not depleted in NSCs.	Seedlings recruited mainly within the crown projection area of adult trees, usually on the protected northern side.	Air temperature differed very little between the northern (protected) and southern (exposed) sides of adult trees, especially in autumn and winter. Southern trees had slightly less negative $\delta^{13}C$ , which led the researchers to suggest tentatively that they may be more prone to desiccation. However, soil moisture content did not differ between northern and southern aspects during the summer.
van de Staaij et al. 1997	<i>Silene vulgaris</i> —a highland population adapted to higher UV-B, and a lowland population adapted to lower UV-B	Greenhouse in the Netherlands	Plants were grown in a greenhouse with either no additional UV-B, a low dose, or a high dose	None reported	The lowland population showed a large reduction in seed-producing flowers and seeds per plant under high UV, but the highland population showed an increase in seeds per plant under high UV. In each population, seed mass and germination were unaffected.	
Velasco & Becerra 2020	Seedlings of nine woody species	Pots in an open grassland in central Chile	Trees were grown together with one of eight different herb species (or control), under either shade cloth or control, and in either a wetter or drier simulated precipitation regime	None reported	The shade treatment had an overwhelmingly large positive impact on the recruitment (survival) of tree seedlings. Water and herb cover had much smaller effects.	

555 **Literature Cited**

- 556  
557 Agyeman, V. K., Swaine, M. D., & Thompson, J. (1999). Responses of tropical forest tree seedlings to  
558 irradiance and the derivation of a light response index. *Journal of Ecology*, 87(5), 815–827.  
559 <https://doi.org/10.1046/j.1365-2745.1999.00400.x>  
560  
561 Akhalkatsi, M., Abdaladze, O., Nakhutsrishvili, G., & Smith, W. K. (2006). Facilitation of Seedling  
562 Microsites by *Rhododendron Caucasicum* Extends the *Betula Litwinowii* Alpine Treeline,  
563 Caucasus Mountains, Republic of Georgia. *Arctic, Antarctic, and Alpine Research*, 38(4), 481–  
564 488. [https://doi.org/10.1657/1523-0430\(2006\)38\[481:FOSMBR\]2.0.CO;2](https://doi.org/10.1657/1523-0430(2006)38[481:FOSMBR]2.0.CO;2)  
565  
566 Bader, M. Y., van Geloof, I., & Rietkerk, M. (2007). High solar radiation hinders tree regeneration above  
567 the alpine treeline in northern Ecuador. *Plant Ecology*, 191(1), 33–45.  
568 <https://doi.org/10.1007/s11258-006-9212-6>  
569  
570 Ball, M. C., Hodges, V. S., & Laughlin, G. P. (1991). Cold-Induced Photoinhibition Limits Regeneration  
571 of Snow Gum at Tree-Line. *Functional Ecology*, 5(5), 663–668. <https://doi.org/10.2307/2389486>  
572  
573 Ballestreri, A. A., Araujo, M. M., Aimi, S. C., Nascimento, N. F. do, Berghetti, Á. L. P., Gasparin, E.,  
574 Tabaldi, L. A., & Zavistanovicz, T. C. (2021). Morphophysiological responses of forest tree  
575 species conducted under different levels of shading in the enrichment of degraded ecosystem.  
576 *Forest Ecology and Management*, 488, 119032. <https://doi.org/10.1016/j.foreco.2021.119032>  
577  
578 Close, D. C., Beadle, C. L., Holz, G. K., & Brown, P. H. (2002). Effect of shade cloth tree shelters on  
579 cold-induced photoinhibition, foliar anthocyanin and growth of *Eucalyptus globulus* and *E. nitens*  
580 seedlings during establishment. *Australian Journal of Botany*, 50(1), 15–20.  
581 <https://doi.org/10.1071/bt01038>

582

583 Egerton, J. J. G., Banks, J. C. G., Gibson, A., Cunningham, R. B., & Ball, M. C. (2000). Facilitation of  
584 Seedling Establishment: Reduction in Irradiance Enhances Winter Growth of Eucalyptus  
585 Pauciflora. *Ecology*, 81(5), 1437–1449. [https://doi.org/10.1890/0012-](https://doi.org/10.1890/0012-9658(2000)081[1437:FOSERI]2.0.CO;2)  
586 [9658\(2000\)081\[1437:FOSERI\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2000)081[1437:FOSERI]2.0.CO;2)

587

588 Gatti, M. G., Campanello, P. I., Villagra, M., Montti, L., & Goldstein, G. (2014). Hydraulic architecture  
589 and photoinhibition influence spatial distribution of the arborescent palm *Euterpe edulis* in  
590 subtropical forests. *Tree Physiology*, 34(6), 630–639. <https://doi.org/10.1093/treephys/tpu039>

591

592 Germino, M. J., & Smith, W. K. (1999). Sky exposure, crown architecture, and low-temperature  
593 photoinhibition in conifer seedlings at alpine treeline. *Plant, Cell & Environment*, 22(4), 407–  
594 415. <https://doi.org/10.1046/j.1365-3040.1999.00426.x>

595

596 Germino, M. J., Smith, W. K., & Resor, A. C. (2002). Conifer seedling distribution and survival in an  
597 alpine-treeline ecotone. *Plant Ecology*, 162(2), 157–168.  
598 <https://doi.org/10.1023/A:1020385320738>

599

600 Gómez-Aparicio, L., Valladares, F., & Zamora, R. (2006). Differential light responses of Mediterranean  
601 tree saplings: Linking ecophysiology with regeneration niche in four co-occurring species. *Tree*  
602 *Physiology*, 26(7), 947–958. <https://doi.org/10.1093/treephys/26.7.947>

603

604 Holly, C., Laughlin, G. P., & Ball, M. C. (1994). Cold-Induced Photoinhibition and Design of Shelters for  
605 Establishment of Eucalypts in Pasture. *Australian Journal of Botany*, 42(2), 139–147.  
606 <https://doi.org/10.1071/bt9940139>

607

608 Howard, M. M., Bae, A., Pirani, Z., Van, N., & Königer, M. (2020). Impairment of chloroplast movement  
609 reduces growth and delays reproduction of *Arabidopsis thaliana* in natural and controlled  
610 conditions. *American Journal of Botany*, *107*(9), 1309–1318. <https://doi.org/10.1002/ajb2.1537>  
611

612 Kothari, S., Montgomery, R. A., & Cavender-Bares, J. (2021). Physiological responses to light explain  
613 competition and facilitation in a tree diversity experiment. *Journal of Ecology*, *109*(5), 2000–  
614 2018. <https://doi.org/10.1111/1365-2745.13637>  
615

616 Külheim, C., Ågren, J., & Jansson, S. (2002). Rapid Regulation of Light Harvesting and Plant Fitness in  
617 the Field. *Science*, *297*(5578), 91–93. <https://doi.org/10.1126/science.1072359>  
618

619 Murray, K. J., Tenhunen, J. D., & Nowak, R. S. (1993). Photoinhibition as a control on photosynthesis  
620 and production of Sphagnum mosses. *Oecologia*, *96*(2), 200–207.  
621 <https://doi.org/10.1007/BF00317733>  
622

623 Ronco, F. (1970). Influence of High Light Intensity on Survival of Planted Engelmann Spruce. *Forest*  
624 *Science*, *16*(3), 331–339. <https://doi.org/10.1093/forestscience/16.3.331>  
625

626 Semchenko, M., Lepik, M., Götzenberger, L., & Zobel, K. (2012). Positive effect of shade on plant  
627 growth: Amelioration of stress or active regulation of growth rate? *Journal of Ecology*, *100*(2),  
628 459–466. <https://doi.org/10.1111/j.1365-2745.2011.01936.x>  
629

630 Slot, M., Wirth, C., Schumacher, J., Mohren, G. M. J., Shibistova, O., Lloyd, J., & Ensminger, I. (2005).  
631 Regeneration patterns in boreal Scots pine glades linked to cold-induced photoinhibition. *Tree*  
632 *Physiology*, *25*(9), 1139–1150. <https://doi.org/10.1093/treephys/25.9.1139>  
633

- 634 Valle, J. C. D., Buide, M. L., Whittall, J. B., Valladares, F., & Narbona, E. (2020). UV radiation increases  
635 phenolic compound protection but decreases reproduction in *Silene littorea*. *PLOS ONE*, *15*(6),  
636 e0231611. <https://doi.org/10.1371/journal.pone.0231611>  
637
- 638 van de Staaij, J. W. M., Bolink, E., Rozema, J., & Ernst, W. H. O. (1997). The impact of elevated UV-B  
639 (280–320 nm) radiation levels on the reproduction biology of a highland and a lowland  
640 population of *Silene vulgaris*. *Plant Ecology*, *128*(1), 173–179.  
641 <https://doi.org/10.1023/A:1009710907336>  
642
- 643 Velasco, N., & Becerra, P. I. (2020). Species-specific effects of the herbaceous layer on recruitment of  
644 woody species under different shading and precipitation conditions. *Forest Ecology and*  
645 *Management*, *460*, 117864. <https://doi.org/10.1016/j.foreco.2020.117864>