1 When and how does photoinhibition matter for plant fitness?

2 Shan Kothari 3 Institut de recherche en biologie végétale, Université de Montréal, Montréal, Québec, Canada, H1X 2B1 Université du Québec à Montréal, Montréal, Québec, Canada, H2X 3Y7 4 5 6 Email: shan.kothari@umontreal.ca 7 8 ____; revision accepted ___ Manuscript received 9 Running head: Photoinhibition and plant fitness 10 11 Abstract 12 The many biophysical factors that shape how plant species sort across environmental gradients may 13 include photoinhibition, which I define as oxidative damage that photosynthetic organisms risk incurring 14 when they absorb excess light energy they cannot safely dissipate. Photoinhibition is seldom explicitly discussed as a potential driver of plant fitness and distributions; here, I aim to show that it can be one. I 15 16 draw on studies showing that natural gradients or experimental manipulations that increase the risk of 17 photoinhibition thereby decrease plant fitness, or favor species with stronger photoprotective adaptations. 18 One implication is that alleviation of photoinhibition may be a common mechanism of facilitation. A 19 large share of this research is set in stressful environments like alpine treelines or drylands, most likely 20 because photoinhibition is most detrimental in the presence of other interacting stress factors. 21 Nevertheless, knowing the specific role of photoinhibition may have practical value—as, for example, in 22 understanding the uses of shading treatments in ecological restoration. 23 24 Much research assumes that photoinhibition diminishes fitness by reducing photosynthesis and causing 25 carbon depletion. Nevertheless, a tension exists in ecological literature on photoinhibition: many kinds of 26 stressful conditions that threaten oxidative damage to photosynthesis also directly constrain the use of 27 carbon for tissue expansion. As a result, conditions that cause photoinhibition may often cause carbon reserves to remain steady or even increase. This fact might be reconciled with photoinhibition's 28 29 demonstrated influence on plant fitness by considering its other consequences. Oxidative damage can

- have consequences far beyond photosynthesis—altering proteins and membranes, blocking phloem
 loading, and triggering cell death. While such effects are harder to measure in the field than
 photosynthesis, doing so may enrich our understanding of photoinhibition's ecological consequences.

 Keywords: fitness; photoinhibition; photoprotection; photosynthesis; reactive oxygen species (ROS); sink
- 35 limitation

Introduction

One goal of plant ecophysiology is to explain how patterns of variation in fitness across environmental gradients emerge from the vast diversity of plant function at the level of tissues and cells. This knowledge is essential for predicting how plant communities will be reshaped by global change factors. Such factors include the quantity and quality of light, which is changing due to decadal trends in cloud cover (Norris et al. 2016) and aerosol emissions (Wild et al. 2009), and at smaller scales due to trends in forest overstory cover (Landuyt et al. 2019). These trends may alter plant fitness because light serves as both a resource essential for photosynthesis and a potential cause of stress due to photoinhibition.

Here, I define photoinhibition narrowly as damage caused by non-thermal energetic consequences of light absorption. Kok (1956) provided early evidence of photoinhibition by showing that algal cultures had lower photosynthesis after exposure to intense light. The study of photoinhibition gained momentum in the 1980s as researchers investigated the structure and function of Photosystem II (PSII), whose D1 protein is particularly susceptible to inactivation by light (Long et al. 1994). Around the same time, new tools like portable chlorophyll fluorometers made it easier to measure PSII efficiency (Schreiber et al. 1975). Photoinhibition is best studied as a result of oxidative damage from photosynthetically active radiation (PAR; 400-700 nm), although it can also be caused by much of the ultraviolet range (UV; Takahashi et al. 2010; Mattila et al. 2023). UV-B can also increase DNA mutation rates by creating pyrimidine dimers (Britt 2004). The effects of UV-related mutational load on fitness are variable (Llorens et al. 2015) and hard to extricate from light's effects on PSII, except perhaps by selectively filtering parts of the light spectrum (Lesser 1996; van de Poll et al. 2001).

In light-exposed leaves, reactive oxygen species (ROS) are produced mainly in chloroplasts and peroxisomes (Foyer & Noctor 2003). In chloroplasts, ROS like singlet oxygen ($^{1}O_{2}$) are generated during the transfer of excitation energy at both PSII and Photosystem I (PSI), rendering both susceptible to damage. Indeed, PSI damage is harder to reverse but also rarer (Sonoike 2011), so lasting photoinhibition

is most often taken to occur when the rate of PSII damage exceeds the rate of repair (Long et al. 1994). This situation can occur when pigments absorb light energy in excess of the plant's ability to dissipate it safely through either photochemical or non-photochemical quenching (e.g. through the xanthophyll cycle; Demmig-Adams & Adams 2006). There remains debate about the main mechanism(s) of this effect. In general, excess light causes the PSII plastoquinone pool to become overreduced, and the resulting limitation in electron transport increases the formation of excited triplet chlorophyll. Next, triplet chlorophyll can interact with O₂ to create ¹O₂ (Mattila et al. 2023), which can either damage PSII (acceptor-side inhibition; Vass 2011) or inhibit repair of PSII that has been damaged directly by light (donor-side inhibition; Hakala et al. 2005; Takahashi & Murata 2008). Stresses like low temperatures or drought tend to exacerbate photoinhibition by reducing photochemical dissipation of light energy, perhaps among other mechanisms (Fig. 1; Hutchison et al. 2000; Takahashi & Murata 2008). Growing evidence suggests that multiple mechanisms contribute to PSII damage under natural conditions, with the balance determined by the amount and spectrum of light and by other environmental conditions like temperature (Kono et al. 2022; Mattila et al. 2023). Acceptor- and donor-side mechanisms often produce different ROS (Kale et al. 2017), which differ in their lifetime, reactivity, and ability to cross membranes (Møller et al. 2007; Mittler 2017). Whether their ecological consequences differ is largely unexplored.

In defining photoinhibition as involving 'damage,' I distinguish it from biochemical mechanisms of photoprotection that prevent damage, which would last longer and be more costly to reverse. The line between sustained biochemical photoprotection and damage is blurry, since it is often ambiguous whether an indicator of reduced electron transport function (like declines in functioning D1 protein) is best interpreted as a result of damage or an adaptive mechanism to *avert* more severe consequences when other factors constrain photochemical dissipation (Adams et al. 2013; Tikkanen et al. 2014). Beyond just the biochemical scale, photoprotective mechanisms include diverse ways to reduce light reaching the chloroplast, dissipate excess energy, or scavenge reactive species (Raven 2011; D'Alessandro 2020), but many of these mechanisms have known drawbacks. For example, all plants grow in environments where

light fluctuates on some time-scale; but while reduced light absorption or non-photochemical quenching (NPQ) induction may be advantageous under high light, they may also reduce photosynthesis when light declines (Raven 2011; De Souza et al. 2022). Plants might thus be expected to evolve towards an optimal level of photoprotection for their typical growth environment.

Although the mechanisms of photoinhibition and photoprotection have become clearer through time, their relevance for population and community ecology remains the subject of debate. Here, I aim to re-evaluate photoinhibition's influence on fitness in the context of our changing understanding of whole-plant function. I focus on land plants, although at times I draw examples from other photosynthetic organisms.

Photoinhibition and source-sink interactions

Given that photoinhibition involves damage to thylakoid membrane complexes, many studies of photoinhibition implicitly assume that its main cost to fitness is to reduce photosynthesis. Indeed, most attempts at quantifying the harm of photoinhibition focus on estimating energetic or other resource-related costs of foregone photosynthesis and protein repair (Murchie & Niyogi 2011; Raven 2011), and using these estimates to project the loss in plant productivity or growth (Ögren & Sjöstrom 1990; Long et al. 1994; Werner et al. 2001; Burgess et al. 2015; Kothari et al. 2021).

But do these costs explain how photoinhibition may reduce fitness? The answer depends on how carbon-limited fitness is. In our high-[CO₂] world, plant growth under moderate stress is often regulated less by carbon assimilation than by direct constraints on tissue expansion or carbon export from leaves (Cabon et al. 2022; Fig. 1). In other words, plants are often sink-limited, particularly under many of the conditions that also cause photoinhibition: the combination of high light and moderate drought, nutrient limitation, or cold temperatures (Körner 2003; Muller et al. 2011). The result often manifests as a buildup of non-structural carbohydrates (NSCs) in tissues (Körner 2003). Elevated NSCs may be mobilized as osmolytes that promote drought tolerance, or as insurance against future disturbance, which are useful for long-term

growth and survival (Sala et al. 2012). However, accumulation of NSCs (especially osmotically inactive ones like starch) suggests that carbon is at least not limiting in the short term.

Although studies of photoinhibition often estimate reductions in photosynthesis, they seldom measure NSC reserves. In those studies that have, hallmarks of photoinhibition are often accompanied by NSC accumulation rather than depletion, consistent with the idea that photoinhibitory conditions are often sink-limiting (Hogewoning & Harbinson 2007; Adams et al. 2013). If the consequences of photoinhibition are described mainly in terms of carbon limitation, in many cases it becomes a puzzle how it could influence fitness (Slot et al. 2005).

These considerations led Adams et al. (2013) to propose that photoinhibition is mainly a *consequence* of growth limitation, not a cause. Accumulation of NSCs causes plants to reorganize their metabolism to downregulate photosynthesis and promote sink activity (White et al. 2016; Fig. 1). Thus, experimental stimulation of source activity or blockage of sink activity can cause many of the physiological hallmarks of photoinhibition, including declines in dark F_v/F_m and D1 activity (Adams et al. 2013). Many plants acclimate by upregulating photoprotection (e.g. NPQ) enough to balance the decline in photochemical quenching, allowing them to avoid damage. Indeed, Adams et al. (2013) question "whether there is any situation in which photoinhibition could be construed as damage or could be limiting plant productivity"—except under conditions seldom found in nature. Without taking a position on the general prevalence of source- vs. sink-limitation, I argue that this challenge should cause us to reconsider *how* photoinhibition matters for plant fitness.

Can excess light reduce fitness?

Here, I try to establish that despite the challenge posed above, photoinhibition can reduce plant fitness. In doing so, I avoid using biochemical or physiological indicators alone. The first reason is the ambiguity about when such indicators are best interpreted as photoinhibition or photoprotection. The second is that

many common indicators—including fluorescence-based measures like (dark- or light-acclimated) F_v/F_m —estimate the efficiency of electron transport, which is not the same as photosynthesis. Because light is both a stressor and an essential resource, an increase in light often causes both a decline in photosynthetic efficiency and an increase in actual photosynthesis. Furthermore, photosynthesis under high light is often limited by RuBisCO kinetics rather than electron transport. The third reason is that, as mentioned earlier, photosynthetic carbon fixation need not always be limiting for fitness.

So how can we show that photoinhibition does (or doesn't) matter? I focus on how excess light affects plant fitness (or proxies) and its consequences, like species distributions, drawing on physiological indicators only in a supporting role. The question is: all else held equal, does increasing available light beyond a certain point cause fitness to decline and/or favor species with stronger photoprotective adaptations (Fig. 2)? The stipulation that all else be held equal is hard to meet exactly, since gradients of light (artificial or natural) are often confounded with factors like air temperature and humidity (Niinemets & Valladares 2004). While these other factors should be controlled as well as possible, studies of light stress need not be done in the complete absence of other abiotic stresses. Indeed, most stresses are only harmful in combination (Mittler 2006; Zandalinas et al. 2021). What *is* important is that the severity of other stresses stays relatively constant across light environments.

From an evolutionary perspective, it would be hard to explain the ubiquity of photoprotective pathways like the xanthophyll cycle(s) if photoinhibition posed no potential threat (Esteban et al. 2009; Goss & Lepetit 2015). Likewise, many lineages have putative light avoidance 'behaviors' like chloroplast movement in plants (Howard et al. 2020) and whole organismal movement in phytoplankton (Raven 2011). Species naturally found in bright environments are often better at avoiding or recovering from photoinhibition (Montgomery et al. 2008; Kothari et al. 2021; Fig. 2A). This pattern holds even in common garden-like environments, which implies that it is not just a result of plasticity. Indeed, organisms in extremely bright, hot environments often have extraordinary mechanisms to avoid and

dissipate excess light (Levin et al. 2021). Mutants of species like *Arabidopsis* also provide a simple system for revealing the importance of photoprotection: under natural conditions, mutants deficient in NPQ produce fewer seeds (Külheim et al. 2002) and mutants deficient in chloroplast movement grow less (Howard et al. 2020). These findings imply that optimal allocation to photoprotection is important for maintaining fitness across light environments.

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Other evidence comes from experiments that grow plants across light environments (Fig. 2B). Researchers have long described how high light can suppress population growth in phytoplankton (Edwards et al. 2015; Croteau et al. 2022). Among land plants, interpretation is often complicated due to potential confounding between light and other microclimatic factors. Appendix S1 summarizes more than 20 case studies that provide evidence that increases in light can reduce plant growth or fitness. Many of these studies implemented treatments to disentangle light from other factors and monitored the microclimate to confirm their effectiveness. For example, Bader et al. (2006) and Baumeister & Callaway (2006) each crossed shading with other treatments to show that high light could account for much of the failure of trees to establish beyond (respectively) tropical and temperate treelines. Elevated daytime leaf temperature may be among the factors most susceptible to confounding with high light since the absorption of shortwave radiation is part of the leaf energy budget (Muir 2019). However, many case studies found that high light is detrimental to survival and growth even in cold environments, where increased leaf temperature would be expected to be beneficial (e.g. Ball et al. 1991; Close et al. 2000; Akhalkatsi et al. 2001; Egerton et al. 2002). Likewise, Germino et al. (2002) found that survival among spruce seedlings at alpine treelines was halved when overtopping grasses were removed, even though the removal treatment ameliorated water and nighttime low-temperature stress. Other studies ruled out other factors by manipulating them directly. For example, Velasco & Becerra (2020) found that artificial shading provided a much stronger benefit than irrigation on tree seedling survival in a semiarid grassland, which suggests that the seedlings were not strongly water-limited and the effect of shading was not due mainly to reduced transpiration.

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A whole-plant perspective on photoinhibition

to alleviate those other stresses.

stressful environments (Cabal et al. 2022).

Many of the case studies in Appendix S1 are from settings like alpine treelines and drylands, where

interacting stresses like low temperature or drought may make photoinhibition more salient. Most studies

on trees were also conducted using seedlings; this may be partly a matter of convenience, but it may also

One implication of this research is that shading may be a common mechanism of facilitation. This point is

again clear in phytoplankton, among which theoretical (Gerla et al. 2011) and empirical (Veraart et al.

2012; Faassen et al. 2015) research suggests that facilitative amelioration of photoinhibition can produce

alternative stable states and allow light-sensitive species to establish. In trees, Kothari et al. (2021) also

neighbors. Much of the research on recruitment at treelines or in canopy gaps emphasizes facilitation

between adult trees and seedlings (Ball et al. 1991; Egerton et al. 2000; Slot et al. 2005). In the context of

Understanding that photoinhibition can reduce fitness can also help explain the uses of certain ecological

combination with other stresses like drought, it may be less expensive to reduce the intensity of light than

interventions. For example, reforestation experiments often find that experimental shading increases

survival (Egerton et al. 2000; Velasco & Becerra 2020). While high light is usually most harmful in

found that fast-growing, shade-intolerant species reduced photoinhibition in their light-sensitive

research guided by the stress-gradient hypothesis (Bertness & Callaway 1994), amelioration of

photoinhibition may be one of the mechanisms through which nurse plants aid their beneficiaries in

reflect a concern that trees are most vulnerable to photoinhibition during recruitment (Germino et al.

So far, I claim there is evidence that photoinhibition can reduce plant fitness. Perhaps in part because photosynthesis is easily measured in the field, ecophysiological studies often explain this influence in terms of reductions in carbon assimilation. However, the critiques described above raise doubts about the generality of such explanations, and it may help to look toward other aspects of plant function. Molecular physiologists have mapped out a much wider range of consequences of excess light-induced ROS generation, which may lead to cellular damage or even death (D'Alessandro et al. 2020)—but primarily in a few model organisms and without yet having demonstrated their importance in nature.

It may be time for a synthesis: Could excess light-induced ROS generation have a role in natural settings beyond its influence on carbon assimilation via electron transport? Besides attacking thylakoid proteins like D1, species like $^{1}O_{2}$ produced along the electron transport chain can also react with the unsaturated lipids in the thylakoid and produce lipid peroxides, which in turn break into reactive carbonyl species (RCS). The products of these cascading reactions can cause further damage and, at high enough levels, trigger pathways towards cell death (Møller et al. 2007; Chan et al. 2012; D'Alessandro et al. 2020). This damage may help explain observations of bleaching and necrosis (Powles 1984; Hogewoning & Harbinson 2007) or leaf abscission (Egerton et al. 2000; Close et al. 2002) under high light. Even at more modest levels, the signaling functions of ROS and their products can initiate other changes beyond the chloroplast. For example, oxidative stress causes callose deposition in phloem-loading complexes, which may have the adaptive role of blocking the spread of pathogens (when present), but also hinders the export of sugars and other compounds from leaves (Demmig-Adams et al. 2014).

These broader consequences of oxidative damage may often matter more than reductions in carbon assimilation. For example, although most research on PSII-inhibiting herbicides in seagrasses and algae focuses on reductions in photosynthesis, other sorts of oxidative damage may cause greater harm (King et al. 2021). Prescott et al. (2020) even suggest that sink-limited plants may dispose of surplus carbon to

avoid feedback inhibition of photosynthesis and the resulting risk of photoinhibition—a proposal which would require that photoinhibition have potential effects besides reducing carbon assimilation.

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Whether any of these adverse consequences occurs depends on whether the plant can acclimate to its environment. In general, ROS and their diverse products are essential as redox signals that coordinate development, signaling, and acclimation to stressful conditions, including upregulation of photoprotection (Mittler 2017). This fact has been taken to imply that elevated ROS production under photoinhibitory conditions is generally not harmful (Foyer 2018). However, the danger may arise when there are runaway spirals of ROS production that plants cannot control. For example, if unrepaired damage to photosynthetic machinery limits photochemical light dissipation, the share of light in excess may rise, which could increase ROS production (Fig. 1) and perhaps create a cycle in which photoinhibition begets more photoinhibition. This possibility is implied by findings that depending on their mechanism, PSIIinhibiting herbicides can steeply enhance ROS production (Fufezan et al. 2002; Chen et al. 2010). ROS and RCS may also inactivate the enzymes that scavenge and detoxify them (D'Alessandro et al. 2020). Plants adapted to bright or fluctuating light often have a strong capacity to acclimate and maintain homeostasis through protective mechanisms like NPQ or ROS scavenging. However, plants adapted to shade or subject to interacting stresses may have more limited capacity (Foyer et al. 1994). Hence, it seems premature to conclude that the physiological consequences of excess light exposure are necessarily benign; such conclusions can only be drawn by observing fitness. Some researchers have suggested that photoinhibition may only matter in unrealistic circumstances, as when plants are beyond their usual niche (Adams et al. 2013). But this is scant reason to dismiss its importance: part of the business of community ecology is to explain why plants don't grow where they don't grow.

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One specific situation in which photoinhibition may reduce fitness other than through energetic costs is during senescence in winter- (and perhaps drought-) deciduous plants. Efficient nutrient resorption during senescence is critical for growth and fitness in following years (May & Killingbeck 1992; El Zein et al.

2011). Senescing leaves are highly vulnerable to light-induced damage (Kar et al. 1993; Merzlyak & Hendry 1994), and the red anthocyanins many species produce in the fall may serve to improve nutrient resorption by screening light (Renner & Zohner 2019). While failures of nutrient resorption could result if photoinhibition causes carbon depletion (Hoch et al. 2001), they might also result from oxidative modifications to proteins involved in breaking down nutrient-containing molecules, or from callose-induced blockage of phloem loading.

On the other hand, one might ask when photoinhibition could be more likely to cause or exacerbate carbon limitation. One example is the combination of high light and low or fluctuating [CO₂], which constrains photochemistry and carbon uptake without directly constraining sink strength (Durchan et al. 2001) and may affect many aquatic producers (Ibelings & Maberly 1998). Likewise, while species adapted to strongly fluctuating light often have high capacity for flexible NPQ (Demmig-Adams & Adams 2006), any damage incurred during transient periods of high light could cause carbon limitation under low light. More speculatively, this question may depend on the relative sensitivity of plants' carbon sources and sinks to environmental stresses. For example, under water limitation, species whose stomatal regulation keeps water potential more stable (i.e. 'isohydric' species) may have lower photosynthesis while more often maintaining the turgor required for tissue growth. This situation may cause carbon depletion, which could worsen if the decline in photochemical dissipation also causes photoinhibition (McDowell et al. 2008). In practice, however, the relationship between stomatal regulation and carbon balance is more complicated (Martínez-Vilalta & Garcia-Forner 2017). As our understanding develops of how plants regulate carbon assimilation and allocation, so too may our understanding of when and how photoinhibition matters.

Considering photoinhibition's role beyond just photosynthesis may require a broadened range of methods.

To establish whether photoinhibition has consequences for fitness, it is important to measure growth or demographic rates and to include light treatments high enough for those consequences to manifest.

Indeed, those studying photoinhibition or light limitation as factors in community assembly should consider exposing plants to light regimes beyond those experienced by (but found within dispersal distance of) existing populations, since a major goal of community ecology is to explain what excludes species beyond their niche (Cadotte & Tucker 2017). Physiological indicators should be chosen and interpreted with care. Measuring NSCs could help establish whether photoinhibition is depleting carbon reserves (Slot et al. 2005; Hogewoning & Harbinson 2007; Adams et al. 2013), but this approach is best treated only as a heuristic of carbon limitation (Weber et al. 2019). To disentangle flexible photoprotection from potential damage, one fluorescence parameter that may be useful is the fraction of light energy not dissipated through either photochemistry or NPQ (sometimes denoted Φ_{NO} ; Demmig-Adams et al. 1996; Kramer et al. 2004; Fig. 1). Measuring the consequences of oxidative damage at the molecular level is tricky under field conditions, but the relatively simple TBARS assay for lipid peroxidation may be a good candidate (Hodges et al. 1999). When these indicators are coincident with variation in fitness, they may provide support for causal claims about photoinhibition's influence.

In summary, we can describe two hypotheses for how photoinhibition may influence plant fitness: (1) by reducing carbon assimilation, and (2) by causing oxidative damage that may lead to cell death (King et al. 2021). These two hypotheses are not mutually exclusive—in fact, they are intertwined—but focusing on the first alone limits our understanding of how photoinhibition affects fitness. These hypotheses may bear different implications under global change: if sink limitation is growing in prevalence (Cabon et al. 2022), focusing on carbon assimilation alone may lead researchers to conclude prematurely that photoinhibition is declining in relevance. Given the scope of human alteration of light regimes, it is essential to understand when and how photoinhibition matters—not just in the leaf, but across the whole plant.

318	Acknowledgements
319	I would like to thank Jeannine Cavender-Bares, Rebecca Montgomery, and Daniel Stanton for useful
320	discussions and for encouraging my interest in photoinhibition, however misguided it may be. I also thank
321	Etienne Laliberté and Alain Paquette for allowing me the time to work on this paper.
322	
323	Author Contributions
324	Shan Kothari was responsible for conceptualizing, writing, and revising this article.
325	
326	Data Availability Statement
327	This article contains no data.

328	Literature Cited						
329	Adams, W. W., Muller, O., Cohu, C. M., & Demmig-Adams, B. (2013). May photoinhibition be a						
330	consequence, rather than a cause, of limited plant productivity? Photosynthesis Research, 117(1),						
331	31–44. https://doi.org/10.1007/s11120-013-9849-7						
332	Akhalkatsi, M., Abdaladze, O., Nakhutsrishvili, G., & Smith, W. K. (2006). Facilitation of Seedling						
333	Microsites by Rhododendron Caucasicum Extends the Betula Litwinowii Alpine Treeline,						
334	Caucasus Mountains, Republic of Georgia. Arctic, Antarctic, and Alpine Research, 38(4), 481-						
335	488. https://doi.org/10.1657/1523-0430(2006)38[481:FOSMBR]2.0.CO;2						
336	Bader, M. Y., van Geloof, I., & Rietkerk, M. (2007). High solar radiation hinders tree regeneration above						
337	the alpine treeline in northern Ecuador. <i>Plant Ecology</i> , 191(1), 33–45.						
338	https://doi.org/10.1007/s11258-006-9212-6						
339	Ball, M. C., Hodges, V. S., & Laughlin, G. P. (1991). Cold-Induced Photoinhibition Limits Regeneration						
340	of Snow Gum at Tree-Line. Functional Ecology, 5(5), 663–668. https://doi.org/10.2307/2389486						
341	Baumeister, D., & Callaway, R. M. (2006). Facilitation by Pinus Flexilis During Succession: A Hierarchy						
342	of Mechanisms Benefits Other Plant Species. <i>Ecology</i> , 87(7), 1816–1830.						
343	https://doi.org/10.1890/0012-9658(2006)87[1816:FBPFDS]2.0.CO;2						
344	Bertness, M. D., & Callaway, R. (1994). Positive interactions in communities. <i>Trends in Ecology &</i>						
345	Evolution, 9(5), 191–193. https://doi.org/10.1016/0169-5347(94)90088-4						
346	Britt, A. B. (2004). Repair of DNA Damage Induced by Solar UV. <i>Photosynthesis Research</i> , 81(2), 105–						
347	112. https://doi.org/10.1023/B:PRES.0000035035.12340.58						
348	Burgess, A. J., Retkute, R., Pound, M. P., Foulkes, J., Preston, S. P., Jensen, O. E., Pridmore, T. P., &						
349	Murchie, E. H. (2015). High-Resolution Three-Dimensional Structural Data Quantify the Impact						
350	of Photoinhibition on Long-Term Carbon Gain in Wheat Canopies in the Field. Plant Physiology						
351	169(2), 1192–1204. https://doi.org/10.1104/pp.15.00722						
352	Cabal, C., Valladares, F., & Martinez-Garcia, R. (2022). The Ecology of Plant Interactions: A Giant with						
353	Feet of Clay. https://doi.org/10.20944/preprints202009.0520.v3						

354	Cabon, A., Kannenberg, S. A., Arain, A., Babst, F., Baldocchi, D., Belmecheri, S., Delpierre, N.,						
355	Guerrieri, R., Maxwell, J. T., McKenzie, S., Meinzer, F. C., Moore, D. J. P., Pappas, C., Rocha,						
356	A. V., Szejner, P., Ueyama, M., Ulrich, D., Vincke, C., Voelker, S. L., Anderegg, W. R. L.						
357	(2022). Cross-biome synthesis of source versus sink limits to tree growth. Science, 376(6594),						
358	758–761. https://doi.org/10.1126/science.abm4875						
359	Cadotte, M. W., & Tucker, C. M. (2017). Should Environmental Filtering be Abandoned? Trends in						
360	Ecology & Evolution, 32(6), 429–437. https://doi.org/10.1016/j.tree.2017.03.004						
361	Chan, T., Shimizu, Y., Pospíšil, P., Nijo, N., Fujiwara, A., Taninaka, Y., Ishikawa, T., Hori, H., Nanba,						
362	D., Imai, A., Morita, N., Yoshioka-Nishimura, M., Izumi, Y., Yamamoto, Y., Kobayashi, H.,						
363	Mizusawa, N., Wada, H., & Yamamoto, Y. (2012). Quality Control of Photosystem II: Lipid						
364	Peroxidation Accelerates Photoinhibition under Excessive Illumination. PLOS ONE, 7(12),						
365	e52100. https://doi.org/10.1371/journal.pone.0052100						
366	Chen, S., Yin, C., Qiang, S., Zhou, F., & Dai, X. (2010). Chloroplastic oxidative burst induced by						
367	tenuazonic acid, a natural photosynthesis inhibitor, triggers cell necrosis in Eupatorium						
368	adenophorum Spreng. Biochimica et Biophysica Acta (BBA) - Bioenergetics, 1797(3), 391–405.						
369	https://doi.org/10.1016/j.bbabio.2009.12.007						
370	Close, D. C., Beadle, C. L., Holz, G. K., & Brown, P. H. (2002). Effect of shadecloth tree shelters on						
371	cold-induced photoinhibition, foliar anthocyanin and growth of Eucalyptus globulus and E. nitens						
372	seedlings during establishment. Australian Journal of Botany, 50(1), 15-20.						
373	https://doi.org/10.1071/bt01038						
374	Croteau, D., Lacour, T., Schiffrine, N., Morin, PI., Forget, MH., Bruyant, F., Ferland, J., Lafond, A.,						
375	Campbell, D. A., Tremblay, JÉ., Babin, M., & Lavaud, J. (2022). Shifts in growth light optima						
376	among diatom species support their succession during the spring bloom in the Arctic. Journal of						
377	Ecology, 110(6), 1356–1375. https://doi.org/10.1111/1365-2745.13874						

378	D'Alessandro, S., Beaugelin, I., & Havaux, M. (2020). Tanned or Sunburned: How Excessive Light						
379	Triggers Plant Cell Death. Molecular Plant, 13(11), 1545–1555.						
380	https://doi.org/10.1016/j.molp.2020.09.023						
381	De Souza, A. P., Burgess, S. J., Doran, L., Hansen, J., Manukyan, L., Maryn, N., Gotarkar, D., Leonelli,						
382	L., Niyogi, K. K., & Long, S. P. (2022). Soybean photosynthesis and crop yield are improved by						
383	accelerating recovery from photoprotection. Science, 377(6608), 851–854.						
384	https://doi.org/10.1126/science.adc9831						
385	Demmig-Adams, B., & Adams III, W. W. (2006). Photoprotection in an ecological context: The						
386	remarkable complexity of thermal energy dissipation. New Phytologist, 172(1), 11–21.						
387	https://doi.org/10.1111/j.1469-8137.2006.01835.x						
388	Demmig-Adams, B., Adams III, W. W., Barker, D. H., Logan, B. A., Bowling, D. R., & Verhoeven, A. S.						
389	(1996). Using chlorophyll fluorescence to assess the fraction of absorbed light allocated to						
390	thermal dissipation of excess excitation. Physiologia Plantarum, 98(2), 253-264.						
391	https://doi.org/10.1034/j.1399-3054.1996.980206.x						
392	Demmig-Adams, B., Stewart, J. J., & Adams, W. W. (2014). Multiple feedbacks between chloroplast and						
393	whole plant in the context of plant adaptation and acclimation to the environment. Philosophical						
394	Transactions of the Royal Society B: Biological Sciences, 369(1640), 20130244.						
395	https://doi.org/10.1098/rstb.2013.0244						
396	Durchan, M., Vácha, F., & Krieger-Liszkay, A. (2001). Effects of severe CO2 starvation on the						
397	photosynthetic electron transport chain in tobacco plants. Photosynthesis Research, 68(3), 203-						
398	213. https://doi.org/10.1023/A:1012917428003						
399	Edwards, K. F., Thomas, M. K., Klausmeier, C. A., & Litchman, E. (2015). Light and growth in marine						
100	phytoplankton: Allometric, taxonomic, and environmental variation. Limnology and						
101	Oceanography, 60(2), 540–552. https://doi.org/10.1002/lno.10033						
102	Egerton, J. J. G., Banks, J. C. G., Gibson, A., Cunningham, R. B., & Ball, M. C. (2000). Facilitation of						
103	Seedling Establishment: Reduction in Irradiance Enhances Winter Growth of Eucalyptus						

404	Pauciflora. <i>Ecology</i> , 81(5), 1437–1449. https://doi.org/10.1890/0012-							
405	9658(2000)081%5b1437:FOSERI%5d2.0.CO;2							
406	El Zein, R., Bréda, N., Gérant, D., Zeller, B., & Maillard, P. (2011). Nitrogen sources for current-year							
407	shoot growth in 50-year-old sessile oak trees: An in situ 15N labeling approach. Tree Physiology							
408	31(12), 1390–1400. https://doi.org/10.1093/treephys/tpr118							
409	Esteban, R., Olano, J. M., Castresana, J., Fernández-Marín, B., Hernández, A., Becerril, J. M., & García-							
410	Plazaola, J. I. (2009). Distribution and evolutionary trends of photoprotective isoprenoids							
411	(xanthophylls and tocopherols) within the plant kingdom. <i>Physiologia Plantarum</i> , 135(4), 379–							
412	389. https://doi.org/10.1111/j.1399-3054.2008.01196.x							
413	Faassen, E. J., Veraart, A. J., Van Nes, E. H., Dakos, V., Lürling, M., & Scheffer, M. (2015). Hysteresis							
414	in an experimental phytoplankton population. Oikos, 124(12), 1617–1623.							
415	https://doi.org/10.1111/oik.02006							
416	Foyer, C. H. (2018). Reactive oxygen species, oxidative signaling and the regulation of photosynthesis.							
417	Environmental and Experimental Botany, 154, 134–142.							
418	https://doi.org/10.1016/j.envexpbot.2018.05.003							
419	Foyer, C. H., Lelandais, M., & Kunert, K. J. (1994). Photooxidative stress in plants. <i>Physiologia</i>							
420	Plantarum, 92(4), 696–717. https://doi.org/10.1111/j.1399-3054.1994.tb03042.x							
421	Foyer, C. H., & Noctor, G. (2003). Redox sensing and signalling associated with reactive oxygen in							
422	chloroplasts, peroxisomes and mitochondria. Physiologia Plantarum, 119(3), 355-364.							
423	https://doi.org/10.1034/j.1399-3054.2003.00223.x							
424	Fufezan, C., Rutherford, A. W., & Krieger-Liszkay, A. (2002). Singlet oxygen production in herbicide-							
425	treated photosystem II. FEBS Letters, 532(3), 407–410. <u>https://doi.org/10.1016/S0014-</u>							
426	<u>5793(02)03724-9</u>							
427	Gerla, D. J., Mooij, W. M., & Huisman, J. (2011). Photoinhibition and the assembly of light-limited							
428	phytoplankton communities. Oikos, 120(3), 359–368. https://doi.org/10.1111/j.1600-							
429	0706.2010.18573.x							

430	Germino, M. J., Smith, W. K., & Resor, A. C. (2002). Conifer seedling distribution and survival in an						
431	alpine-treeline ecotone. Plant Ecology, 162(2), 157–168.						
432	https://doi.org/10.1023/A:1020385320738						
433	Goss, R., & Lepetit, B. (2015). Biodiversity of NPQ. Journal of Plant Physiology, 172, 13–32.						
434	https://doi.org/10.1016/j.jplph.2014.03.004						
435	Hakala, M., Tuominen, I., Keränen, M., Tyystjärvi, T., & Tyystjärvi, E. (2005). Evidence for the role of						
436	the oxygen-evolving manganese complex in photoinhibition of Photosystem II. Biochimica et						
437	Biophysica Acta (BBA) - Bioenergetics, 1706(1), 68–80.						
438	https://doi.org/10.1016/j.bbabio.2004.09.001						
439	Hoch, W. A., Zeldin, E. L., & McCown, B. H. (2001). Physiological significance of anthocyanins during						
440	autumnal leaf senescence. Tree Physiology, 21(1), 1–8. https://doi.org/10.1093/treephys/21.1.1						
441	Hodges, D. M., DeLong, J. M., Forney, C. F., & Prange, R. K. (1999). Improving the thiobarbituric acid-						
442	reactive-substances assay for estimating lipid peroxidation in plant tissues containing anthocyanin						
443	and other interfering compounds. Planta, 207(4), 604–611.						
444	https://doi.org/10.1007/s004250050524						
445	Hogewoning, S. W., & Harbinson, J. (2007). Insights on the development, kinetics, and variation of						
446	photoinhibition using chlorophyll fluorescence imaging of a chilled, variegated leaf. Journal of						
447	Experimental Botany, 58(3), 453-463. https://doi.org/10.1093/jxb/erl219						
448	Howard, M. M., Bae, A., Pirani, Z., Van, N., & Königer, M. (2020). Impairment of chloroplast movement						
449	reduces growth and delays reproduction of Arabidopsis thaliana in natural and controlled						
450	conditions. American Journal of Botany, 107(9), 1309–1318. https://doi.org/10.1002/ajb2.1537						
451	Hutchison, R. S., Groom, Q., & Ort, D. R. (2000). Differential Effects of Chilling-Induced						
452	Photooxidation on the Redox Regulation of Photosynthetic Enzymes. Biochemistry, 39(22),						
453	6679–6688. https://doi.org/10.1021/bi0001978						

454 Ibelings, B. W., & Maberly, S. C. (1998). Photoinhibition and the availability of inorganic carbon restrict photosynthesis by surface blooms of cyanobacteria. Limnology and Oceanography, 43(3), 408-455 456 419. https://doi.org/10.4319/lo.1998.43.3.0408 457 Kale, R., Hebert, A. E., Frankel, L. K., Sallans, L., Bricker, T. M., & Pospíšil, P. (2017). Amino acid 458 oxidation of the D1 and D2 proteins by oxygen radicals during photoinhibition of Photosystem II. Proceedings of the National Academy of Sciences, 114(11), 2988–2993. 459 460 https://doi.org/10.1073/pnas.1618922114 Kar, M., Streb, P., Hertwig, B., & Feierabend, J. (1993). Sensitivity to photodamage increases during 461 462 senescence in excised leaves. Journal of Plant Physiology, 141(5), 538–544. https://doi.org/10.1016/S0176-1617(11)80453-0 463 464 King, O. C., Smith, R. A., Warne, M. S. J., Merwe, J. P. van de, Connolly, R. M., & Brown, C. J. (2021). 465 Combined impacts of photosystem II-inhibiting herbicides and light availability on seagrass and 466 marine microalgae. Marine Ecology Progress Series, 668, 215–230. https://doi.org/10.3354/meps13717 467 468 Kok, B. (1956). On the inhibition of photosynthesis by intense light. *Biochimica et Biophysica Acta*, 469 21(2), 234–244. https://doi.org/10.1016/0006-3002(56)90003-8 470 Kono, M., Miyata, K., Matsuzawa, S., Noguchi, T., Oguchi, R., Suzuki, Y., Terashima, I., Kono, M., 471 Miyata, K., Matsuzawa, S., Noguchi, T., Oguchi, R., Suzuki, Y., & Terashima, I. (2022). Mixed 472 population hypothesis of the active and inactive PSII complexes opens a new door for photoinhibition and fluorescence studies: An ecophysiological perspective. Functional Plant 473 Biology, 49(10), 917–925. https://doi.org/10.1071/FP21355 474 475 Körner, C. (2003). Carbon limitation in trees. *Journal of Ecology*, 91(1), 4–17. 476 https://doi.org/10.1046/j.1365-2745.2003.00742.x 477 Kothari, S., Montgomery, R. A., & Cavender-Bares, J. (2021). Physiological responses to light explain 478 competition and facilitation in a tree diversity experiment. Journal of Ecology, 109(5), 2000– 479 2018. https://doi.org/10.1111/1365-2745.13637

480	Kramer, D. M., Johnson, G., Kiirats, O., & Edwards, G. E. (2004). New Fluorescence Parameters for the						
481	Determination of QA Redox State and Excitation Energy Fluxes. Photosynthesis Research, 79(2),						
482	209–218. https://doi.org/10.1023/B:PRES.0000015391.99477.0d						
483	Külheim, C., Ågren, J., & Jansson, S. (2002). Rapid Regulation of Light Harvesting and Plant Fitness in						
484	the Field. Science, 297(5578), 91–93. https://doi.org/10.1126/science.1072359						
485	Landuyt, D., De Lombaerde, E., Perring, M. P., Hertzog, L. R., Ampoorter, E., Maes, S. L., De Frenne,						
486	P., Ma, S., Proesmans, W., Blondeel, H., Sercu, B. K., Wang, B., Wasof, S., & Verheyen, K.						
487	(2019). The functional role of temperate forest understorey vegetation in a changing world.						
488	Global Change Biology, 25(11), 3625–3641. https://doi.org/10.1111/gcb.14756						
489	Laughlin, D. C., & Messier, J. (2015). Fitness of multidimensional phenotypes in dynamic adaptive						
490	landscapes. Trends in Ecology & Evolution, 30(8), 487–496.						
491	https://doi.org/10.1016/j.tree.2015.06.003						
492	Lesser, M. (1996). Acclimation of phytoplankton to UV-B radiation: Oxidative stress and photoinhibition						
493	of photosynthesis are not prevented by UV-absorbing compounds in the dinoflagellate						
494	Prorocentrum micans. Marine Ecology Progress Series, 132, 287–297.						
495	https://doi.org/10.3354/meps132287						
496	Levin, G., Kulikovsky, S., Liveanu, V., Eichenbaum, B., Meir, A., Isaacson, T., Tadmor, Y., Adir, N., &						
497	Schuster, G. (2021). The desert green algae Chlorella ohadii thrives at excessively high light						
498	intensities by exceptionally enhancing the mechanisms that protect photosynthesis from						
499	photoinhibition. The Plant Journal, 106(5), 1260–1277. https://doi.org/10.1111/tpj.15232						
500	Llorens, L., Badenes-Pérez, F. R., Julkunen-Tiitto, R., Zidorn, C., Fereres, A., & Jansen, M. A. K. (2015).						
501	The role of UV-B radiation in plant sexual reproduction. Perspectives in Plant Ecology,						
502	Evolution and Systematics, 17(3), 243–254. https://doi.org/10.1016/j.ppees.2015.03.001						
503	Long, S. P., Humphries, S., & Falkowski, P. G. (1994). Photoinhibition of Photosynthesis in Nature.						
504	Annual Review of Plant Physiology and Plant Molecular Biology, 45(1), 633-662.						
505	https://doi.org/10.1146/annurev.pp.45.060194.003221						

506	Martinez-Vilalta, J., & Garcia-Forner, N. (2017). Water potential regulation, stomatal behaviour and							
507	hydraulic transport under drought: Deconstructing the iso/anisohydric concept. Plant, Cell &							
508	Environment, 40(6), 962–976. https://doi.org/10.1111/pce.12846							
509	Mattila, H., Mishra, S., Tyystjärvi, T., & Tyystjärvi, E. (2023). Singlet oxygen production by							
510	photosystem II is caused by misses of the oxygen evolving complex. New Phytologist, 237(1),							
511	113–125. https://doi.org/10.1111/nph.18514							
512	May, J. D., & Killingbeck, K. T. (1992). Effects of Preventing Nutrient Resorption on Plant Fitness and							
513	Foliar Nutrient Dynamics. <i>Ecology</i> , 73(5), 1868–1878. https://doi.org/10.2307/1940038							
514	McDowell, N., Pockman, W. T., Allen, C. D., Breshears, D. D., Cobb, N., Kolb, T., Plaut, J., Sperry, J.,							
515	West, A., Williams, D. G., & Yepez, E. A. (2008). Mechanisms of plant survival and mortality							
516	during drought: Why do some plants survive while others succumb to drought? New Phytologist,							
517	178(4), 719–739. https://doi.org/10.1111/j.1469-8137.2008.02436.x							
518	Merzlyak, M. N., & Hendry, G. a. F. (1994). Free radical metabolism, pigment degradation and lipid							
519	peroxidation in leaves during senescence. Proceedings of the Royal Society of Edinburgh, Section							
520	B: Biological Sciences, 102, 459–471. https://doi.org/10.1017/S0269727000014482							
521	Mittler, R. (2006). Abiotic stress, the field environment and stress combination. <i>Trends in Plant Science</i> ,							
522	11(1), 15–19. https://doi.org/10.1016/j.tplants.2005.11.002							
523	Mittler, R. (2017). ROS Are Good. Trends in Plant Science, 22(1), 11–19.							
524	https://doi.org/10.1016/j.tplants.2016.08.002							
525	Møller, I. M., Jensen, P. E., & Hansson, A. (2007). Oxidative Modifications to Cellular Components in							
526	Plants. Annual Review of Plant Biology, 58(1), 459–481.							
527	https://doi.org/10.1146/annurev.arplant.58.032806.103946							
528	Montgomery, R. A., Goldstein, G., & Givnish, T. J. (2008). Photoprotection of PSII in Hawaiian							
529	lobeliads from diverse light environments. Functional Plant Biology, 35(7), 595–605.							
530	Muir, C. D. (2019). tealeaves: An R package for modelling leaf temperature using energy budgets. AoB							
531	PLANTS, 11(6), plz054. https://doi.org/10.1093/aobpla/plz054							

532	Muller, B., Pantin, F., Génard, M., Turc, O., Freixes, S., Piques, M., & Gibon, Y. (2011). Water deficits						
533	uncouple growth from photosynthesis, increase C content, and modify the relationships between						
534	C and growth in sink organs. Journal of Experimental Botany, 62(6), 1715–1729.						
535	https://doi.org/10.1093/jxb/erq438						
536	Murchie, E. H., & Niyogi, K. K. (2011). Manipulation of Photoprotection to Improve Plant						
537	Photosynthesis. <i>Plant Physiology</i> , 155(1), 86–92. https://doi.org/10.1104/pp.110.168831						
538	Niinemets, Ü., & Valladares, F. (2004). Photosynthetic Acclimation to Simultaneous and Interacting						
539	Environmental Stresses Along Natural Light Gradients: Optimality and Constraints. Plant						
540	Biology, 6(3), 254–268. https://doi.org/10.1055/s-2004-817881						
541	Norris, J. R., Allen, R. J., Evan, A. T., Zelinka, M. D., O'Dell, C. W., & Klein, S. A. (2016). Evidence for						
542	climate change in the satellite cloud record. <i>Nature</i> , 536(7614), 72–75.						
543	https://doi.org/10.1038/nature18273						
544	Ögren, E., & Sjöström, M. (1990). Estimation of the effect of photoinhibition on the carbon gain in leaves						
545	of a willow canopy. <i>Planta</i> , 181(4), 560–567. https://doi.org/10.1007/BF00193011						
546	Powles, S. B. (1984). Photoinhibition of Photosynthesis Induced by Visible Light. <i>Annual Review of</i>						
547	Plant Physiology, 35(1), 15-44. https://doi.org/10.1146/annurev.pp.35.060184.000311						
548	Prescott, C. E., Grayston, S. J., Helmisaari, HS., Kaštovská, E., Körner, C., Lambers, H., Meier, I. C.,						
549	Millard, P., & Ostonen, I. (2020). Surplus Carbon Drives Allocation and Plant-Soil Interactions.						
550	Trends in Ecology & Evolution, 35(12), 1110–1118. https://doi.org/10.1016/j.tree.2020.08.007						
551	Raven, J. A. (2011). The cost of photoinhibition. <i>Physiologia Plantarum</i> , 142(1), 87–104.						
552	https://doi.org/10.1111/j.1399-3054.2011.01465.x						
553	Renner, S. S., & Zohner, C. M. (2019). The occurrence of red and yellow autumn leaves explained by						
554	regional differences in insolation and temperature. New Phytologist, 224(4), 1464-1471.						
555	https://doi.org/10.1111/nph.15900						
556	Sala, A., Woodruff, D. R., & Meinzer, F. C. (2012). Carbon dynamics in trees: Feast or famine? <i>Tree</i>						
557	Physiology, 32(6), 764-775. https://doi.org/10.1093/treephys/tpr143						

558	Schreiber, U., Groberman, L., & Vidaver, W. (1975). Portable, solid-state fluorometer for the						
559	measurement of chlorophyll fluorescence induction in plants. Review of Scientific Instruments,						
560	46(5), 538–542. https://doi.org/10.1063/1.1134252						
561	Sonoike, K. (2011). Photoinhibition of photosystem I. <i>Physiologia Plantarum</i> , 142(1), 56–64.						
562	https://doi.org/10.1111/j.1399-3054.2010.01437.x						
563	Slot, M., Wirth, C., Schumacher, J., Mohren, G. M. J., Shibistova, O., Lloyd, J., & Ensminger, I. (2005).						
564	Regeneration patterns in boreal Scots pine glades linked to cold-induced photoinhibition. Tree						
565	Physiology, 25(9), 1139–1150. https://doi.org/10.1093/treephys/25.9.1139						
566	Takahashi, S., Milward, S. E., Yamori, W., Evans, J. R., Hillier, W., & Badger, M. R. (2010). The Solar						
567	Action Spectrum of Photosystem II Damage. Plant Physiology, 153(3), 988–993.						
568	https://doi.org/10.1104/pp.110.155747						
569	Takahashi, S., & Murata, N. (2008). How do environmental stresses accelerate photoinhibition? <i>Trends in</i>						
570	Plant Science, 13(4), 178–182. https://doi.org/10.1016/j.tplants.2008.01.005						
571	Tikkanen, M., Mekala, N. R., & Aro, EM. (2014). Photosystem II photoinhibition-repair cycle protects						
572	Photosystem I from irreversible damage. Biochimica et Biophysica Acta (BBA) - Bioenergetics,						
573	1837(1), 210–215. https://doi.org/10.1016/j.bbabio.2013.10.001						
574	Van De Poll, W. H., Eggert, A., Buma, A. G. J., & Breeman, A. M. (2001). Effects of UV-B-Induced						
575	DNA Damage and Photoinhibition on Growth of Temperate Marine Red Macrophytes: Habitat-						
576	Related Differences in UV-B Tolerance. Journal of Phycology, 37(1), 30–38.						
577	https://doi.org/10.1046/j.1529-8817.2001.037001030.x						
578	Vass, I. (2011). Role of charge recombination processes in photodamage and photoprotection of the						
579	photosystem II complex. <i>Physiologia Plantarum</i> , 142(1), 6–16. https://doi.org/10.1111/j.1399-						
580	<u>3054.2011.01454.x</u>						
581	Velasco, N., & Becerra, P. I. (2020). Species-specific effects of the herbaceous layer on recruitment of						
582	woody species under different shading and precipitation conditions. Forest Ecology and						
583	Management, 460, 117864. https://doi.org/10.1016/j.foreco.2020.117864						

584	Veraart, A. J., Faassen, E. J., Dakos, V., van Nes, E. H., Lürling, M., & Scheffer, M. (2012). Recovery						
585	rates reflect distance to a tipping point in a living system. <i>Nature</i> , 481(7381), 357–359.						
586	https://doi.org/10.1038/nature10723						
587	Weber, R., Gessler, A., & Hoch, G. (2019). High carbon storage in carbon-limited trees. New Phytologist						
588	222(1), 171–182. https://doi.org/10.1111/nph.15599						
589	Werner, C., Ryel, R. J., Correia, O., & Beyschlag, W. (2001). Effects of photoinhibition on whole-plant						
590	carbon gain assessed with a photosynthesis model. Plant, Cell & Environment, 24(1), 27-40.						
591	https://doi.org/10.1046/j.1365-3040.2001.00651.x						
592	White, A. C., Rogers, A., Rees, M., & Osborne, C. P. (2016). How can we make plants grow faster? A						
593	source-sink perspective on growth rate. Journal of Experimental Botany, 67(1), 31-45.						
594	https://doi.org/10.1093/jxb/erv447						
595	Wild, M. (2009). Global dimming and brightening: A review. Journal of Geophysical Research:						
596	Atmospheres, 114(D10). https://doi.org/10.1029/2008JD011470						
597	Zandalinas, S. I., Sengupta, S., Fritschi, F. B., Azad, R. K., Nechushtai, R., & Mittler, R. (2021). The						
598	impact of multifactorial stress combination on plant growth and survival. New Phytologist,						
599	230(3), 1034–1048. https://doi.org/10.1111/nph.17232						

Figures

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Figure 1: (A) The balance between carbon sources and sinks may be compared to liquid being poured (photosynthesis) into a funnel (phloem transport) draining into a flask (carbon sinks). The carbon that flows into the flask is available to be consumed by metabolism and growth. When the spout is stopped (blockage of phloem transport) or the flask is smaller (limited sink capacity), carbon can build up in source tissues. Often, this buildup causes feedback inhibition of photosynthesis (dashed arrows), reducing the inflow rate. This representation is adapted from Adams et al. (2013). (B) Feedback inhibition or external constraints on light use (e.g. low temperature) reduce the electron transport rate at a given light level (red) compared to a healthy plant under favorable conditions (blue). As a result, the fraction of absorbed light used to power electron transport (Φ_{PSII}) declines and a greater fraction is in excess. (C) Absorbed light energy can be partitioned into three quantum yields— Φ_{PSII} , Φ_{NPO} , and Φ_{NO} . Φ_{NPO} is the fraction dissipated through NPQ. Φ_{NO} is the fraction that undergoes non-regulated dissipation—including through pathways that produce ROS—and generally does not decline below ~0.16 (dashed line; Kono et al. 2022). Under internal or external constraints to electron transport, Φ_{PSII} declines, but the resulting share of increase in Φ_{NPO} vs. Φ_{NO} depends on how much the plant can induce NPQ, allowing a range of scenarios (red line). When evaluating the risk of damage, it may help to consider whether Φ_{NO} is elevated rather than whether Φ_{PSII} is reduced.

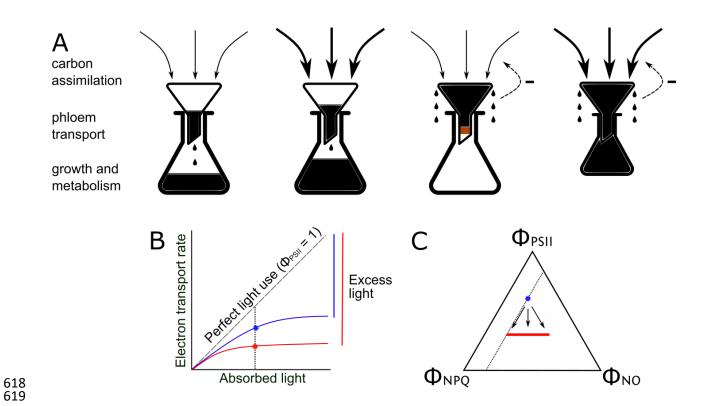


Figure 2: Two potential approaches to test the importance of photoinhibition and photoprotective adaptations for fitness, similar to the general 'likelihood' and 'vital rates' approaches described by Laughlin & Messier (2015). I consider two species (S1 and S2), depicting S1 with more yellowish leaves and steeper leaf angles to imply stronger photoprotective adaptations—high NPQ by carotenoids and reduced light absorption at midday, respectively. For simplicity, I assume these traits are not plastic. (A) One approach is to observe whether photoprotective traits are correlated consistently with environmental gradients in the risk of photoprotection. As with all observational data, causal interpretation requires care to rule out competing explanations. For example, S1 may dominate under high light because S2 is filtered out by abiotic stresses like photoinhibition, outcompeted by S1, or a combination. Nevertheless, if species found in brighter sites consistently have greater constitutive photoprotective adaptations, it would imply that those adaptations help maintain fitness under high light. (B) Another approach is to monitor vital rates across gradients of light. Here, S1 has its greatest growth rates at high light, while S2's growth is reduced at high light levels. Dashed lines separate individuals in each treatment to clarify that they are not directly interacting. This kind of experiment can be done with one or multiple species/genotype(s). If there are multiple, researchers can test whether more lightsensitive plants have fitness optima at lower light levels.

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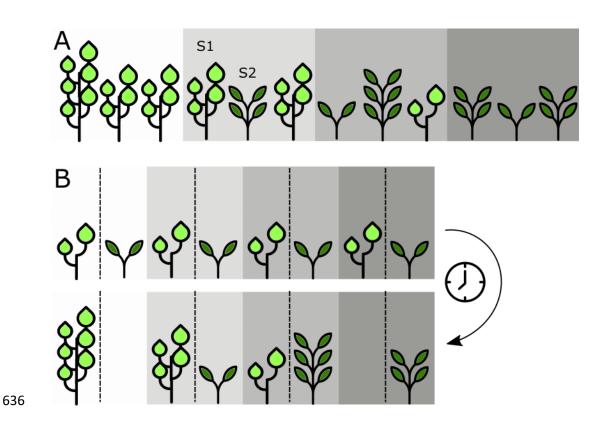
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Appendix S1

Table S1: A brief summary of studies that provide evidence that photoinhibition due to excess light can have negative effects on plant fitness. These studies all report declines in some fitness-related variable (survival, growth, or reproductive output) in conditions with high light. Most of these studies are experimental, but I include some studies that report only distributions of existing individuals if there is a strong case that absences are due to mortality rather than other factors like dispersal limitation. In general, I omit studies that only provide physiological evidence that photoinhibition is occurring without testing whether it influences fitness or resulting population/community-level consequences. I also omit studies that only compare fitness in across coarse natural gradients (e.g. understory vs. gaps) because such gradients often include changes in multiple environmental variables and cannot be used to isolate the importance of photoinhibition. However, I include a few examples where such contrasts are accompanied by physiological and/or microclimatic measurements that allow variation in fitness to be attributed more convincingly to the effects of light. This table should not be considered a formal evidence synthesis, since I only aim to summarize various studies that appear to show some positive evidence that photoinhibition can reduce fitness.

Citation	Species	Ecosystem or setting	Treatment	Physiological response	Fitness response	Notes
Agyeman et al. 2003	Seedlings of 16 west African tree species	Shadehouses in Ghana	Trees were grown in one of six shadehouses with varying degrees of shade.	None reported	In many (but not all) species, whole-plant relative growth rate was highest at irradiances below the maximum growth irradiance. Shade-tolerant species showed the greatest inhibition of growth at high irradiance.	More severe shading treatments tended to reduce maximum air temperature and slightly increased relative humidity.
Akhalkatsi et al. 2006	Betula litwinowii seedlings	Alpine treelines in the country of Georgia	The number of seedlings was surveyed across a number of plots within sites that varied in slope, aspect, elevation, and overstory cover of <i>B. litwinowii</i> or the shrub <i>Rhododendron</i> caucasium	Seedlings in sun-exposed microsites had more red anthocyanic leaves.	More sheltered microsites beneath shrub or tree cover, or on the north-facing walls of soil depressions, had greater <i>B. litwinowii</i> seedling abundance.	The sheltered sites with greater seedling abundance had lower air and soil temperatures than more exposed sites.
Bader et al.	Seedlings of	Alpine treelines	The number of sprouts	None reported	Out of the six species in the	Microclimatic monitoring
2006	six (in the	in northern	was surveyed along		observational survey, four were	revealed that the forest buffered

	observational component) or eight (in the experimental component) tree species	Ecuador	transects that crossed the treeline from the cloud forest into the páramo. In addition, some seedlings were experimentally transplanted into either the forest or the páramo, and the latter were subjected to shading by shadecloth, clearing of neighbors, or both treatments		almost completely absent beyond the treeline but the other two showed dramatic increases in abundance. In the experimental component, survival was greatest in the forest, then in the páramo under shadecloth, then in páramo controls. Clearing of neighbors also had a negative effect on survival.	temperature extremes, but the four páramo treatments were similar. However, cleared and unshaded plots did have higher soil temperature and apparently lower soil moisture. The authors note several additional observations that suggest a role for photoinhibition. For example, the species that survive best in the páramo appear to have photoprotective adaptations like reflective leaves or specialized pigments. Also, in many species the light-exposed parts of the leaves were the only ones to show apparent damage.
Ball et al. 1991	Eucalyptus pauciflora seedlings	Alpine treelines in southeastern Australia	Plants were naturally growing on slopes in aspects either more protected or more exposed to sunlight during the winter; for physiological measurements, some trees were also shaded artificially	Trees on exposed sites had lower rates of photosynthesis under light-limited conditions, but not under high light.	Juvenile trees tended to be distributed close to the canopy margin and along an arc to the better-protected south of the canopy. Juvenile trees in exposed areas were often stunted.	Trees on more exposed sites did also tend to have slightly lower nighttime leaf temperature.
Ballestreri et al. 2021	Four subdominant or understory tree species	Restoration experiment in the Atlantic Forest of southern Brazil	Trees were grown in the varying degrees of shade provided by one of four overstory tree species	Deeper shading caused increases in chlorophyll content and dark-acclimated F_{ν}/F_{m} , and decreases in stomatal density. All species appeared to show greater photosynthetic capacity under deeper shade.	One species (Eugenia involucrata) had a major increase in mortality under the greatest light level. Many species showed greatest growth in height or stem diameter under partial shade.	Because the treatment here involves interactions with neighbors, the outcome may result not just from shading but also from other sorts of mechanisms of interactions among neighboring plants.
Baumeister & Callaway 2006	Pseudotsuga menziesii and Ribes cereum seedlings	Forest- grassland ecotones in Montana, United States	The number of natural recruits was surveyed in relation to wind exposure whether they were growing under an adult <i>Pinus flexilis</i> crown. In one experiment, seedlings were planted under <i>P</i> .	None reported	Natural seedlings were disproportionately found growing under <i>P. flexilis</i> crowns. In the first experiment, survival was increased by shade and by protection from wind. In the second experiment, shade had a strong primary influence on survival and seedling height,	Plant-available N and P were not significantly different between P. flexilis crowns and the open. Litter depth was greater under the crowns. Soil moisture was lower at 10 cm under P. flexilis and in shade treatments, and equivalent at 20 cm. Soil moisture was also

			flexilis or in the open, and at windward or leeward sites. In a separate experiment, seedlings were grown in the open under one of six treatments that include shade, increased snow, and wind protection		and additional benefits of snow accumulation and wind protection for survival only appeared in the shade. In both experiments, <i>Ribes</i> mortality in the open was highest during the late summer, but <i>Pseudotsuga</i> mortality was highest over the winter.	greater in spring and early summer in the snowdrift plots. Based on the seasonal pattern of mortality, the authors suggest that the facilitative effect of shade on <i>Ribes</i> (but not <i>Pseudotsuga</i>) had more to do with moisture than photoinhibition.
Close et al. 2002	Eucalyptus nitens and Eucalyptus globulus seedlings	Alpine treelines in Tasmania, Australia	Plants were either left exposed or surrounded with open-top shadecloth shelters for the first fourteen weeks after planting	Unshaded <i>E. globulus</i> developed high leaf anthocyanin content, as well as severe leaf abscission and severe but transient declines in dark-acclimated F _v /F _m . Unshaded <i>E. nitens</i> had the same responses, but much more moderate. Shaded trees showed slight increases in anthocyanins and declines in dark-acclimated Fv/Fm after the shadecloth was removed.	Unshaded <i>E. globulus</i> suffered severe leaf abscission and had high mortality and reduced height growth compared to shaded <i>E. globulus</i> . However, unshaded <i>E. nitens</i> had much more modest levels of abscission and greater height growth than shaded <i>E. nitens</i> .	The researchers measured air temperature and light quality and found that the shelters had minimal effects on both.
del Valle et al. 2020	Silene littorea	Greenhouse and experimental garden in Mediterranean southern Spain	Plants were grown either under a methacrylate filter that transmitted most UV or a polycarbonate filter that excluded most UV	Plants under UV exclusion produced less anthocyanins and phenolics. They also had greater midday F_{ν}/F_{m} , but the similar pre-dawn F_{ν}/F_{m} .	Plants under UV exclusion had the same ovule and seed production per flower, but considerably higher pollen production and seed set.	
Egerton et al. 2000	Eucalyptus pauciflora seedlings	Treeless pasture in southeastern Australia	Plants were grown on either the exposed or sheltered sides of vertical screens that intercepted half of incoming light	Trees on the exposed side of the screens had lower dark-acclimated and midday $F_{\rm v}/F_{\rm m}$ and lowered photosynthetic light-response curves. These trees also lost much of their leaf area to abscission during the winter.	Exposed trees had lower shoot and total biomass after the winter.	The researchers measured air temperature and soil moisture. They found that the shelters had no significant influence on air temperature, but caused a small increase of soil moisture on the exposed side.
Gatti et al. 2014	Saplings of the palm Euterpe edulis	Shadehouse and various sites in subtropical northern Argentina	Plants were grown in either high or low light in shadehouses or in natural canopy understories/gaps	Seedlings grown under higher irradiance had a smaller relative decline in F_{ν}/F_{m} and faster recovery. High growth irradiance did not affect stem water potential but did cause	In the shadehouse, relatively high irradiance treatments are associated with somewhat lower growth and irradiance. In natural gaps, survival was much lower but growth was much	In this study, the subtle but perhaps important influence of the treatments on water relations makes it hard to uniquely pinpoint an effect of photoinhibition, since high

				substantial decreases in leaf water potential and increases in plant Huber value.	higher than in understories.	mortality at high irradiance could have been exacerbated by water deficits.
Germino & Smith 1999	Picea engelmannii and Abies lasiocarpa seedlings	Alpine treelines in Wyoming, United States	The position of plants was surveyed along four distinct transects. For physiological measurements, some potted plants were also experimentally subjected to nighttime warming (transparent plastic sheets), daytime shading, or both	Potted A. lasiocarpa seedlings subjected to daytime shading had higher saturated photosynthetic rates than unshaded seedlings, both comparing shading to control and shading + warming to just warming. There was no major effect in P. engelmannii. Among natural seedlings, those in exposed environments had more inclined and clustered needles.	Both species were almost entirely found in the one transect with high tree cover. <i>A. lasiocarpa</i> in particular was largely missing from highly open microsites.	The authors measured light spectral quality and found that it was unaffected by the treatments.
Germino et al. 2002	Picea engelmannii and Abies lasiocarpa seedlings	Alpine treelines in Wyoming, United States	Naturally occurring seedlings were surveyed repeatedly for survival as a function of factors that influence their exposure to stress. For some naturally occuring seedlings, neighboring grasses were removed. Some seedlings were also artifically sown among several sites with varying exposure to high light and other potential stress factors.	None reported	P. engelmannii seedlings (but not saplings or emergents) had greater survival close to the cover of the tree island. P. engelmannii emergents had greater mortality on more exposed south and east-facing slopes. In general, survival was greater when there were features upwind or overhead, or when trees were surrounded by moss or overtopped by grass.	Neighbor removal alleviated water and low temperature stress but still increased mortality, which the authors attribute to photoinhibition due to increased light. Neighbor removal also caused greater wind stress.
Gómez- Aparicio et al. 2006	Two evergreen and two deciduous late- successional tree species	Montane botanical garden in Mediterranean southeastern Spain	Plants were growing in pots either under full sunlight or under mild or severe shading treatments imposed by shadecloth.	Under shade, all species increased chlorophyll content and decreased carotenoid content. Shade also caused increases in F_{ν}/F_{m} throughout the day, including before dawn. The two deciduous species had lowered photosynthetic lightresponse curves under high light, while the two evergreen species showed the opposite	One deciduous species (<i>Acer opalus</i>) experienced a very high rate of mortality at full sunlight, which was reduced by two-thirds under even a moderate shading treatment. No other species experienced negative survival or growth (total dry mass) consequences from high light.	Soil moisture was held constant across treatments. The moderate shade treatment had no major effect on air temperature, but the more intense shade treatment tended to reduce temperature substantially.

Holly et al. 1994	Eucalyptus polyanthemos seedlings	Treeless pasture in southeastern Australia	Plants were either grown in the open or in a series of shelters that excluded various fractions of incident light	pattern. The authors report many other changes in plant allocation and physiology that are less relevant here. Trees had the highest pre-dawn F _v /F _m under 50% or 30% of natural light, both in midwinter and in spring.	Trees showed the greatest growth in height under 50% or 30% of natural light.	The researchers measured humidity, air temperature, and wind speed. While they found that while most shelter designs had minimal effects on humidity and temperature, shelters reduced wind speed in proportion to their shading intensity.
Howard et al. 2020	Arabidopsis thaliana—an NPQ-deficient mutant, a chloroplast movement-impaired mutant, three mutants deficient in chloroplast movement as well as other aspects of physiological function, and a wild-type	Growth chambers and an experimental garden in Massachusetts, United States	Plants were grown either outdoors or in a growth chamber with light fluctuating on an hour-long cycle	Under natural conditions, the chloroplast movement-impaired mutant had higher dark-acclimated F_{ν}/F_{m} than the wild-type.	Under natural conditions, the chloroplast movement-impaired movement had smaller rosettes, smaller and fewer leaves, fewer siliques, fewer secondary inflorescences, and earlier bolting than the wild-type. The NPQ-deficient mutant was largely unaffected.	monsty.
Kothari et al. 2021	Eight temperate, winter- deciduous broadleaf tree species in the first eight years of growth	Densely planted tree diversity experiment in Minnesota, United States	Plants were grown either in monocultures, where they were generally unshaded, or in bicultures with faster-growing conifers that provided substantial shade	Plants grown in bicultures had substantially higher pre-dawn F_v/F_m and lower production of photoprotective pigments. The most shade-tolerant species had the greatest increase in pre-dawn F_v/F_m in biculture. At least in one species (<i>Tilia americana</i>), they also showed high photosynthetic rates under high light levels and delayed leaf senescence.	Two species (<i>T. americana</i> and <i>Acer negundo</i>) grew faster with larger neighbors; most of the others grew slower.	Because the treatment here involves interactions with neighbors, the outcome may result not just from shading but also from other sorts of mechanisms of interactions among neighboring plants. Trees under the shade of larger neighbors generally had more negative pre-dawn leaf water potential, but less negative midday water potential.

Külheim et al. 2002	Arabidopsis thaliana— two NPQ- deficient mutants and a wild-type	Growth chambers and an experimental garden in northern Sweden	Plants were grown in growth chambers with constant and fluctuating light conditions, as well as in the field.	The two mutants had lower midday F_{ν}/F_m under field conditions, especially under high light.	In the field and under fluctuating light, the two mutants had fewer seeds and fruits per plant. Under constant light, there was no effect.	
Murray et al. 1993	Multiple species of Sphagnum mosses	Moist mountain slopes in northern Alaska	At one site, all shade-producing vascular plants were removed in several plots, which were compared to control plots. At another site without high vascular plant cover, shadecloth shelters were set up and compared to control plots. In addition, physiological measurements were taken in the laboratory from various Arctic and temperate mosses.	Mosses under high light had lower dark-acclimated F_{ν}/F_m and lower photosynthetic capacity than those under lower light.	At both sites with shade experiments, mosses under shade (from shadecloth or vascular plants) had much greater growth in length.	Shaded treatments tended to have somewhat lower moss surface temperature.
Ronco 1970	Picea engelmannii seedlings	Nursery in the Central Rocky Mountains	Trees were grown in one of three shade treatments	Unshaded seedlings had much lower photosynthetic light-response curves than fully shaded seedlings. Unshaded and partially shaded seedlings had no major differences in their water deficit.	The paper relies on a 1961 research note by the same author, which is not widely available, to support the claim that light influences seedling survival. It reports that trees growing under full sun suffered from chlorosis and had greater mortality than those under partial shade. It also reports that drought, frost heaving, and herbivory are not major causes of mortality.	
Semchenko et al. 2012	46 herbaceous species from temperate grasslands	Common garden in Estonia	Plants were grown either in full sunlight or in three shading treatments of varying intensity	None reported	Moderate shade increased total plant biomass relative to full sunlight	The shading treatments moderately buffered air temperature and increased relative humidity, particularly the more intense ones. The authors attribute the positive effect of shade on growth to dynamic regulation of growth

Slot et al. 2005	Pinus sylvestris juveniles	Glades with low tree density within forests in western Siberia	Plants were naturally growing on slopes in aspects either more protected or more exposed to sunlight during the winter. Some seedlings were also either left exposed or artifically shaded following removal of adult trees.	In the smaller size class across the growing season, more naturally exposed trees had lower pre-dawn and midday F_v/F_m and lower photosynthetic rates at equivalent light availability. These exposed small trees had greater non-photochemical quenching, xanthophyll pigment to chlorophyll ratios, and xanthophyll de-epoxidation state. Similar results held when comparing artificially exposed and shaded trees. The researchers found that more exposed and photoinhibited trees were not depleted in	Seedlings recruited mainly within the crown projection area of adult trees, usually on the protected northern side.	rather than amelioration of abiotic stress. However, it is unclear how this is possible, considering that plants cannot build tissue with material they have not acquired. Air temperature differed very little between the northern (protected) and southern (exposed) sides of adult trees, especially in autumn and winter. Southern trees had slightly less negative δ^{13} C, which led the researchers to suggest tentatively that they may be more prone to desiccation. However, soil moisture content did not differ between northern and southern aspects during the summer.
Velasco & Becerra 2020	Seedlings of nine woody species	Pots in an open grassland in central Chile	Trees were grown together with one of eight different herb species (or control), under either shadecloth or control, and in either a wetter or drier simulated precipitation regime	NSCs. None reported	The shade treatment had an overwhelmingly large positive impact on the recruitment (survival) of tree seedlings. Water and herb cover had much smaller effects.	

647 648	Literature Cited						
649	Agyeman, V. K., Swaine, M. D., & Thompson, J. (1999). Responses of tropical forest tree seedlings to						
650	irradiance and the derivation of a light response index. <i>Journal of Ecology</i> , 87(5), 815–827.						
651	https://doi.org/10.1046/j.1365-2745.1999.00400.x						
652	Akhalkatsi, M., Abdaladze, O., Nakhutsrishvili, G., & Smith, W. K. (2006). Facilitation of Seedling						
653	Microsites by Rhododendron Caucasicum Extends the Betula Litwinowii Alpine Treeline,						
654	Caucasus Mountains, Republic of Georgia. Arctic, Antarctic, and Alpine Research, 38(4), 481–						
655	488. https://doi.org/10.1657/1523-0430(2006)38[481:FOSMBR]2.0.CO;2						
656	Bader, M. Y., van Geloof, I., & Rietkerk, M. (2007). High solar radiation hinders tree regeneration above						
657	the alpine treeline in northern Ecuador. <i>Plant Ecology</i> , 191(1), 33–45.						
658	https://doi.org/10.1007/s11258-006-9212-6						
659	Ball, M. C., Hodges, V. S., & Laughlin, G. P. (1991). Cold-Induced Photoinhibition Limits Regeneration						
660	of Snow Gum at Tree-Line. Functional Ecology, 5(5), 663–668. https://doi.org/10.2307/2389486						
661	Ballestreri, A. A., Araujo, M. M., Aimi, S. C., Nascimento, N. F. do, Berghetti, Á. L. P., Gasparin, E.,						
662	Tabaldi, L. A., & Zavistanovicz, T. C. (2021). Morphophysiological responses of forest tree						
663	species conducted under different levels of shading in the enrichment of degraded ecosystem.						
664	Forest Ecology and Management, 488, 119032. https://doi.org/10.1016/j.foreco.2021.119032						
665	Baumeister, D., & Callaway, R. M. (2006). Facilitation by Pinus Flexilis During Succession: A Hierarchy						
666	of Mechanisms Benefits Other Plant Species. <i>Ecology</i> , 87(7), 1816–1830.						
667	https://doi.org/10.1890/0012-9658(2006)87[1816:FBPFDS]2.0.CO;2						
668	Close, D. C., Beadle, C. L., Holz, G. K., & Brown, P. H. (2002). Effect of shadecloth tree shelters on						
669	cold-induced photoinhibition, foliar anthocyanin and growth of Eucalyptus globulus and E. nitens						
670	seedlings during establishment. Australian Journal of Botany, 50(1), 15–20.						
671	https://doi.org/10.1071/bt01038						
672	Egerton, J. J. G., Banks, J. C. G., Gibson, A., Cunningham, R. B., & Ball, M. C. (2000). Facilitation of						
673	Seedling Establishment: Reduction in Irradiance Enhances Winter Growth of Eucalyptus						

674	Pauciflora. Ecology, 81(5), 1437–1449. https://doi.org/10.1890/0012-
675	9658(2000)081[1437:FOSERI]2.0.CO;2
676	Gatti, M. G., Campanello, P. I., Villagra, M., Montti, L., & Goldstein, G. (2014). Hydraulic architecture
677	and photoinhibition influence spatial distribution of the arborescent palm Euterpe edulis in
678	subtropical forests. <i>Tree Physiology</i> , 34(6), 630–639. https://doi.org/10.1093/treephys/tpu039
679	Germino, M. J., & Smith, W. K. (1999). Sky exposure, crown architecture, and low-temperature
680	photoinhibition in conifer seedlings at alpine treeline. Plant, Cell & Environment, 22(4), 407-
681	415. https://doi.org/10.1046/j.1365-3040.1999.00426.x
682	Germino, M. J., Smith, W. K., & Resor, A. C. (2002). Conifer seedling distribution and survival in an
683	alpine-treeline ecotone. Plant Ecology, 162(2), 157–168.
684	https://doi.org/10.1023/A:1020385320738
685	Gómez-Aparicio, L., Valladares, F., & Zamora, R. (2006). Differential light responses of Mediterranean
686	tree saplings: Linking ecophysiology with regeneration niche in four co-occurring species. Tree
687	Physiology, 26(7), 947–958. https://doi.org/10.1093/treephys/26.7.947
688	Holly, C., Laughlin, G. P., & Ball, M. C. (1994). Cold-Induced Photoinhibition and Design of Shelters for
689	Establishment of Eucalypts in Pasture. Australian Journal of Botany, 42(2), 139–147.
690	https://doi.org/10.1071/bt9940139
691	Howard, M. M., Bae, A., Pirani, Z., Van, N., & Königer, M. (2020). Impairment of chloroplast movement
692	reduces growth and delays reproduction of Arabidopsis thaliana in natural and controlled
693	conditions. American Journal of Botany, 107(9), 1309–1318. https://doi.org/10.1002/ajb2.1537
694	Kothari, S., Montgomery, R. A., & Cavender-Bares, J. (2021). Physiological responses to light explain
695	competition and facilitation in a tree diversity experiment. Journal of Ecology, 109(5), 2000-
696	2018. https://doi.org/10.1111/1365-2745.13637
697	Külheim, C., Ågren, J., & Jansson, S. (2002). Rapid Regulation of Light Harvesting and Plant Fitness in
698	the Field. Science, 297(5578), 91–93. https://doi.org/10.1126/science.1072359

699	Murray, K. J., Tennunen, J. D., & Nowak, R. S. (1993). Photoinnibition as a control on photosynthesis
700	and production of Sphagnum mosses. Oecologia, 96(2), 200–207.
701	https://doi.org/10.1007/BF00317733
702	Ronco, F. (1970). Influence of High Light Intensity on Survival of Planted Engelmann Spruce. Forest
703	Science, 16(3), 331–339. https://doi.org/10.1093/forestscience/16.3.331
704	Semchenko, M., Lepik, M., Götzenberger, L., & Zobel, K. (2012). Positive effect of shade on plant
705	growth: Amelioration of stress or active regulation of growth rate? Journal of Ecology, 100(2),
706	459–466. https://doi.org/10.1111/j.1365-2745.2011.01936.x
707	Slot, M., Wirth, C., Schumacher, J., Mohren, G. M. J., Shibistova, O., Lloyd, J., & Ensminger, I. (2005).
708	Regeneration patterns in boreal Scots pine glades linked to cold-induced photoinhibition. Tree
709	Physiology, 25(9), 1139–1150. https://doi.org/10.1093/treephys/25.9.1139
710	Valle, J. C. D., Buide, M. L., Whittall, J. B., Valladares, F., & Narbona, E. (2020). UV radiation increase
711	phenolic compound protection but decreases reproduction in Silene littorea. PLOS ONE, 15(6),
712	e0231611. https://doi.org/10.1371/journal.pone.0231611
713	Velasco, N., & Becerra, P. I. (2020). Species-specific effects of the herbaceous layer on recruitment of
714	woody species under different shading and precipitation conditions. Forest Ecology and
715	Management, 460, 117864. https://doi.org/10.1016/j.foreco.2020.117864