

1 **Discovering the rules of plant biogeography using a trait-based approach**

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10 11 **Abstract**

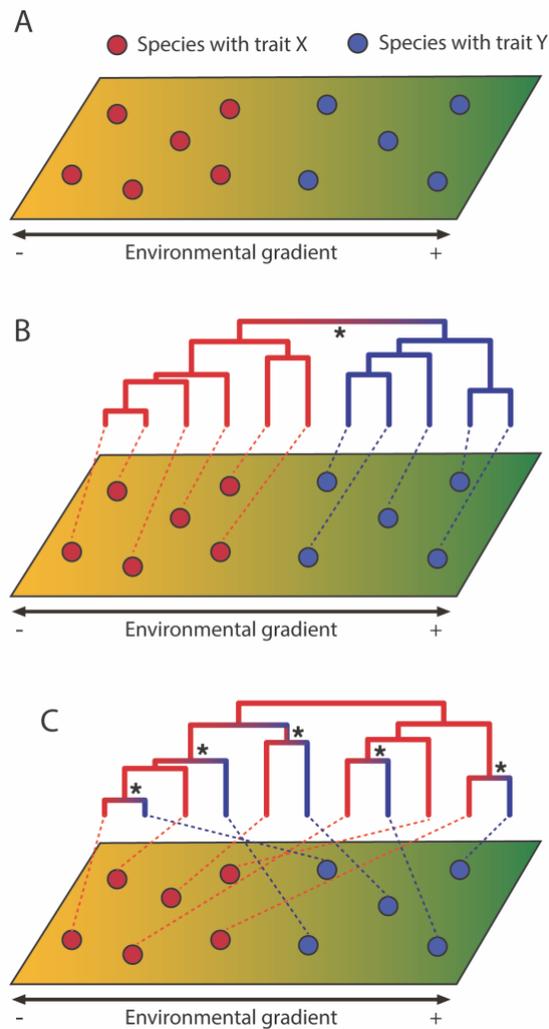
12 Lineage-specific traits determine how plants interact with their surrounding environment. As
13 different species may find similar phenotypic solutions through evolution to tolerate, persist in
14 and invade environments with certain characteristics, some traits may become more common in
15 certain types of habitats. These general patterns of geographical trait distribution point towards
16 the existence of some rules in how plants diversify in space over time. Trait-environment
17 correlation analyses are ways to discover general rules in plant biogeography by quantifying to
18 what extent unrelated lineages have similar evolutionary responses to a given type of habitat. In
19 this synthesis, I give a short historical overview on trait-environment correlation analyses, from
20 some key observations from classic naturalists to modern approaches using trait evolution
21 models, large phylogenies, and massive datasets of traits and distributions. I discuss some
22 limitations of modern approaches, including the need for more realistic models, the lack of data
23 from tropical areas, and the necessary focus on trait scoring that goes beyond macro-
24 morphology. Overcoming these limitations will allow the field to explore new questions related
25 to trait lability and niche evolution and to better set apart rules and exceptions in how plants
26 diversify in space over time.

27 **Key words:** environmental variables, phylogenetic comparative methods, trait evolution

28 29 **Introduction: lineage specific trait as determinants of plant distribution**

30 Lineage-specific traits control how plants interact with their physical environment and
31 can modulate their geographical distribution in at least three different ways. First, they can allow
32 plant lineages to survive in an environment that, due to physiological constraints, they would not
33 be able to tolerate otherwise (Good, 1931). For example, species native to closed canopy biomes
34 tend to have larger leaf areas (Givnish, 1988) and larger seeds to store nutrients for germination
35 (Foster & Janson, 1985) due to photosynthesis limitations in shady habitats. Second, lineage-

36 specific traits can facilitate lineages to persist and reproduce in certain environments by allowing
 37 them to better compete for resources or survive against natural enemies. For instance, spines tend
 38 to be more common in open habitats where grassing herbivores are numerous (Charles-
 39 Dominique et al., 2016) and higher diversity of chemical compounds is observed in tropical
 40 rainforests where plants are under constant pressure from pathogens (Kursar et al., 2009). Third,
 41 lineage-specific traits can increase chances of lineages to invade new environments by
 42 facilitating events of long-distance dispersal and the establishment of new populations from just
 43 a few individuals. Such traits include, for instance, capacity for self-fertilization (Pannel et al.,
 44 2015) and certain types of seed dispersal strategies that increase lineage vagility over
 45 evolutionary time (Onstein et al., 2019; Vasconcelos et al., 2021).



46
 47 **Figure 1.** (A) Schematic representation of the geographical distribution of eleven species across
 48 an environmental gradient (- to +) and which present two different traits (X and Y). Each trait is
 49 more common in one of the extremes of the environmental gradient. Over evolutionary time, this
 50 pattern can be formed through: (B) common descent of sympatric species; or (C)
 51 convergent evolutionary responses of unrelated lineages. The second scenario (C) provides

52 significantly more robust evidence for trait-environment correlation in an explicit evolutionary
53 framework due to the larger number of empirical replicates (marked with asterisk).

54

55 The crucial role that traits play in modulating plant distribution may lead some traits to
56 become more common in certain types of habitats, as many different species find similar
57 phenotypic solutions through their evolution to tolerate, persist in and invade environments with
58 certain characteristics (Figure 1A). Similar phenotypic solutions may arise from common
59 descent, when closely related species inherit a similar suite of beneficial traits to exist in one
60 given type of habitat (Figure 1B). For instance, most species of montane lupines (*Lupinus*,
61 Fabaceae) are perennial and belong to a clade that invaded the Andes and diversified *in situ*,
62 meaning that there are few events of elevation transition linked to the evolution of perennial life
63 history strategy in that group (Drummond et al., 2012). However, similar phenotypic solutions
64 can also arise in distantly related species through parallelisms and convergences (Figure 1C; e.g.
65 Donoghue et al., 2022). For instance, the way in which we recognize biomes often refer to traits
66 that are commonly found across several unrelated lineages that occur under a similar temperature
67 and precipitation. The biome classification of “broadleaf rainforests”, for example, refer to a
68 plant trait (“broadleaves”) that is common in warmer forests that receive a lot of rain throughout
69 the year, and “shrublands” indicate drier or cooler areas that are dominated by a particular life
70 form (“shrubs”). In this case, independent events of environment transitions may appear
71 correlated with the acquisition of a similar phenotypic characteristic through the evolution of
72 independent lineages, even if that biome has a disjunct distribution around the world (e.g.
73 savannas and rainforests; Pennington and Hughes, 2014; Eiserhardt et al., 2017).

74 Distantly related lineages having similar evolutionary responses to the same component
75 of the physical environment (such as climate, soil, and topography) point towards the existence
76 of some general rules in how plants diversify in space over time. With modern tools, it is
77 possible to quantify to what extent unrelated lineages have similar evolutionary responses to a
78 given physical environment, and thus how general these general rules are, through analyses of
79 trait-environment correlations. In this synthesis, I give a brief overview of how trait-environment
80 correlations have been historically noted and quantified and discuss some avenues for future
81 research based on current limitations of modern approaches.

82

83 **A brief historical overview on trait-based approaches to plant biogeography**

84 The history of observation that plant trait distribution correlate with aspects of the
85 physical environment goes back to before the proposition of the evolutionary theory. Von
86 Humboldt and Bonpland (1807) provided arguably the most popular account of his time on how
87 vegetative traits respond to environmental temperature, by observing that alpine plants in tropical
88 mountains have similar vegetative characteristics as those found in the temperate zones of
89 Europe. These observations were later mentioned by Darwin (1859, Origin of Species, Chapter

90 XI) to support his theory of natural selection, as evidence that unrelated plant lineages (e.g.
91 species of different genera and families) may acquire similar forms when faced with similar
92 environmental challenges, no matter where they are on the globe. In the first half of the 20th
93 century, key further contributions to these observations came from the works of Arber (1920),
94 who described similar anatomical patterns in roots and stems of aquatic plants for different
95 families of flowering plants; and Raunkiaer (1934) who proposed a categorization for life forms
96 in plants in relation to how growing buds are protected during unfavorable seasons. In the second
97 half of the 20th century, Stebbins (1950, 1974) contributions were remarkable for linking the
98 physical environment not only to the convergent evolution of vegetative traits, but also
99 reproductive traits such as flower and fruits. He suggested that the physical environment may
100 often cause an indirect effect on the spatial distribution of plant traits that depend on animal
101 behavior, because the distribution of animals themselves (e.g. pollinators and dispersers) are also
102 impacted by characteristics of the physical environment. It is important to note that most of the
103 trait-environment correlation hypotheses developed during this period were based mainly on
104 morphological descriptions, natural history observations, or simple statistics and did not account
105 for common ancestry among species. Though most patterns were described in the context of the
106 evolutionary theory, at that time there were no tools available to quantify how specific or general
107 these patterns were across many lineages and in an explicit evolutionary framework – that is,
108 when considering common ancestry among lineages in the analyses (Felsenstein, 1985).

109 The end of the 20th century brought the popularization of computers, the global
110 positioning system (“GPS”) and molecular sequencing techniques. These new technologies
111 allowed quantifying trait-environment correlations using more accurate accounts of species
112 geographical distributions (see also Colli-Silva et al., 2020) and to consider the impact of shared
113 descentance on trait evolution through molecular-based phylogenies (Felsenstein, 1988; Webb
114 and Donoghue, 2005; Webb et al., 2008) and phylogenetic comparative methods (Felsenstein,
115 1985; Donoghue, 1989). Some noteworthy findings of these early stages of quantitative trait-
116 environment correlations using comparative methods include a latitudinal gradient of seed size
117 due to a correlation with vegetation type (Moles et al., 2007), the correlation between seed and
118 seedling morphology and degree of canopy opening (Zanne et al., 2005), and the evolution of
119 defense traits in relation to soil variables (Fine et al., 2004). Studies of this period have typically
120 used simple trait evolution models (Pagel 1994) or regressions of phylogenetic independent
121 contrasts (Felsenstein, 1985) to measure trait-environment correlations.

122

123 **Modern approaches to trait-environment correlations**

124 The use of more realistic approaches for measuring trait-environment correlations were
125 generally hampered by the lack of global datasets and new models that only became widely
126 available, or popularized, in the last 15 years. Existing databases of molecular sequences (e.g.
127 NCBI, 2022) and occurrence points (e.g. GBIF, 2022), and newly available environmental layers
128 (Karger et al., 2017; Brown et al., 2018), as well as collaborative initiatives to score massive trait

129 datasets (e.g. Wright et al., 2004; Kattge et al., 2011; Díaz et al., 2016), have allowed many trait-
130 environment correlation hypotheses to be tested at a global scale and in an explicitly
131 evolutionary framework (e.g. Moles 2018; Bruehlheide et al., 2018; Sinnott-Armstrong et al.,
132 2021). The use of broader datasets in terms of both taxonomic and geographic scope is important
133 because defining rules requires generalizations that work for as many lineages as possible, and
134 quantifying that is only possible when data from many plant groups are combined in a single
135 analytical framework. Inferences can be dubious when derived from single events (Maddison
136 and FitzJohn, 2015), so to understand which patterns are rules and which are exceptions multiple
137 natural replicates of the same type of event are needed (Figure 1C) (e.g. replicated radiations;
138 Donoghue et al., 2022). The possibility of reconstructing large phylogenies (e.g. Zanne et al.,
139 2014, 2018; Beaulieu and O'Meara, 2018), or using multiple phylogenies that present the pattern
140 of interest (e.g. Simon et al., 2009, Vasconcelos et al., 2020; 2021), means that an adequate
141 number of independent evolutionary transitions of the same type (e.g. multiple habitat shifts
142 and/or multiple transitions between trait states) can be achieved more easily.

143 With the need for increasing the number of independent replicates of a certain trait-
144 environment association also comes the need for more realistic trait evolution models that can
145 incorporate the heterogeneity of evolutionary processes across the tree of life. For example, new
146 extensions of hidden-Markov models allow transition rates between states of a discrete trait to
147 vary across a phylogeny (Boyko and Beaulieu 2021, 2022), which would be expected if the
148 dynamics of trait evolution differ among clades, a compelling assumption especially in larger
149 phylogenies (Beaulieu et al., 2013). Phylogenetic regression methods that allow the error term to
150 be modeled according to different assumptions of how continuous traits evolve (Ho and Ané,
151 2014) and shift detection methods that allow the parameters of continuous trait evolution models
152 to vary across the phylogeny (Khabbazian et al., 2016; Uyeda and Harmon, 2014) also allowed
153 for more biologically realistic pictures of continuous trait evolution. Similarly, models that allow
154 for the joint evolution of discrete and continuous traits (Tribble et al., 2021; Boyko et al., 2022)
155 allow for traits and environmental variables to influence one another throughout evolution in
156 cases where they are correlated.

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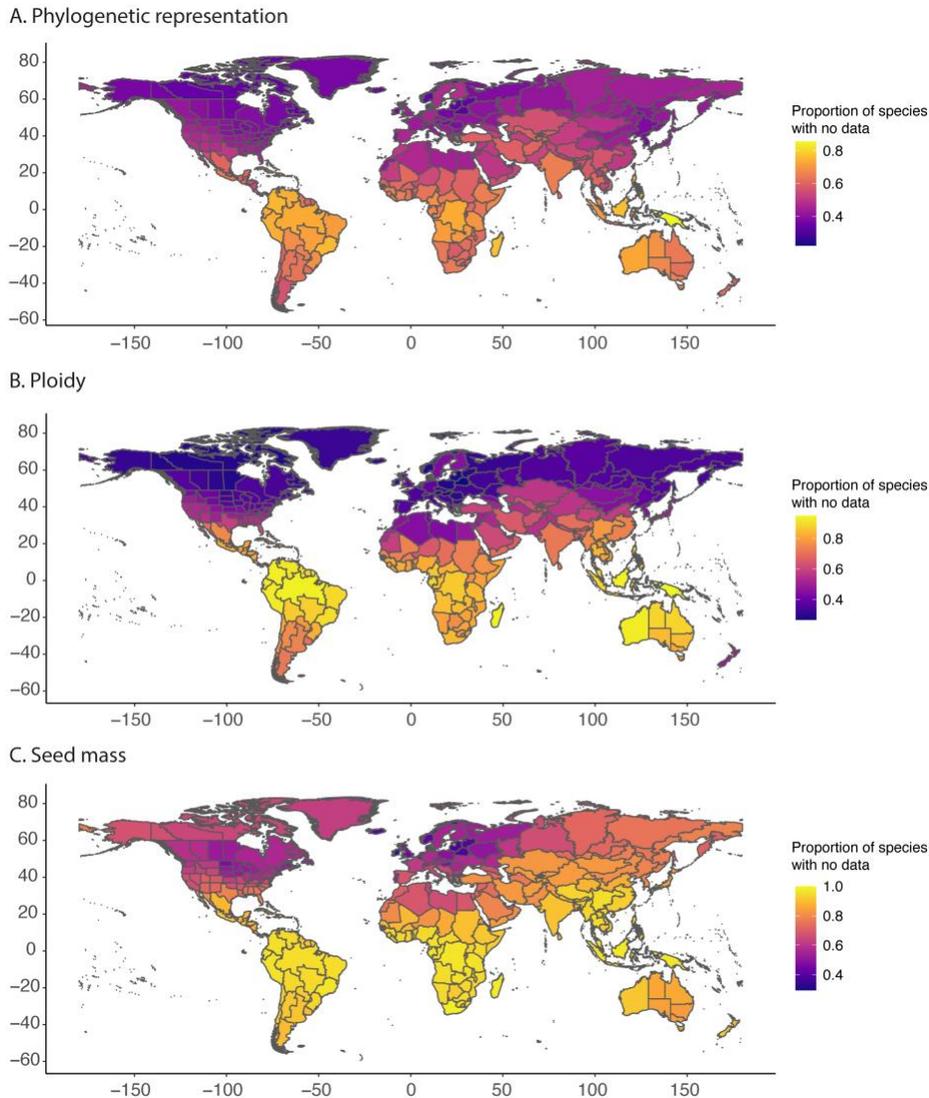
158 **Limitations of current approaches**

159 There are exciting ways in which trait-environment correlations can be used to
160 understand plant biodiversity with the development of more realistic models and increasing
161 availability of trait, distribution, and phylogenetic datasets. However, there are also many
162 limitations in current approaches that must be tackled by future studies aiming at quantifying
163 trait-environment correlations as the field moves forward. I highlight three of these limitations
164 below.

165 First, we need to keep working on the development of more realistic models. For
166 instance, models that simultaneously account for the differential dynamics of speciation and
167 extinction rates as well as trait and environment evolution would be more realistic, as all these

168 processes affect how traits become common in one type of environment (Vasconcelos et al.
169 2022). A common problem in model development is that more realistic models tend also to be
170 more complex (i.e., with more parameters to be estimated) and then face identifiability problems
171 (Louca and Pennell, 2020), power issues (Davis et al., 2013), and computational limitations
172 (Maliet and Morlon, 2022). Many of the current available approaches also frequently require
173 data transformations that may not be realistic. For example, the discretization of traits and
174 environmental variables that are clearly continuous in nature (e.g., elevation) and the lack of
175 appropriate ways to simplify multivariate traits in an explicit phylogenetic framework (Uyeda et
176 al., 2015). Solving these limitations will increase our flexibility in asking different types of
177 questions due to the use of more realistic datasets and assumptions related to how correlated
178 evolution works.

179 Second, we must fill the gaps in datasets of phylogenies, traits, and distributions from
180 poorly known groups, especially in tropical areas. Though large online databases of molecular
181 data, geographical distribution, and traits are exceptional resources for global analyses, they tend
182 to cover a higher proportion of the diversity of temperate regions (Figure 2; see also Collen et al.,
183 2008; Cornwell et al., 2019). The fact that data richness (Collen et al., 2008; Cornwell et al.,
184 2019) and taxonomic accuracy (Freeman and Pennell, 2021) is still skewed towards the poles
185 may lead to biased interpretations of results from large scale analyses. This means that to
186 understand, for example, how plant lineages have moved in space over time or how they adapted
187 to specific habitats, even if we try to include all the available data, results may be weighted for
188 patterns observed in temperate habitats. Global generalities in trait-environment correlations will
189 only be properly documented when data from the whole globe is included in the analyses
190 (Cornwell et al., 2019). Until sampling in the tropics matches the sampling in the temperate
191 regions, efforts on understanding the role of traits on the spatial diversification of plants at a
192 global scale will remain preliminary at best. Future studies that seek to understand these general
193 patterns should seek not only theoretical and methodological advancement, but also in filling this
194 fundamental lack of data from tropical regions.



195

196 **Figure 2.** The latitudinal bias in large datasets of phylogenetic representation and traits. (A)
 197 Molecular data for phylogenetic reconstruction and (B,C) two traits commonly used in trait-
 198 environment correlation analyses: (B) ploidy and (C) seed mass. Distribution data comes from
 199 POWO (2022). Other data comes from, respectively, (A) Smith and Brown (2018), (B) Rice et
 200 al., (2019), and (C) Maitner et al., (2018). Code and details for plotting maps are available in
 201 github.com/tncvasconcelos/synthesis.

202

203 Third, we should focus on improving available datasets of phylogenies, traits, and
 204 distributions not only in quantity, but also in quality. Much of the large-scale analyses on trait-
 205 environment correlations has so far been focused on macro-morphological traits that can be
 206 readily scored from herbarium collections, taxonomic descriptions, or that are commonly
 207 measured from plots of forestry surveys. Though form and function are linked, and macro-
 208 morphology can tell us a lot about plant adaptations to their environment (e.g. Donoghue et al.,

209 2022), traits related to seed germination (e.g. Tudela-Isanta et al., 2018), cytotype (e.g. Rice et
210 al., 2020), below ground organs (e.g. Laliberté 2017; Carmona et al., 2021), defensive chemicals
211 (e.g. Hahn et al., 2019), phenology (e.g. Staggemeier et al., 2010; Fernández-Martínez et al.,
212 2019), and anatomy (e.g. Alcantara et al., 2018) are also crucial to understand spatial
213 diversification of plants, and comprehensive datasets for these are still scarce. To understand the
214 role of the physical environment on the evolution of traits that are primarily associated with
215 animal interactions, such as pollination or dispersal strategies, we must also have a better
216 understanding of pollinator and disperser spatial distributions, as well as details of these
217 interactions, and this data is also scant for most plant groups (see also Weber and Agrawal, 2012;
218 Dellinger, 2020). Much of this data can be only collected through carefully designed experiments
219 or field observations (Sinnott-Armstrong et al., 2022) that can be expensive, risky, and time-
220 consuming, and for that reason often avoided by research groups.

221

222 **New biological questions and shifts in paradigm**

223 It is perhaps worth noting that the progress in the field discussed in here seems to be
224 mostly technical – that is, based on the development of new datasets and methods and not
225 necessarily on a new set of biological questions. A focus on technical progress is not necessarily
226 a bad thing. In a way, trait-environment correlation studies have always aimed to answer the
227 simple question of “why plants are where they are and why they look how they look” that existed
228 for centuries. The advantage of modern approaches is that they allow a more biologically
229 realistic picture of how correlated evolution works, and so can lead to more satisfactory answers
230 to these questions. However, it is also possible to argue that technical developments in studies of
231 trait-environment correlations have allowed for a completely new set of biological questions to
232 be asked. For instance, the parameterization of some evolutionary processes in new extensions of
233 trait evolution models have changed the way in which we investigate trait-environment
234 correlations. One example is the focus on evolutionary rates. Measuring rates have opened the
235 possibility to explore questions related to the role of the environment on trait lability (e.g. Lovo
236 et al., 2021) and the role of traits on niche conservatism and niche evolution (e.g. Smith &
237 Beaulieu, 2009; López-Jurado et al., 2019; Qiu et al., 2019; Baniaga et al., 2020; Vasconcelos et
238 al., 2021), rather than merely testing if support for a trait-environment correlation exists or not.
239 Previously established generalities are also challenged when old hypotheses are confronted with
240 new data that often come from poorly studied groups and areas (e.g. Vasconcelos et al., 2019;
241 Vasconcelos et al., 2020). Increasing the number of empirical replicates may shift paradigms in
242 terms of rules and exceptions of how plants diversify in space (e.g. Igea and Tanentzap, 2020;
243 Sun et al., 2020). In that way, overcoming current limitations in data and methods will allow the
244 field to explore new questions and to better set apart rules and exceptions in trait-based
245 approaches to plant biogeography.

246

247 **Acknowledgments**

248 Several people have contributed to clarify my thoughts on trait-environment correlations over the
249 last few years, and I would like to thank especially James Boyko, Jeremy Beaulieu, and Brian
250 O'Meara for discussion and/or suggestions that helped improve this manuscript. Funding comes
251 from NSF grant DEB-1916558.

252

253 **Data Availability Statement**

254 Data used for generating the maps in Figure 2 was retrieved from the literature cited in its
255 caption. More details are provided in <https://github.com/tncvasconcelos/synthesis>.

256

257 **Literature Cited**

258 Alcantara, S., Ree, R. H., & Mello-Silva, R. (2018). Accelerated diversification and functional
259 trait evolution in Velloziaceae reveal new insights into the origins of the campos rupestres'
260 exceptional floristic richness. *Annals of Botany*, 122(1), 165-180.

261 Arber, A. R. (1920). *Water plants: a study of aquatic angiosperms*. University Press.

262 Baniaga, A. E., Marx, H. E., Arrigo, N., & Barker, M. S. (2020). Polyploid plants have faster
263 rates of multivariate niche differentiation than their diploid relatives. *Ecology Letters*, 23(1), 68-
264 78.

265 Beaulieu, J. M., O'Meara, B. C., & Donoghue, M. J. (2013). Identifying hidden rate changes in
266 the evolution of a binary morphological character: the evolution of plant habit in campanulid
267 angiosperms. *Systematic Biology*, 62(5), 725-737.

268 Beaulieu, J. M., & O'Meara, B. C. (2018). Can we build it? Yes we can, but should we use it?
269 Assessing the quality and value of a very large phylogeny of campanulid angiosperms. *American*
270 *Journal of Botany*, 105(3), 417-432.

271 Boyko, J. D., & Beaulieu, J. M. (2021). Generalized hidden Markov models for phylogenetic
272 comparative datasets. *Methods in Ecology and Evolution*, 12(3), 468-478.

273 Boyko, J., & Beaulieu, J. (2022). Reducing the biases in false correlations between discrete
274 characters. Preprint at: 10.32942/osf.io/e2kj8

275 Boyko, J., O'Meara, B., & Beaulieu, J. (2022). Jointly Modeling the Evolution of Discrete and
276 Continuous Traits. Preprint at: 10.32942/osf.io/fb8k7

277 Brown, J. L., Hill, D. J., Dolan, A. M., Carnaval, A. C., & Haywood, A. M. (2018). PaleoClim,
278 high spatial resolution paleoclimate surfaces for global land areas. *Scientific Data*, 5(1), 1-9.

279 Bruelheide, H., Dengler, J., Purschke, O., Lenoir, J., Jiménez-Alfaro, B., Hennekens, S. M., et al.
280 (2018). Global trait–environment relationships of plant communities. *Nature Ecology &*
281 *Evolution*, 2(12), 1906-1917.

282 Carmona, C. P., Bueno, C. G., Toussaint, A., Träger, S., Díaz, S., Moora, M., et al. (2021). Fine-
283 root traits in the global spectrum of plant form and function. *Nature*, 597(7878), 683-687.

284 Charles-Dominique, T., Davies, T. J., Hempson, G. P., Bezeng, B. S., Daru, B. H., Kabongo, R.
285 M., et al. (2016). Spiny plants, mammal browsers, and the origin of African
286 savannas. *Proceedings of the National Academy of Sciences*, 113(38), E5572-E5579.

287 Collen, B., Ram, M., Zamin, T., & McRae, L. (2008). The tropical biodiversity data gap:
288 addressing disparity in global monitoring. *Tropical Conservation Science*, 1(2), 75-88.

289 Colli-Silva, M., Reginato, M., Cabral, A., Forzza, R. C., Pirani, J. R., & Vasconcelos, T. N. C.
290 (2020). Evaluating shortfalls and spatial accuracy of biodiversity documentation in the Atlantic
291 Forest, the most diverse and threatened Brazilian phytogeographic domain. *Taxon*, 69(3), 567-
292 577.

293 Cornwell, W. K., Pearse, W. D., Dalrymple, R. L., & Zanne, A. E. (2019). What we (don't) know
294 about global plant diversity. *Ecography*, 42(11), 1819-1831.

295 Darwin, C. (1859). *On the origin of species*. Routledge.

296 Davis, M. P., Midford, P. E., & Maddison, W. (2013). Exploring power and parameter
297 estimation of the BiSSE method for analyzing species diversification. *BMC Evolutionary*
298 *Biology*, 13(1), 1-11.

299 Dellinger, A. S. (2020). Pollination syndromes in the 21st century: where do we stand and where
300 may we go? *New Phytologist*, 228(4), 1193-1213.

301 Díaz, S., Kattge, J., Cornelissen, J. H., Wright, I. J., Lavorel, S., Dray, S., et al. (2016). The
302 global spectrum of plant form and function. *Nature*, 529(7585), 167-171.

303 Donoghue, M. J. (1989). Phylogenies and the analysis of evolutionary sequences, with examples
304 from seed plants. *Evolution*, 43(6), 1137-1156.

305 Donoghue, M. J., Eaton, D. A., Maya-Lastra, C. A., Landis, M. J., Sweeney, P. W., Olson, M. E.,
306 et al. (2022). Replicated radiation of a plant clade along a cloud forest archipelago. *Nature*
307 *Ecology & Evolution*, 1-12.

308 Drummond, C. S., Eastwood, R. J., Miotto, S. T., & Hughes, C. E. (2012). Multiple continental
309 radiations and correlates of diversification in *Lupinus* (Leguminosae): testing for key innovation
310 with incomplete taxon sampling. *Systematic Biology*, 61(3), 443-460.

311 Eiserhardt, W. L., Couvreur, T. L., & Baker, W. J. (2017). Plant phylogeny as a window on the
312 evolution of hyperdiversity in the tropical rainforest biome. *New Phytologist*, 214(4), 1408-1422.

313 Felsenstein, J. (1985). Phylogenies and the comparative method. *The American*
314 *Naturalist*, 125(1), 1-15.

315 Felsenstein, J. (1988). Phylogenies from molecular sequences: inference and reliability. *Annual*
316 *Review of Genetics*, 22(1), 521-565.

317 Fernández-Martínez, M., Pearse, I., Sardans, J., Sayol, F., Koenig, W. D., LaMontagne, J. M., et
318 al. (2019). Nutrient scarcity as a selective pressure for mast seeding. *Nature Plants*, 5(12), 1222-
319 1228.

320 Fine, P. V., Mesones, I., & Coley, P. D. (2004). Herbivores promote habitat specialization by
321 trees in Amazonian forests. *Science*, 305(5684), 663-665.

322 Foster, S., & Janson, C. H. (1985). The relationship between seed size and establishment
323 conditions in tropical woody plants. *Ecology*, 66(3), 773–780. <https://doi.org/10.2307/1940538>

324 Freeman, B. G., & Pennell, M. W. (2021). The latitudinal taxonomy gradient. *Trends in Ecology*
325 *& Evolution*, 36(9), 778-786.

326 GBIF: Global Biodiversity Information Facility (2022), GBIF Home Page. Available from:
327 <https://www.gbif.org> [22 August 2022].

328 Good, R. O. (1931). A theory of plant geography. *New Phytologist*, 149-171.

329 Givnish, T. J. (1988). Adaptation to sun and shade: a whole-plant perspective. *Functional Plant*
330 *Biology*, 15(2), 63-92.

331 Hahn, P. G., Agrawal, A. A., Sussman, K. I., & Maron, J. L. (2019). Population variation,
332 environmental gradients, and the evolutionary ecology of plant defense against herbivory. *The*
333 *American Naturalist*, 193(1), 20-34.

334 Igea, J., & Tanentzap, A. J. (2020). Angiosperm speciation cools down in the tropics. *Ecology*
335 *Letters*, 23(4), 692-700.

336 Ho, L. S., Tung & Ané, C. (2014). A linear-time algorithm for Gaussian and non-Gaussian trait
337 evolution models. *Systematic biology*, 63(3), 397-408.

338 Karger, D. N., Conrad, O., Böhner, J., Kawohl, T., Kreft, H., Soria-Auza, R. W., et al. (2017).
339 Climatologies at high resolution for the earth’s land surface areas. *Scientific data*, 4(1), 1-20.

340 Kattge, J., Diaz, S., Lavorel, S., Prentice, I. C., Leadley, P., Bönisch, G., et al. (2011). TRY—a
341 global database of plant traits. *Global change biology*, 17(9), 2905-2935.

342 Khabbazian, M., Kriebel, R., Rohe, K., & Ane, C. (2016). Fast and accurate detection of
343 evolutionary shifts in Ornstein–Uhlenbeck models. *Methods in Ecology and Evolution*, 7(7),
344 811-824.

345 Kursar, T. A., Dexter, K. G., Lokvam, J., Pennington, R. T., Richardson, J. E., Weber, M. G., et
346 al. (2009). The evolution of antiherbivore defenses and their contribution to species coexistence
347 in the tropical tree genus *Inga*. *Proceedings of the National Academy of Sciences*, 106(43),
348 18073-18078.

349 Laliberté, E. (2017). Below-ground frontiers in trait-based plant ecology. *New Phytologist*,
350 213(4), 1597-1603.

351 López-Jurado, J., Mateos-Naranjo, E., & Balao, F. (2019). Niche divergence and limits to
352 expansion in the high polyploid *Dianthus broteri* complex. *New Phytologist*, 222(2), 1076-1087.

353 Louca, S., & Pennell, M. W. (2020). Extant timetrees are consistent with a myriad of
354 diversification histories. *Nature*, 580(7804), 502-505.

355 Lovo, J., Alcantara, S., Vasconcelos, T. N., Sajo, M. D. G., Rudall, P. J., Prenner, G., ... &
356 Mello-Silva, R. (2021). Evolutionary lability in floral ontogeny affects pollination biology in
357 Trimezieae. *American Journal of Botany*, 108(5), 828-843.

358 Maddison, W. P., & FitzJohn, R. G. (2015). The unsolved challenge to phylogenetic correlation
359 tests for categorical characters. *Systematic Biology*, 64(1), 127-136.

360 Maitner, B. S., Boyle, B., Casler, N., Condit, R., Donoghue, J., Durán, S. M., et al. (2018). The
361 bien r package: A tool to access the Botanical Information and Ecology Network (BIEN)
362 database. *Methods in Ecology and Evolution*, 9(2), 373-379

363 Maliet, O., & Morlon, H. (2022). Fast and accurate estimation of species-specific diversification
364 rates using data augmentation. *Systematic Biology*, 71(2), 353-366.

365 Moles, A. T., Ackerly, D. D., Tweddle, J. C., Dickie, J. B., Smith, R., Leishman, M. R., ... &
366 Westoby, M. (2007). Global patterns in seed size. *Global Ecology and Biogeography*, 16(1),
367 109-116.

368 Moles, A. T. (2018). Being John Harper: Using evolutionary ideas to improve understanding of
369 global patterns in plant traits. *Journal of Ecology*, 106(1), 1-18.

370 NCBI: National Center for Biotechnology Information. Bethesda (MD): National Library of
371 Medicine (US), National Center for Biotechnology Information; [1988] – [22 August 2022].
372 Available from: <https://www.ncbi.nlm.nih.gov/>

373 Onstein, R. E., Kissling, W. D., Chatrou, L. W., Couvreur, T. L., Morlon, H., & Sauquet, H.
374 (2019). Which frugivory-related traits facilitated historical long-distance dispersal in the custard
375 apple family (Annonaceae)? *Journal of Biogeography*, 46(8), 1874-1888.

376 Qiu, F., Baack, E. J., Whitney, K. D., Bock, D. G., Tetreault, H. M., Rieseberg, L. H., &
377 Ungerer, M. C. (2019). Phylogenetic trends and environmental correlates of nuclear genome size
378 variation in *Helianthus* sunflowers. *New Phytologist*, 221(3), 1609-1618.

379 Pagel, M. (1994). Detecting correlated evolution on phylogenies: a general method for the
380 comparative analysis of discrete characters. *Proceedings of the Royal Society of London. Series*
381 *B: Biological Sciences*, 255(1342), 37-45.

382 Pannell, J. R., Auld, J. R., Brandvain, Y., Burd, M., Busch, J. W., Cheptou, P. O., et al. (2015).
383 The scope of Baker's law. *New Phytologist*, 208(3), 656-667.

384 Pennington, R. T., & Hughes, C. E. (2014). The remarkable congruence of New and Old World
385 savanna origins. *New Phytologist*, 204(1), 4-6.

386 POWO: Plants of the World Online. (2022) Available from: <https://powo.science.kew.org/>
387 [Accessed February 2022]

388 Raunkiaer, C. (1934). *The life forms of plants and statistical plant geography; being the*
389 *collected papers of C. Raunkiaer*. The Clarendon Press, Oxford, UK.

390 Rice, A., Šmarda, P., Novosolov, M., Drori, M., Glick, L., Sabath, N., et al. (2019). The global
391 biogeography of polyploid plants. *Nature Ecology & Evolution*, 3(2), 265-273.

392 Simon, M. F., Grether, R., de Queiroz, L. P., Skema, C., Pennington, R. T., & Hughes, C. E.
393 (2009). Recent assembly of the Cerrado, a neotropical plant diversity hotspot, by in situ
394 evolution of adaptations to fire. *Proceedings of the National Academy of Sciences*, 106(48),
395 20359-20364.

396 Sinnott-Armstrong, M. A., Donoghue, M. J., & Jetz, W. (2021). Dispersers and environment
397 drive global variation in fruit colour syndromes. *Ecology Letters*, 24(7), 1387-1399.

398 Sinnott-Armstrong, M. A., Deanna, R., Pretz, C., Liu, S., Harris, J. C., Dunbar-Wallis, A., et al
399 (2022). How to approach the study of syndromes in macroevolution and ecology. *Ecology and*
400 *Evolution*, 12(3), e8583.

401 Smith, S. A., & Beaulieu, J. M. (2009). Life history influences rates of climatic niche evolution
402 in flowering plants. *Proceedings of the Royal Society B: Biological Sciences*, 276(1677), 4345-
403 4352.

404 Smith, S. A., & Brown, J. W. (2018). Constructing a broadly inclusive seed plant
405 phylogeny. *American Journal of Botany*, 105(3), 302-314.

406 Staggemeier, V. G., Diniz-Filho, J. A. F., & Morellato, L. P. C. (2010). The shared influence of
407 phylogeny and ecology on the reproductive patterns of Myrteae (Myrtaceae). *Journal of*
408 *Ecology*, 98(6), 1409-1421.

409 Stebbins, G. L. (1950). *Variation and evolution in plants*. Columbia University Press, USA.

410 Stebbins, G. L. (1974). *Flowering plants: evolution above the species level*. Harvard University
411 Press, USA.

412 Sun, M., Folk, R. A., Gitzendanner, M. A., Soltis, P. S., Chen, Z., Soltis, D. E., & Guralnick, R.
413 P. (2020). Recent accelerated diversification in rosids occurred outside the tropics. *Nature*
414 *Communications*, 11(1), 1-12.

415 Tribble, C. M., May, M. R., Jackson-Gain, A., Zenil-Ferguson, R., Specht, C. D., & Rothfels, C.
416 J. (2021). Unearthing modes of climatic adaptation in underground storage organs across
417 Liliales. bioRxiv. Available at: 10.1101/2021.09.03.458928

418 Tudela-Isanta, M., Fernández-Pascual, E., Wijayasinghe, M., Orsenigo, S., Rossi, G., Pritchard,
419 H. W., & Mondoni, A. (2018). Habitat-related seed germination traits in alpine habitats. *Ecology*
420 *and Evolution*, 8(1), 150-161.

421 Uyeda, J. C., & Harmon, L. J. (2014). A novel Bayesian method for inferring and interpreting the
422 dynamics of adaptive landscapes from phylogenetic comparative data. *Systematic Biology*, 63(6),
423 902-918.

424 Uyeda, J. C., Caetano, D. S., & Pennell, M. W. (2015). Comparative analysis of principal
425 components can be misleading. *Systematic Biology*, 64(4), 677-689.

426 Vasconcelos, T. N., Chartier, M., Prenner, G., Martins, A. C., Schönenberger, J., Wingler, A., &
427 Lucas, E. (2019). Floral uniformity through evolutionary time in a species-rich tree lineage. *New*
428 *Phytologist*, 221(3), 1597-1608.

429 Vasconcelos, T. N., Alcantara, S., Andriano, C. O., Forest, F., Reginato, M., Simon, M. F., &
430 Pirani, J. R. (2020). Fast diversification through a mosaic of evolutionary histories characterizes
431 the endemic flora of ancient Neotropical mountains. *Proceedings of the Royal Society*
432 *B*, 287(1923), 20192933.

433 Vasconcelos, T., Boyko, J. D., & Beaulieu, J. M. (2021). Linking mode of seed dispersal and
434 climatic niche evolution in flowering plants. *Journal of Biogeography*.

435 Vasconcelos, T., O'Meara, B. C., & Beaulieu, J. M. (2022). Retiring “cradles” and “museums”
436 of biodiversity. *The American Naturalist*, 199(2), 194-205.

437 Von Humboldt, A., & Bonpland, A. (1807). *Essay on the Geography of Plants*. (2010 English
438 translation) University of Chicago Press, USA.

439 Webb, C. O., Ackerly, D. D., & Kembel, S. W. (2008). Phylocom: software for the analysis of
440 phylogenetic community structure and trait evolution. *Bioinformatics*, 24(18).

441 Webb, C. O., & Donoghue, M. J. (2005). Phylomatic: tree assembly for applied
442 phylogenetics. *Molecular Ecology Notes*, 5(1), 181-183.

443 Weber, M. G., & Agrawal, A. A. (2012). Phylogeny, ecology, and the coupling of comparative
444 and experimental approaches. *Trends in Ecology & Evolution*, 27(7), 394-403.

445 Wright, I. J., Reich, P. B., Westoby, M., Ackerly, D. D., Baruch, Z., Bongers, F., ... & Villar, R.
446 (2004). The worldwide leaf economics spectrum. *Nature*, 428(6985), 821-827.

447 Zanne, A. E., Chapman, C. A., & Kitajima, K. (2005). Evolutionary and ecological correlates of
448 early seedling morphology in East African trees and shrubs. *American Journal of Botany*, 92(6),
449 972-978.

450 Zanne, A. E., Tank, D. C., Cornwell, W. K., Eastman, J. M., Smith, S. A., FitzJohn, R. G., ... &
451 Beaulieu, J. M. (2014). Three keys to the radiation of angiosperms into freezing
452 environments. *Nature*, 506(7486), 89-92.

453 Zanne, A. E., Pearse, W. D., Cornwell, W. K., McGlenn, D. J., Wright, I. J., & Uyeda, J. C.
454 (2018). Functional biogeography of angiosperms: life at the extremes. *New Phytologist*, 218(4),
455 1697-1709.