

# Biorestoror: A Framework for Synthetic Succession with a Qualitative System-Level Illustration

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**Abstract:** Ecosystem restoration in severely degraded or soil-absent environments requires approaches that operate independently of natural soils and make effective use of locally available resources. The Biorestoror platform introduces synthetic succession as a systems-based engineering framework structured in sequential functional phases and aligned with in situ resource utilization (ISRU) principles, understood here as the use of locally available resources independent of terrestrial or extraterrestrial setting. Its core element, the soil initiator, is a prototype soilless, mineral-dominated substrate engineered to support early soil-like functionality under low-organic conditions by integrating mineral components, porous carbon scaffolds, and targeted microbial inoculants. In the initial phase, locally available mineral materials are configured to achieve targeted physical and geochemical properties, including porosity, pH buffering, and potential nutrient accessibility. These are combined with microorganisms associated with mineral nutrient mobilization, such as rock-solubilizing bacteria (RSB), to support biological colonization. Subsequent phases introduce vegetation and plant-associated microbial communities, enabling organic matter accumulation and increasing system complexity. A qualitative engineering test of a defined prototype configuration was conducted to assess system-level behavior rather than to isolate causal mechanisms. The prototype supported plant establishment and continued growth in a mineral-dominated substrate without compost, humus, or conventional fertilizers. Observations included stabilization of substrate pH, visible structural changes, and growth of two successive plant generations (*Sinapis alba* and *Phaseolus vulgaris*). The inclusion of a cultivated species in the second generation is consistent with multi-stage plant establishment under the tested prototype conditions. As no control treatments or quantitative chemical or microbiological analyses were included, these findings should be interpreted as qualitative indicators of system-level feasibility, not as evidence of specific underlying mechanisms or validated process-level performance. Further controlled studies are required to resolve component contributions, process dynamics, reproducibility, scalability, and operational performance.

**Keywords:** synthetic succession; synthetic pedogenesis; biochar scaffold; rock-solubilizing bacteria; mineral-dominated substrate; soil-like functionality; ecological restoration; in situ resource utilization

## I. Introduction

Biorestoror presents a systems-based framework for synthetic pedogenesis, in which soil is treated not as a material input but as an emergent, engineerable function arising from controlled pedogenetic processes. The framework integrates mineral components forming a soilless substrate, targeted microbial activation, and functional carbon scaffolds within a structured, staged system design, forming a configuration potentially applicable in environments where natural soil formation is absent or operationally infeasible, including terrestrial restoration and in situ resource utilization (ISRU) contexts. The theoretical foundation and system-level conceptual design of the framework are described in an earlier preprint [1], and were subsequently presented in poster form at an international conference focused on dryland systems [2].

Severely degraded, arid, or soil-absent environments—including mine tailings, exposed bedrock, desert crusts, and extraterrestrial regolith analogues—pose a fundamental challenge to ecological restoration, sustainable agriculture, and bioregenerative life-support systems. In such contexts, natural soil formation is extremely slow or suppressed due to minimal organic matter, limited biological activity, unfavorable pH conditions, and poor water retention. Conventional restoration approaches typically assume the presence of residual soil functionality and are therefore ineffective where soil is biologically inactive or entirely absent [3].

47 Comparable constraints arise in controlled environments and extraterrestrial settings, where soils do not exist by  
48 definition and must be functionally generated from mineral parent materials. This perspective is directly relevant to  
49 in-situ resource utilization (ISRU), where the mobilization of nutrients and the establishment of biologically  
50 functional interfaces from locally available mineral resources represent a critical limitation for long-duration  
51 missions and closed-loop life-support architectures.

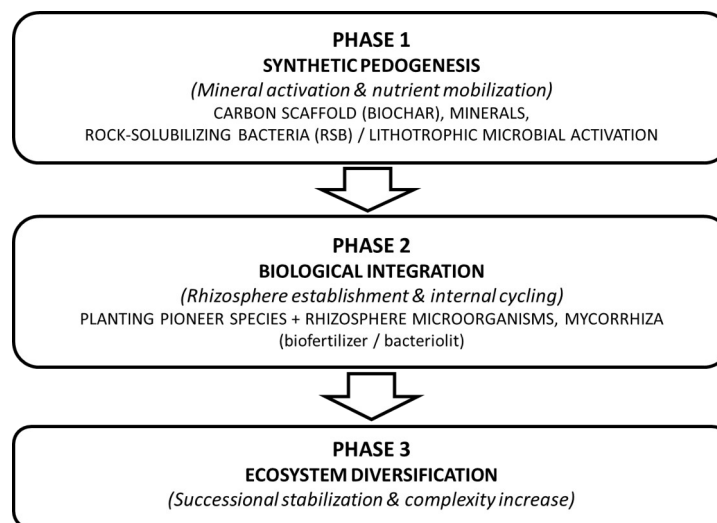
52 In natural systems, early soil development occurs during primary succession, when initially sterile mineral  
53 substrates are colonized by microorganisms, leading to the formation of a proto-soil (protopedon). A central driver  
54 of this process is the activity of rock-solubilizing and lithotrophic microorganisms, which mobilize essential  
55 nutrients through mineral weathering long before significant organic matter accumulation occurs [4,5]. In  
56 engineered analogues of this process, early microbial colonization may be supported by transient, low-intensity  
57 organic stimulation acting as a mild activator, without constituting a sustained external carbon source.

58 Most existing restoration technologies focus on accelerating secondary succession and therefore rely on pre-  
59 existing soil functionality. However, plant establishment depends not on soil as a material entity, but on a set of  
60 functional services, including mechanical support, access to water and nutrients, gas exchange, and a biologically  
61 active rhizosphere. These services can be delivered through engineered systems, effectively decoupling plant growth  
62 from natural soils [6].

63 Building on this functional view, synthetic succession has been proposed as a controlled analogue of natural  
64 primary succession. Rather than improving degraded soils, it focuses on the deliberate creation of a synthetic parent  
65 material designed to support microbial activation, mineral weathering, and early aggregation. Biorestorer  
66 implements this concept as a controllable pedogenetic platform, whose core element is a soil initiator—an  
67 engineered mineral–biochar substrate providing water retention, pH buffering, nutrient mobilization, and  
68 rhizosphere support.

69 Within this framework, biochar is not treated merely as a stable carbon additive, but as an active microbial–  
70 mineral interface. The concept of the charosphere describes the unique microbial habitat created by biochar,  
71 characterized by elevated microbial density, altered community composition, and intensified biochemical activity  
72 relative to the surrounding mineral matrix [7,8]. Biochar particles provide porous microenvironments that promote  
73 close spatial coupling between microorganisms and mineral surfaces, and have been associated in the literature with  
74 processes such as localized mineral weathering, ion exchange, and nutrient mobilization at the mineral–biochar  
75 substrate, particularly during early stages of soil formation.

76 A key design feature of the Biorestorer system is the use of dual-temperature biochar. Lower-temperature  
77 biochar (~500–600 °C) supports microbial activation and early-stage biological processes through higher surface  
78 reactivity and porosity, while higher-temperature biochar (~750–800 °C) contributes long-term structural stability  
79 and buffering capacity due to its chemical persistence [9]. Together, these components enable the transition from a  
80 mineral-dominated substrate to a biologically active protopedon.  
81



82

83 **Figure 1. Graphical abstract illustrating synthetic succession in the Biorestorer concept.**

84 Although biochar is used here as a reference implementation, the Biorestorer framework is not intrinsically  
85 dependent on biochar as a specific material. From a systems perspective, biochar functions as a carbonaceous  
86 structural scaffold, whose role may be fulfilled by alternative carbon-based or carbon-rich substrates providing  
87 comparable porosity, surface reactivity, chemical stability, and microbial hosting capacity. Functional analogues  
88 may include engineered porous carbons, pyrolyzed organic residues, charred polymer-derived materials, or  
89 carbonized waste streams produced under controlled thermal or chemical conditions. Provided that such materials  
90 offer a stable porous architecture, suitable surface chemistry for microbial attachment, and resistance to rapid  
91 degradation, they can serve as effective microbial–mineral interfaces within the Biorestorer system, thereby  
92 extending its applicability to resource-constrained and ISRU-oriented bioregenerative life support architectures.

93 The objective of this paper is to describe the Biorestorer system architecture, outline its conceptual pedogenetic  
94 framework, and present an illustrative assessment of early soil-like development and plant establishment on a  
95 predominantly mineral substrate. By framing soil as an emergent functional interface rather than a prerequisite  
96 material, this work contributes to scalable strategies for environmental restoration and bioregenerative life-support  
97 systems operating under resource-constrained conditions.

## 98 **II. Materials and Methods**

99 The processes described in the conceptual framework, including microbially induced weathering and nutrient  
100 mobilization, are not directly evaluated in this study and are presented as theoretical components of the system  
101 design.

### 102 **A. Theoretical Framework of the Biorestorer System**

103 The Biorestorer system is based on the premise that essential plant nutrients are inherently present within  
104 mineral parent materials and that a key limitation in soil-absent or severely degraded environments is nutrient  
105 inaccessibility rather than absolute scarcity. In natural systems, nutrient availability emerges through long-term  
106 mineral weathering mediated by physical, chemical, and biological processes [4,5]. Where functional soil is absent,  
107 these processes are limited, resulting in mineral substrates that may be chemically rich but biologically inaccessible.

108 Rather than compensating for this limitation through external fertilizers or organic amendments, the Biorestorer  
109 approach aims to create conditions consistent with access to nutrients associated with mineral substrates by  
110 promoting conditions consistent with early pedogenetic development. This approach is conceptually aligned with  
111 natural primary succession, in which nutrient cycling emerges through microbially influenced mineral  
112 transformations prior to significant organic matter accumulation [4,10]. Accordingly, mineral components are  
113 treated as potential nutrient reservoirs rather than inert fillers.

114 The Biorestorer soil initiator (a soilless substrate), integrates interdependent components:

- 115 1) mineral parent material,
- 116 2) rock-solubilizing and lithotrophic microorganisms,
- 117 3) dual-temperature biochar serving as a structural and biochemical scaffold,
- 118 4) a controlled water regime,
- 119 5) Staged System Design and Operational Phases.

120 Together, these components establish a framework for synthetic pedogenesis in which mineral composition,  
121 microbial activity, porosity, moisture dynamics, and pH buffering are treated as controllable design variables.

122 An illustrative assessment of system behavior under a defined framework, including parallel system pathways, is  
123 presented in the following sections.

### 124 **B. Mineral Components as Synthetic Parent Material**

125 The mineral fraction of the Biorestorer substrate functions as a synthetic parent material analogous to bedrock or  
126 unconsolidated deposits in natural soil formation [5]. A range of silicate and carbonate rocks may be used, including  
127 basalt, andesite, dolomite, granodiorite, limestone-derived materials, regolith analogues, and other locally available  
128 mineral resources, depending on terrestrial or extraterrestrial application context.

129 These substrates contain essential macro- and micronutrients, including calcium, magnesium, potassium,  
130 phosphorus, iron, and trace elements bound within their crystal lattices [5].

131 Although often considered inert on operational timescales, mineral-bound nutrients become bioavailable through  
132 microbially mediated weathering, particularly within acidic or chelating microenvironments [4,10]. The Biorestorer  
133 system is explicitly designed to promote such microenvironments, enabling mineral dissolution and nutrient release  
134 without external fertilization. While mineral composition influences pH dynamics and weathering rates, it does not

134 alter the fundamental role of the mineral fraction as a long-term nutrient source, allowing adaptation to local  
135 geological conditions through material substitution.

### 136 **C. Rock-Solubilizing and Lithotrophic Microorganisms**

137 Rock-solubilizing bacteria (RSB) and functionally similar lithotrophic microorganisms form the core biological  
138 component of the Biorestorator system. These organisms mobilize nutrients directly from mineral substrates through  
139 the production of organic acids, chelating compounds, and redox-active metabolites, facilitating the release of  
140 phosphorus, calcium, magnesium, iron, and trace elements [4,11].

141 In natural primary succession, such microorganisms establish the first biologically accessible nutrient pools  
142 under low-carbon conditions [4,10]. Biorestorator deliberately integrates this function into the engineered substrate,  
143 treating microbial mineral weathering as a conceptual system component rather than an emergent byproduct. The  
144 system does not depend on high organic inputs, enabling pedogenetic processes to proceed under low-carbon  
145 conditions characteristic of arid, degraded, or extraterrestrial analogue environments.

### 146 **D. Dual-Temperature Biochar as Structural and Biochemical Scaffold**

147 Within the Biorestorator system, biochar functions primarily as a structural and biochemical scaffold rather than as  
148 a deliberately added nutrient source. Biochars produced at different pyrolysis temperatures exhibit distinct and  
149 complementary properties. Lower-temperature biochar (~500–600 °C) provides higher surface functionality and  
150 cation exchange capacity, supporting microbial colonization, nutrient retention, and early rhizosphere development  
151 [6]. Higher-temperature biochar (~750–800 °C) offers greater chemical stability and long residence times,  
152 contributing to long-term structural integrity and buffering capacity [9].

153 The combined use of these biochar fractions balances early biological activation with long-term system stability.  
154 In addition, biochar provides microhabitats that buffer moisture, pH fluctuations, and temperature extremes,  
155 enhancing microbial survival and activity during early synthetic pedogenesis [7,8,12].

156 Importantly, these roles are defined functionally rather than materially. While biochar represents a convenient  
157 and well-characterized carbon scaffold, its structural and biochemical functions within the Biorestorator framework  
158 are in principle substitutable by alternative carbon-based or carbon-rich scaffolds that provide comparable porosity,  
159 surface reactivity, and environmental buffering.

### 160 **E. Water Regime and Process Activation**

161 Water availability is a critical control variable within the Biorestorator system, as microbial mineral weathering  
162 and nutrient mobilization are strongly water-dependent [4]. The system is designed to operate under intermittent  
163 wetting regimes, reflecting hydrological conditions typical of arid and semi-arid environments. Episodic water  
164 inputs activate microbial metabolism, promote localized mineral dissolution, and support aggregation while limiting  
165 nutrient leaching and system destabilization [13].

166 Within synthetic pedogenesis, the water regime functions as an engineered process parameter regulating  
167 microbial activity, reaction kinetics, and system progression toward a biologically active protopedon. As plants are  
168 introduced, they become integral system components, contributing to nutrient and carbon cycling through root  
169 exudation and rhizosphere structuring. Early vegetation thus reinforces synthetic pedogenesis and supports the  
170 transition from an externally activated system toward a partially self-sustaining bioregenerative loop.

### 171 **F. Staged System Design and Operational Phases**

172 The Biorestorator framework is structured as a staged system design in which synthetic pedogenesis is approached  
173 as a sequence of functionally distinct but interconnected phases. Rather than representing discrete experimental  
174 treatments, these phases define operational states of the system characterized by evolving interactions between  
175 mineral, microbial, and plant components, conceptually aligned with processes described in natural soil  
176 development and ecological succession [4,5,10].

177 In the initial phase, mineral components, carbon scaffolds, and microbial inoculants are combined to establish a  
178 baseline substrate with controlled physical and geochemical properties, including porosity, moisture retention, and  
179 pH buffering. This phase focuses on enabling microbial colonization and early system activation under low-organic  
180 conditions, consistent with early-stage mineral–microbe interactions observed in primary succession environments  
181 [4,10].

182 Subsequent phases involve the introduction of vegetation and the emergence of plant–microbe–substrate  
183 interactions. Root development and biomass turnover contribute to internal carbon inputs and localized modification

184 of substrate conditions. Microbial inputs may be maintained or supplemented across phases as part of the integrated  
185 system design, without isolating their individual effects.

186 The transition between phases is guided by operational indicators rather than predefined thresholds. These  
187 indicators may include stabilization of substrate conditions and the emergence of observable biological activity,  
188 without implying specific causal mechanisms.

189 This staged approach provides a structured framework for examining system behavior across configurations  
190 while maintaining flexibility in implementation.

191

### III. Illustrative Study Methodology

#### A. Scope and Study Design

192 The study was designed as a minimal feasibility assessment of the Biorestor framework under a defined set of  
193 conditions. Its objective was not to test specific mechanisms or comparative performance, but to examine whether  
194 the system can maintain functional continuity across staged transitions and support observed plant establishment in a  
195 mineral-dominated substrate.  
196

197 The experiment was not intended as an optimization study, comparative agronomic trial, or quantitative  
198 performance evaluation. Instead, it was designed as an illustrative, system-level assessment of the framework under  
199 a defined configuration. Observed plant growth was interpreted as an indication that the system configuration may  
200 be compatible with the transition from synthetic pedogenesis (Phase I) to biological integration (Phase II), but not as  
201 evidence of underlying mechanisms or their relative contributions. Quantitative observations were collected as  
202 contextual indicators of system stability and were not used as primary success criteria.

203 This reflects the study design, which was not intended as a comparative or hypothesis-testing experiment, but as  
204 an illustrative assessment of system behavior under a defined configuration. The results should therefore be  
205 interpreted as preliminary qualitative observations within the scope of the framework.

#### B. Experimental Setup

206 The experiment was conducted under controlled indoor conditions using a total substrate volume of 32 L.  
207 Moisture levels were maintained at values sufficient to support microbial activity, within a range reported in the  
208 literature to be associated with mineral weathering processes [4]. No external fertilization or organic amendments  
209 were applied at any stage.

210 The substrate consisted predominantly of mineral components with minimal organic matter inputs. Organic  
211 material was not intentionally introduced and was present only in negligible amounts associated with mineral  
212 fractions, consistent with experimental approaches focusing on early pedogenetic processes under low-carbon  
213 conditions [14].

214 Substrate (soil initiator), composition (v/v):

- 215 1) 70 % basalt aggregate (1–3 mm fraction), commercially available construction-grade jointing gravel,
- 216 2) 20 % dolomite rubble of natural origin, included as a mineral component with buffering capacity,
- 217 3) 10 % biochar (KBRO-600-ZZ Biograda, produced from digestate derived from kitchen waste, high ash  
218 content).
- 219

220 The initial microbial population was not assumed to originate from the biochar component. Instead, dolomite of  
221 natural origin was included as part of the substrate, which may contain naturally occurring microbial communities.  
222 In addition, a commercial microbial inoculant was applied during experimental phase 2.

223 Waste-derived biochar was selected to reflect circular resource use under constrained biomass conditions. While  
224 such materials are sometimes regarded as lower-grade in conventional agronomy due to elevated ash content, this  
225 mineral fraction represents an inherent component of biochar rather than an externally added amendment. In the  
226 present system, it may represent a potential source of buffering capacity and mineral-associated nutrients; however,  
227 its functional contribution under the tested conditions cannot be resolved. The feedstock choice also supports  
228 scalability through high-volume waste streams.

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232

Parameter	Unit	Value
Production temperature	°C	~500
pH (H <sub>2</sub> O)	–	8.9
Ash content	% (dry basis)	42.5
Total organic carbon (TOC)	% (dry basis)	34.2
Total nitrogen (N)	% (dry basis)	2.5
Phosphorus (as P <sub>2</sub> O <sub>5</sub> )	% (dry basis)	2.44
Potassium (as K <sub>2</sub> O)	% (dry basis)	1.83
Calcium (Ca)	% (dry basis)	10.3
Iron (Fe)	mg/kg	5310

233 **Table 1. Physicochemical Properties of the Biochar Sample (KBRO-500-ZZ)**

234 The biochar component was characterized by an accredited laboratory (ALS Czech Republic). The analyzed  
 235 sample (KBRO-500-ZZ), produced at ~500 °C, provides an indicative characterization of the mineral ash fraction  
 236 and associated nutrient content of a biochar derived from the same feedstock as that used in the system (KBRO-600-  
 237 ZZ). While differences in pyrolysis temperature (500 °C vs 600 °C) may influence specific physicochemical  
 238 properties, this analysis provides contextual insight into the mineral characteristics of the biochar component. The  
 239 complete laboratory report is provided in Supplementary Material S1.

240 The resulting substrate was mineral-dominated in composition and function, incorporating biochar as a stable  
 241 carbon scaffold, with only negligible incidental organic material associated with the dolomite fraction and no added  
 242 humus, compost, or externally supplied NPK fertilizers.

243 Substrate pH was monitored during Phase I at weeks 2, 3, and 4 using semi-quantitative colorimetric drop tests  
 244 (1:5 w/w substrate-to-distilled-water suspension; 30 min equilibration) as operational system indicators of substrate  
 245 condition rather than quantitative analytical measurements. Iron was assessed using colorimetric drop tests in week  
 246 4 of Phase I as a qualitative indicator of system state under the tested conditions. During Phase II, both pH and iron  
 247 presence were re-evaluated prior to green manuring to monitor system conditions as part of phase-transition  
 248 assessment under plant growth.

### 249 **C. Pre-treatment and System Activation**

250 To initiate synthetic pedogenesis, the basalt and biochar fractions were mixed at a 1:1 ratio, and the biochar  
 251 component was pre-activated within the mixture by soaking for 24 hours in 2 L of activation solution consisting of  
 252 20 mL household vinegar diluted in tap water (corresponding to approximately 1 % vinegar solution, ~0.08 % acetic  
 253 acid). The mixture was not rinsed, allowing all soluble compounds to remain within the system. This pre-treatment  
 254 was intended to activate the biochar surface and provide a transient stimulus for microbial activation and mineral  
 255 surface reactivity without inducing sustained acidification, consistent with approaches used to trigger early-stage  
 256 microbial activity in mineral substrates [4].

### 257 **D. Phase I: Synthetic Pedogenesis**

258 Following substrate preparation, the system was incubated for five weeks without vegetation to allow for initial  
 259 biological colonization and early substrate transformation processes. Comparable timescales for the onset of  
 260 microbially mediated mineral transformations under controlled conditions have been reported in the literature [4,14].

261 The system was monitored qualitatively for physical stability and visually observable structural changes. These  
 262 observations describe visible structural changes but do not allow inference about pedogenetic processes or  
 263 underlying mechanisms.

### 264 **E. Phase II: Biological Integration, Test Groups, and Replication Structure**

265 After completion of Phase I, and following confirmation of stable substrate pH and a positive qualitative iron  
 266 response as operational indicators compatible with Phase II, the experiment was divided into two parallel test groups  
 267 using *Sinapis alba* (white mustard) as a fast-growing, non-pioneer test species. The aim was to qualitatively observe  
 268 plant establishment under mineral-dominated conditions without direct mechanistic inference [3].

269 The two groups represent distinct operational scenarios rather than comparative experimental variants. In both  
 270 groups, independent pots served as biological replicates, with the pot constituting the experimental unit.

271 1) Test Group 1 – Phenological Growth:

272 5 independent pots (n = 5), each containing 1 mustard plant, were cultivated through a full phenological cycle

273 without biomass removal.  
274 2) Test Group 2 – Green Manuring:  
275 5 independent pots (n = 5), containing 13 mustard plants in total, were cultivated for biomass production. After  
276 harvest, above-ground biomass was reincorporated into the substrate as green manure. A subsequent bean  
277 generation was then grown in the amended substrate: first sowing in 3 pots (n = 3), followed by repeated sowing  
278 in 5 pots (n = 5) containing 8 bean plants in total.  
279 In both groups, Bacteriolit (Marcel Mézy) microbial inoculant was applied at 1 g per 3.2 L pot to support early  
280 rhizosphere microbial activity and initiate internal nutrient cycling [4].  
281 Although replicated cultivation units were used within each phase, no uninoculated negative control was  
282 included; replication therefore supports internal consistency assessment only, not comparative causal inference.

#### 283 **F. Observations and Interpretation Criteria**

284 The primary outcome of the study was the visual observation of sustained plant growth on a mineral-dominated  
285 substrate lacking conventional organic soil inputs.

286 Supporting observations included:

- 287 1) substrate pH (measured after two weeks and monitored thereafter),
- 288 2) qualitative colorimetric indication of iron presence (drop-based spot test),
- 289 3) visual evidence of structural changes,
- 290 4) above-ground green biomass at harvest.

291 Substrate pH and qualitative iron presence were monitored primarily as operational indicators guiding the  
292 transition from Phase I (synthetic pedogenesis) to Phase II (biological integration), rather than as measurements of  
293 nutrient dynamics or evidence of underlying mechanisms.

294 Biomass data were recorded exclusively for contextual comparison and were not interpreted as indicators of  
295 system performance or optimization.

296 The study was interpreted as consistent with early soil-like system behavior if the following conditions were  
297 observed:

- 298 1) structural stability of the mineral substrate,
- 299 2) absence of system collapse or extreme pH drift,
- 300 3) sustained growth of *Sinapis alba* in both test groups.

301 These criteria are intended as qualitative reference points for interpreting system behavior within the scope of an  
302 illustrative study.

303

## IV. Observations

### 304 **A. Phase I: Synthetic Pedogenesis Outcomes**

305 During the five-week incubation period without vegetation, the mineral-dominated substrate remained  
306 structurally stable, showing no collapse, excessive compaction, or component segregation. Visual inspection  
307 revealed localized structural changes, particularly in regions with higher biochar content.

308 Substrate pH stabilized at approximately 7.5 following the incubation period and remained stable throughout  
309 Phase I and subsequent plant growth. No progressive acidification or alkalization was observed, indicating  
310 physicochemical conditions compatible with plant establishment under the tested conditions.

311 Qualitative iron spot tests produced a positive colorimetric response, indicating the presence of iron under the  
312 tested conditions. This observation is descriptive only and does not provide quantitative or mechanistic information.

313 Taken together, these observations indicate observable structural changes in the substrate during the incubation  
314 period, consistent with early-stage soil-like structural features (in a descriptive sense).

315 However, in the absence of control treatments and quantitative chemical or microbiological analyses, these  
316 results should be interpreted as qualitative indicators rather than evidence of defined mechanisms.



317

318 **Figure 2. Substrate structure at different stages of the experiment:** (left) freshly prepared substrate; (center)  
319 after Phase I (synthetic pedogenesis); (right) after Phase II (2 weeks of green manuring). Visual differences in  
320 substrate structure are observable across stages.

#### 321 **B. Phase II: Plant Establishment and System Response**

322 Following transition to Phase II, *Sinapis alba* successfully germinated and exhibited sustained growth in both  
323 test groups under mineral-dominated conditions, without the addition of external fertilizers or organic amendments.

324 In the phenological growth group, plants completed their growth cycle without biomass removal. Continuous  
325 development without visible signs of severe nutrient limitation or system instability provided a qualitative indication  
326 of sustained plant–substrate interaction under the given conditions.

327 In the green manuring group, harvested *Sinapis alba* biomass was incorporated into the substrate. The system  
328 remained structurally stable following biomass incorporation, with no visible signs of anaerobic collapse or  
329 excessive compaction.

330 A second plant generation (*Phaseolus vulgaris*, bush bean cultivar ‘Sonesta’) successfully germinated and  
331 exhibited sustained early growth in the green manuring group without additional substrate modification or nutrient  
332 inputs. This observation is consistent with the possibility of internal system reinforcement following biomass  
333 incorporation.



334

335 **Figure 3. Test groups.** (left) Test group 2 (green-manuring); (right) Test group 1 (phenology). Representative plant  
 336 height of approximately 127 cm observed at 63 days after sowing (28 Sep 2025).

337 **C. Supporting Observations**

338 In Test Group 2 (green manuring), above-ground green biomass was recorded at harvest to enable contextual  
 339 comparison with existing studies. Mean biomass yield reached approximately 14 g/L of substrate volume (46 g per  
 340 pot). These values were used exclusively as supporting indicators of sustained plant growth under mineral substrate  
 341 conditions and were not interpreted as measures of system performance, productivity, or optimization.

342 Throughout Phase II, no signs of system collapse, severe nutrient limitation, or pathological plant responses were  
 343 observed. Root systems visibly penetrated and structured the substrate, further indicating functional plant–substrate  
 344 integration.

345 **D. Summary of Observations**

346 The study indicates that the Biorestorer system, under the tested configuration, maintained structural integrity  
 347 and supported plant establishment in a mineral-dominated substrate without the addition of compost, humus, or  
 348 conventional fertilizers.

349 Specifically, the system exhibited the following observable outcomes:

- 350 1) structural stability of the mineral-dominated substrate throughout both experimental phases, without  
 351 collapse, excessive compaction, or component segregation,
- 352 2) stable substrate pH (~7.5) under both incubation and plant growth conditions,
- 353 3) qualitative colorimetric responses consistent with iron presence,
- 354 4) sustained plant growth of *Sinapis alba* under mineral-dominated conditions,
- 355 5) successful establishment of a second plant generation (*Phaseolus vulgaris*) following biomass  
 356 incorporation,
- 357 6) absence of visible system instability, severe nutrient limitation, or pathological plant responses during the  
 358 observation period.

359 These observations are consistent with observable system-level properties such as structural stability and  
360 sustained plant growth under the given conditions.

361 The observed system behavior is consistent with the qualitative interpretation criteria defined for this study  
362 and supports further investigation of the Biorestorer framework under controlled and comparative experimental  
363 conditions.

364

## V. Discussion

365 The conceptual framework is defined independently of material-specific nutrient inputs, whereas the  
366 implemented system included mineral-associated nutrient sources as part of the substrate configuration.

367 Plant establishment and sustained growth were observed in a mineral-dominated substrate under the tested  
368 system configuration. These observations describe system behavior under the tested conditions but do not constitute  
369 evidence for the framework or its underlying assumptions.

370 The mineral ash fraction, as an inherent component of biochar resulting from the pyrolysis process, is  
371 structurally associated with the carbon matrix rather than present as a freely available amendment [15,16]. Elemental  
372 analysis of the biochar indicated the presence of mineral-associated nutrients (e.g., K<sub>2</sub>O 1.83 %, P<sub>2</sub>O<sub>5</sub> 2.44 %),  
373 although these forms are not directly equivalent to readily available fertilizer inputs and do not imply immediate  
374 nutrient accessibility under the tested conditions [15,16].

375 Accordingly, this work does not determine the origin of nutrients supporting plant growth or distinguish between  
376 mineral-associated availability and microbially mediated mobilization. Any apparent consistency with processes  
377 described in the conceptual framework is therefore non-mechanistic and does not imply validation of underlying  
378 assumptions.

### A. Nutrient Accessibility Versus Nutrient Addition

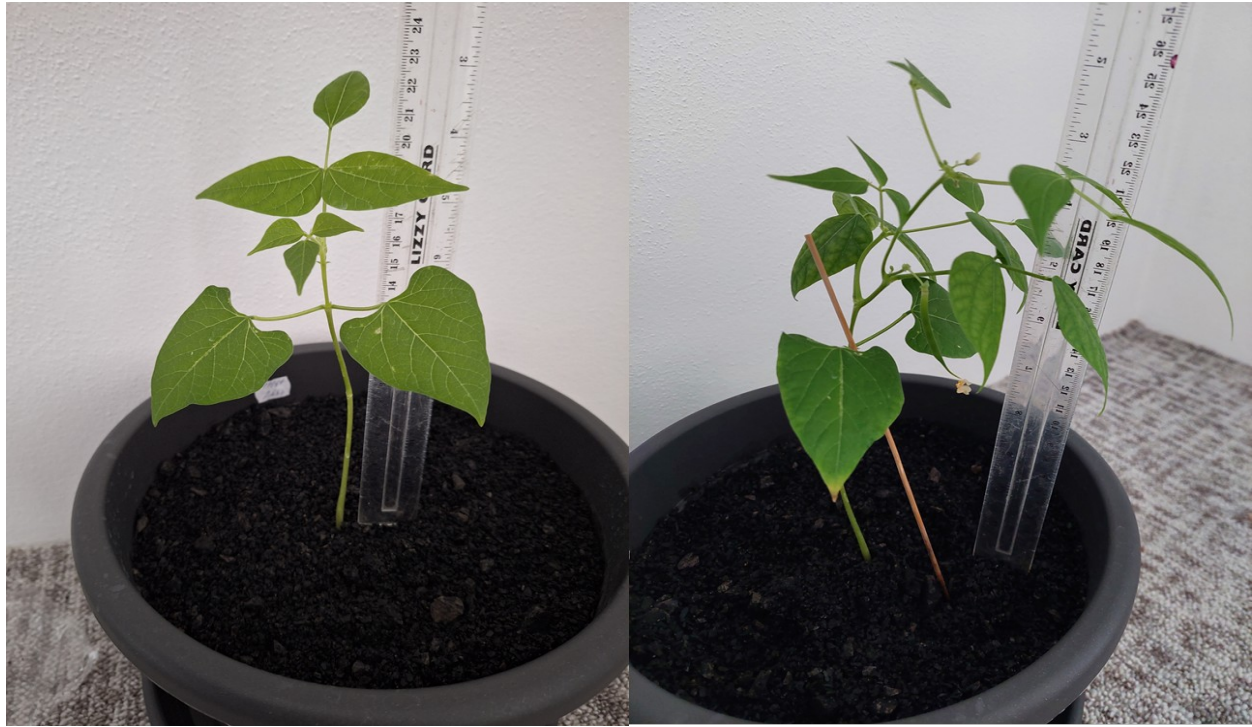
379 A key implication of these observations is the distinction between nutrient accessibility and external nutrient  
380 addition. The absence of compost, humus, or conventional fertilizers indicates that plant growth occurred without  
381 external organic inputs. However, the substrate was not chemically inert and included mineral-associated nutrient  
382 sources, particularly within the biochar component, which contained an ash fraction.

383 Plant establishment therefore indicates that conditions compatible with plant growth were present within the  
384 system in the absence of external inputs; however, their specific sources and relative contributions cannot be  
385 resolved without control treatments and quantitative analysis. Observed stabilization of substrate pH (~7.5) and  
386 qualitative colorimetric detection of iron are consistent with the presence of mineral-associated components under  
387 the given conditions, but do not allow determination of nutrient availability or underlying processes.

388 An initial *Phaseolus vulgaris* sowing (3 plants) was affected by external operational factors (pest infestation and  
389 subsequent phytotoxicity from treatment), leading to premature loss despite prior establishment. These effects were  
390 treated as external constraints unrelated to substrate performance, and the system was re-sown (8 plants) to allow  
391 continued qualitative observation.

392 Mechanical support structures in plants, including stem base thickening and anchoring root development,  
393 represent adaptive responses influenced by environmental conditions such as mechanical stress, water availability,  
394 light regime, and resource availability [17–19]. Under conditions of low mechanical loading and stable water  
395 availability, plants may preferentially allocate biomass to vertical growth and photosynthetic tissues.

396 Plant morphology was not used as a standalone diagnostic indicator of nutrient status, as similar structural traits  
397 may arise from multiple interacting environmental drivers, including light availability and mechanical loading [20–  
398 22]. Under the tested conditions, morphology cannot be unambiguously attributed to nutrient-related limitations.  
399



400

401 **Figure 4. Phaseolus vulgaris during Phase II.** *Limited mechanical anchoring observed under substrate conditions*  
 402 *(same plant, winter window-light conditions; 16 November and 6 December 2025).*



403

404 **Figure 5. Phaseolus vulgaris during Phase II.** *Following re-sowing after loss of the initial sowing, images show*  
 405 *the same plants on 10 January and 30 January 2026, from vegetative growth to flowering and early pod formation*  
 406 *under substrate conditions. Plant architecture and shoot development are consistent with low mechanical loading*  
 407 *and winter window-light conditions.*

408 **B. Role of Plants in Synthetic Pedogenesis and Cycle Closure**

409 Root penetration and rhizosphere development were observed alongside structural changes in the substrate.  
410 These observations coincide with localized variation in substrate conditions. Green manuring provided an internal,  
411 plant-derived organic input within the system.

412 Following plant establishment, root exudation and biomass turnover can act as internal carbon inputs for  
413 microbial processes, supporting system-level carbon cycling without reliance on external organic amendments. The  
414 successful establishment of a second plant generation following green manuring indicates system continuity beyond  
415 a single growth cycle under the tested conditions [23].

416 Importantly, these processes did not result in a conventional amendment-based system. Organic inputs remained  
417 internally generated and limited in scale, preserving the mineral-dominated character of the substrate within the  
418 system-level framework.

419 **C. System Stability and Engineered Control**

420 The absence of system collapse, extreme pH drift, or structural failure highlights the importance of engineered  
421 control variables within the Biorestorator framework. Controlled water availability, mineral composition, and biochar  
422 scaffolding created a bounded operational space in which biological processes could proceed without destabilizing  
423 feedbacks. This controlled yet biologically active state differentiates synthetic pedogenesis from both uncontrolled  
424 natural succession and conventionally managed growth substrates lacking dynamic biological feedbacks [14].

425 While only a single biochar fraction (~600 °C) was used in the illustrative study, a laboratory characterization  
426 was performed on a comparable biochar produced at ~500 °C, indicating the presence of a substantial mineral ash  
427 fraction and associated nutrient content. While not identical to the material used in the experiment, this analysis  
428 provides contextual information on the potential nutrient pool of biochar-associated mineral fractions.

429 The complementary functions of low- and high-temperature biochars in balancing early activation and long-term  
430 structural stability are therefore proposed at the conceptual system level rather than resolved experimentally in this  
431 study [9].

432 The observed system stability should be interpreted as a qualitative outcome under the tested configuration rather  
433 than as evidence of specific controlling mechanisms. However, Phase I conditions (pre-vegetation incubation)  
434 provide an internal temporal reference for subsequent observations in Phase II, allowing limited qualitative  
435 assessment of system stability across experimental phases. No substantial qualitative changes in pH or semi-  
436 quantitative colorimetric iron response were observed between these phases, consistent with stable physicochemical  
437 conditions over time, without implying specific underlying processes.

438 Literature reports indicate that untreated mineral substrates or early-stage systems lacking developed organic  
439 matter typically exhibit limited nutrient accessibility and constrained plant establishment without external inputs  
440 [4,5,10].

441 **D. Scope and Limitations**

442 The study was intentionally limited in scope and designed as an exploratory, system-level assessment of the  
443 Biorestorator framework under a defined configuration. The framework is inherently defined at the system level,  
444 integrating multiple interacting components and sequential developmental phases, and is therefore not readily  
445 reducible to single-variable experimental validation. Accordingly, the objective of this study was not to isolate  
446 individual mechanisms or quantify performance, but to examine whether the integrated system can sustain  
447 functional continuity under defined conditions.

448 Within this scope, the absence of control treatments, statistical hypothesis testing, and comparative experimental  
449 design reflects the exploratory nature of the study rather than an omission in experimental design. Consequently, the  
450 results should not be interpreted as indicators of agronomic productivity, system efficiency, or validated functional  
451 relationships. No uninoculated control treatment was included, and therefore causal attribution of observed  
452 outcomes to specific system components is not possible.

453 The study provides an initial feasibility-oriented reference point for the Biorestorator framework rather than a  
454 component-resolved or mechanistic evaluation. A rigorous evaluation of individual components or phase-specific  
455 contributions would require a structured experimental design with multiple control and comparison treatments  
456 across different stages of system development, which was beyond the scope of the present work.

457 Key aspects not addressed in this study—including microbial community composition, nutrient fluxes, and long-  
458 term dynamics—represent priorities for future work aimed at evaluating system performance, robustness, and  
459 applicability across different mineral substrates and operational contexts.

## 460 E. Implications for Restoration and Bioregenerative Systems

461 Within the limited scope of this study, the observations indicate a possible conceptual pathway for initiating soil-  
462 like functionality in environments where restoration approaches based on secondary succession may be ineffective.  
463 By framing soil formation as a staged and partially controllable process, the Biorestorer concept provides a  
464 conceptual link between ecological succession theory and engineered bioregenerative systems [3].

465

## VI. Conclusions

466 This work presents Biorestorer as a systems-based framework for synthetic pedogenesis, offering a novel  
467 perspective on soil formation as an engineered process. The framework is based on mineral components forming a  
468 soilless substrate combined with a carbon scaffold and targeted microbial activation within a structured, staged  
469 system design, representing a configuration with potential applicability in terrestrial restoration and in situ resource  
470 utilization (ISRU) contexts. By reframing soil not as a material input but as an emergent functional interface guided  
471 through staged system design, the framework integrates interacting biological and mineral components within a  
472 structured succession-based approach. Within this context, pedogenetic processes are treated as contributing  
473 elements of system development rather than primary objects of validation, aligning the framework with established  
474 concepts of ecological succession while extending them into engineered environments.

475 The illustrative study was designed to examine whether the integrated system can sustain observable plant  
476 growth under a defined configuration. Within this scope, plant establishment and continued growth were observed  
477 under mineral-dominated conditions without external organic inputs. Substrate stability, sustained growth without  
478 compost or fertilizers, and the emergence of a second plant generation are consistent with continued plant–substrate  
479 interaction under the tested conditions.

480 The use of waste-derived, ash-rich biochar in the illustrative study further highlights the adaptability of the  
481 Biorestorer framework to suboptimal and heterogeneous material inputs. Rather than relying on idealized or highly  
482 processed substrates, the framework is compatible with circular resource use, where locally available minerals and  
483 waste-derived carbon materials can be integrated as functional system components. This flexibility suggests  
484 conceptual relevance for resource-constrained environments and indicates possible future applicability in degraded  
485 terrestrial systems as well as extraterrestrial settings, where ISRU principles may be relevant.

486 Notably, the successful establishment of a second plant generation (*Phaseolus vulgaris*), a cultivated species  
487 rather than a classical pioneer plant, indicates that plant establishment was not limited to a single species within the  
488 tested configuration. This observation should be interpreted cautiously and does not imply agronomic performance  
489 or nutrient sufficiency under field conditions.

490 In the absence of control treatments and quantitative analyses, these findings remain qualitative and do not allow  
491 attribution to specific processes or mechanisms. The study therefore does not aim to validate underlying  
492 mechanisms, but to assess whether the framework is operationally feasible at the system level. As such, it is not  
493 structured as a comparative or hypothesis-testing experiment.

494 Taken together, the observations provide an initial system-level reference point for the Biorestorer framework  
495 and are consistent with its conceptual viability under the tested conditions, while identifying clear priorities for  
496 further controlled investigation. Future work will focus on quantitative characterization, inclusion of control  
497 treatments, and evaluation of longer-term system dynamics to assess performance, scalability, and applicability in  
498 both terrestrial restoration and bioregenerative life-support contexts.

499

## 500 Declaration on the Use of Generative AI and AI-Assisted Technologies

501 During the preparation of this manuscript, the author used generative AI tools for language editing, stylistic  
502 refinement, structural organization, and clarification of terminology. AI-assisted tools were not used to generate  
503 original scientific concepts, experimental data, analyses, interpretations, or conclusions. All scientific content,  
504 methodological decisions, data, interpretations, and final conclusions were developed, reviewed, and approved by  
505 the author, who takes full responsibility for the integrity and accuracy of the manuscript.

506

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## Výsledok

Matrica: PEVNÁ VZORKA

Názov vzorky

BIOCHAR-KBRO-ZZ-500

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Číslo vzorky

PR24D1475001

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Dátum odberu/čas odberu

22.10.2024

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Parameter	Kód metódy	LOQ	Jednotka	Výsledok	NM	Výsledok	NM	Výsledok	NM
<b>Ostatné</b>									
Neštandardný	I-ANNEX-WET	-	-	výsledky v prílohe	---	---	---	---	---
<b>Fyzikálne parametre</b>									
Merná hmotnosť	S-SGRAV-GR	0.01	g/cm3	2.01	± 10.0%	---	---	---	---
pH (H2O)	S-PHH2O-ELE	1.0	-	8.9	± 1.7%	---	---	---	---
Popol bezvodý A(d) pri 815 °C	I-ASH815GRS	0.10	% suš.	42.5	± 5.0%	---	---	---	---
Popol pôvodný A(ar) pri 815 °C	I-ASH815GRS	0.10	%	40.8	± 5.0%	---	---	---	---
Voda analytická M(ad)	I-WA-GR	0.50	%	2.32	± 24.6%	---	---	---	---
Voda celková M(ar)	I-WT-CC	0.50	%	3.88	---	---	---	---	---
Voda hrubá M(ex)	I-WG-GR	0.50	%	1.61	± 28.8%	---	---	---	---
Sušina pri 105 °C	S-DRY-GRCI	0.10	%	97.0	± 5.0%	---	---	---	---
<b>Anorganické parametre</b>									
Celkový organický uhlík (TOC)	S-TOC-TC-IR	0.10	% suš.	34.2	± 20.0%	---	---	---	---
Celkový uhlík	S-TOC-TC-IR	0.010	% suš.	36.0	± 16.2%	---	---	---	---
Dusík pôvodný N(ar)	I-ELEM-TCDS	0.10	%	2.41	± 15.2%	---	---	---	---
Dusíkaté látky v sušine	I-ELEM-TCDS	0.10	% suš.	2.50	± 15.2%	---	---	---	---
Kyslík bezvodý O(d)	I-ELEM-TCDS	5.0	% suš.	16.2	---	---	---	---	---
Kyslík pôvodný O(ar)	I-ELEM-TCDS	5.0	%	15.6	---	---	---	---	---
Síra spaliteľná bezvodá S(d)	I-ELEM-TCDS	0.10	% suš.	0.24	± 31.5%	---	---	---	---
Síra spaliteľná pôvodná S(ar)	I-ELEM-TCDS	0.10	%	0.23	± 32.5%	---	---	---	---
Celkový anorganický uhlík (TIC)	S-TOC-TC-IR	0.010	% suš.	1.80	± 15.0%	---	---	---	---
Uhlíčitany	S-TOC-TC-IR	0.050	% suš.	8.98	± 15.0%	---	---	---	---
<b>Celkové kovy / Hlavné kationy</b>									
Ca	I-CA-ICPS	0.000010	% suš.	10.3	± 20.0%	---	---	---	---
Mg	I-MG-ICPS	0.0000050	% suš.	0.464	± 20.0%	---	---	---	---
S	I-S-ICPS	5.0	mg/kg suš.	1430	± 20.0%	---	---	---	---
K ako K2O	I-K-ICPS	0.00060	% suš.	1.83	± 20.0%	---	---	---	---
P ako P2O5	I-P-ICPS	0.0023	% suš.	2.44	± 20.0%	---	---	---	---
<b>extrahovateľné kovy / hlavné kationy</b>									
Ag	S-METAXHB1	0.50	mg/kg	<0.50	---	---	---	---	---
As	S-METAXHB1	0.50	mg/kg	0.76	± 20.0%	---	---	---	---
Ba	S-METAXHB1	0.20	mg/kg	90.3	± 20.0%	---	---	---	---
Be	S-METAXHB1	0.010	mg/kg	0.143	± 20.0%	---	---	---	---
Cd	S-METAXHB1	0.40	mg/kg	<0.40	---	---	---	---	---
Celkový fosfor	S-METAXHB1	5.0	mg/kg	15100	± 20.0%	---	---	---	---
Co	S-METAXHB1	0.20	mg/kg	3.81	± 20.0%	---	---	---	---
Cr	S-METAXHB1	0.50	mg/kg	33.7	± 20.0%	---	---	---	---
Cu	S-METAXHB1	1.0	mg/kg	62.4	± 20.0%	---	---	---	---
Fe	S-METAXHB1	10	mg/kg	5310	± 20.0%	---	---	---	---
Hg	S-METAXHB1	0.20	mg/kg	<0.20	---	---	---	---	---
Li	S-METAXHB1	1.0	mg/kg	9.9	± 20.0%	---	---	---	---
Mn	S-METAXHB1	0.50	mg/kg	209	± 20.0%	---	---	---	---
Mo	S-METAXHB1	0.40	mg/kg	1.14	± 20.0%	---	---	---	---
Ni	S-METAXHB1	1.0	mg/kg	15.2	± 20.0%	---	---	---	---
Pb	S-METAXHB1	1.0	mg/kg	9.2	± 20.0%	---	---	---	---
Sb	S-METAXHB1	0.50	mg/kg	<0.50	---	---	---	---	---
Sn	S-METAXHB1	1.0	mg/kg	3.1	± 20.0%	---	---	---	---
Sr	S-METAXHB1	0.10	mg/kg	104	± 20.0%	---	---	---	---
Tl	S-METAXHB1	0.50	mg/kg	<0.50	---	---	---	---	---
V	S-METAXHB1	0.10	mg/kg	5.71	± 20.0%	---	---	---	---
Zn	S-METAXHB1	3.0	mg/kg	173	± 20.0%	---	---	---	---
<b>Polycyklické aromatické uhľovodíky (PAU)</b>									
Naftalén	S-PAHGMS05	0.010	mg/kg suš.	1.12	± 30.0%	---	---	---	---
Acenaftylén	S-PAHGMS05	0.010	mg/kg suš.	<0.130	---	---	---	---	---