

Sex differential effects of developmental heat stress on life-history and reproductive traits

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1 Abstract

2 Global warming has led to increased mean global temperatures with projections suggesting
3 continued warming throughout this century, posing an escalating threat to biological systems
4 worldwide. Ectotherms are most vulnerable to this change as heat stress conditions can have
5 severe implications on their development, mating interactions, and fitness. However, the sex-
6 specific effects of developmental temperature on these factors remain poorly understood. Here,
7 we conducted an experiment using the decorated cricket, *Gryllobates sigillatus*, to disentangle
8 sex-specific effects of developmental temperature on life-history, pre- and post-copulatory and
9 fitness-related traits. Crickets were reared at two temperatures: a control temperature (27 °C)
10 and a heat-stress temperature (36 °C) and their development was monitored. Post sexual
11 maturation, mating assays were conducted in a full-factorial mating design, and after successful
12 fertilization, reproductive output was quantified. We found that heat stress accelerated
13 development to adulthood differentially in both sexes. Furthermore, we found that female
14 developmental temperature affected mating latency, with heat-stressed females exhibiting
15 longer latencies, suggesting that females exert control over this trait. Conversely, male
16 developmental temperature affected courtship and copulation success. Heat-stressed males
17 were less willing to initiate courtship and largely unsuccessful at transferring the
18 spermatophore during mating. Post-copulation, heat stressed females took longer to feed on the
19 nuptial gift. Moreover, the feeding time was also significantly predicted by the wet mass of
20 both males and females, suggesting that gift consumption is driven by a combination of male-
21 provided quality and the female's own energetic requirements. Lastly, we found that heat-
22 stressed females exhibited significantly reduced fecundity and fertility, highlighting

23 downstream reproductive costs of developmental thermal stress. Thus, our study highlights
24 how developmental temperature alters life history, fertility, and can have sex-specific impact
25 on shared mating traits.

26 **Keywords:** developmental thermal stress, mating behaviour, thermal plasticity, sex-specific
27 effects, climate change

28 1. Introduction

29 Global warming has been leading to greater temperature variation and prolonged heatwaves.
30 Ectotherms depend on the environmental temperature to regulate their body temperatures and
31 are thus sensitive to external thermal changes. External thermal conditions influence various
32 traits in ectotherms, like morphology, life history, behaviour, and physiology (Angilletta et al.,
33 2003; Schulte et al., 2015; Audzijonyte et al., 2016; Abram et al., 2017). Ectotherms function
34 within a permissive temperature range, where physiological and behavioural processes
35 maintain homeostasis efficiently, and once a specific critical temperature is exceeded, they
36 enter a stressful temperature range in which disruptive processes (such as cellular damage and
37 metabolic imbalances) overtake the physiological processes that maintain homeostasis (Ørsted
38 et al., 2022). Changes in ectotherm physiology due to environmental temperature ultimately
39 affect life-history, morphological, behavioural, and reproductive traits (Kim et al., 2017;
40 Ohlberger, 2017; Porcelli et al., 2017; Michelangeli et al., 2018). All of these together
41 contribute to an individual's fitness and capacity to survive in variable environments. As heat
42 stress has cumulative effects with exposure time within a stressful temperature range, it is
43 important to understand how long-term heat stress impacts these traits.

44 Thermal stress during development can lead to lifelong alterations in insect morphology,
45 metabolism and resource allocation (Leith et al. 2021; Neven, 2000). Changes in resource
46 allocation, along with metabolic changes, may alter the rate at which insects develop into
47 adults. Previous studies with different insects have shown that development rate is affected by
48 changes in temperature (Garrad et al., 2015; Kong et al., 2024). Faster development may lead
49 to a shift in energy investment, with a focus on maturing faster at the expense of adult nutrient
50 reserve accumulation. According to Atkinson (1994), the temperature-size rule states that, for
51 most ectotherms, an increase in temperature during development results in a reduction in body
52 size, while a decline in temperature leads to an increase in adult body size. The relationship
53 between the temperature at which organisms develop and their size is most thoroughly

54 understood in arthropods (Hirst et al., 2015, Horne et al., 2015), especially in insects of the
55 order Diptera (French et al., 1998). However, in some insect species, including the cricket
56 *Acanthogryllus asiaticus* and the Asian lady beetle *Harmonia axyridis*, individuals reared at
57 high temperatures have been found to be heavier than those reared at low temperatures (Singh
58 et al., 2020; Knapp & Nedvěd, 2013). Thus, the extent to which the temperature-size rule
59 applies across other insect orders remains understudied.

60 The ability to reproduce may be hindered by poor-quality gametes resulting from heat stress
61 (Porcelli et al., 2017; Gruntenko et al., 2003), with thermal effects on fertility manifesting well
62 before survival is compromised. Research has shown that the thermal fertility limit is reached
63 long before the lethal thermal limit, underscoring that reproductive failure is among the earliest
64 and most ecologically consequential costs of heat stress (Parratt et al., 2021; van Heerwaarden
65 and Sgrò, 2021). Heat stress significantly impairs male fertility through reduced sperm
66 production, decreased sperm motility, morphological sperm abnormalities, and reduced sperm
67 density (Hansen, 2009; Nguyen et al., 2013; Hurley et al., 2018; Sales et al., 2018; Domenech
68 and Fricke, 2023), and once a critical temperature threshold is crossed, males may become
69 entirely infertile (Araripe et al., 2004; Sales et al., 2021). Female reproductive efficiency can
70 be equally vulnerable. Heat stress is known to cause oocyte degeneration, defective
71 vitellogenesis, and reduced ovary size which can collectively result in decreased fecundity
72 (Krebs and Loeschke, 1994; Gandara and Barbosa, 2022; Kirk Green et al., 2019). Since female
73 fecundity and fertility represents the primary bottleneck through which thermal stress can
74 translate into reduced population reproductive output, understanding how developmental
75 temperature specifically shapes female reproduction is essential for assessing the full
76 reproductive cost of climate warming in ectotherms.

77 Reproductive success under heat stress may, in addition, be influenced by mating interactions.
78 Mating interactions include mate attraction, courtship display, copulation, and post-mating
79 behaviours (Ombuya et al., 2025). These mating interactions can be strongly influenced by
80 both developmental temperature and environmental conditions (Leith et al., 2021; Macchiano
81 et al., 2023). Several studies have demonstrated how temperature influences various aspects of
82 mating interactions in several species (Patton and Krebs, 2001; Augustin et al., 2022; Katsuki
83 and Miyatake, 2009; Bhaisare and Chaudhary, 2023; Smith et al., 2025). The role of these
84 behaviours is highly significant when determining the reproductive outcome, which directly
85 influences reproductive fitness. Although many researchers have studied the effects of
86 temperature on reproductive traits, sex-specific consequences of thermal stress on mating

87 behaviour remain comparatively understudied, with males and females rarely examined
88 independently within the same experimental framework (Dougherty et al., 2023; Reinhold and
89 Rizvi, 2026). Since mating interactions are shaped by contributions from both sexes, explicitly
90 disentangling the effects of temperature on each partner's behavioural contribution is essential
91 for understanding how heat stress impacts matings to ultimately shape reproductive fitness.

92 The decorated cricket, *Grylloides sigillatus*, is a cosmopolitan species that serves as a model
93 organism for studying thermal effects due to its tropical and subtropical distribution and the
94 ease with which mating behaviours can be observed. The species is typically found in the
95 temperature range of 20 °C to 30 °C (Smith and Thomas, 1988). During their development,
96 these crickets pass through seven nymphal instar stages, after which they undergo the final
97 moult and become adults (Kong et al., 2025). In *Grylloides sigillatus*, mating behaviour starts
98 with the male producing a distant calling song to attract the female. As the female gets closer,
99 this calling song changes into a softer courtship song. If interested, the female mounts the male,
100 and the male then initiates copulation (Figure 1). While copulating, the male transfers a
101 spermatophore, which is composed of two parts: a gelatinous spermatophylax and a sperm
102 containing ampulla. The female removes the spermatophylax and starts feeding on it
103 immediately after mating, while sperm transfer occurs via the ampulla (Sakaluk, 1984). At this
104 post-copulatory stage, the male guards the female (Figure 1). When the female finishes
105 consuming the gift, she then bends to retrieve the ampulla left behind in the genital papilla and
106 feeds on it. We selected *Grylloides sigillatus* as an appropriate model species for investigating
107 the influence of temperature on several attributes for both sexes, given its well-characterized
108 life history and its relevance as a subtropical ectotherm likely to experience pronounced
109 developmental sensitivity to elevated temperatures.

110 In this study, we investigate the sex-specific effects of developmental heat stress on life-history,
111 morphology, mating traits and fertility in *Grylloides sigillatus*. We reared crickets at two
112 temperatures: a heat-stress treatment (36 °C; Kong et al., 2024) and a control treatment (27 °C;
113 Rizvi et al., 2025) throughout development. We recorded the time to adulthood and the body
114 mass of adults. Following sexual maturation, we conducted no-choice mating trials to
115 disentangle sex-specific developmental temperature effects on pre- and post-copulatory mating
116 interactions. To test the effect of developmental temperature on reproductive output, female
117 fecundity and hatching success was assessed after copulation. We expected the development to
118 be faster and body mass to increase under the heat-stress treatment, as observed in previous
119 studies (Kong et al., 2024). We also hypothesized that the temperature treatment for the males

120 would affect the courtship behaviour and copulation time, while the female temperature
121 treatment would affect traits such as mating latency, mating success, nuptial gift feeding time,
122 and fecundity. Furthermore, we hypothesized that male and female temperature treatments
123 might interact in order to influence some shared mating traits (such as mating latency and
124 mating success) and the reproductive output. Our study provides evidence on sex-specific
125 developmental responses to thermal stress, offering critical insights into how organisms
126 navigate the reproductive challenges imposed by rapid climate change.

127 **2. Methods**

128 **2.1. Animal Rearing**

129 To disentangle the effects of developmental temperature on mating behaviour and fertility, two
130 experimental treatments, one of high-temperature (36 °C) individuals and another of control-
131 temperature (27 °C) individuals, were developed. Laboratory population of *Grylloides sigillatus*
132 originated from a stock of approximately 200 individuals (adults and subadults) sourced from
133 an insect breeder firm in April 2022 (ReptilienKosmos, Germany). Crickets were maintained
134 in a laboratory room at 27 ± 1 °C on a 12:12 light: dark cycle. They were provided with ad
135 libitum food (Nekton cricket breeding concentrate), water in a glass vial (22 ml) plugged with
136 cotton wool, and egg cartons for shelter. Individuals for the experiment were sourced from the
137 12th generation of the lab population. Plastic cups filled with moist cotton wool were placed
138 inside the population boxes to collect eggs for 1 week. Eggs were then counted and placed in
139 separate incubators: one at 27 °C with a 12L:12D photoperiod and the other at 36 °C with a
140 12L:12D photoperiod to develop the treatments.

141 Once eggs started hatching, the number of hatchlings was counted. The hatchlings were placed
142 into separate boxes (16.9 cm × 10.5 cm × 7.4 cm) with egg carton pieces, a vial of water (13mL)
143 with cotton plug, and some food (Nekton cricket breeding concentrate). The hatchlings were
144 then placed under the same environmental conditions in which they had hatched to complete
145 further development. In order to maintain virgin populations, male and female nymphs were
146 segregated as the ovipositor started to become visible in the females. Separation was conducted
147 by administering a 1-minute carbon dioxide (CO₂) shock to knock out the crickets, then
148 transferring them to segregated boxes (16.9 cm × 10.5 cm × 7.4 cm).

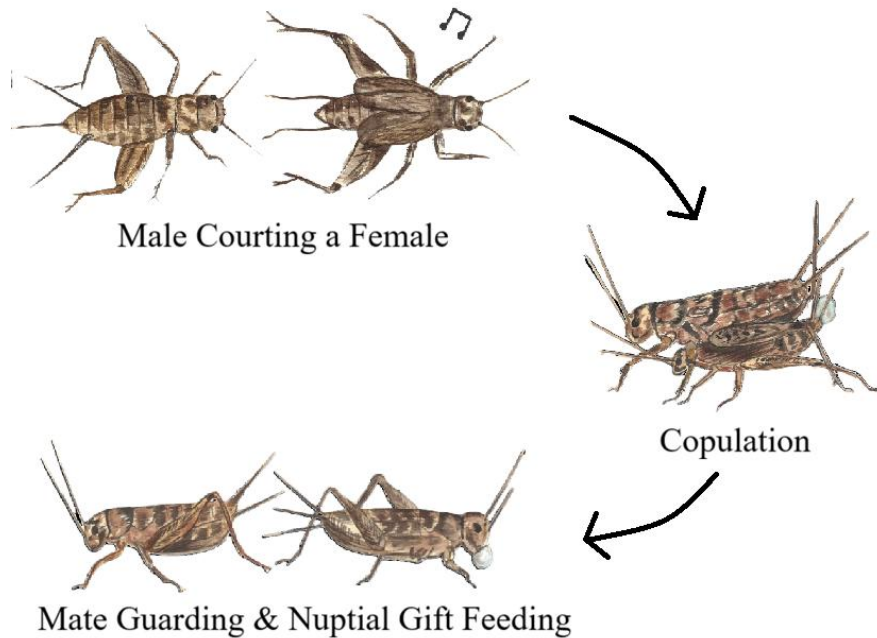
149 Each box contained between 3 to 6 individuals at a time. Development of a total of 300
150 individuals (27°C males: 90, 27°C females: 81, 36°C males: 61, 36°C females: 61) was
151 monitored. Once the males had moulted into adults, they were marked individually by placing
152 a small dot of paint marker on their abdomen to identify them during mating trials. Post
153 marking, the males were placed in a measuring cup, and their weights were measured using an
154 electronic balance (Kern 770). Females were not marked, but were measured just before the
155 mating trials and then placed directly into the mating arena.

156 **2.2. Mating Trials**

157 Mating trials were conducted in a full factorial design (Figure 2) and at a constant
158 environmental temperature of 27 °C. Individuals reared at 36 °C were given up to 1 hour to
159 acclimatize to the ambient temperature of 27 °C before the mating trials began. Additionally, a
160 subset of heat stressed males and females (reared at 36 °C) were mated at an ambient
161 temperature of 36 °C to tease apart the effects of developmental temperature and environmental
162 temperature during mating. This allowed us to assess whether behavioural responses observed
163 in heat-stressed individuals reflected lasting developmental effects or were instead modulated
164 by the ambient thermal conditions experienced during the trial itself. The females from the
165 treatment boxes were knocked down using carbon dioxide, weighed, and then placed
166 individually in mating arenas (16.9 cm x 10.5 cm x 7.4 cm) with an egg carton piece. The
167 marked males were individually placed in different cups after knocking them down with carbon
168 dioxide. All individuals were then transferred to a dark room with red light, as crickets are
169 insensitive to red light. A marked male was randomly selected and introduced into the mating
170 arena. Once the male started singing, the stopwatch (ATP digital stopwatch TIM1166) was
171 started, and the time taken for the female to mount the male after the first singing was recorded
172 as a measure of mating latency. Immediately after the first mounting, the stopwatch was
173 resumed to measure mating duration (the time the female remained mounted on the male until
174 the transfer of the spermatophore). The stopwatch was paused whenever the female dismounted
175 and resumed when she mounted again; these timestamps were recorded. Once sperm transfer
176 was complete, the stopwatch was resumed when the female began feeding on the nuptial gift
177 and stopped once she finished.

178 Each pair was given a maximum of 15 minutes to initiate mating, after which the partners were
179 changed. Partners were changed by replacing the focal male with a different-marked male in
180 the mating arena with the female. If an individual was unable to mate successfully despite

181 trying with five different partners, they were not used for any further trials. All mating trials
 182 were conducted at least 1 week after the final moulting, to ensure the individuals were sexually
 183 receptive.



184 **Figure 1:** Simplified illustration of mating system of *Gryllodes sigillatus*

185

		Male Rearing Temperature	
		27°C	36°C
Female Rearing Temperature	27°C		
	36°C		

186

187 **Figure 2:** Full-factorial mating crosses conducted at an ambient temperature of 27 °C

188 **2.3. Female Fecundity and Fertility Measurements**

189 After a mating trial was successfully completed, the male was removed from the arena to
 190 prevent further matings. The females were given a piece of egg carton, food (Nekton cricket

191 breeding concentrate), a water vial (13mL) with a cotton plug, and an egg pad (a cup containing
192 damp cotton wool) to serve as an oviposition substrate. All the females were placed in
193 incubators at 27 °C with a 12L:12D photoperiod for egg laying. Each female was kept in the
194 incubator for 3 weeks during egg-laying. The eggs were collected from the pads weekly and
195 counted under a stereo microscope (Leica MS5, 1.6x magnification), and the pads were then
196 replaced with fresh ones. The collected eggs were counted, covered with wet cotton, and placed
197 in separately labelled boxes (16.9 cm × 10.5 cm × 7.4 cm). They were then kept in the incubator
198 for hatching at an ambient temperature of 27 °C and a photoperiod of 12L:12D. Only the eggs
199 laid by the females during the first two weeks were incubated. Eggs laid in the third week were
200 counted, then frozen overnight and discarded. Once hatchlings emerged, they were counted,
201 frozen, and discarded. Each box containing eggs was kept for up to 3 weeks to detect any signs
202 of hatching. If no hatchlings were obtained within 3 weeks, they were discarded. In boxes from
203 which hatchlings were obtained, the boxes were kept in the incubator for a period of 1 week
204 post the final hatching, after which the remaining eggs were discarded.

205 **2.4. Statistical Analyses**

206 **2.4.1. Development Time and Wet Mass**

207 Statistical analysis was performed in R version 4.5.2 (R Core Team 2025). Data processing and
208 the conversion of group-level observations to individual-level records were conducted using
209 *tidyverse* (Wickham et al., 2019), with duration calculated via *lubridate* (Grolemund and
210 Wickham, 2011). To analyze development speed, we conducted survival analysis using *survival*
211 (Therneau 2026) and *survminer* (Kassambara et al., 2025). Specifically, to resolve issues of
212 monotone likelihood and provide stable estimates for temperature effects, we used Firth's
213 Penalized Likelihood Cox Regression via the *coxphf* package (Heinze et al., 2023). This
214 allowed for the calculation of Hazard Ratios (HR) to compare the relative rate of adult
215 emergence between rearing temperatures and sexes. For adult wet mass, a linear model was
216 used to check for the effects of temperature, sex, and their interaction. The model was tested
217 using the *DHARMA* (Hartig, 2024) package to simulate residual diagnostics to assess normality,
218 homoscedasticity, and the presence of outliers. Post-hoc pairwise comparison for both
219 development time and wet mass was conducted using the *emmeans* package (Lenth and
220 Piaskowski, 2025). Data visualization was conducted using *ggplot2* (Wickham 2016),
221 *patchwork* (Pederson, 2025), *ggsignif* (Eltze et al., 2021), and *ggpubr* (Kassambara 2025).

222 2.4.2. Mating Trials and Reproductive Output

223 All statistical analyses were performed in R version 4.5.2 (R Core Team 2025), with packages
224 loaded via *pacman* (Rinker and Kurkiewicz, 2017) to ensure a streamlined workflow. The
225 analyses were based on 462 mating trials conducted across all experimental treatments. We fit
226 generalized linear mixed-effects models, linear mixed models using *lme4* (Bates et al., 2015)
227 or *glmmTMB* (Brooks et al., 2017), or Bayesian models using *brms* (Bürkner, 2017), depending
228 on the data and error structure. For all models (unless stated otherwise), male developmental
229 temperature, female developmental temperature, and their two-way interaction were included
230 as fixed effects, while individual ID was included as a random effect. Male and female wet
231 mass were included as covariates in all relevant analyses. All models were tested using the
232 *DHARMA* (Hartig, 2024) package to simulate residual diagnostics to assess normality,
233 homoscedasticity, and the presence of outliers. The effect of thermal stress on the probability
234 of male courtship was analysed using a Bayesian model with a Bernoulli distribution. Since
235 mating latency data (in seconds) were right-skewed, we applied a log transformation to
236 improve the normality of the residuals and stabilize the variances. For mating duration data (in
237 seconds), the data were rank-transformed prior to analysis to accommodate non-normality and
238 outliers. To test for correlation among some factors in each pair, a Pearson correlation test was
239 used.

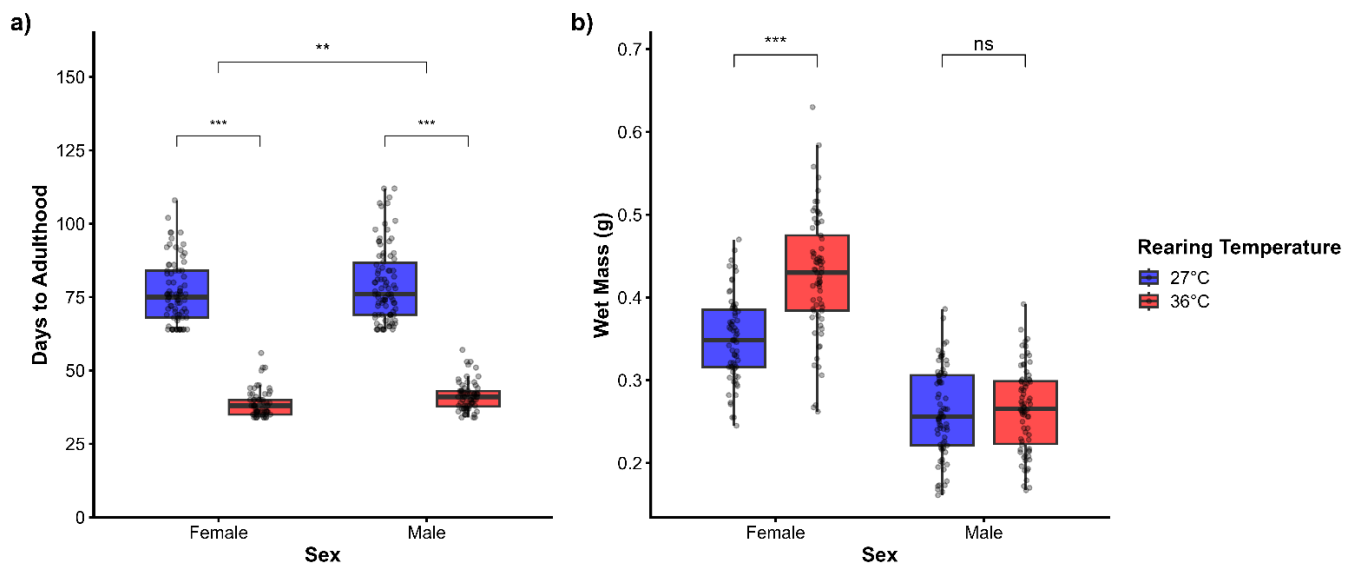
240 For post-copulatory traits like nuptial gift feeding duration, female fecundity and fertility, due
241 to insufficient data from the other two crosses as a result of high mating failure, only the 27 °C
242 males x 36 °C females and 27 °C males x 27 °C females crosses were considered. Feeding
243 duration was analysed using female developmental temperature as a predictor, with female and
244 male wet mass as covariates. As the individuals were not repeated, random effects were not
245 included in the model. Furthermore, using the *car* (Fox and Weisberg, 2019) package, Levene's
246 Test was conducted to assess homogeneity of variance across the two female treatments. We
247 fitted a negative binomial generalized linear model to analyse the number of eggs laid (in two
248 weeks), with female developmental temperature as the main predictor and scaled female wet
249 mass as covariate. We modelled egg hatching success for week 1, week 2, and the total of both
250 using beta-binomial generalized linear models fitted with *glmmTMB*. To test the effects of
251 environmental temperature during mating, we used data from the 36 °C male x 36°C female
252 crosses conducted at an ambient temperature of 36 °C and 27 °C. Data visualization was
253 performed using *ggplot2* (Wickham 2016), *ggsignif* (Eltze et al., 2021), *ggpattern* (FC and
254 Davis, 2026), *patchwork* (Pederson, 2025), and *ggpubr* (Kassambara 2025). Post hoc

255 comparisons of main level effects and interactions were performed using the function emmeans
256 in the ‘emmeans’ package (Lenth and Piaskowski, 2025).

257 3. Results

258 3.1. Effect of Rearing Temperature on Development Time and Wet Mass

259 Temperature significantly affected the time required for hatchlings to mature into adults.
260 Crickets reared at 36 °C developed nearly twice as fast as those at 27°C (Firth’s Cox PH: HR
261 = 1911.69, $p < 0.001$, Figure 3a, ST 1). The median development time was 39 days (95% CI:
262 38–40) at 36°C compared to 76 days (95% CI: 74–77) at 27°C. Sex also played a significant
263 role in developmental rate (HR = 0.72, $p < 0.01$, ST 1); males exhibited a slower developmental
264 pace than females at both temperature treatments. At 27°C, males reached adulthood at a
265 median of 76 days (95% CI: 74–80) compared to 75 days for females (95% CI: 72–77). At
266 36°C, this gap widened, with males requiring a median of 41 days (95% CI: 39–42) versus 38
267 days for females (95% CI: 36–40). Developmental temperature had a significant effect on
268 female wet mass ($\beta = 0.077$, SE = 0.009, $t = 7.82$, $p < 0.001$, Figure 3b, ST 2), with females
269 reared at 36°C being significantly heavier than those reared at 27°C. In contrast, developmental
270 temperature had no significant effect on male wet mass ($t = 0.372$, $p = 0.710$, Figure 3b, ST 2).

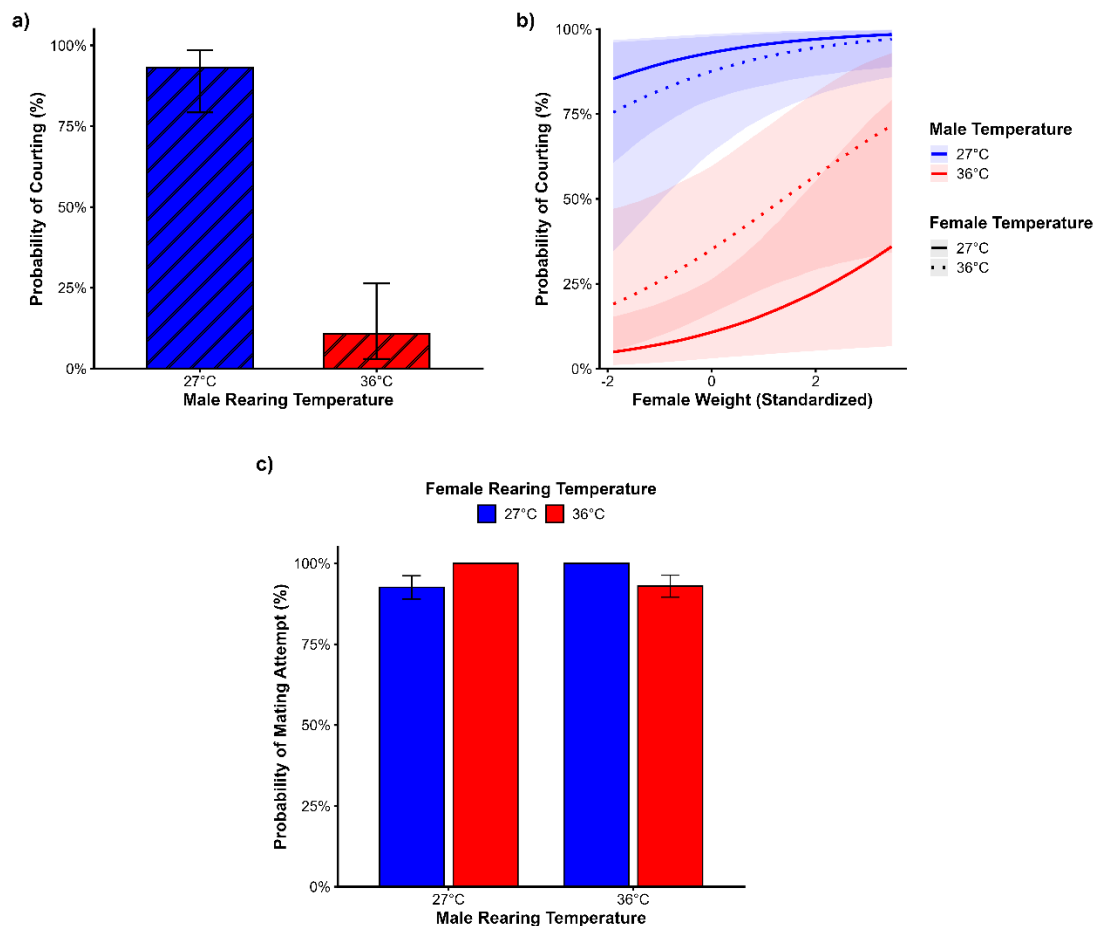


271

272 **Figure 1:** Effect of rearing temperature on (a) time taken to reach adulthood and (b) wet mass. Females
273 showed a significant increase in wet mass with higher developmental temperature. Horizontal bars
274 indicate the median.

275 3.2. Effect of Developmental Temperature on Courtship

276 Male developmental temperature had a strong negative effect on the probability of courting (β
 277 = 0.45, 95% credible interval [CI] = [-7.06, -2.99], Figure 4a), with males reared at 36 °C being
 278 less likely to court females than those reared at 27 °C. On the other hand, female wet mass had
 279 a positive effect on the probability of being courted ($\beta = 0.45$, 95% credible interval [CI] =
 280 [0.04, 0.87], Figure 4b), with females having higher wet mass being more likely to be courted
 281 by males. There was no significant effect of male wet mass, female developmental temperature,
 282 or the interaction between the two temperature treatments on courtship probability (ST 3).
 283 Additionally, male courtship initiation led to female mounting in 95.7% of cases across all
 284 treatment groups ($p < 0.001$). Fisher's exact test confirmed that the high probability of an
 285 attempt following courtship did not differ significantly across the four temperature treatments
 286 ($p = 0.14$, Figure 4c)



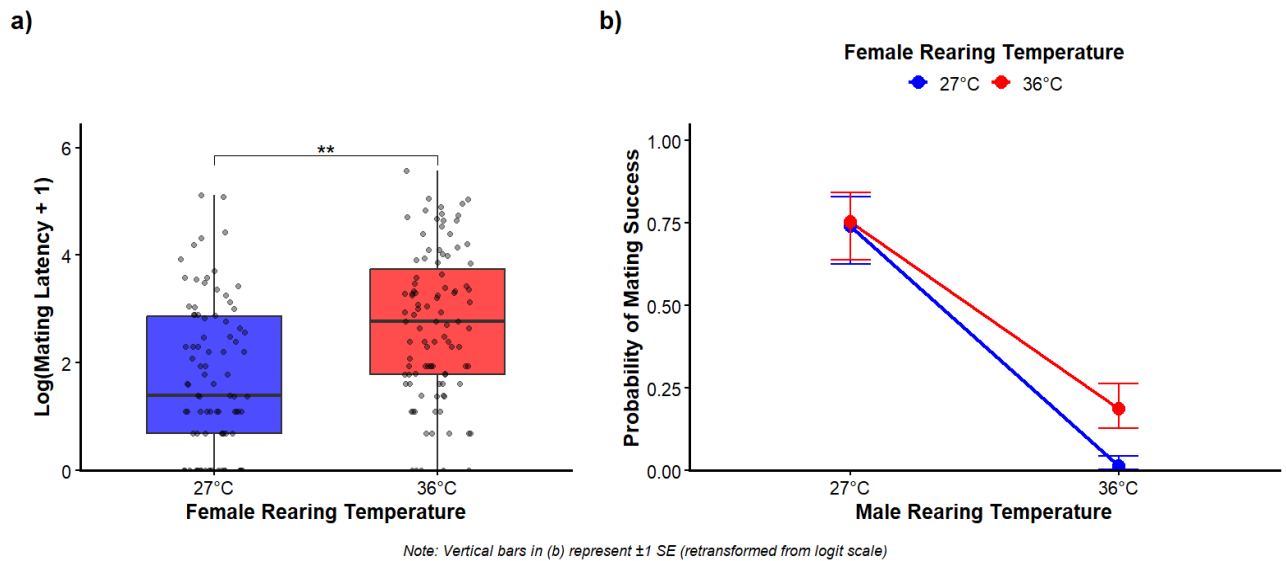
287

288 **Figure 2:** Effect of developmental temperature on courtship behaviour and mating attempt. (a) Males
 289 developed at 36 °C show a decreased probability of courting females as compared to those developed
 290 at 27 °C. Solid bars represent the posterior mean probability calculated by the Bayesian model. Vertical
 291 error bars represent the 95% Credible Intervals (CI), indicating the range with 95% probability in

292 *which the true population means reside. (b) Heavier females experience a higher probability of being*
293 *courted by males than lighter females within the 15-minutes observation period. (c) Probability of a*
294 *female mounting a male across all temperature treatments post-male courtship initiation. Vertical error*
295 *bars indicate ± 1 SE.*

296 **3.4. Effect of Developmental Temperature on Mating Latency and Mating** 297 **Success**

298 Mating latency was significantly affected by female temperature treatment ($\beta = 0.99$, SE =
299 0.36, $p < 0.01$, Figure 5a), with females reared at 36 °C exhibiting longer mating latencies
300 compared to those reared at 27 °C. Male and female wet mass, male developmental
301 temperature, and the interaction between the male and female treatments had no significant
302 effect on the mating latency (ST 4). Mating success was significantly influenced by male
303 developmental temperature ($\beta = - 5.26$, SE = 1.36, $p < 0.001$, Figure 5b), with males reared at
304 27 °C being much more successful in transferring a spermatophore to the female than those
305 reared at 36 °C. Although as a main effect female developmental temperature did not have a
306 significant effect on mating success ($p = 0.92$), a significant interaction was observed between
307 the male and female temperature treatments ($\beta = 2.66$, SE = 1.32, $p = 0.04$, Figure 5b), with
308 males reared at 36 °C being more successful in transferring spermatophore to females reared
309 at 36 °C, than with females reared at 27 °C. Male and female weights did not significantly
310 affect mating success (ST 5). There were also no detectable effects of any predictors on mating
311 duration.

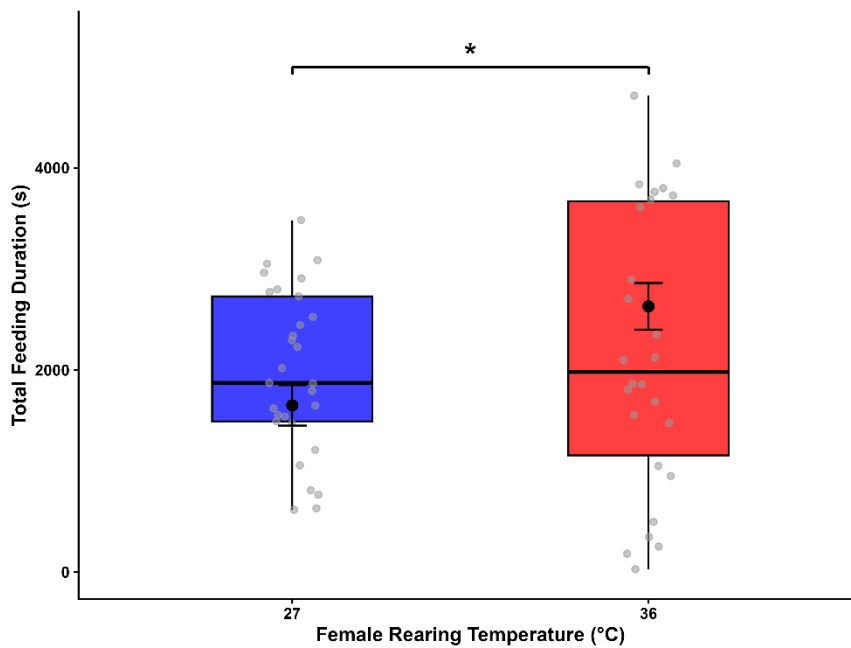


312

313 **Figure 3:** Effect of developmental temperature on mating latency and mating success. (a) Females
 314 reared at 36 °C exhibit longer mating latencies than those reared at 27 °C. Horizontal bars represent
 315 the median log-transformed mating latency. (b) Males developed at 36°C were more successful in
 316 mating with females developed at 36°C than those developed at 27°C. Overall, males developed at 36°C
 317 were less successful in sperm transfer than those developed at 27°C. Asymmetry between the lines
 318 represents an interaction effect.

319 3.5. Effect of Female Developmental Temperature on Nuptial Gift Feeding 320 Duration

321 Female developmental temperature significantly influenced feeding duration, with females
 322 reared at 36°C taking longer to feed on the nuptial gift than those reared at 27°C ($\beta = 1004.90$,
 323 $SE = 404.10$, $t = 2.49$, $p = 0.016$, Figure 6). Furthermore, feeding duration was significantly
 324 affected by both female and male mass. Female wet mass was a significant negative predictor
 325 ($\beta = -6.47$, $SE = 2.60$, $t = -2.49$, $p = 0.016$, S1), whereas male wet mass was a significant
 326 positive predictor ($\beta = 7.91$, $SE = 2.91$, $t = 2.71$, $p = 0.009$, S1). A post-hoc Levene's test
 327 indicated that the variance in feeding behaviour was also significantly affected by temperature
 328 ($F_{1,53} = 7.88$, $p < 0.01$) as females raised at 36 °C showed greater variance in their feeding
 329 duration than those raised at 27 °C.

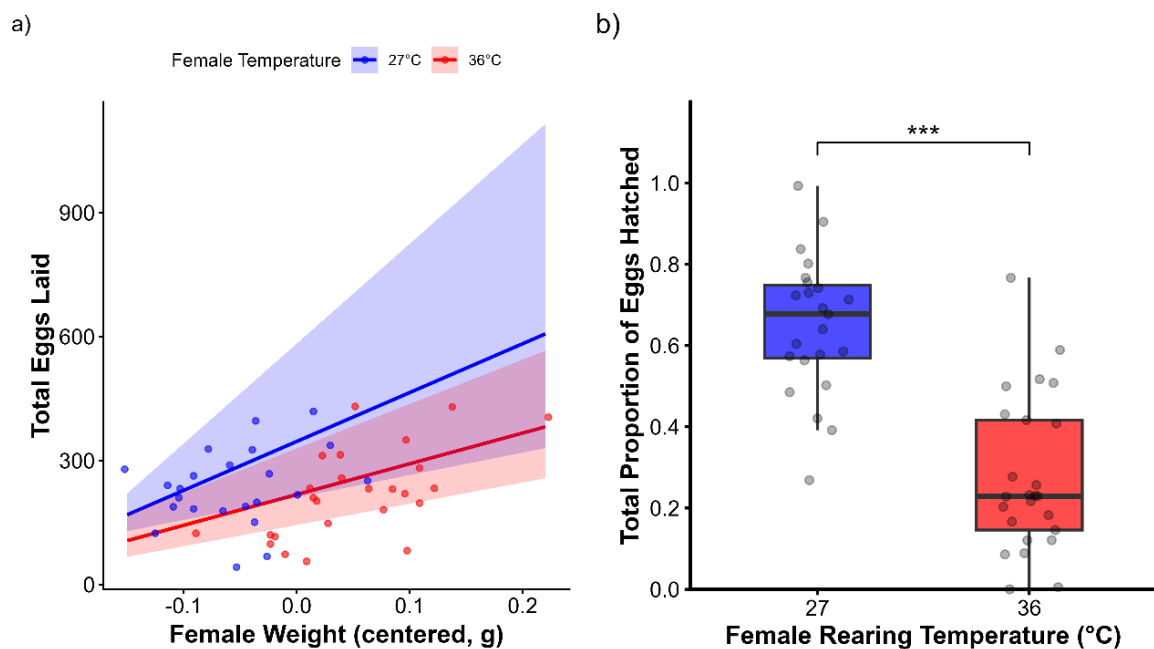


330

331 **Figure 6:** Total nuptial-gift feeding duration between female developmental temperature treatments
 332 ($n=55$). Plot shows that heat stressed females took longer to feed than control ones. Boxplots show the
 333 interquartile range and median, grey points represent raw data and black points with error bars indicate
 334 model adjusted means with 95% CI.

335 3.6. Effect of Female Developmental Temperature on Fecundity and Fertility

336 Developmental temperature significantly affected female fecundity, with females reared at
 337 36°C laying fewer eggs than control females reared at 27°C ($\beta = -0.46$, $z = -2.71$, $p = 0.006$,
 338 Figure 7a). Female wet mass was also a significant positive predictor of fecundity ($\beta = 3.46$, z
 339 $= 3.21$, $p = 0.001$, Figure 7a), indicating that heavier females produced more eggs. Female
 340 developmental temperature also had a significant effect on fertility ($\beta = -1.23$, $SE = 0.32$, $p <$
 341 0.001 , Figure 7b, ST 6), with females reared at 27 °C having greater fertility than those reared
 342 at 36 °C. Additionally, eggs laid in the first and second week post-mating by females reared at
 343 27 °C had a higher success of hatching (Week 1: $\beta = -1.05$, $SE = 0.41$, $p < 0.01$, S2, ST 6; Week
 344 2: $\beta = -1.32$, $SE = 0.42$, $p < 0.01$, S2, ST 6) compared to eggs laid by females reared at 36 °C.



345

346 **Figure 7:** Effect of developmental temperature on female reproductive output (n=49). a) Plot indicates
 347 a significant negative effect of female rearing temperature on fecundity, independent of the observed
 348 variation in female body weight. The regression lines and shaded 95% confidence intervals are derived
 349 from a negative binomial generalized linear model, showing predicted egg counts as a function of
 350 female body weight b) Females reared at 27 °C have greater fertility than those reared at 36 °C when
 351 successfully mated with control males. Horizontal bars indicate the median proportion of eggs hatched,
 352 and boxes indicate the interquartile range.

353 3.7. Effect of Environmental Temperature on Mating Behaviour

354 Mating trial temperature had no detectable effects on probability of male courtship, mating
 355 latency, probability of mating attempt, mating duration and copulation success in heat stressed
 356 individuals (reared at 36 °C). The only two traits where we detected a significant effect of trial
 357 temperature were number of mating attempts and nuptial gift feeding duration. There was an
 358 increased mating attempt frequency at 36°C compared to 27°C ($z = 2.56$, $p = 0.01$, S3). Neither
 359 male nor female wet mass significantly influenced the number of mating attempts (ST 7).
 360 Females mated at 36 °C had shorter feeding duration than those mated at 27 °C ($t_{15.2} = -2.43$, p
 361 $= 0.03$, S4). Additionally, male wet mass positively affected feeding duration ($\beta = 536.39$, $p =$
 362 0.03 , S5), while female weight had no detectable effect (ST 8).

363 4. Discussion

364 In this study, we investigated the sex-specific effects of developmental temperature on life-
365 history traits, morphological traits, mating behaviour, and reproductive output in *Gryllodes*
366 *sigillatus*. We found that both male and female rearing temperatures affected development rate
367 and mating behavioural traits differentially. Our results also show that developmental
368 temperature affected body mass and reproductive output of females. Lastly, we could tease
369 apart the effects of developmental and environmental temperature on mating behaviour in heat
370 stressed crickets, showing that environmental temperature can also alter certain mating traits.

371 4.1. Effect of Developmental Temperature on Life History and 372 Adult Morphology

373 Developmental temperature strongly influenced life-history and body mass trajectories in
374 *Gryllodes sigillatus*. The crickets developed faster when reared at 36 °C than at 27 °C, a
375 common pattern among many ectothermic organisms (Lee et al., 2013; Ruthsatz et al., 2018;
376 Ciordia et al., 1963; Lazo and Pita, 2012; Paul, 1998; Kong et al., 2024). Additionally, across
377 both temperature treatments, females developed faster than males. This pattern aligns with
378 broader evidence for sex-specific developmental scaling in insects, where females prioritize
379 faster maturation to enhance reproductive opportunity (Degen et al., 2015). Thermal
380 performance curves (TPCs) show how rates of biological processes increase exponentially with
381 temperature up to a critical thermal optimum, beyond which performance rapidly declines,
382 highlighting how an ectotherm's body temperature affects its performance and fitness (Sinclair
383 et al., 2016). The accelerated development at 36 °C captures the ascending phase of the curve,
384 and as Kong et al. (2024) showed, beyond 36 °C, the development of *G. sigillatus* becomes
385 impaired; it is likely that 36 °C represents the critical thermal optimum for these crickets.

386 The temperature-size rule states that ectotherms developing under higher temperatures are
387 relatively smaller due to a disproportionate effect of temperature on the growth and
388 development rates (Atkinson, 1994). However, our results show an exception to this rule,
389 female *G. sigillatus* were larger when reared at 36°C compared to 27°C. While large body size
390 generally correlates with increased fitness in ectotherms (Kingsolver and Huey, 2008), this
391 result suggests selection for size may intensify under thermal stress, a pattern previously noted
392 in other orthopterans (Walters and Hassall, 2006). Our finding of increased female mass at
393 36°C partially aligns with Kong et al. (2024), however, we found no significant treatment effect

394 on male mass. This discrepancy likely stems from differences in measurement timing; we
395 measured mass within two weeks of adult eclosion, whereas Kong et al. (2024) averaged
396 measurements over four weeks. Given that insect body mass is dynamic and known to fluctuate
397 post-eclosion (Sturm, 2024), the lack of a significant difference in males may reflect different
398 rates of mass loss between treatments early in adulthood. Ultimately, the sex-specific response
399 in body mass suggests divergent selection pressures on male and female reproductive strategies
400 under thermal stress.

401 **4.2. Effect of Developmental Temperature on Mating Behaviour**

402 We found that developmental temperature significantly affected various aspects of mating
403 behaviour. Males reared at 36 °C were less likely to court than those reared at 27 °C. The
404 reduced probability of courtship initiation in males reared at 36 °C suggests that developmental
405 thermal stress impairs the motivational or physiological threshold for reproductive behaviour.
406 Decreased courtship frequency after heat shock exposure was also observed in *Drosophila*,
407 though these effects were much smaller (Smith et al., 2025; Patton and Krebs, 2001). The
408 intensity of courtship behaviours increases as temperature rises (Rosenthal et al., 2019), thereby
409 increasing the organism's energy expenditure. Previous studies in ectotherms have shown that
410 heat stress increases energy requirements for somatic maintenance (Klepsatel et al, 2016;
411 Harding et al., 2023; Sokolova, 2021). It is likely that in *Gryllodes sigillatus* there is a trade-
412 off in resource allocation between somatic maintenance and reproductive effort, similar to a
413 study in *Gryllus bimaculatus* (Ratz et al., 2024). Consequently, reduced courtship can severely
414 affect population viability by reducing mating frequency. A decreased motivation to court can
415 also impact the intensity of sexual selection, as the cost of finding partners for females can
416 increase, which may lead to females choosing lower quality partners as long as they court.

417 Furthermore, our study indicated that female body mass had a considerable impact on mating
418 preferences of males. Heavier females were preferred for courtship by the males over lighter
419 ones. The preference or inclination implies that despite the presence of heat stress, there
420 remains some form of sexual selection to maximize reproductive success. Male insects prefer
421 heavier or larger females because they are more likely to be gravid (Bonduriansky and Brooks,
422 1998; Bonduriansky, 2001) or more fecund (Honěk, 1993). Thus, it is likely that, in *Gryllodes*
423 *sigillatus*, female body mass serves as an honest signal of reproductive potential, enabling
424 males to assess female quality and choose whom to court.

425 Contrary to our prediction, our findings reveal that female developmental temperature was the
426 only factor affecting mating latency. Females exposed to high-temperature conditions showed
427 increased mating latency, possibly indicating a shift in the threshold of mate acceptance.
428 Mating latency has been widely studied as a proxy for female mate choice, and a delay could
429 represent a longer assessment period. While rearing temperatures can modulate mate
430 preference and choosiness (Westerman and Monteiro, 2016; Grace and Shaw, 2004), the
431 observed delay may also reflect the high energetic costs of mating for females under thermal
432 stress. This supports the hypothesis that females, as the higher-investing sex, maintain control
433 over mating decisions (Trivers, 1972; Andersson and Iwasa, 1996). Moreover, heat stress may
434 even reinforce this asymmetry by linking the timing of acceptance to female condition.
435 Although mating latency increased, females had a 95.7% probability of mounting the courting
436 males. This could be due to a decreased female mate preference, which occurs in such no-
437 choice experiments (Dougherty and Shuker, 2015). In a natural setting where females can leave
438 the male, this increased latency might translate into total rejection rather than delayed
439 acceptance. While testing for differences in mating latency between females that mated only
440 once and those given repeated opportunities post-hoc, we found no effect of the number of
441 opportunities or development temperature of either sex (ST 9). This suggests that while our
442 experimental constraints may have lowered female choosiness, their underlying selectivity
443 remained robust.

444 A key feature determining successful mating in this species is the male's transfer of the
445 spermatophore to the female. During mating, alignment between the male and female is critical,
446 enabling the male to transfer his spermatophore into the genital opening of the female, an act
447 that demands precise neuromuscular control (Snell and Killian, 2000). We observed that heat
448 stressed males were frequently unable to transfer their spermatophores which negatively
449 affected mating success. Changes in copulatory behaviours due to heat stress have been
450 previously examined in beetles (Vasudeva et al., 2018). Additionally, heat stress has been
451 shown to alter locomotor activity (Bello et al., 2025), and reduced male motor performance at
452 elevated rearing temperatures may further compromise the physical coordination required for
453 successful spermatophore placement and transfer. Although the developmental temperature of
454 females did not independently affect successful mating, there was an interaction between the
455 developmental temperatures of both sexes. Copulations involving heat-stressed males with
456 heat-stressed females were more successful compared to copulations with control females.
457 Heat exposure might have induced plasticity in females, altering their behaviour during

458 mounting in response to males. Therefore, it is likely that heat-stressed females aligned better
459 with heat-stressed males than control females did. This means that copulation success might be
460 less influenced by the male's performance and more by the physical compatibility between the
461 two individuals, as determined by their thermal histories during development. Nevertheless,
462 further research is needed to identify the precise factors that lead to alignment for successful
463 copulation, and how they are impacted by thermal stress. Our results emphasize the detrimental
464 effects of rising temperatures on mating behaviour, particularly male vulnerability to heat
465 stress, which could drive widespread mating failure, reducing population reproductive output.

466 Our results indicate that both developmental temperature and body weight are significant
467 determinants of nuptial gift feeding duration in this species. The significant positive effect of
468 male weight on feeding duration likely supports the hypothesis that larger males provide
469 higher-quality gifts, which may extend ampulla attachment and enhance fertilization success
470 (Fedorka and Mousseau, 2002). Additionally, larger males may guard females for longer
471 durations, which has been shown to directly extend female feeding time on the nuptial gift and
472 enhance ampulla retention (Haneke-Reinders et al., 2020; Rizvi and Reinhold, 2026). However,
473 due to the mixed evidence on the correlation of mass and mate-guarding duration, these
474 possibilities can only be speculated at this stage. 36°C-reared females fed longer despite their
475 greater mass, likely reflecting heat-induced nutritional deficits (lower lipid reserves despite
476 larger size; Enriquez & Visser 2023) and impaired digestive efficiency (enzyme inhibition;
477 Grčić et al. 2023). This compensatory feeding behaviour addresses the persistent metabolic
478 costs of developmental stress.

479 **4.3. Effect of Developmental Temperature on Reproductive Output**

480 Our results show that developmental heat stress significantly reduced female fecundity,
481 consistent with broader patterns of thermal trade-offs in ectotherms. While high temperatures
482 often accelerate metabolic processes, they can also impose physiological costs, such as reduced
483 allocation to reproductive tissue (Siegle et al., 2022; Gremion et al., 2025). The positive
484 relationship we observed between female mass and egg production is well-documented across
485 numerous insect species; larger females generally possess greater lipid and protein reserves,
486 allowing them to translate superior nutritional status into higher fecundity (Bateman et al.,
487 2001; Lease and Wolf, 2011). However, in our case, although females reared at 36°C were
488 heavier, they paradoxically exhibited lower fecundity over the two-week observation period.
489 This finding suggests a significant physiological trade-off: while high developmental

490 temperatures may promote increased body mass, they may also impair reproductive output.
491 This could occur by shifting energy away from egg production toward somatic maintenance or
492 by altering the maturation rate of oocytes (Awde et al., 2023; Schou et al., 2021).

493 There was also a marked decline in female fertility under heat-stress conditions. Hatching
494 success for eggs produced by heat stressed females during both the first and second weeks after
495 mating, was lower. This persistent reduction suggests that developmental heat stress can induce
496 long-lasting physiological damage that cannot be recovered during the adult stage. Effects of
497 heat stress on mosquito and *Drosophila* fertility (Pekřanská et al., 2025; Kirk Green et al.,
498 2019) have been previously observed, with heat-stressed females exhibiting reduced fertility.
499 A low hatching rate could be attributed to changes in the structure and function of sperm-storing
500 organs (Farrow et al., 2021), resulting in reduced sperm numbers or malfunctioning of the
501 biochemical processes that sustain their vitality. Another possible factor leading to reduced
502 fertility is maternal influence on oocyte quality, whereby elevated developmental temperature
503 induces apoptosis in the germline. Such effects of developmental heat stress have been
504 observed in *Drosophila melanogaster* (Gandara and Barbosa, 2022), where females reared at
505 warm temperatures exhibit impaired oocyte quality and increased germline cyst death and in
506 pufferfish (Lee et al., 2009), where high temperature treatment during gonadal development
507 led to complete germ cell degeneration.

508 Through our experimental design, we were interested in disentangling sex-specific effects on
509 fitness-related traits, however the high rate of unsuccessful copulations involving the heat-
510 stressed males resulted in insufficient data to determine any paternal effects (98% and 76% of
511 the matings failed between 36°C males x 27°C females and 36°C males x 36°C females,
512 respectively). Given that male sperm quality is a critical determinant of reproductive success
513 (Wang and Gunderson, 2022), examining the independent effects of male developmental
514 temperature on sperm function and hatching success represents an important and necessary
515 extension of this work. Thermally induced reductions in male fertilisation capacity would
516 directly diminish male fitness by reducing paternity and simultaneously impose indirect fitness
517 costs on females that invest energetic resources into mating and egg production without the
518 benefit of successful fertilisation, thereby affecting overall population viability.

519 **4.4. Effect of Environmental Temperature on Mating Behaviour**

520 The mating behavioural repertoire of heat-stressed individuals was largely robust to
521 environmental thermal variation except the number of times a pair attempted to mate and the

522 nuptial gift feeding time by females. The frequency of mating attempts increased by 88.42%
523 when mating was carried out under heat-stress conditions. While locomotion and activity levels
524 in insects typically rise with temperature (Hannigan et al., 2023; Suverkropp et al., 2001;
525 Taylor, 1963), this heat-induced hyperactivity may impair the coordination required for
526 successful copulation, necessitating repeated mating attempts. Furthermore, elevated
527 temperatures increase metabolic rates (Riemer et al., 2018), which may have led to faster
528 feeding under heat stress as females attempt to meet the greater energetic demands of a high-
529 temperature environment. Consistent with our previous results (see 4.2) male body mass also
530 played a significant role as females mated to heavier males fed longer.

531 **4.5. Conclusion**

532 To summarise, our study provides evidence that both male and female developmental
533 temperatures modulate life-history traits, morphology, mating behaviours, and reproductive
534 output. Such studies are becoming increasingly important for disentangling sex-specific effects
535 on reproduction, as the two sexes are subject to different selective pressures and physiological
536 constraints that shape their distinct responses to environmental stress. As global temperatures
537 continue to rise, understanding how heat stress experienced during development translates into
538 impaired adult reproductive function becomes increasingly urgent, particularly for ectotherms
539 whose life-history traits and mating behaviours are inextricably linked to their thermal
540 environment. Our findings, therefore, contribute to a growing body of evidence that climate
541 warming poses a significant threat to the reproductive integrity of ectotherm populations,
542 shaping reproductive success, population fitness, and the evolutionary dynamics of sexual
543 selection.

544 **Acknowledgements**

545 We would like to acknowledge the contribution of Annika Sengewald for helping with the
546 caretaking of the experimental population. We are grateful to Sonja Schindler for helping with
547 data collection and providing technical support. We also thank Vanshika Dhiman for designing
548 the illustrations used in Figures 1 and 2.

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552 (lead) **DS**: Conceptualisation, Data Curation, Methodology, Software, Formal Analysis,
553 Investigation, Project Administration, Visualisation (lead), Writing - Original Draft, Writing -
554 Review and Editing (supporting) **KR**: Resources, Funding Acquisition, Supervision
555 (supporting), Writing - Review and Editing (supporting)

556 **Conflict of Interest Declaration**

557 The authors declare no competing interests.

558 **Funding**

559 This project was supported by funds from the Department of Evolutionary Biology, Bielefeld
560 University (awarded to DS) and the Deutscher Akademischer Austauschdienst (DAAD)
561 Doctoral Programmes in Germany (awarded to TR).

562 **References**

- 563 Abram, P. K., Boivin, G., Moiroux, J., & Brodeur, J. (2017). Behavioural effects of temperature on
564 ectothermic animals: unifying thermal physiology and behavioural plasticity. *Biol Rev*, 92: 1859-
565 1876. <https://doi.org/10.1111/brv.12312>
- 566 Ahlmann-Eltze, C., & Patil, I. (2021). *ggsignif: R package for displaying significance brackets for*
567 *'ggplot2'*. PsyArXiv. <https://doi.org/10.31234/osf.io/7awm6>
- 568 Ana Caroline, P. G., & Drummond-Barbosa, D. (2022). Warm and cold temperatures have distinct
569 germline stem cell lineage effects during *Drosophila* oogenesis. *Development*, 149(5): dev200149.
570 <https://doi.org/10.1242/dev.200149>
- 571 Andersson, M., & Iwasa, Y. (1996). Sexual selection. *Trends in Ecology & Evolution*, 11(2), 53-58.
572 [https://doi.org/10.1016/0169-5347\(96\)81042-1](https://doi.org/10.1016/0169-5347(96)81042-1)
- 573 Angilletta Jr., M. J., & Dunham, A. E. (2003). The Temperature-Size Rule in Ectotherms: Simple
574 Evolutionary Explanations May Not Be General. *The American Naturalist*, 162(3): 332-342.
575 <https://doi.org/10.1086/377187>
- 576 Araripe, L., Klaczko, L., Moreteau, B., & David, J. (2004). Male sterility thresholds in a tropical
577 cosmopolitan drosophilid, *Zaprionus indianus*. *Journal of Thermal Biology*, 29(2): 73-80.
578 <https://doi.org/10.1016/j.jtherbio.2003.11.006>
- 579 Atkinson, D. (1994). Temperature and organism size—A biological law for ectotherms? *Advances*
580 *in Ecological Research*, 25: 1-58. [https://doi.org/10.1016/S0065-2504\(08\)60212-3](https://doi.org/10.1016/S0065-2504(08)60212-3)
- 581 Audzijonyte, A., Fulton, E., Haddon, M., Helidoniotis, F., Hobday, A. J., Kuparinen, A.,
582 Morrongiello, J., Smith, A. D., Upston, J., & Waples, R. S. (2016). Trends and management
583 implications of human-influenced life-history changes in marine ectotherms. *Fish Fish*, 17: 1005-
584 1028. <https://doi.org/10.1111/faf.12156>

- 585 Augustin, J., Bourgeois, G., Brodeur, J., & Boivin, G. (2022). Low and high temperatures decrease
586 the mating success of an egg parasitoid and the proportion of females in the population. *Journal of*
587 *Thermal Biology*, 110: 103382. <https://doi.org/10.1016/j.jtherbio.2022.103382>
- 588 Bateman, P.W., Gilson, L.N. & Ferguson, J.W.H. (2001), Investment in Mate Guarding May
589 Compensate for Constraints on Ejaculate Production in the Cricket *Gryllodes sigillatus*. *Ethology*,
590 107: 1087-1098. <https://doi.org/10.1046/j.1439-0310.2001.00756.x>
- 591 Bates, D., Maechler, M., Bolker, B., & Walker, S. (2015). Fitting linear mixed-effects models using
592 lme4. *Journal of Statistical Software*, 67(1), 1–48. <https://doi.org/10.18637/jss.v067.i01>
- 593 Beckers, O. M., Murphey, K. J., Pease, J. R., & Norman, N. (2019). Parallel plasticity of mating
594 songs and preferences in the field cricket *Gryllus rubens*. *Ethology*, 125: 476–484.
595 <https://doi.org/10.1111/eth.12872>
- 596 Bello, Z. M., Aisha, A., Sambo, Z. S., Ademola, A. O., Isa, A. S., Olatomide, O. D., & Ochai, J.
597 (2025). Heat Stress-Induced Oxidative Stress and Locomotor Deficit in *Drosophila Melanogaster*.
598 *Journal of Experimental and Clinical Anatomy*, 22(2): 329–332.
599 <https://doi.org/10.4314/jeca.v22i2.18>
- 600 Bhaisare, L. Y., & Chaudhary, D. D. (2023). Mate Guarding Behaviour in Response to Temperature
601 in Parthenium Beetle *Zygogramma bicolorata* Pallister. *Indian Journal of Entomology*, 86(4):
602 1223–1228. <https://doi.org/10.55446/IJE.2023.1511>
- 603 Bonduriansky, R., & Brooks, R. (2011). Male antler flies (*Protophila litigata*; Diptera:
604 Piophilidae) are more selective than females in mate choice. *Canadian Journal of Zoology*, 76:
605 1277-1285. <https://doi.org/10.1139/z98-069>
- 606 Bonduriansky, R. (2001). The evolution of male mate choice in insects: a synthesis of ideas and
607 evidence. *Biological Reviews*, 76(3): 305–339. <https://doi.org/10.1017/S1464793101005693>
- 608 Brooks, M. E., Kristensen, K., van Benthem, K. J., Magnusson, A., Berg, C. W., Nielsen, A., Skaug,
609 H. J., Maechler, M., & Bolker, B. M. (2017). glmmTMB balances speed and flexibility among
610 packages for zero-inflated generalized linear mixed modeling. *The R Journal*, 9(2), 378–400.
611 <https://doi.org/10.32614/RJ-2017-066>
- 612 Bürkner, P.-C. (2017). brms: An R package for Bayesian multilevel models using Stan. *Journal of*
613 *Statistical Software*, 80(1), 1–28. <https://doi.org/10.18637/jss.v080.i01>
- 614 Canal Domenech, B., & Fricke, C. (2023). Developmental heat stress interrupts spermatogenesis
615 inducing early male sterility in *Drosophila melanogaster*. *Journal of Thermal Biology*, 114: 103589.
616 <https://doi.org/10.1016/j.jtherbio.2023.103589>
- 617 Ciordia, H., & Bizzell, W. E. (1963). The Effects of Various Constant Temperatures on the
618 Development of the Free Living-Stages of Some Nematode Parasites of Cattle. *The Journal of*
619 *Parasitology*, 49(1): 60–63. <https://doi.org/10.2307/3275675>
- 620 Costa, S. G., Magalhães, S., & Rodrigues, L. R. (2023). Multiple mating rescues offspring sex ratio
621 but not productivity in a haplodiploid exposed to developmental heat stress. *Functional Ecology*,
622 37(5): 1291-1303. <https://doi.org/10.1111/1365-2435.14303>
- 623 Daly, M. (1978). The Cost of Mating. *The American Naturalist*, 112(986): 771–774.
624 <http://www.jstor.org/stable/2460052>

625 Degen, T., Hovestadt, T., Mitesser, O., & Hölker, F. (2015). High Female Survival Promotes
626 Evolution of Protogyny and Sexual Conflict. *PLoS ONE*, 10(3): e0118354.
627 <https://doi.org/10.1371/journal.pone.0118354>

628 Dougherty, L. R., & Shuker, D. M. (2015). The effect of experimental design on the measurement
629 of mate choice: a meta-analysis. *Behavioral Ecology*, 26(2): 311–319.
630 <https://doi.org/10.1093/beheco/aru125>

631 Dougherty, L. R., Frost, F., Maenpaa, M. I., Rowe, M., Cole, B. J., Vasudeva, R., Pottier, P.,
632 Schultner, E., Macartney, E. L., Lindenbaum, I., Smith, J. L., Carazo, P., Graziano, M., Weaving,
633 H., Domenech, B. C., Berger, D., Meena, A., Bishop, T. R., Noble, D. W. A., Price, T. A. (2023). A
634 systematic map of studies testing the relationship between temperature and animal reproduction.
635 *Ecological Solutions and Evidence*, 5(1): e12303. <https://doi.org/10.1002/2688-8319.12303>

636 Enriquez, T., & Visser, B. (2023). The importance of fat accumulation and reserves for insect
637 overwintering. *Current opinion in insect science*, 60, 101118.
638 <https://doi.org/10.1016/j.cois.2023.101118>

639 Farrow, R. A., Deeming, D. C., & Eady, P. E. (2021). Male and female developmental temperature
640 modulate post-copulatory interactions in a beetle. *Journal of Thermal Biology*, 103: 103155.
641 <https://doi.org/10.1016/j.jtherbio.2021.103155>

642 FC, M., Davis, T., & ggplot2 authors. (2026). *ggpattern: 'ggplot2' pattern geoms* (Version 1.3.1) [R
643 package]. <https://doi.org/10.32614/CRAN.package.ggpattern>

644 Fedorka, K. M., & Mousseau, T. A. (2002). Nuptial gifts and the evolution of male body size.
645 *Evolution*, 56(3): 590–596. <https://doi.org/10.1111/j.0014-3820.2002.tb01369.x>

646 Fox, J., & Weisberg, S. (2019). *An R companion to applied regression* (3rd ed.). Sage.
647 <https://www.john-fox.ca/Companion/>

648 French, V., Feast, M., & Partridge, L. (1998). Body size and cell size in *Drosophila*: The
649 developmental response to temperature. *Journal of Insect Physiology*, 44(11): 1081-1089.
650 [https://doi.org/10.1016/S0022-1910\(98\)00061-4](https://doi.org/10.1016/S0022-1910(98)00061-4)

651 Garrad, R., Booth, D. T., & Furlong, M. J. (2016). The effect of rearing temperature on
652 development, body size, energetics and fecundity of the diamondback moth. *Bulletin of*
653 *Entomological Research*, 106(2): 175–181. <https://doi.org/10.1017/S000748531500098X>

654 Geiselhardt, S., Otte, T., & Hilker, M. (2012). Looking for a similar partner: Host plants shape
655 mating preferences of herbivorous insects by altering their contact pheromones. *Ecology Letters*,
656 15(9): 971-977. <https://doi.org/10.1111/j.1461-0248.2012.01816.x>

657 Gemeno, C., & Haynes, K. F. (2001). Impact of Photoperiod on the Sexual Behavior of the Black
658 Cutworm Moth (Lepidoptera: Noctuidae). *Environmental Entomology*, 30(2): 189-195.
659 <https://doi.org/10.1603/0046-225X-30.2.189>

660 Grace, J. L., & Shaw, K. L. (2004). Effects of developmental environment on signal-preference
661 coupling in a Hawaiian cricket. *Evolution*, 58(7): 1627–1633. <https://doi.org/10.1111/j.0014-3820.2004.tb01744.x>

662

663 Grčić, A., Ilijin, L., Filipović, A., Matic, D., Mrdaković, M., Todorović, D., Vlahović, M., &
664 Perić-Mataruga, V. (2023). Digestive enzyme activity and macromolecule content in the

665 hemolymph of differentially adapted *Lymantria dispar* L. populations after short-term increases in
666 ambient temperature *Environmental research*, 236(Pt 1), 116461.
667 <https://doi.org/10.1016/j.envres.2023.116461>

668 Gremion, A., Thakur, M. P., & Martínez-De León, G. (2025). Reproductive Costs Increase with
669 Longer Extreme Heat Events in Collembola. *Ecology and evolution*, 15(7), e71775.
670 <https://doi.org/10.1002/ece3.71775>

671 Grieve, I., & Pilakouta, N. (2026). Heatwaves impair female but not male fertility in a subsocial
672 insect. *Biol Lett*, 22(1): 20250665. <https://doi.org/10.1098/rsbl.2025.0665>

673 Grolemund, G., & Wickham, H. (2011). Dates and times made easy with lubridate. *Journal of*
674 *Statistical Software*, 40(3), 1–25. <https://www.jstatsoft.org/v40/i03/>

675 Gruntenko, N. E., Bownes, M., Terashima, J., Sukhanova, M. Z., & Raushenbach, I. Y. (2003). Heat
676 stress affects oogenesis differently in wild-type *Drosophila virilis* and a mutant with altered juvenile
677 hormone and 20-hydroxyecdysone levels. *Insect Molecular Biology*, 12: 393-404.
678 <https://doi.org/10.1046/j.1365-2583.2003.00424.x>

679 Haneke-Reinders, M., Mazur, AP., Zyma, O. Ramm, S, & Reinhold, K. (2020). Disentangling a
680 shared trait: male control over mate guarding duration revealed by a mate exchange
681 experiment. *Behav Ecol Sociobiol* 74, 54. <https://doi.org/10.1007/s00265-020-02832-1>

682 Hannigan, S., Nendel, C., & Krull, M. (2023). Effects of temperature on the movement and feeding
683 behaviour of the large lupine beetle, *Sitona gressorius*. *J Pest Sci*, 96: 389–402.
684 <https://doi.org/10.1007/s10340-022-01510-7>

685 Hansen, P. J. (2009). Effects of heat stress on mammalian reproduction. *Philos Trans R Soc Lond B*
686 *Biol Sci*, 364(1534): 3341–3350. <https://doi.org/10.1098/rstb.2009.0131>

687 Harding, L., Jackson, A. L., & Payne, N. (2022). Energetic costs increase with faster heating in an
688 aquatic ectotherm. *Conservation Physiology*, 11(1): coad042.
689 <https://doi.org/10.1093/conphys/coad042>

690 Hartig, F. (2024). *DHARMA: Residual diagnostics for hierarchical (multi-level / mixed) regression*
691 *models* (Version 0.4.7) [R package]. <https://doi.org/10.32614/CRAN.package.DHARMA>

692 Heinze, G., Ploner, M., Jiricka, L., & Steiner, G. (2023). *coxphf: Cox regression with Firth's*
693 *penalized likelihood* (Version 1.13.4) [R package]. <https://doi.org/10.32614/CRAN.package.coxphf>

694 Hirst, A. G., Horne, C. R., & Atkinson, D. (2015). Equal temperature–size responses of the sexes
695 are widespread within arthropod species. *Proceedings of the Royal Society B: Biological Sciences*,
696 282(1820), 20152475. <https://doi.org/10.1098/rspb.2015.2475>

697 Honěk, A. (1993). Intraspecific Variation in Body Size and Fecundity in Insects: A General
698 Relationship. *Oikos*, 66(3): 483–492. <https://doi.org/10.2307/3544943>

699 Horne, C. R., Hirst, A. G., & Atkinson, D. (2015). Temperature-size responses match latitudinal-
700 size clines in arthropods, revealing critical differences between aquatic and terrestrial species.
701 *Ecology Letters*, 18(4), 327-335. <https://doi.org/10.1111/ele.12413>

702 Hurley, L. L., McDiarmid, C. S., Friesen, C. R., Griffith, S. C., & Rowe, M. (2018). Experimental
703 heatwaves negatively impact sperm quality in the zebra finch. *Proc Biol Sci*, 285(1871): 20172547.
704 <https://doi.org/10.1098/rspb.2017.2547>

705 Iossa, G., Maury, C., Fletcher, R. M., & Eady, P. E. (2019). Temperature-induced developmental
706 plasticity in *Plodia interpunctella*: Reproductive behaviour and sperm length. *Journal of*
707 *Evolutionary Biology*, 32(7): 675–682. <https://doi.org/10.1111/jeb.13447>

708 Iossa, G. (2019). Sex-Specific Differences in Thermal Fertility Limits. *Trends in Ecology &*
709 *Evolution*, 34(6): 490-492. <https://doi.org/10.1016/j.tree.2019.02.016>

710 Jørgensen, L. B., Ørsted, M., Malte, H., Wang, T., & Overgaard, J. (2022). Extreme escalation of
711 heat failure rates in ectotherms with global warming. *Nature*, 611(7934): 93-98.
712 <https://doi.org/10.1038/s41586-022-05334-4>

713 Kassambara, A., Kosinski, M., & Biecek, P. (2025). *survminer: Drawing survival curves using*
714 *'ggplot2'* (Version 0.5.1) [R package]. <https://doi.org/10.32614/CRAN.package.survminer>

715 Kassambara, A. (2025). *ggpubr: 'ggplot2' based publication ready plots* (Version 0.6.2) [R
716 package]. <https://doi.org/10.32614/CRAN.package.ggpubr>

717 Katsuki, M., & Miyatake, T. (2009). Effects of temperature on mating duration, sperm transfer and
718 remating frequency in *Callosobruchus chinensis*. *Journal of Insect Physiology*, 55(2): 113-116.
719 <https://doi.org/10.1016/j.jinsphys.2008.10.012>

720 Kim, B., Park, K., & Rhee, K. (2013). Heat stress response of male germ cells. *Cell. Mol. Life Sci.*,
721 70: 2623–2636. <https://doi.org/10.1007/s00018-012-1165-4>

722 Kim, S-Y., Costa, M. M., Esteve-Codina, A., & Velando, A. (2017). Transcriptional mechanisms
723 underlying life-history responses to climate change in the three-spined stickleback. *Evol Appl*, 10:
724 718–730. <https://doi.org/10.1111/eva.12487>

725 Kingsolver, J., & Huey, R. (2008). Size, temperature, and fitness: three rules. *Evolutionary Ecology*
726 *Research*, 10: 251-268. <https://doi.org/10.17615/jtzc-v174>

727 Kirk Green, C., Moore, P. J., & Sial, A. A. (2019). Impact of heat stress on development and fertility
728 of *Drosophila suzukii* Matsumura (Diptera: Drosophilidae). *Journal of Insect Physiology*, 114: 45-
729 52. <https://doi.org/10.1016/j.jinsphys.2019.02.008>

730 Klepsatel, P., Gáliková, M., Xu, Y., & Kühnlein, R. P. (2016). Thermal stress depletes energy
731 reserves in *Drosophila*. *Scientific Reports*, 6(1): 33667. <https://doi.org/10.1038/srep33667>

732 Knapp, M., & Nedvěd, O. (2013). Gender and Timing during Ontogeny Matter: Effects of a
733 Temporary High Temperature on Survival, Body Size and Colouration in *Harmonia axyridis*. *PLOS*
734 *ONE*, 8(9): e74984. <https://doi.org/10.1371/journal.pone.0074984>

735 Kong, J. D., Ritchie, M. W., Vadboncoeur, É., MacMillan, H. A., & Bertram, S. M. (2025).
736 Growth, development, and life history of a mass-reared edible insect, *Grylloides sigillatus*
737 (Orthoptera: Gryllidae). *Journal of Economic Entomology*, 118(3), 1093-1103.
738 <https://doi.org/10.1093/jee/toaf073>

739 Kong, J. D., Vadboncoeur, É., Bertram, S. M., & MacMillan, H. A. (2024). Temperature-
740 dependence of life history in an edible cricket: Implications for optimising mass-rearing. *Current*
741 *Research in Insect Science*, 7: 100109. <https://doi.org/10.1016/j.cris.2025.100109>

742 Krebs, R. A., & Loeschcke, V. (1994). Effects of exposure to short-term heat stress on fitness
743 components in *Drosophila melanogaster*. *Journal of Evolutionary Biology*, 7(1): 39–49.
744 <https://doi.org/10.1046/j.1420-9101.1994.7010039.x>

- 745 Lazo, C. S., & Pita, I. M. (2012). Effect of temperature on survival, growth and development of
746 *Mytilus galloprovincialis* larvae. *Aquaculture Research*, 43: 1127-1133.
747 <https://doi.org/10.1111/j.1365-2109.2011.02916.x>
- 748 Lease, H. M., & Wolf, B. O. (2011). Lipid content of terrestrial arthropods in relation to body
749 size, phylogeny, life history stage, and sex. *Physiological Entomology*, 36(1), 29–38.
750 <https://doi.org/10.1111/j.1365-3032.2010.00767.x>
- 751 Lee, K. H., Yamaguchi, A., Rashid, H., Kadomura, K., Yasumoto, S., & Matsuyama, M. (2009).
752 Germ Cell Degeneration in High-Temperature Treated Pufferfish, *Takifugu rubripes*. *Sex Dev*, 3(4):
753 225–232. <https://doi.org/10.1159/000228723>
- 754 Lee, W-S., Monaghan, P., & Metcalfe, N. B. (2013). Experimental demonstration of the growth
755 rate–lifespan trade-off. *Proc Biol Sci*, 280(1752): 20122370.
756 <https://doi.org/10.1098/rspb.2012.2370>
- 757 Leith, N. T., Macchiano, A., Moore, M. P., & Fowler-Finn, K. D. (2021). Temperature impacts all
758 behavioral interactions during insect and arachnid reproduction. *Current Opinion in Insect Science*,
759 45: 106-114. <https://doi.org/10.1016/j.cois.2021.03.005>
- 760 Lenth, R., & Piaskowski, J. (2025). *emmeans: Estimated marginal means, aka least-squares means*
761 (Version 2.0.0) [R package]. <https://doi.org/10.32614/CRAN.package.emmeans>
- 762 Li, X., Jia, X., Xiang, H., Diao, H., Yan, Y., Wang, Y., & Ma, R. (2019). The Effect of Photoperiods
763 and Light Intensity on Mating Behavior and Reproduction of *Grapholita molesta* (Lepidoptera:
764 Tortricidae). *Environmental Entomology*, 48(5): 1035-1041. <https://doi.org/10.1093/ee/nvz066>
- 765 Macchiano, A., Miller, E., Agali, U., Ola-Ajose, A., & Fowler-Finn, K. D. (2023). Developmental
766 temperature alters the thermal sensitivity of courtship activity and signal–preference relationships,
767 but not mating rates. *Oecologia*, 202(1), 97–111. <https://doi.org/10.1007/s00442-023-05376-z>
- 768 Michelangeli, M., Goulet, C. T., Kang, H. S., Wong, B. B. M., & Chapple, D. G. (2018). Integrating
769 thermal physiology within a syndrome: Locomotion, personality and habitat selection in an
770 ectotherm. *Funct Ecol*, 32: 970–981. <https://doi.org/10.1111/1365-2435.13034>
- 771 Neven, L. G. (2000). Physiological responses of insects to heat. *Postharvest Biology and*
772 *Technology*, 21(1): 103-111. [https://doi.org/10.1016/S0925-5214\(00\)00169-1](https://doi.org/10.1016/S0925-5214(00)00169-1)
- 773 Nguyen, T. M., Bressac, C., & Chevrier, C. (2013). Heat stress affects male reproduction in a
774 parasitoid wasp. *Journal of Insect Physiology*, 59(3): 248-254.
775 <https://doi.org/10.1016/j.jinsphys.2012.12.001>
- 776 Ohlberger, J. (2013). Climate warming and ectotherm body size – from individual physiology to
777 community ecology. *Funct Ecol*, 27: 991-1001. <https://doi.org/10.1111/1365-2435.12098>
- 778 Ombuya, A., Guo, J., & Liu, W. (2025). Insect Mating Behaviors: A Review of the Regulatory Role
779 of Neuropeptides. *Insects*, 16(5). <https://doi.org/10.3390/insects16050506>
- 780 Parratt, S. R., Walsh, B. S., Metelmann, S., et al. (2021). Temperatures that sterilize males better
781 match global species distributions than lethal temperatures. *Nat. Clim. Chang.*, 11: 481–484.
782 <https://doi.org/10.1038/s41558-021-01047-0>

783 Patton, Z. J., & Krebs, R. A. (2001). The Effect of Thermal Stress on the Mating Behavior of
784 Three Drosophila Species. *Physiological and Biochemical Zoology*, 74(1): 118-124.
785 <https://doi.org/10.1086/323327>

786 Paul, J. M. (1998). Development of Larvae of the Golden King Crab *Lithodes Aequispinus*
787 (Anomura: Lithodidae) Reared at Different Temperatures. *Journal of Crustacean Biology*, 19(1):
788 42-45. <https://doi.org/10.2307/1549544>

789 Pedersen, T. (2025). *patchwork: The composer of plots* (Version 1.3.2) [R package].
790 <https://doi.org/10.32614/CRAN.package.patchwork>

791 Pekl'anská, M., van Heerwaarden, B., Hoffmann, A. A., Nouzová, M., Šíma, R., & Ross, P. A.
792 (2025). Elevated developmental temperatures below the lethal limit reduce *Aedes aegypti* fertility.
793 *J Exp Biol*, 228(3): JEB249803. <https://doi.org/10.1242/jeb.249803>

794 Pembury Smith, M. Q., Latkova, L., & Snook, R. R. (2025). Elevated temperatures have sex-
795 specific effects on nuptial gift behavior. *Behavioral Ecology*, 36(4).
796 <https://doi.org/10.1093/beheco/ara049>

797 Porcelli, D., Gaston, K. J., Butlin, R. K., & Snook, R. R. (2017). Local adaptation of reproductive
798 performance during thermal stress. *Journal of Evolutionary Biology*, 30(2): 422-429.
799 <https://doi.org/10.1111/jeb.13018>

800 Rankin, D. J., & Kokko, H. (2007). Do males matter? The role of males in population dynamics.
801 *Oikos*, 116(2): 335-348. <https://doi.org/10.1111/j.0030-1299.2007.15451.x>

802 R Core Team. (2025). *R: A language and environment for statistical computing*. R Foundation for
803 Statistical Computing. <https://www.R-project.org/>

804 Rebar, D., & Rodríguez, R. L. (2015). Insect mating signal and mate preference phenotypes covary
805 among host plant genotypes. *Evolution*, 69(3): 602-610. <https://doi.org/10.1111/evo.12604>

806 Reinhold, K., & Rizvi, T. (2026). The significance of disentangling shared mating traits: A review
807 of empirical approaches. *Behav Ecol Sociobiol* 80(3) <https://doi.org/10.1007/s00265-025-03681-6>

808 Riemer, K., Anderson-Teixeira, K. J., Smith, F. A., Harris, D. J., & Morgan Ernest, S. K. (2018).
809 Body size shifts influence effects of increasing temperatures on ectotherm metabolism. *Global*
810 *Ecology and Biogeography*, 27(8): 958-967. <https://doi.org/10.1111/geb.12757>

811 Rinker, T. W., & Kurkiewicz, D. (2017). *pacman: Package management for R* (Version 0.5.0) [R
812 package]. <http://github.com/trinker/pacman>

813 Rizvi, T., Tuni, C., & Reinhold, K. (2026). From mating to sperm storage: density-dependent
814 plasticity in pre- and post-copulatory shared mating traits. *Evolution*,
815 qpag076. <https://doi.org/10.1093/evolut/qpag076>

816 Rizvi, T., & Reinhold, K. (2026). Developmental density shapes adult mate guarding strategies in
817 an invertebrate [Preprint]. *EcoEvoRxiv*. <https://doi.org/10.32942/X26M12>

818 Rosenthal, M. F., & Elias, D. O. (2019). Nonlinear changes in selection on a mating display
819 across a continuous thermal gradient. *Proc Biol Sci*, 286(1907): 20191450.
820 <https://doi.org/10.1098/rspb.2019.1450>

- 821 Ruthsatz, K., Peck, M. A., Dausmann, K. H., Sabatino, N. M., & Glos, J. (2018). Patterns of
822 temperature induced developmental plasticity in anuran larvae. *Journal of Thermal Biology*, 74:
823 123-132. <https://doi.org/10.1016/j.jtherbio.2018.03.005>
- 824 Sakaluk, S. K. (1984). Male Crickets Feed Females to Ensure Complete Sperm Transfer. *Science*,
825 223(4636): 609-610. <https://doi.org/10.1126/science.223.4636.609>
- 826 Sales, K., Vasudeva, R., & Gage, M. J. G. (2021). Fertility and mortality impacts of thermal stress
827 from experimental heatwaves on different life stages and their recovery in a model insect. *R Soc*
828 *Open Sci.*, 8(3): 201717. <https://doi.org/10.1098/rsos.201717>
- 829 Sales, K., Vasudeva, R., Dickinson, M. E., Godwin, J. L., Lumley, A. J., Michalczyk, Ł.,
830 Hebberecht, L., Thomas, P., Franco, A., & Gage, M. J. (2018). Experimental heatwaves compromise
831 sperm function and cause transgenerational damage in a model insect. *Nature Communications*,
832 9(1): 4771. <https://doi.org/10.1038/s41467-018-07273-z>
- 833 Schou, M. F., Bonato, M., Engelbrecht, A., Brand, Z., Svensson, E. I., Melgar, J., Muvhali, P. T.,
834 Cloete, S. W. P., & Cornwallis, C. K. (2021). Extreme temperatures compromise male and female
835 fertility in a large desert bird. *Nature communications*, 12(1), 666. [https://doi.org/10.1038/s41467-](https://doi.org/10.1038/s41467-021-20937-7)
836 [021-20937-7](https://doi.org/10.1038/s41467-021-20937-7)
- 837 Siegle, M. R., Taylor, E. B., & O'Connor, M. I. (2022). Heat Wave Intensity Drives Sublethal
838 Reproductive Costs in a Tidepool Copepod. *Integrative organismal biology (Oxford,*
839 *England)*, 4(1), obac005. <https://doi.org/10.1093/iob/obac005>
- 840 Sinclair, B. J., Marshall, K. E., Sewell, M. A., Levesque, D. L., Willett, C. S., Slotsbo, S., Dong, Y.,
841 Harley, C. D. G., Marshall, D. J., Helmuth, B. S., & Huey, R. B. (2016). Can we predict ectotherm
842 responses to climate change using thermal performance curves and body temperatures? *Ecol Lett*,
843 19: 1372-1385. <https://doi.org/10.1111/ele.12686>
- 844 Smith, R. L., & Thomas, W. (1988). Southwestern Distribution and Habitat Ecology of *Gryllodes*
845 *supplicans*. *American Entomologist*, 34(4): 186-191. <https://doi.org/10.1093/besa/34.4.186>
- 846 Snell, L., & Killian, K. (2000). The role of cercal sensory feedback during spermatophore transfer
847 in the cricket, *Acheta domesticus*. *Journal of Insect Physiology*, 46(6), 1017-1032.
848 [https://doi.org/10.1016/S0022-1910\(99\)00213-9](https://doi.org/10.1016/S0022-1910(99)00213-9)
- 849 Sokolova, I. (2021). Bioenergetics in environmental adaptation and stress tolerance of aquatic
850 ectotherms: linking physiology and ecology in a multi-stressor landscape. *J Exp Biol*, 224.
851 <https://doi.org/10.1242/jeb.236802>
- 852 Suverkropp, B. P., Bigler, F., & Van Lenteren, J. C. (2001). Temperature influences walking speed
853 and walking activity of *Trichogramma brassicae* (Hym., Trichogrammatidae). *Journal of Applied*
854 *Entomology*, 125: 303-307. <https://doi.org/10.1046/j.1439-0418.2001.00546.x>
- 855 Taylor, L. R. (1963). Analysis of the Effect of Temperature on Insects in Flight. *Journal of Animal*
856 *Ecology*, 32(1): 99-117. <https://doi.org/10.2307/2520>
- 857 Therneau, T. (2026). *A package for survival analysis in R* (Version 3.8-6) [R package].
858 <https://CRAN.R-project.org/package=survival>

859 Trivers, R. L. (1972). Parental investment and sexual selection. In B. Campbell (Ed.), *Sexual*
860 *selection and the descent of man, 1871–1971* (pp. 139–179). Aldine.
861 <https://www2.nau.edu/~shuster/shustercourses/BIO%20698/Literature/Trivers%201972.pdf>

862 van Heerwaarden, B., & Sgrò, C. M. (2021). Male fertility thermal limits predict vulnerability to
863 climate warming. *Nat Commun*, 12: 2214. <https://doi.org/10.1038/s41467-021-22546-w>

864 Vasudeva, R., Deeming, D. C., & Eady, P. E. (2018). Larval developmental temperature and ambient
865 temperature affect copulation duration in a seed beetle. *Behaviour*, 155(1): 69-82.
866 <https://doi.org/10.1163/1568539X-00003479>

867 Walters, R. J., & Hassall, M. (2006). The Temperature-Size Rule in Ectotherms: May a General
868 Explanation Exist after All? *The American Naturalist*, 167(4): 510-523.
869 <https://doi.org/10.1086/501029>

870 Wang, W. W., & Gunderson, A. R. (2022). The Physiological and Evolutionary Ecology of Sperm
871 Thermal Performance. *Frontiers in physiology*, 13, 754830.
872 <https://doi.org/10.3389/fphys.2022.754830>

873 Westerman, E., & Monteiro, A. (2016). Rearing Temperature Influences Adult Response to Changes
874 in Mating Status. *PLOS ONE*, 11(2): e0146546. <https://doi.org/10.1371/journal.pone.0146546>

875 Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L.D., François, R., Grolemund, G.,
876 Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T.L., Miller, E., Bache, S.M., Müller, K., Ooms,
877 J., Robinson, D., Seidel, D.P., Spinu, V., Takahashi, K., Vaughan, D., Wilke, C., Woo, K., & Yutani,
878 H. (2019). “Welcome to the tidyverse.” *Journal of Open Source Software*, 4(43),
879 1686. [doi:10.21105/joss.01686](https://doi.org/10.21105/joss.01686).

880 Wickham, H. (2016). *ggplot2: Elegant graphics for data analysis*. Springer-Verlag.

881 Zhang, S., Cao, Z., Wang, Q., Zhang, F., & Liu, T. (2013). Exposing eggs to high temperatures
882 affects the development, survival and reproduction of *Harmonia axyridis*. *Journal of Thermal*
883 *Biology*, 39: 40-44. <https://doi.org/10.1016/j.jtherbio.2013.11.007>

884 Ørsted, M., Jørgensen, L. B., & Overgaard, J. (2022). Finding the right thermal limit: a framework
885 to reconcile ecological, physiological and methodological aspects of CTmax in ectotherms. *J Exp*
886 *Biol*, 225(19): jeb244514. <https://doi.org/10.1242/jeb.244514>

887

SUPPLEMENTARY MATERIAL

888

889

890 *ST 1: Summary of Firth's Penalized Cox Proportional Hazards Regression model for the effect of*
 891 *temperature and sex on development rate. 95% CI and p-values calculated by Profile Likelihood.*

Variable	Coefficient (β)	HR ($e\beta$)	95% CI	p - value
Temperature (36 °C)	7.556	1911.69	[258.55, 244,893.6]	< 0.001
Sex (Male)	- 0.333	0.716	[0.569, 0.902]	0.005

892

893 *ST 2: Summary of Linear model for the effect of temperature, sex, and their interaction on body mass,*
 894 *and post-hoc analysis of differences within sex.*

Section	Comparison / Variable	Estimate	Std. Error	t-ratio	p-value
Model Coefficients	(Intercept: Female 27°C)	0.350	0.007	49.31	< 0.001
	Temperature (36°C)	+0.078	0.010	7.83	< 0.001
	Sex (Male)	-0.089	0.010	-9.13	< 0.001
	Interaction (Temperature × Sex)	-0.074	0.014	-5.41	< 0.001
Post-Hoc (Within Sex)	Females: 27°C vs 36°C	-0.078	0.009	-7.83	< 0.001
	Males: 27°C vs 36°C	-0.003	0.009	-0.37	0.710

895

896 *ST 3: Summary of posterior estimates from Bayesian Generalized Linear Mixed model for the effect of*
 897 *male rearing temperature, female rearing temperature, and their interaction on the probability of male*
 898 *initiating courtship.*

Predictor	Estimate (Mean)	Est. Error	Lower 95% CI	Upper 95% CI	Rhat
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Intercept	2.65	0.74	1.34	4.23	1.00
Female Temperature (36°C)	-0.63	1.01	-2.65	1.36	1.00
Male Temperature (36°C)	-4.80	1.03	-7.06	-2.99	1.00
Female Weight (z)	0.45	0.21	0.04	0.87	1.00
Male Weight (z)	-0.47	0.30	-1.08	0.09	1.00
Female Temperature × Male Temperature	2.17	1.26	-0.23	4.74	1.00

899

900 *ST 4: Summary of Linear Mixed Effects model for the effect of male rearing temperature, female rearing*
901 *temperature, and their interaction on log-transformed mating latency.*

Type	Predictor	Estimate	Std. Error	df	t value	p-value
Fixed Effects	(Intercept)	1.587	0.215	96.88	7.37	< 0.001
	Male Temperature (36°C)	0.297	0.326	92.46	0.91	0.364
	Female Temperature (36°C)	0.990	0.357	100.79	2.77	0.007
	Male Weight (z)	-0.153	0.103	177.38	-1.48	0.141
	Female Weight (z)	-0.056	0.132	108.75	-0.42	0.674
	Male Temperature × Female Temperature	-0.054	0.451	95.16	-0.12	0.905
	Random Effects	Groups	Variance	Std. Dev.		
Female ID		0.3026	0.5501			
Male ID		0.0000	0.0000			
Residual		1.6062	1.2673			

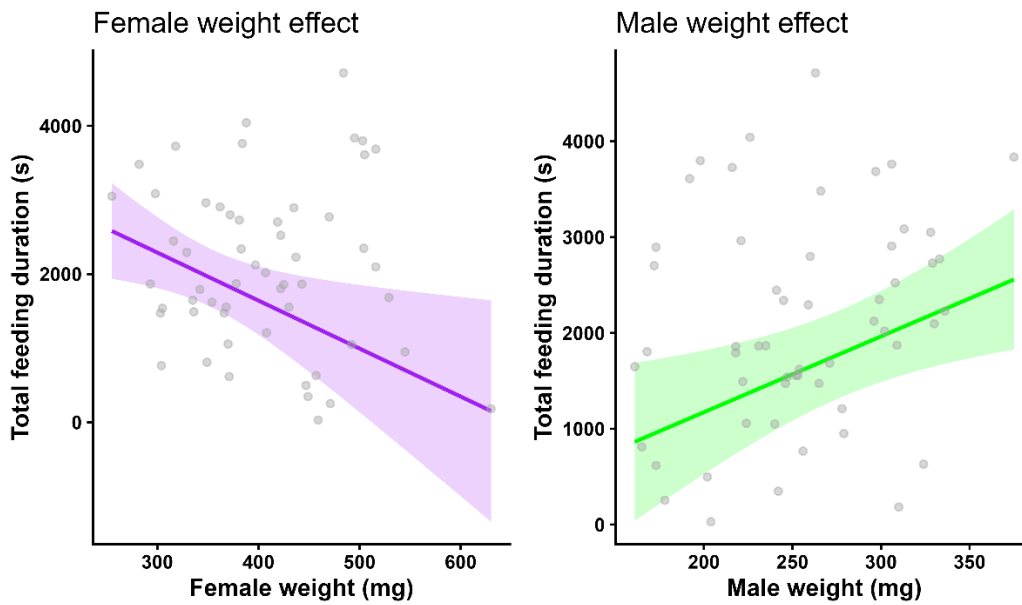
902 **ST 5:** Summary of Generalized Linear Mixed model for the effect of male rearing temperature, female
 903 rearing temperature, and their interaction on mating success.

Type	Predictor	Estimate (Log-Odds)	Std. Error	z value	p-value
Fixed Effects	(Intercept)	1.052	0.533	1.973	0.048
	Male Temperature (36°C)	-5.256	1.363	-3.858	< 0.001
	Female Temperature (36°C)	0.071	0.674	0.106	0.916
	Male Weight (z)	0.061	0.222	0.276	0.782
	Female Weight (z)	0.209	0.261	0.802	0.422
	Male Temperature × Female Temperature	2.665	1.324	2.013	0.044
Random Effects	Groups	Variance	Std. Dev.		
	Male ID	1.001	1.000		
	Female ID	< 0.001	< 0.001		

904

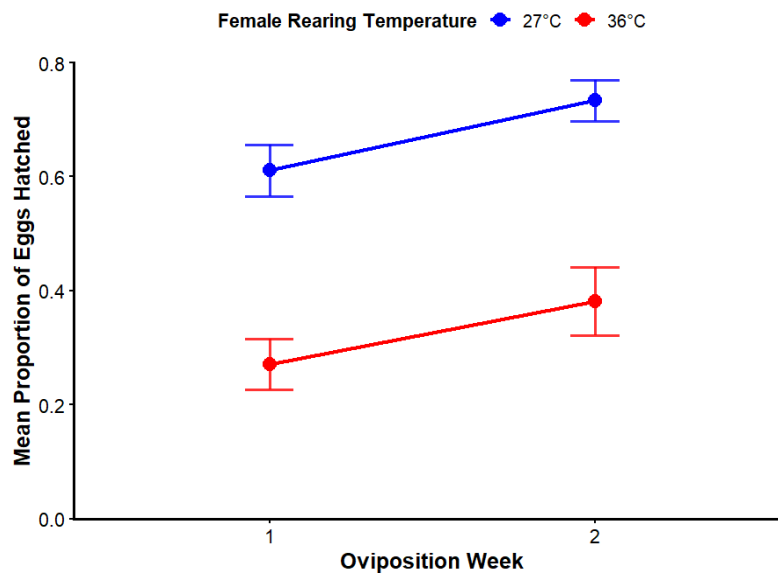
905

Model-adjusted body mass effects



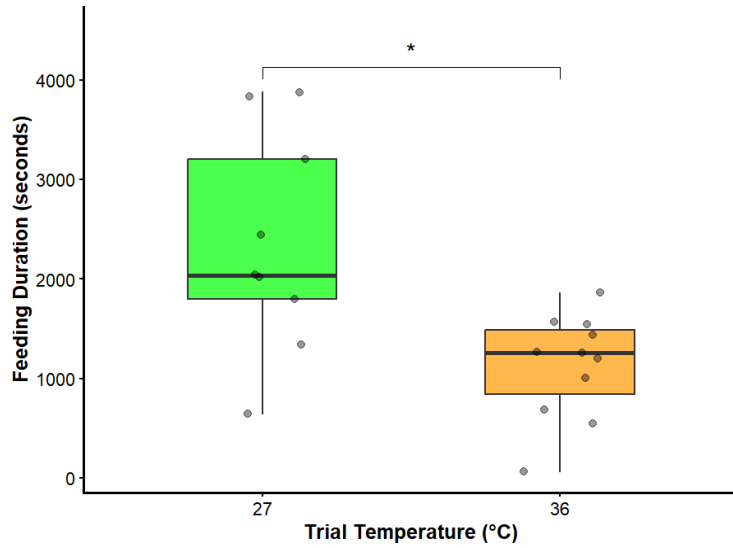
906

907 **S 1:** Model-adjusted relationships between body mass and total feeding duration. The left panel shows
908 the effect of female weight, and the right panel shows the effect of male weight. Grey points indicate
909 individual observations, coloured lines represent fitted linear model predictions, and shaded bands
910 show 95% confidence intervals. The models were adjusted for the remaining covariates in the full linear
911 model.



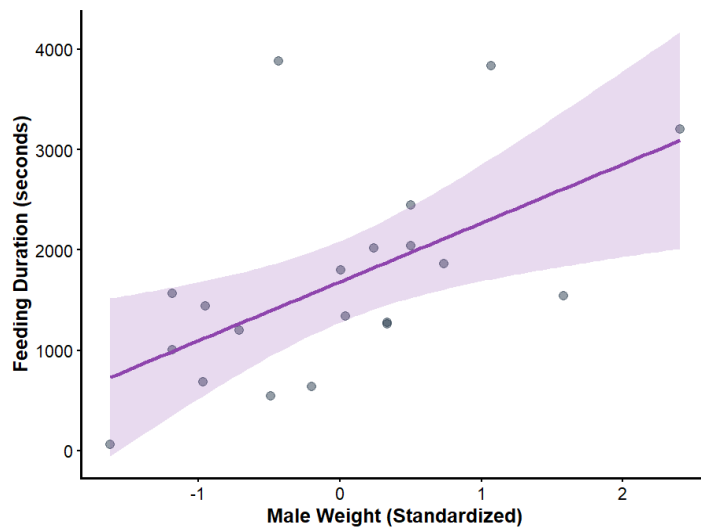
912

913 **S 2:** Effect of female developmental temperature on fertility (week-wise). Eggs laid by females reared
914 at 27 °C in the first- and second-week post-mating showed greater success of hatching than eggs laid
915 by females reared at 36 °C. Vertical error bars represent the standard error (\pm SE), while points
916 represent the mean.



924

925 *S 4: Effect of trial temperature on feeding duration of nuptial gift. At a higher ambient temperature of*
 926 *36 °C, females fed on the nuptial gift much faster than at 27 °C. Horizontal bars indicate the median*
 927 *feeding duration, and boxes indicate the interquartile range.*



928

929 *S 5: Effect of male weight on the feeding duration of the nuptial gift by females. Females mated with*
 930 *heavier males show an increase in feeding duration compared to those mated with lighter males.*

931 *ST 7: Summary of Negative Binomial Generalized Linear Mixed Model for the effect of trial*
 932 *temperature on the number of mating attempts*

Type	Predictor	Estimate (Log-Count)	Std. Error	z value	p-value
Fixed Effects	(Intercept)	0.412	0.183	2.246	0.025

	Trial Temperature (36°C)	0.634	0.248	2.555	0.011
	Male Weight (z)	0.118	0.111	1.065	0.287
	Female Weight (z)	0.009	0.132	0.069	0.945
Random Effects	Groups	Variance	Std. Dev.		
	Female ID	0.0977	0.3125		
	Male ID	< 0.001	< 0.001		

933

934 *ST 8: Summary of Linear Mixed-Effects Model for the effect of trial temperature on the feeding duration*
935 *of nuptial gift*

Type	Predictor	Estimate	Std. Error	df	t value	p-value
Fixed Effects	(Intercept)	2274.65	314.21	14.78	7.239	< 0.001
	Trial Temperature (36°C)	-1098.01	452.85	15.18	-2.425	0.028
	Male Weight (z)	536.39	230.32	15.80	2.329	0.034
	Female Weight (z)	-212.56	142.97	2.91	-1.487	0.236
Random Effects	Groups	Variance	Std. Dev.			
	Male ID	623,700	789.70			
	Female ID	< 0.001	< 0.001			
	Residual	59,040	243.00			

936

937 **ST 9:** Summary of Linear mixed effects model for the effect of male rearing temperature, female rearing
 938 temperature, their interaction, and female experience on log-transformed mating latency.

Type	Predictor	Estimate	Std. Error	df	t value	p-value
Fixed Effects	(Intercept)	1.972	0.258	73.13	7.634	< 0.001
	Male Temperature (36°C)	0.193	0.366	76.89	0.527	0.600
	Female Temperature (36°C)	0.684	0.359	78.90	1.905	0.060
	First Success (Yes)	-0.414	0.297	133.05	-1.395	0.165
	Male Weight (z)	-0.243	0.110	153.14	-2.207	0.029
	Female Weight (z)	-0.138	0.133	90.63	-1.036	0.303
	Female Temperature × Male Temperature	0.107	0.465	77.11	0.229	0.819
	Random Effects	Groups	Variance	Std. Dev.		
Female ID		0.1712	0.4138			
Male ID		0.0000	0.0000			