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Plastic shifts in thermal preference and thermoregulation strategy across ontogeny in an invasive fly

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ABSTRACT

Behavioural thermoregulation allows ectotherms to escape extreme or seek optimal temperatures. Its precision can impact survival and fitness under changing conditions and its plasticity can be an adaptive strategy when the plasticity of thermal limits is insufficient to buffer against warming. We explore the developmental plasticity of behavioural thermoregulation strategies using the invasive fly *Drosophila suzukii*, including *Wolbachia*-infected individuals, and long-term exposure to contrasting thermal regimes across multiple generations. We measured the plasticity of its thermal preference (Tp) and used its mean (thermal need) and variance (thermoregulation precision) to assess thermoregulation strategies. Typically, mean Tp increased with developmental temperature as well as the precision in temperature selection. Tp differed between life stages (higher in larvae and females than in males), reflecting different thermal needs. *Wolbachia* infection was associated with a reduction of Tp in adults but an increase in larvae, associated with a shift of the thermoregulation strategy with higher precision at intermediate Tp. Modulation of Tp may represent a mechanism for coping with changing or novel environmental conditions, opening new perspectives as to whether plasticity in Tp is adaptive under natural conditions, whether such plasticity facilitates colonisation or persistence during range expansion, and whether Tp plasticity itself evolves during range expansion.

INTRODUCTION

Ectotherms are particularly vulnerable to changes in temperature as their body temperature often matches the ambient or surface temperature with direct effect on their physiology (Angilletta, 2009). Altered temperature regimes such as those induced by climate change are already impacting species leading to shifts in distribution and extinctions (Pech et al., 2017; Renner and Zohner, 2018; Harvey et al., 2020). Ectotherms are threatened by temperatures exceeding their thermal limits across all latitudes (Sunday et al., 2014), especially during heat waves (Morley et al., 2019). Plastic responses constitute the main process by which ectotherms can adjust their tolerance level to heat waves in the short term (Kingsolver and Buckley, 2018; Buckley and Kingsolver, 2021). However, the level of plasticity of thermal limits (e.g., Gunderson et al., 2017; Fey et al., 2019), and of other thermal performance traits such as optimal temperature (Sinclair et al., 2016; Kingsolver and Buckley, 2017; Johnson et al., 2023), typically are insufficient to buffer against warming (Burton and Einum 2025).

Ectotherms can, alternatively, use behavioural thermoregulation to escape overheating (Leith et al., 2024) or to find optimal temperatures within heterogeneous habitats (Martin and Huey, 2008), which makes them more likely to survive (Danks, 1978; Ma and Ma, 2022). The precision of thermoregulation could, therefore, impact survival and fitness under changing conditions (Pincebourde and Woods, 2020; Ma et al., 2021; Deconninck et al., 2025). For example, a thermoregulation strategy that allows an organism to be highly precise when selecting a temperature that is optimal for a physiological process (e.g., development, egg maturation) can increase fitness directly. Alternatively, a strategy that allows an organism to be highly precise at selecting a temperature, when it approaches potentially deleterious temperature, can increase survival, and hence fitness, indirectly. In ectotherms, these thermoregulation strategies have been traditionally considered to be fixed at the species level (May, 1979; Lahondère, 2023), but some plasticity has been reported in various traits related

to behavioural thermoregulation (Gvoždík, 2012; Llewelyn et al., 2017). These levels of plasticity can also vary across ontogeny and may be linked to shifts in thermoregulation strategies, as reported in *D. melanogaster* (Deconninck et al., 2025). For example, thermoregulation precision may vary with temperature depending on the costs associated with sublethal temperature avoidance and the benefits of reaching the optimal temperature. This recent conceptual development of behavioural thermoregulation plastic responses (Deconninck et al., 2025) remains to be applied more broadly to infer the extent to which behavioural plasticity may help ectotherms to adapt rapidly to varying temperature conditions, including resistance or tolerance to extreme heat.

In this study, we explore the developmental plasticity of thermoregulation strategies in the invasive fly *Drosophila suzukii* (Mastumura, 1931). This pest species of soft fruit shows various seasonal adaptations (Tait et al., 2020, 2021), exploits different habitats (e.g., crops, woodland, urban areas; Ulmer et al., 2024), has invaded temperate and tropical areas (Walsh et al., 2011), has plastic thermal limits (Raynaud-Berton et al., 2024) and is an important example of a successful invader that is benefiting from climate change (Ward and Masters, 2007; Diez et al., 2012; Cornelissen et al., 2019; Little et al., 2020; Robinson et al., 2020; Skendžić et al., 2021; Deconninck et al., 2026). We used thermal preference (Tp) mean and variance as proxies of behavioural thermoregulation strategies using the framework developed by Deconninck et al. (2025). In this framework, the mean Tp reflects the thermal need (higher Tp likely reflecting a shift of optimum temperature of physiological processes to warmer temperature; Martin and Huey, 2015), the variance of Tp reflects the precision (the lower the variance, the higher the precision) and their relationship allows deciphering of behavioural thermoregulation strategies (Deconninck et al., 2025). We measured the Tp of *D. suzukii* across ontogeny (larvae, young and old adults), including males, virgin and fertile females, infected or not by the endosymbiont *Wolbachia*, under constant and fluctuating

temperature regimes across four generations. Temperature regimes were used to induce strong plastic responses allowing assessment of their impact on the behavioural strategy, particularly on thermoregulation precision. Contrasting results have been reported in the literature on the direction of the effects of temperature, life stage and *Wolbachia* infection on Tp (Yamamoto and Ohba, 1984; Krstevska and Hoffmann, 1994; McDaniel et al., 1995; Rajpurohit and Schmidt, 2016; MacLean et al., 2019; Truitt et al., 2019; Strunov et al., 2023). We hypothesised that life stage, sex and age would have a strong influence on Tp, predicting that larvae and females would seek higher temperatures to accelerate their development or egg maturation (Cohet and David, 1978; Gandara and Drummond-Barbosa, 2022; Grainger and Levine, 2022), and males would have lower Tp because of lower sterility threshold (Jørgensen et al., 2006; van Heerwaarden and Sgrò, 2021; Ørsted et al., 2024). We also assessed the stability of thermal responses across successive generations and expected Tp and behavioural strategy to be consistent among the four generations tested (Paranjpe et al., 2013; Castañeda et al., 2019).

METHODS

Origin and maintenance of experimental flies

Drosophila suzukii flies originated from a field collection of infested raspberries in Rennes, France (48°7'2.158"N, 1°40'40.053"W), in October 2020, by ECOBIO Laboratory (University of Rennes). One hundred and ninety isofemale lines were produced and nine of them were infected by *Wolbachia* as assessed by specific PCR assays with *Wolbachia*-specific primers targeting the *wsp* gene (81F/691R) following Braig et al. (1998). These infected lines were mixed in cages, and the population amplified. Tetracycline treatment was used to obtain separate lines differing by the presence (W+) or absence (W-) of *Wolbachia* following Hague et al. (2021). The microbiota of W- flies was restored by rearing W- flies

on a substrate inoculated with the faeces of W+ flies for three generations following Strunov et al. (2022), as used by Deconninck et al. (2024). The two lines were reared in 100 mL plastic bottles containing ~30 mL of standard cornmeal diet and incubated at 20°C, LD 12:12 h and 75% RH (STRADER, EV1300, Angers, France). At least 30 bottles of 50–200 flies per line were used for continuous maintenance. The *Wolbachia* infection status of the two lines was verified prior to the experiments using diagnostic PCR (Braig et al., 1998).

Temperature regimes

Groups of around a hundred 4-to-5-day-old adults from the source lines were placed for 5 d at 20°C in 100 mL bottles containing oviposition medium. Adults were then removed and bottles with eggs were randomly distributed between five temperature treatments: three constant regimes (16, 20, 24°C) and two fluctuating regimes differing in their thermal variance (18–22 and 16–24°C, both with mean temperature of 20°C) under L:D 12:12 h and 75% RH (climatic chambers, STRADER, Angers, France) (Figure 1A). In the fluctuating regimes, the coldest and warmest temperatures were reached at midnight and midday, respectively, and maintained for almost 11.5 hours (the transition between the two temperatures occurred over 30 min). Populations were maintained for four generations. A subsample of ~500 adult flies from each temperature treatment, ranging from the first to the last emergences, were distributed in eight bottles with fresh oviposition medium to produce the next generation.

Thermal gradient apparatus

Thermal preference (Tp) was measured in a linear thermal gradient (13–28 °C), using an apparatus similar to those described by Takeuchi et al. (2009), Goda et al. (2014) and Deconninck et al. (2025). The system consisted of an aluminium slab (500 × 250 × 20 mm)

heated at one end and cooled at the other using 45W Peltier devices (Quick-cool®, Germany) controlled by Arduino units (UNO R3). Water cooling prevented overheating of the Peltier elements and the apparatus was maintained in a climate-controlled laboratory at 22°C.

Ten lanes (400 × 10 × 5 mm) were machined into the slab and covered with transparent Makrolon® strips to prevent escape while allowing position tracking. Adults were introduced through small openings that were sealed during trials, whereas larval lanes were filled with a homogeneous gel (distilled water, sucrose 50 g.L⁻¹, agar 7g.L⁻¹) to permit movement. Insects' positions were recorded every 5 min using a camera (Kodak PixPro AZ422) mounted above the apparatus.

Before experiments, temperatures were calibrated in each lane using T-type thermocouples connected to HOBO data loggers (UX120-014M, ONSET). Temperatures were recorded every 30 sec for 60 min at 4-cm intervals, and linear regression were used to predict temperature from position. Gradients were highly linear (all R² > 0.99) and stable over time and between assays.

Thermal preference measurement

We use two units of the gradient apparatus described above, with all electrical parameters kept constant throughout the study, to measure the Tp of wandering third-instar larvae (L3), young adults (3 day-old) and old adults (13 day-old), including males, virgin and fertile females and W– and W+ individuals. We focused on L3 as earlier stages were delicate to manipulate in our design, and whether they exhibit similar thermoregulation behaviour remains unknown. The life stage, sex and reproductive status of adults were grouped under 'Life stage' (LifeS) for analyses and included seven levels: larvae (L3), 3- and 13-day-old virgin females (Fem_V_3d; Fem_V_13d), 3- and 13-day-old fertile females (Fem_F_3d;

Fem_F_13d), and 3- and 13-day-old males (Male_3d; Male_13d) (Table 1; Figure 1B). Measurements were made on individuals from generations 1 to 4 (G1–G4; Figure 1A).

To assess the range of T_p at the population level, 50 individuals with a unique combination of variables were introduced into five lanes (10×5 replicates; Figure 1C). In total, about 14,000 individuals were measured (50×7 life stages $\times 5$ temperature regimes $\times 2$ *Wolbachia* infection status $\times 4$ generations). The lanes were considered as independent replicates because no communication or contact was possible among individuals from different lanes. Before starting the assays, the gradient was switched on 15 min to allow stabilisation. Adult flies or larvae were placed in the centre of the lane at a temperature of $\sim 20^\circ\text{C}$ (through the hole). The lanes were photographed every 5 min over 60 min to document individual positions. Initial trial runs indicated that the movements of individuals were much less frequent after about 20 min. Therefore, we considered the T_p of every individual to be the temperature along the thermal gradient at their position after 45 min, according to common practice in *Drosophila* T_p assessment (position after 37 ± 14 min; see Deconninck et al., 2025 for review). All T_p assays were run within the same daily time window (between 0900 and 1200), to minimise any influence of circadian rhythm on T_p , and light was homogeneous to avoid any attraction toward a given direction (Dillon et al., 2009). All flies were handled with a mouth aspirator to avoid confounding effects of CO_2 anaesthesia (Nilson et al., 2006; Milton and Partridge, 2008).

Statistical analyses

Data were analysed in R version 4.5.3 (R Core Team, 2021). Thermal preference (T_p) was analysed using linear mixed-effects models fitted using the lmer function from the *lme4* package (Bates et al., 2015). The model included temperature treatment, generation (G1–G4), *Wolbachia* infection status, life stage, and all pairwise interactions among these factors as

fixed effects. Gradient and lane nested within gradient system were included as random effects to account for the experimental design. Higher-order interactions were evaluated but were not retained because they substantially increased model complexity without improving model support ($\Delta AIC = 89.4$), produced non-estimable marginal means for sparse factor combinations, and did not alter the biological interpretation of the results. Model-estimated contrasts were obtained from estimated marginal means (EMMs) using the *emmeans* package (Lenth and Piaskowski 2026). Pairwise treatment-versus-control contrasts were calculated using 20 °C (temperature treatment), L3 (life stage), or *Wolbachia*-uninfected (W-) individuals as the reference level, depending on the factor analysed. Estimated contrasts (ΔT_p relative to the reference level) and their standard errors were extracted and visualised as point estimates with 95% confidence intervals.

To identify thermoregulation strategies, we quantified the mean T_p , used as an index of thermal need, and the variance in T_p among individuals within each experimental lane, used as an index of thermoregulatory precision (following Deconninck et al., 2025). Each lane contained a group of 10 individuals sharing the same combination of developmental temperature, life stage and *Wolbachia* infection status from the same generation. For each lane, we calculated the mean T_p and variance of T_p . We then averaged these lane-level means and within-lane variances across the five replicate lanes corresponding to each developmental temperature treatment. This approach preserves the biological interpretation of within-lane behavioural variation while allowing lane to be treated as the experimental replicate in subsequent analyses (see below). The relationship between mean T_p and within-lane T_p variance was first explored using a LOESS smoothing procedure (span = 10) within each life stage $\times$ *Wolbachia* combination, following Deconninck et al. (2025). This non-parametric approach allowed visual exploration of potential thermoregulation strategies within each life stage, without imposing a predefined functional form while accounting for

the broad variation in T_p induced by developmental temperature treatments. We then formally tested these relationships using second-order polynomial linear models including mean T_p , *Wolbachia* infectious status, life stage, and their interactions as fixed effects. The significance of the Linear (β_1) and quadratic (β_2) components was used to classify the relationship for each life stage $\times$ *Wolbachia* combination as linear, U-shaped, inverted U-shaped, or non-significant. These analyses on thermoregulation strategies were performed on pooled data from generations G1–G4 to increase the power of the test, and because generation had only a weak effect. We were mostly interested in the variation induced by temperature treatments to assess thermoregulation strategies of each life stage, but the thermoregulation strategies can also be assessed within each temperature treatment following the same procedure to explore the variation induced by ontogeny (Figure S3).

RESULTS

Important T_p plasticity

All variables and most of their interactions had significant effects on T_p , with temperature treatment, life stage and *Wolbachia* having the strongest effects (all $p < 0.0001$; Table 2; Figure S1). Temperature treatment effect depended on life stage with larval T_p being much less influenced (Figure 2A) but most adults selecting lower temperatures when developed at 16 °C and higher temperatures when developed at 24 °C (mean $\pm$ se: 19.0 ± 0.1 and 20.5 ± 0.1 °C, respectively; Figure 2A). Individuals that developed under the low fluctuating regime (18–22 °C) had T_p similar to those developed at constant 20 °C (both 19.9 ± 0.1 °C) but selected lower T_p under the high fluctuating regime (16–24 °C), similar to the constant 16 °C regime (19.1 ± 0.1 °C; Figure 2A). Regarding life stage, males had the lowest T_p (18.8 ± 0.1 °C both 3 d and 13 d), virgin females had intermediate T_p (3 d: 20.2 ± 0.1 ; 13 d: 19.2 ± 0.1 °C), and larvae and fertile females had the highest T_p (L3: 20.6 ± 0.1 ; 3 d: 20.9 ± 0.1 ; 13 d:

20.5 $\pm$ 0.1 °C, respectively) (Figure S1, 2A, 2B). *Wolbachia*-infected individuals generally had reduced Tp compared to non-infected individuals (W–: 20.1 $\pm$ 0.0°C; W+: 19.6 $\pm$ 0.0 °C), but the effect was driven by adults having lower Tp when infected, while larvae had a higher Tp when infected (Figure 2C). Finally, Tp decreased from 20.0 $\pm$ 0.1 °C in G1 to 19.6 $\pm$ 0.1 °C in G3 but further returned to values equivalent to those in G1 (G4: 19.8 $\pm$ 0.1; Figure 2D).

Distinct behavioural thermoregulation strategies among life stages

Only a subset of life stages showed significant relationships between Tp variance (reflecting precision) and Tp mean (reflecting thermal need), and *Wolbachia* infection appeared to change the shape of this relationship (Figure 3, Table S1). W– larvae, old fertile females and old males had a lower Tp variance at high mean Tp, suggesting a ‘precision at high temperature’ strategy (Deconninck et al., 2025). W– young males and W+ old males had a low Tp variance at low and high mean Tp, suggesting a ‘precision at extreme’ strategy (inverted U-shape; Deconninck et al., 2025). W+ larvae had a high Tp variance at low and high mean Tp, suggesting a ‘precision at optimum’ strategy (U-shape; Deconninck et al., 2025).

DISCUSSION

Behavioural thermoregulation can buffer environmental change before ectotherms reach their thermal limits. However, understanding its adaptive significance requires integrating multiple sources of phenotypic plasticity rather than considering Tp alone (Gvoždík, 2012). Here, we show that Tp in *D. sukuzii* is highly plastic, responding to developmental temperature, life stage and *Wolbachia* infection, with strong interactions among these factors. Despite this plasticity, thermoregulatory strategies were relatively conserved across ontogeny and were generally characterised by “precision at extreme” (low Tp variance near deleterious

temperatures). A notable exception was the contrasting effect of *Wolbachia* on larval and adult thermoregulation, suggesting different underlying mechanisms. Overall, our results indicate that predicting thermoregulatory behaviour in the field requires knowledge of an individual's thermal history, life stage, reproductive and infection status.

Multiple drivers of thermal preference plasticity

The Tp of *D. sukukii* was highly plastic, responding to all variables tested, with temperature, life stage (including ontogeny and sex) and *Wolbachia* having the strongest effects.

Increasing developmental temperature generally increased Tp, a pattern already reported in other ectotherms, including seahorses, shrimps, killifish and spiders (Podrabsky et al., 2008; Alfaro et al., 2013; Reiser et al., 2014; Mascaró et al., 2019). Studies in *Drosophila* have likewise found positive effects of temperature on Tp (Yamamoto and Ohba, 1984; McDaniel et al., 1995), although others reported no effect (MacLean et al., 2019) or reduced Tp at temperatures above the thermal optimum (Krstevska and Hoffmann, 1994). Our results suggest that the inconsistencies between studies may reflect differences in sex, age and reproductive status, all of which modulated the magnitude, and sometimes the direction, of the temperature effect. Exposure to stressful temperatures beyond the thermal optimum, as well as thermal hardening, is also expected to reduce Tp (McDaniel et al., 1995; Dillon et al., 2009).

The Tp of *D. sukukii* responded differently to constant and fluctuating temperature regimes. Low amplitude fluctuations (18–22°C) produced the same Tp as a constant temperature with the same mean (20 °C), consistent with findings in *D. melanogaster* (Deconninck et al., 2025). In contrast, high-amplitude fluctuations (16–24 °C) reduced Tp to levels similar to those observed after development at a constant 16 °C. One possible explanation is a lower Tp protects against potentially harmful high temperatures, as suggested

for killifish (Podrabsky et al., 2008), although *D. sukii* tolerates temperatures well above 30 °C (Enriquez and Colinet, 2017). Alternatively, because *D. sukii* that developed at a constant 24 °C selected a higher T_p than those reared under high fluctuations, the lower end of the fluctuating regime may drive the reduction in T_p . This is consistent with recent findings reporting that the thermal optimum for survival shifts from ~25 to 18 °C under constant versus similarly fluctuating temperatures (Raynaud-Berton et al., 2024), suggesting that the lower T_p is an adaptive response to improved performance at lower mean temperature.

In *Drosophila*, both T_p and its plasticity generally differ between larvae and adults (MacLean et al., 2019). In *D. sukii*, larvae had higher T_p than adults overall, although similar to fertile females. Selecting warmer temperatures allows faster larval development, providing a competitive advantage and reducing predation risk before emergence (Nunney 1990; Grainger and Levine, 2022; but see also ‘grow now, pay later’ hypothesis of Metcalfe and Monaghan (2001)). Although larval sex could not be determined, the low variance in T_p suggests that males and females display similar thermoregulatory behaviour at this stage. Adult females had higher T_p than males, particularly after females had mated. Fertile females may select higher temperatures to accelerate oogenesis (Cohet and David, 1978; Gandara and Drummond-Barbosa, 2022), and to optimise larval fitness as shown in Destierdt et al. (2026). Conversely, virgin females and older adults may prefer cooler temperatures to reduce metabolic costs and increase longevity (Marshall and Sinclair, 2010). Males had the lowest T_p , consistent with evidence that male fertility is more sensitive to elevated temperatures than female fertility (Jørgensen et al., 2006; van Heerwaarden and Sgrò, 2021; Ørsted et al., 2024).

Wolbachia-infected adults exhibited a lower T_p , consistent with findings in *D. melanogaster*, where multiple *Wolbachia* strains reduced adult T_p by 2–8 °C (Truitt et al., 2019), arguing against the hypothesis that *Wolbachia* effects on T_p reflect confounding

factors (Strunov et al., 2023). A lower T_p could be a self-medicating behaviour to reduce bacterial proliferation (“behavioural chill”; Fedorka et al., 2016) and thereby limit fitness costs associated with *Wolbachia* over-proliferation (Strunov et al., 2013a,b; 2024). In contrast, infected larvae exhibited a slightly higher T_p (up to 0.8 °C), a pattern not previously reported that could reflect *Wolbachia*-mediated manipulation of host behaviour (see thermoregulation strategy section below).

Finally, the generation had only a minor influence on *D. sukii* T_p . In ectotherms, T_p often displays low to moderate heritability (see Bestion et al. (2023) for an example in lizards), whereas maternal effects may contribute more strongly to variation in T_p (Paranjpe et al., 2013). In *D. melanogaster*, bet-hedging limits the heritability of T_p , supposedly reflecting adaptation to seasonal environments (Kain et al., 2015). We propose that a similar strategy applies to *D. sukii* whose life cycle is particularly adapted to seasonal variations (De Ro et al., 2025). This prediction could be evaluated by analysing inter-individual variation in T_p across generations and under different selection regimes (see also Deconninck et al., 2025).

Thermoregulation strategy: Be precise when approaching thermal extremes

The increase in mean T_p associated with warmer developmental temperatures was generally accompanied by more precise temperature selection (lower T_p variance), consistent with the “precision at extremes” and “precision at high temperature” strategies described by Deconninck et al. (2025). This pattern is consistent with studies showing that behavioural thermoregulation in herbivorous insects primarily aims to avoid overheating rather than maximise reproductive performance (Leith et al., 2024). In contrast, spider mites appear to rely more on physiological than behavioural mechanisms to tolerate heat because their leaf surface habitat provides limited thermal heterogeneity (Caillon et al., 2014; Ma et al., 2021).

Fruit flies, by comparison, can readily move or fly to cooler microhabitats and are expected to benefit from behavioural avoidance of sublethal heat stress, particularly because fertility declines at temperatures below those affecting survival (Jørgensen et al., 2006; Parratt et al., 2021; van Heerwaarden and Sgrò, 2021; Ørsted et al., 2024). More generally, Tp variance was lower in *D. suzukii* than reported for *D. melanogaster* (Deconninck et al., 2025), even in life stages that showed little evidence of plastic thermoregulatory strategies. This consistently high precision may contribute to the invasion success of *D. suzukii* by allowing exploitation of fine scale thermal refuges, although direct evidence is still lacking. Alternatively, the weak relationship between Tp mean and Tp variance could reflect limited plasticity in *D. suzukii* or to the relatively permissive thermal regimes used here.

Regardless of *Wolbachia* infection, larvae preferred warmer temperatures than any other life stage. This is consistent with their ecology, as larvae develop within fruit that can reach temperatures well above ambient air temperature under solar radiation (Woolf and Ferguson, 2000; Saudreau et al., 2009), and their developmental optimum ($\sim 28^{\circ}\text{C}$) exceeds that of adults ($\sim 24\text{--}27^{\circ}\text{C}$) (Sánchez-Ramos et al., 2019; Winkler et al., 2021; Baser et al., 2025; Raynaud-Berton et al., 2024). Although larvae are confined to their host fruit, they may exploit within-fruit thermal heterogeneity to avoid overheating or approach their thermal optimum (Kührt et al., 2005; Saudreau et al., 2007, 2009, 2011). Notably, *Wolbachia* not only increased larval Tp, opposite to its effect in adults, but also shifted thermoregulation towards a “precision at optimum” strategy (Deconninck et al., 2025), with infected larvae selecting intermediate temperatures more precisely than uninfected larvae. Assuming that Tp reflects the thermal optimum for performance (Martin and Huey, 2008), this behaviour could enhance larval foraging and development, which is the main function of thermoregulation behaviour reported in reptiles (Ladyman et al., 2003; Taylor et al., 2021). Alternatively, *Wolbachia* may manipulate larval thermoregulation to favour its own proliferation by promoting the selection

of warmer microhabitats (Fedorka et al., 2016; Strunov et al., 2013a,b). Although speculative, this contrasts with the proposed “behavioural chill” response in adults and suggests that Wolbachia-host interactions may differ fundamentally between larval and adult stages.

CONCLUSIONS

Drosophila suzukii exhibited marked plasticity in Tp, with all tested variables influencing its expression. Although we could not directly assess the fitness consequences of this plasticity, modulation of Tp may help individuals to cope with extreme or novel environments (Gvoždík, 2012; Ma et al., 2021). In *D. suzukii* and other drosophilids, behavioural thermoregulation plasticity may therefore have contributed to range expansion and invasive success (Deconninck et al., 2026). Whether Tp measured in artificial thermal gradients reflects thermoregulatory behaviour in more complex, natural conditions remains uncertain. Nevertheless, our approach of inferring thermoregulatory strategies from both the mean and variance of Tp provides a useful conceptual framework. Future studies should determine whether Tp plasticity is adaptive under natural conditions, facilitates colonisation and persistence during range expansion, and itself evolves during range expansion, as predicted by theoretical models (Eriksson and Rafajlović, 2022; Usui et al., 2023; Swaegers et al., 2024).

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AUTHOR CONTRIBUTIONS

GD, VF and SP conceived the ideas and designed methodology; GD collected and analysed the data and led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

REFERENCES

- Alfaro C, Figueroa DP, Torres H, Veloso C, Venegas F, Canals L, Canals M. 2013 Effect of thermal acclimation on preferred temperatures in two mygalomorph spiders inhabiting contrasting habitats. *Physiol. Entomol.* **38**, 20–25. <https://doi.org/10.1111/j.1365-3032.2012.00853.x>
- Angilletta Jr. MJ. 2009 *Thermal Adaptation: A Theoretical and Empirical Synthesis*. Oxford University Press. <https://doi.org/10.1093/acprof:oso/9780198570875.001.1>

- Baser N, Rossini L, Anfora G, Temel KM, Gualano S, Garone E, Santoro F. 2025 Thermal development, mortality, and fertility of an Apulian strain of *Drosophila suzukii* at different temperatures. *Insects* **16**, 60. <https://doi.org/10.3390/insects16010060>
- Bates D, Mächler M, Bolker B, Walker S. 2015 Fitting linear mixed-effects models using lme4. *J. Stat. Softw.* **67**, 1–48. <https://doi.org/10.18637/jss.v067.i01>
- Bestion E, San-Jose LM, Di Gesu L, Richard M, Sinervo B, Côte J, Calvez O, Guillaume O, Cote J. 2023 Plastic responses to warmer climates: A semi-natural experiment on lizard populations. *Evolution* **77**, 1634–1646. <https://doi.org/10.1093/evolut/qpad070>
- Braig HR, Zhou W, Dobson SL, O’Neill SL. 1998 Cloning and characterization of a gene encoding the major surface protein of the bacterial endosymbiont *Wolbachia pipientis*. *J. Bacteriol.* **180**, 2373–2378. <https://doi.org/10.1128/JB.180.9.2373-2378.1998>
- Buckley LB, Kingsolver JG. 2021 Evolution of thermal sensitivity in changing and variable climates. *Annu. Rev. Ecol. Evol. Syst.* **52**, 563–586. <https://doi.org/10.1146/annurev-ecolsys-011521-102856>
- Burton T, Einum S. 2025 High capacity for physiological plasticity occurs at a slow rate in ectotherms. *Ecol. Lett.* **28**, 1–9. <https://doi.org/10.1111/ele.70046>
- Caillon R, Suppo C, Casas J, Arthur Woods H, Pincebourde S. 2014 Warming decreases thermal heterogeneity of leaf surfaces: Implications for behavioural thermoregulation by arthropods. *Funct. Ecol.* **28**, 1449–1458. <https://doi.org/10.1111/1365-2435.12288>
- Castañeda LE, Romero-Soriano V, Mesas A, Roff DA, Santos M. 2019 Evolutionary potential of thermal preference and heat tolerance in *Drosophila subobscura*. *J. Evol. Biol.* **32**, 818–824. <https://doi.org/10.1111/jeb.13483>
- Cohet Y, David J. 1978 Control of the adult reproductive potential by preimaginal thermal conditions - A study in *Drosophila melanogaster*. *Oecologia* **36**, 295–306. <https://doi.org/10.1007/BF00348055>

415 Cornelissen B, Neumann P, Schweiger O. 2019 Global warming promotes biological
 416 invasion of a honey bee pest. *Glob. Chang. Biol.* **25**, 3642–3655.
 417 <https://doi.org/10.1111/gcb.14791>
 418 Danks H V. 1978 Modes of seasonal adaptation in the insects: I. Winter survival. *Can.*
 419 *Entomol.* **110**, 1167–1205. <https://doi.org/10.4039/Ent1101167-11>
 420 De Ro M, Devos T, Berkvens N, Casteels H, Bonte J, Colinet H, De Clercq P. 2025 Cold
 421 tolerance of European populations of *Drosophila suzukii* varies among seasonal
 422 phenotypes. *J. Therm. Biol.* **132**, 104251. <https://doi.org/10.1016/j.jtherbio.2025.104251>
 423 Deconninck G, Larges J, Henri H, Beaugiard L, Foray V, Pincebourde S. 2024 *Wolbachia*
 424 improves the performance of an invasive fly after a diet shift. *J. Pest Sci. (2004)*. **97**,
 425 2087–2099. <https://doi.org/10.1007/s10340-023-01739-w>
 426 Deconninck G, Meyer N, Colinet H, Pincebourde S. 2025 Thermal preference plasticity in
 427 ectotherms: integrating temperature affinity and thermoregulation precision. *Am. Nat.*
 428 **203**, 247–260. <https://doi.org/10.1086/736575>
 429 Deconninck G *et al.* 2026 A roadmap to key traits of invasive Drosophilidae. *Biol. Rev.*
 430 <https://doi.org/10.1002/brev.70148>
 431 Destierdt W, Deconninck G, Crespo JE, Moyer E, Foray V, Chabrierie O, Pincebourde P.
 432 2026 Temperature overrides nutritional cues for optimal oviposition decision in a
 433 polyphagous invasive insect. *J. Exp. Biol.* **229**, jeb251743.
 434 <https://doi.org/10.1242/jeb.251743>
 435 Diez JM *et al.* 2012 Will extreme climatic events facilitate biological invasions? *Front.*
 436 *Ecol. Environ.* **10**, 249–257. <https://doi.org/10.1890/110137>
 437 Dillon ME, Wang G, Garrity PA, Huey RB. 2009 Thermal preference in *Drosophila*. *J.*
 438 *Therm. Biol.* **34**, 109–119. <https://doi.org/10.1016/j.jtherbio.2008.11.007>

Enriquez T, Colinet H. 2017 Basal tolerance to heat and cold exposure of the spotted wing drosophila, *Drosophila suzukii*. *PeerJ* **5**, e3112. <https://doi.org/10.7717/peerj.3112>

Eriksson M, Rafajlović M. 2022 The role of phenotypic plasticity in the establishment of range margins. *Philos. Trans. R. Soc. B* **377**. <https://doi.org/10.1098/rstb.2021.0012>

Fedorka KM, Kutch IC, Collins L, Musto E. 2016 Cold temperature preference in bacterially infected *Drosophila melanogaster* improves survival but is remarkably suboptimal. *J. Insect Physiol.* **93–94**, 36–41. <https://doi.org/10.1016/j.jinsphys.2016.08.005>

Fey SB *et al.* 2019 Opportunities for behavioral rescue under rapid environmental change. *Glob. Chang. Biol.* **25**, 3110–3120. <https://doi.org/10.1111/gcb.14712>

Gandara ACP, Drummond-Barbosa D. 2022 Warm and cold temperatures have distinct germline stem cell lineage effects during *Drosophila* oogenesis. *Development* **149**. <https://doi.org/10.1242/dev.200149>

Goda T, Leslie JR, Hamada FN. 2014 Design and analysis of temperature preference behavior and its circadian rhythm in *Drosophila*. *J. Vis. Exp.* 83, e51097. <https://doi.org/10.3791/51097>

Grainger TN, Levine JM. 2022 Rapid evolution of life-history traits in response to warming, predation and competition: A meta-analysis. *Ecol. Lett.* **25**, 541–554. <https://doi.org/10.1111/ele.13934>

Gunderson AR, Dillon ME, Stillman JH. 2017 Estimating the benefits of plasticity in ectotherm heat tolerance under natural thermal variability. *Funct. Ecol.* **31**, 1529–1539. <https://doi.org/10.1111/1365-2435.12874>

Gvoždík L. 2012 Plasticity of preferred body temperatures as means of coping with climate change? *Biol. Lett.* **8**, 262–265. <https://doi.org/10.1098/rsbl.2011.0960>

- Hague MTJ, Woods HA, Cooper BS. 2021 Pervasive effects of *Wolbachia* on host activity. *Biol. Lett.* **17**, rsbl.2021.0052. <https://doi.org/10.1098/rsbl.2021.0052>
- Harvey JA, Heinen R, Gols R, Thakur MP. 2020 Climate change-mediated temperature extremes and insects: From outbreaks to breakdowns. *Glob. Chang. Biol.* **26**, 6685–6701. <https://doi.org/10.1111/gcb.15377>
- Johnson CA, Ren R, Buckley LB. 2023 Temperature sensitivity of fitness components across life cycles drives insect responses to climate change. *Am. Nat.* **202**, 753–766. <https://doi.org/10.1086/726896>
- Jørgensen KT, Sørensen JG, Bundgaard J. 2006 Heat tolerance and the effect of mild heat stress on reproductive characters in *Drosophila buzzatii* males. *J. Therm. Biol.* **31**, 280–286. <https://doi.org/10.1016/j.jtherbio.2005.11.026>
- Kingsolver JG, Buckley LB. 2017 Quantifying thermal extremes and biological variation to predict evolutionary responses to changing climate. *Philos. Trans. R. Soc. B Biol. Sci.* **372**, 20160147. <https://doi.org/10.1098/rstb.2016.0147>
- Kingsolver JG, Buckley LB. 2018 How do phenology, plasticity, and evolution determine the fitness consequences of climate change for montane butterflies? *Evol. Appl.* **11**, 1231–1244. <https://doi.org/10.1111/eva.12618>
- Krstevska B, Hoffmann AA. 1994 The effects of acclimation and rearing conditions on the response of tropical and temperate populations of *Drosophila melanogaster* and *D. simulans* to a temperature gradient (Diptera: Drosophilidae). *J. Insect Behav.* **7**, 279–288. <https://doi.org/10.1007/BF01989735>
- Kührt U, Samietz J, Dorn S. 2005 Thermoregulation behaviour in codling moth larvae. *Physiol. Entomol.* **30**, 54–61. <https://doi.org/10.1111/j.0307-6962.2005.00431.x>
- Lahondère C. 2023 Recent advances in insect thermoregulation. *J. Exp. Biol.* **226**. <https://doi.org/10.1242/jeb.245751>

- Ladyman M, Bonnet X, Lourdaïs O, Bradshaw D, Naulleau G. 2003 Gestation, thermoregulation, and metabolism in a viviparous snake, *Vipera aspis*: Evidence for fecundity-independent costs. *Physiol. Biochem. Zool.* **76**, 497–510. <https://doi.org/10.1086/376420>
- Leith NT, Miller EA, Fowler-Finn KD. 2024 Thermoregulation enhances survival but not reproduction in a plant-feeding insect. *Funct. Ecol.* **38**, 1344–1356. <https://doi.org/10.1111/1365-2435.14546>
- Lenth R, Piaskowski J. 2026 emmeans: Estimated Marginal Means, aka Least-Squares Means. R package version 2.0.3, <https://rvlenth.github.io/emmeans/>.
- Little CM, Chapman TW, Hillier NK. 2020 Plasticity is key to success of *Drosophila suzukii* (Diptera: Drosophilidae) invasion. *J. Insect Sci.* **20**, 5. <https://doi.org/10.1093/jisesa/ieaa034>
- Llewelyn J, Macdonald S, Hatcher A, Moritz C, Phillips BL. 2017 Thermoregulatory behaviour explains countergradient variation in the upper thermal limit of a rainforest skink. *Oikos* **126**, 748–757. <https://doi.org/10.1111/oik.03933>
- Ma C-S, Ma G, Pincebourde S. 2021 Survive a warming climate: Insect responses to extreme high temperatures. *Annu. Rev. Entomol.* **66**, 163–184. <https://doi.org/10.1146/annurev-ento-041520-074454>
- Ma G, Ma C Sen. 2022 Potential distribution of invasive crop pests under climate change: incorporating mitigation responses of insects into prediction models. *Curr. Opin. Insect Sci.* **49**, 15–21. <https://doi.org/10.1016/j.cois.2021.10.006>
- MacLean HJ, Sørensen JG, Kristensen TN, Loeschcke V, Beedholm K, Kellermann V, Overgaard J. 2019 Evolution and plasticity of thermal performance: An analysis of variation in thermal tolerance and fitness in 22 *Drosophila* species. *Philos. Trans. R. Soc. B Biol. Sci.* **374**, 20180548. <https://doi.org/10.1098/rstb.2018.0548>

- Marshall KE, Sinclair BJ. 2010 Repeated stress exposure results in a survival-reproduction trade-off in *Drosophila melanogaster*. *Proc. R. Soc. B Biol. Sci.* **277**, 963–969. <https://doi.org/10.1098/rspb.2009.1807>
- Martin TL, Huey RB. 2008 Why ‘suboptimal’ is optimal: Jensen’s inequality and ectotherm thermal preferences. *Am. Nat.* **171**, 102–118. <https://doi.org/10.1086/527502>
- Mascaró M, Horta JL, Diaz F, Paschke K, Rosas C, Simões N. 2019 Effect of a gradually increasing temperature on the behavioural and physiological response of juvenile *Hippocampus erectus*: Thermal preference, tolerance, energy balance and growth. *J. Therm. Biol.* **85**, 102406. <https://doi.org/10.1016/j.jtherbio.2019.102406>
- May ML. 1979 Insect thermoregulation. *Annu. Rev. Entomol.* **24**, 313–349. <https://doi.org/10.1146/annurev.en.24.010179.001525>
- McDaniel R, Hostert EE, Seager RD. 1995 Acclimation and adaptive behavior of *Drosophila robusta* and *D. tripunctata* adults in response to combined temperature and desiccation stress. *Am. Midl. Nat.* **133**, 52. <https://doi.org/10.2307/2426347>
- Metcalfe NB, Monaghan P. 2001 Compensation for a bad start: grow now, pay later? *Trends Ecol. Evol.* **16**, 254–260. [https://doi.org/10.1016/S0169-5347\(01\)02124-3](https://doi.org/10.1016/S0169-5347(01)02124-3)
- Milton CC, Partridge L. 2008 Brief carbon dioxide exposure blocks heat hardening but not cold acclimation in *Drosophila melanogaster*. *J. Insect Physiol.* **54**, 32–40. <https://doi.org/10.1016/j.jinsphys.2007.08.001>
- Morley SA, Peck LS, Sunday JM, Heiser S, Bates AE. 2019 Physiological acclimation and persistence of ectothermic species under extreme heat events. *Glob. Ecol. Biogeogr.* **28**, 1018–1037. <https://doi.org/10.1111/geb.12911>
- Nilson TL, Sinclair BJ, Roberts SP. 2006 The effects of carbon dioxide anesthesia and anoxia on rapid cold-hardening and chill coma recovery in *Drosophila melanogaster*. *J. Insect Physiol.* **52**, 1027–1033. <https://doi.org/10.1016/j.jinsphys.2006.07.001>

- Nunney L. 1990 *Drosophila* on oranges: colonization, competition, and coexistence. *Ecology* **71**, 1904–1915. <https://doi.org/10.2307/1937598>
- Ørsted M, Willot Q, Olsen AK, Kongsgaard V, Overgaard J. 2024 Thermal limits of survival and reproduction depend on stress duration: A case study of *Drosophila suzukii*. *Ecol. Lett.* **27**, 1–13. <https://doi.org/10.1111/ele.14421>
- Paranjpe DA, Bastiaans E, Patten A, Cooper RD, Sinervo B. 2013 Evidence of maternal effects on temperature preference in side-blotched lizards: implications for evolutionary response to climate change. *Ecol. Evol.* **3**, 1977–1991. <https://doi.org/10.1002/ece3.614>
- Parratt SR, Walsh BS, Metelmann S, White N, Manser A, Bretman AJ, Hoffmann AA, Snook RR, Price TAR. 2021 Temperatures that sterilize males better match global species distributions than lethal temperatures. *Nat. Clim. Chang.* **11**, 481–484. <https://doi.org/10.1038/s41558-021-01047-0>
- Pecl GT *et al.* 2017 Biodiversity redistribution under climate change: Impacts on ecosystems and human well-being. *Science*. **355**, 1389. <https://doi.org/10.1126/science.aai9214>
- Pincebourde S, Casas J. 2015 Warming tolerance across insect ontogeny: influence of joint shifts in microclimates and thermal limits. *Ecology* **96**, 986–997. <https://doi.org/10.1890/14-0744.1>
- Pincebourde S, Woods HA. 2020 There is plenty of room at the bottom: microclimates drive insect vulnerability to climate change. *Curr. Opin. Insect Sci.* **41**, 63–70. <https://doi.org/10.1016/j.cois.2020.07.001>
- Podrabsky JE, Clelen D, Crawshaw LI. 2008 Temperature preference and reproductive fitness of the annual killifish *Austrofundulus limnaeus* exposed to constant and fluctuating temperatures. *J. Comp. Physiol. A Neuroethol. Sensory, Neural, Behav. Physiol.* **194**, 385–393. <https://doi.org/10.1007/s00359-008-0313-7>

- R Core Team. 2021 R: A language and environment for statistical computing. Austria, Vienna.
- Rajpurohit S, Schmidt PS. 2016 Measuring thermal behavior in smaller insects: A case study in *Drosophila melanogaster* demonstrates effects of sex, geographic origin, and rearing temperature on adult behavior. *Fly (Austin)*. **10**, 149–161.
<https://doi.org/10.1080/19336934.2016.1194145>
- Raynaud-Berton B, Gibert P, Suppo C, Pincebourde S, Colinet H. 2024 Modelling thermal reaction norms for development and viability in *Drosophila suzukii* under constant, fluctuating and field conditions. *J. Therm. Biol.* **123**, 103891.
<https://doi.org/10.1016/j.jtherbio.2024.103891>
- Reiser S, Herrmann JP, Temming A. 2014 Thermal preference of the common brown shrimp (*Crangon crangon*, L.) determined by the acute and gravitational method. *J. Exp. Mar. Bio. Ecol.* **461**, 250–256. <https://doi.org/10.1016/j.jembe.2014.08.018>
- Renner SS, Zohner CM. 2018 Climate change and phenological mismatch in trophic interactions among plants, insects, and vertebrates. *Annu. Rev. Ecol. Evol. Syst.* **49**, 165–182. <https://doi.org/10.1146/annurev-ecolsys-110617-062535>
- Robinson TB, Martin N, Loureiro TG, Matikina P, Robertson MP. 2020 Double trouble: the implications of climate change for biological invasions. *NeoBiota* **62**, 463–487.
<https://doi.org/10.3897/neobiota.62.55729>
- Sánchez-Ramos I, Fernández CE, González-Núñez M. 2019 Comparative analysis of thermal performance models describing the effect of temperature on the preimaginal development of *Drosophila suzukii*. *J. Pest Sci. (2004)*. **92**, 523–541.
<https://doi.org/10.1007/s10340-018-1030-9>

586 Saudreau M, Marquier A, Adam B, Monney P, Sinoquet H. 2009 Experimental study of
587 fruit temperature dynamics within apple tree crowns. *Agric. For. Meteorol.* **149**, 362–
588 372. <https://doi.org/10.1016/j.agrformet.2008.09.001>

589 Saudreau M, Marquier A, Adam B, Sinoquet H. 2011 Modelling fruit-temperature
590 dynamics within apple tree crowns using virtual plants. *Ann. Bot.* **108**, 1111–1120.
591 <https://doi.org/10.1093/aob/mcr054>

592 Saudreau M, Sinoquet H, Santin O, Marquier A, Adam B, Longuenesse JJ, Guilioni L,
593 Chelle M. 2007 A 3D model for simulating the spatial and temporal distribution of
594 temperature within ellipsoidal fruit. *Agric. For. Meteorol.* **147**, 1–15.
595 <https://doi.org/10.1016/j.agrformet.2007.06.006>

596 Sinclair BJ *et al.* 2016 Can we predict ectotherm responses to climate change using thermal
597 performance curves and body temperatures? *Ecol. Lett.* **19**, 1372–1385.
598 <https://doi.org/10.1111/ele.12686>

599 Skendžić S, Zovko M, Živković IP, Lešić V, Lemić D. 2021 Effect of climate change on
600 introduced and native agricultural invasive insect pests in Europe. *Insects* **12**, 1–21.
601 <https://doi.org/10.3390/insects12110985>

602 Strunov AA, Ilinskii YY, Zakharov IK, Kiseleva E V. 2013a Effect of high temperature on
603 survival of *Drosophila melanogaster* infected with pathogenic strain of *Wolbachia*
604 bacteria. *Russ. J. Genet. Appl. Res.* **3**, 435–443.
605 <https://doi.org/10.1134/S2079059713060099>

606 Strunov A, Kiseleva E, Gottlieb Y. 2013b Spatial and temporal distribution of pathogenic
607 *Wolbachia* strain wMelPop in *Drosophila melanogaster* central nervous system under
608 different temperature conditions. *J. Invertebr. Pathol.* **114**, 22–30.
609 <https://doi.org/10.1016/j.jip.2013.05.001>

- Strunov A, Lerch S, Blanckenhorn WU, Miller WJ, Kapun M. 2022 Complex effects of environment and *Wolbachia* infections on the life history of *Drosophila melanogaster* hosts. *J. Evol. Biol.* **35**, 788–802. <https://doi.org/10.1111/jeb.14016>
- Strunov A, Schoenherr C, Kapun M. 2023 *Wolbachia* has subtle effects on thermal preference in highly inbred *Drosophila melanogaster* which vary with life stage and environmental conditions. *Sci. Rep.* **13**, 1–10. <https://doi.org/10.1038/s41598-023-40781-7>
- Strunov A, Schönherr C, Kapun M. 2024 *Wolbachia* effects on thermal preference of natural *Drosophila melanogaster* are influenced by host genetic background, *Wolbachia* type, and bacterial titer. *Environ. Microbiol.* **26**, e16579. <https://doi.org/10.1111/1462-2920.16579>
- Sunday JM, Bates AE, Kearney MR, Colwell RK, Dulvy NK, Longino JT, Huey RB. 2014 Thermal-safety margins and the necessity of thermoregulatory behavior across latitude and elevation. *Proc. Natl. Acad. Sci. U. S. A.* **111**, 5610–5615. <https://doi.org/10.1073/pnas.1316145111>
- Swaegers J, De Cupere S, Gaens N, Lancaster LT, Carbonell JA, Sánchez Guillén RA, Stoks R. 2024 Plasticity and associated epigenetic mechanisms play a role in thermal evolution during range expansion. *Evol. Lett.* **8**, 76–88. <https://doi.org/10.1093/evlett/qrac007>
- Tait G, Cabianca A, Grassi A, Pfab F, Oppedisano T, Puppato S, Mazzoni V, Anfora G, Walton VM. 2020 *Drosophila suzukii* daily dispersal between distinctly different habitats. *Entomol. Gen.* **40**, 25–37. <https://doi.org/10.1127/entomologia/2019/0876>
- Tait G *et al.* 2021 *Drosophila suzukii* (Diptera: Drosophilidae): A decade of research towards a sustainable integrated pest management program. *J. Econ. Entomol.* **114**, 1950–1974. <https://doi.org/10.1093/jee/toab158>

Takeuchi KI *et al.* 2009 Changes in temperature preferences and energy homeostasis in dystroglycan mutants. *Science* **323**, 1740–1743. <https://doi.org/10.1126/science.1165712>

Taylor EN *et al.* 2021 The thermal ecology and physiology of reptiles and amphibians: A user's guide. *J. Exp. Zool. Part A Ecol. Integr. Physiol.* **335**, 13–44. <https://doi.org/10.1002/jez.2396>

Truitt AM, Kapun M, Kaur R, Miller WJ. 2019 *Wolbachia* modifies thermal preference in *Drosophila melanogaster*. *Environ. Microbiol.* **21**, 3259–3268. <https://doi.org/10.1111/1462-2920.14347>

Ulmer R *et al.* 2024 Urban ecology of *Drosophila suzukii*. *Urban Ecosyst.* **27**, 1983–2004. <https://doi.org/10.1007/s11252-024-01554-w>

Usui T *et al.* 2023 The evolution of plasticity at geographic range edges. *Trends Ecol. Evol.* **38**, 831–842. <https://doi.org/10.1016/j.tree.2023.04.004>

van Heerwaarden B, Sgrò CM. 2021 Male fertility thermal limits predict vulnerability to climate warming. *Nat. Commun.* **12**, 2214. <https://doi.org/10.1038/s41467-021-22546-w>

Walsh DB, Bolda MP, Goodhue RE, Dreves AJ, Lee J, Bruck DJ, Walton VM, O'Neal SD, Zalom FG. 2011 *Drosophila suzukii* (Diptera: Drosophilidae): Invasive pest of ripening soft fruit expanding its geographic range and damage potential. *J. Integr. Pest Manag.* **2**, 3–9. <https://doi.org/10.1603/IPM10010>

Ward NL, Masters GJ. 2007 Linking climate change and species invasion: An illustration using insect herbivores. *Glob. Chang. Biol.* **13**, 1605–1615. <https://doi.org/10.1111/j.1365-2486.2007.01399.x>

Winkler A, Jung J, Kleinhenz B, Racca P. 2021 Estimating temperature effects on *Drosophila suzukii* life cycle parameters. *Agric. For. Entomol.* **23**, 361–377. <https://doi.org/10.1111/afe.12438>

659 Woolf AB, Ferguson IB. 2000 Postharvest responses to high fruit temperatures in the field.
660 *Postharvest Biol. Technol.* **21**, 7–20. [https://doi.org/10.1016/S0925-5214\(00\)00161-7](https://doi.org/10.1016/S0925-5214(00)00161-7)
661 Yamamoto A, Ohba S. 1984 Temperature preferences of eleven *Drosophila* species from
662 Japan: the relationship between preferred temperature and some ecological
663 characteristics in their natural habitats. *Zoolog. Sci.* **1**, 631–640.
664

665 **TABLES**

666 **Table 1** Variables considered to induce plasticity in *D. suzukii* thermal preference

Environmental	
Temperature treatment	Constant (16, 20, 24°C) Fluctuating (18-22, 16-24°C)
Life stage	
Age	Larvae (L3) 3-day-old adult (D3) 13-day-old adult (D13)
Sex	Larvae (None) Male (M) Female (F)
Reproductive status	Larvae (L3) Virgin female (Female_V) Fertile female (Female_F) Male (Male)
Life stage (LifeS; integrative variable of Sex, Age and Reproductive status)	Larvae (L3) 3-day-old virgin females (Fem_V_3d) 3-day-old fertile females (Fem_F_3d) 13-day-old virgin females (Fem_V_13d) 13-day-old fertile females (Fem_F_13d) 3-day-old males (Male_3d) 13-day-old males (Male_13d)
Symbionts	
<i>Wolbachia</i> infection	Uninfected (W-) Infected (W+)
Generation	
Generation	G1, G2, G3, G4

667

668

Table 2 Effects of temperature treatment, generation, *Wolbachia* infection status, life stage and their pairwise interactions on thermal preference (Tp) in *Drosophila suzukii*, estimated using linear mixed-effects models with gradient and lane nested within gradient included as random intercepts.

	Chisq	Df	<i>p</i> -value	
Temperature treatment (E_temp)	392.27	4	<2.2e-16	***
Generation (Gen)	25.52	3	1.2e-05	***
<i>Wolbachia</i> (Wol)	78.97	1	<2.2e-16	***
Life stage (LifeS)	1046.12	6	<2.2e-16	***
E_temp × Gen	40.76	12	5.4e-05	***
E_temp × Wol	11.09	4	0.02561	*
E_temp × LifeS	179.25	24	<2.2e-16	***
Gen × Wol	0.49	3	0.9207	
Gen × LifeS	86.07	18	7.3e-11	***
Wol × LifeS	57.01	6	1.8e-10	***

FIGURES

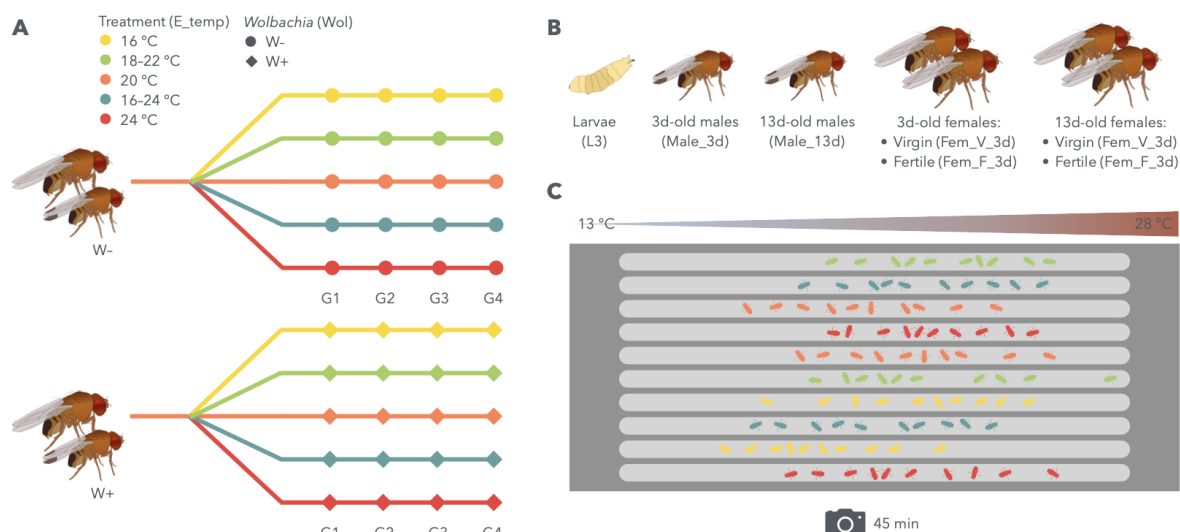


Figure 1 Experimental design. (A) From two source lines of *Drosophila suzukii*, *Wolbachia*-uninfected (W-; circle) and infected (W+; diamond), initially raised at 20 °C, five subpopulations of each were allocated to different temperature regimes (E_temp: 16, 18–22, 20, 16–24 and 24 °C; yellow, green, orange, blue and red, respectively). The populations were maintained for four generations. (B) We measured thermal preference (Tp) among different life stages: larvae (L3), young and old males (Male_3d, Male_13d) and young and old females, virgin and fertile (Fem_V_3d, Fem_V_13d, Fem_F_3d, Fem_F_13d), for both W- and W+ populations. (C) Tp measure consisted in placing 10 individuals with a unique combination of variables within a lane of a thermal gradient made of aluminium and with temperature ranging from 13 to 28 °C. A total of 50 individuals was measured for each variable combination. Photographs were taken every 5 min for 60 min and the position at 45 min was recorded to assess Tp (see Deconninck et al. (2025) for review on Tp measurement in *Drosophila*).

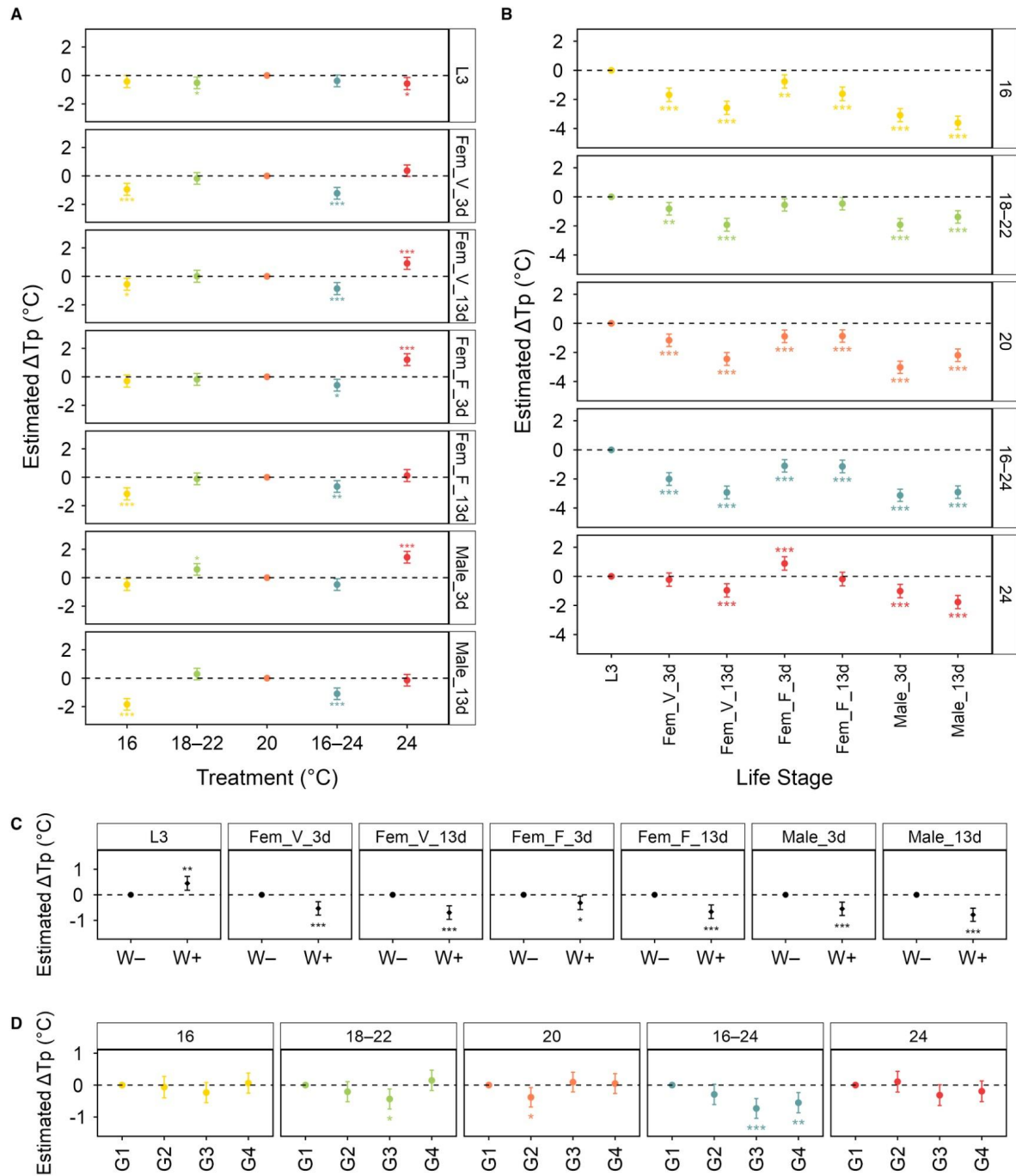
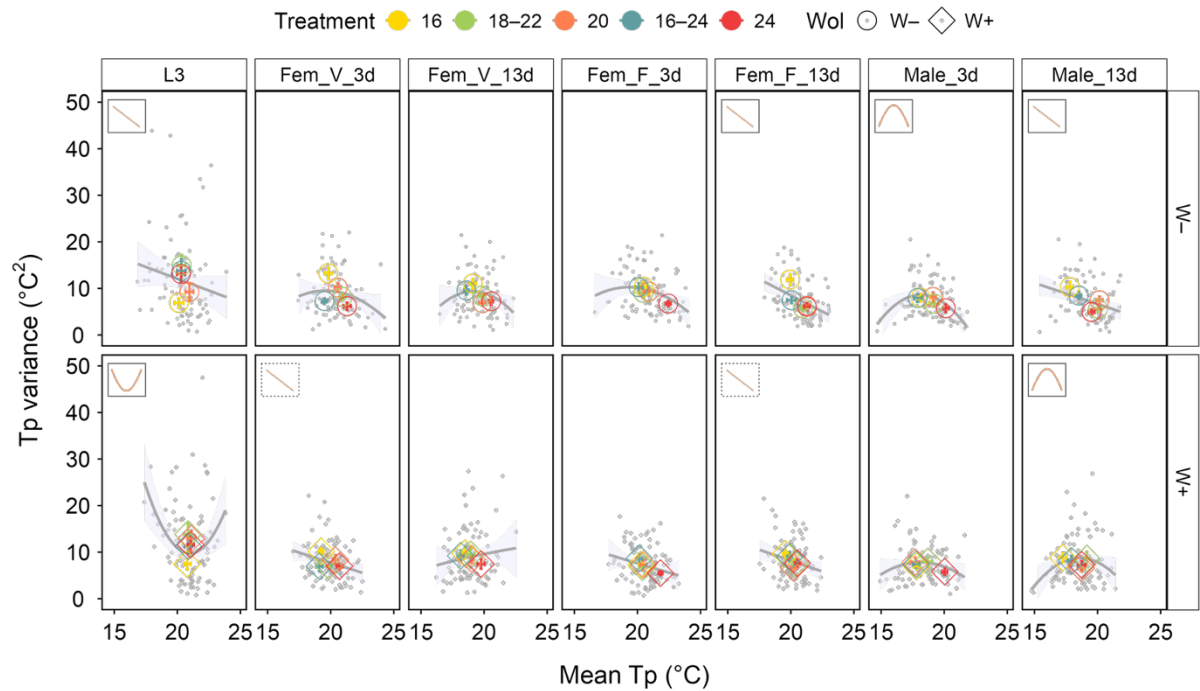


Figure 2 Context-dependent effects of temperature treatment, life stage, generation and *Wolbachia* infection on mean thermal preference (Tp) in *Drosophila sukuzii*. Estimated contrasts were obtained from linear mixed-effects models using estimated marginal means (EMMs). Panels show the estimated effects of (A) temperature treatment within life stages, (B) life stage within temperature treatments, (C) *Wolbachia* infection within life stages, and (D) generation within temperature treatments. Contrasts are expressed relative to the

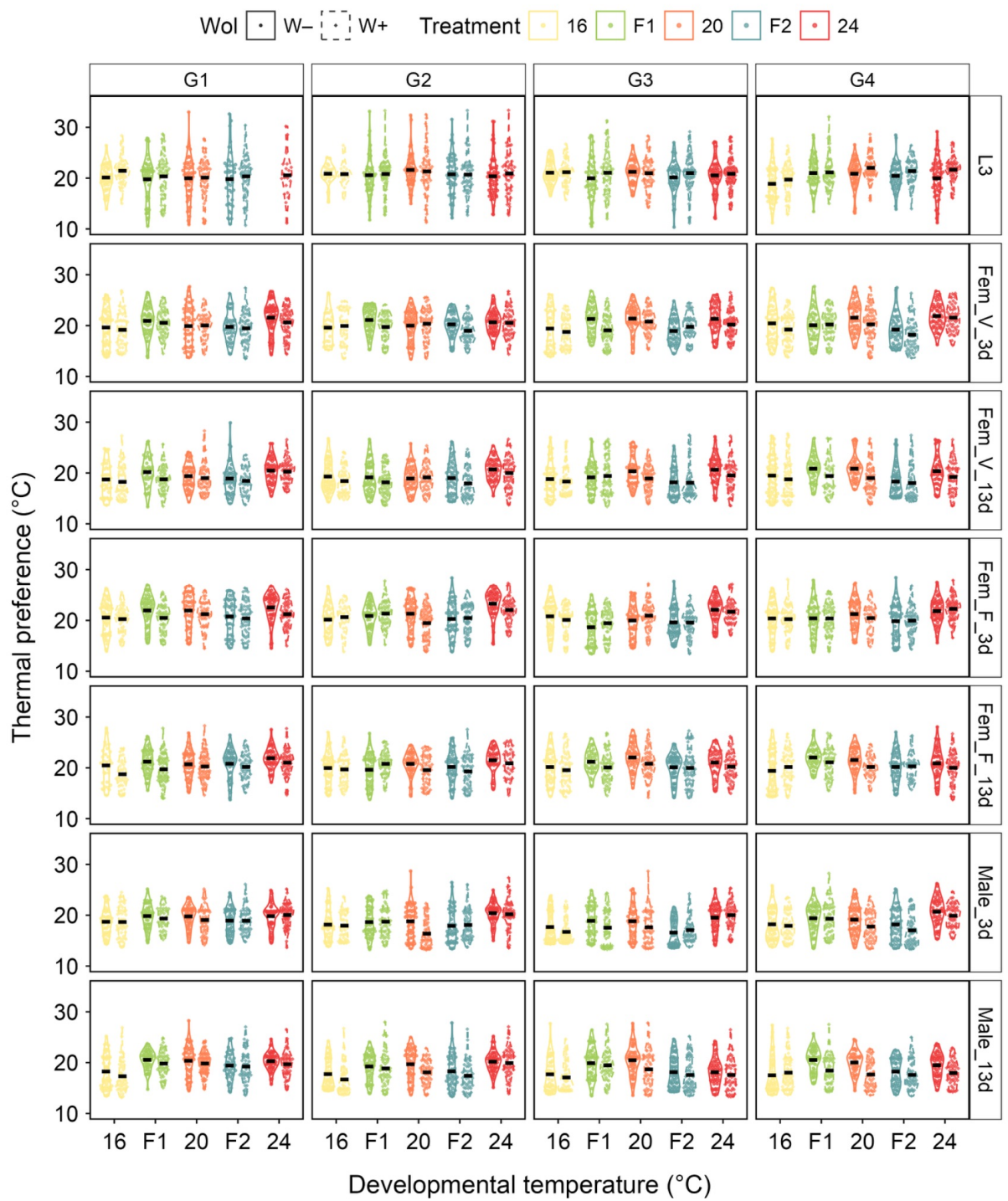
8 appropriate reference level for each panel (20 °C for temperature, L3 for life stage, W– for
9 *Wolbachia* infection status, and G1 for generation). Points indicate estimated contrasts (ΔT_p ,
10 °C), error bars indicate 95% confidence intervals, and asterisks denote significant differences
11 from the corresponding reference level (*: $p < 0.05$; **: $p < 0.01$; ***: $p < 0.001$).



13

14 **Figure 3** Thermal preference (Tp) mean and variance patterns, with plastic responses of Tp
15 across life stages and *Wolbachia* infection status in *Drosophila sukukii*. Relationship between
16 Tp mean and Tp variance of Tp across temperature treatments (constant regimes: 16, 20 and
17 24°C, in yellow, orange and red; fluctuating regimes: 18–22 and 16–24°C, in green and blue),
18 life stages (L3: third larval stage; Fem_V_3d, Fem_V_13d, Fem_F_3d, Fem_F_13d: 3d and
19 13d-old virgin and fertile females; Male_3d, Male_13d: 3d and 13d-old males) and
20 *Wolbachia* infection status (W–: uninfected, circle; W+: infected, diamond). Grey points
21 represent averaged lane-level observations (n = 5 lanes for each temperature treatment and
22 generation). Coloured symbols indicate treatment-level mean values ($\pm$ SE) of Tp mean and
23 Tp variance. Analyses were conducted using pooled data from generations G1 to G4. Fitted
24 linear, quadratic or loess (span 10) regressions are shown as solid lines with 95% confidence
25 bands. Symbols indicate inferred behavioural thermoregulation strategies following
26 Deconninck et al. (2025) when the relationship between Tp variance and Tp mean was

27 statistically significant (or marginally significant, shown as dashed symbols; see Table S1 for
28 details).



30

31 **Figure S1** Thermal preference (Tp) across temperature treatments, generations, life stages
32 and *Wolbachia* infection status in *Drosophila sukuzii*. Points represent mean Tp ($\pm$ SE) for
33 each combination of variables. Colours indicate temperature treatment (constant: 16, 20 or
34 24°C, in yellow, orange and red; fluctuating regimes: F1 = 18-22 and F2 = 16-24°C, in green

and blue) and symbols indicate infection status (W–: uninfected, circle; W+: *Wolbachia*-infected, diamond). Panels are arranged by life stage (rows) and generation (columns). Tp measurements at G1, L3, W+, 24 °C could not be obtained due to technical constraints.

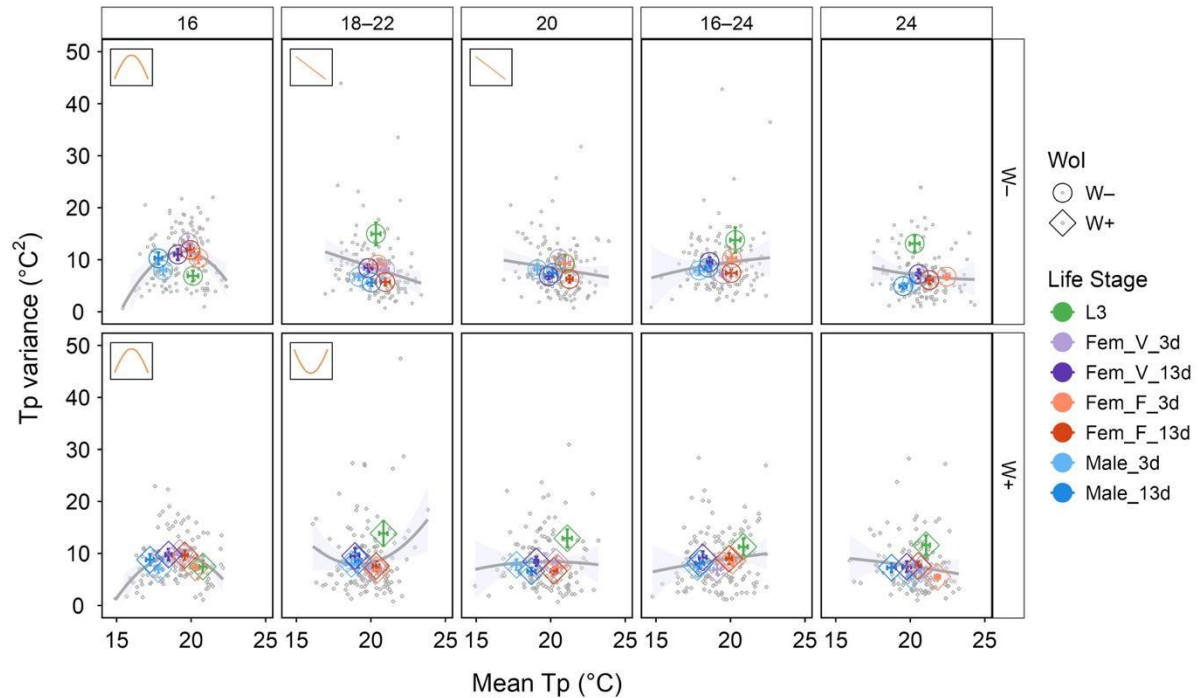


Figure S2 Thermal preference (Tp) mean and variance patterns, with plastic responses of Tp across temperature treatments and *Wolbachia* infection status in *Drosophila suzukii*. Relationship between Tp mean and Tp variance of Tp across temperature treatments (constant regimes: 16, 20 and 24°C, in yellow, orange and red; fluctuating regimes: 18–22 and 16–24°C, in green and blue), life stages (L3: third larval stage; Fem_V_3d, Fem_V_13d, Fem_F_3d, Fem_F_13d: 3d and 13d-old virgin and fertile females; Male_3d, Male_13d: 3d and 13d-old males) and *Wolbachia* infection status (W–: uninfected, circle; W+: infected, diamond). Grey points represent averaged lane-level observations (n = 5 lanes for each temperature treatment and generation). Coloured symbols indicate treatment-level mean values ($\pm$ SE) of Tp mean and Tp variance. Analyses were conducted using pooled data from generations G1 to G4. Fitted linear, quadratic or loess (span 10) regressions are shown as

solid lines with 95% confidence bands. Symbols indicate inferred behavioural thermoregulation strategies following Deconninck et al. (2025) when the relationship between Tp variance and Tp mean was statistically significant (see Table S2 for details). Interpretation: Life stage is used to induce plasticity. For a given temperature treatment, the mean and variance of Tp co-vary during ontogeny. The inverted U-shape relationship between mean and variance of Tp indicates that the life stage with the highest mean Tp is also the one with highest precision (e.g., W+ larvae at 16 °C).

Table S1 Group-specific linear (β_1) and quadratic (β_2) effects of mean Tp on Tp variance for each life stage, with inferred relationship shape (when $p < 0.05$).

Wol	LifeS	β_1	β_1 <i>p</i> -value	β_2	β_2 <i>p</i> -value	Inferred shape
W–	L3	-75.84	0.0019	32.12	0.1953	Linear decreasing
	Fem_V_3d	-1.35	0.9679	-34.01	0.1803	ns
	Fem_V_13d	-25.75	0.2888	-58.42	0.1219	ns
	Fem_F_3d	-7.48	0.8057	-32.02	0.1340	ns
	Fem_F_13d	-101.63	0.0244	20.04	0.6112	Linear decreasing
	Male_3d	-89.76	0.0012	-68.20	0.0043	Inverted U ($\cap$)
	Male_13d	-65.26	0.0105	-21.65	0.4390	Linear decreasing
W+	L3	-194.52	0.0000	152.74	0.0000	U-shape
	Fem_V_3d	-46.73	0.0739	-20.75	0.5870	Marginal - Linear decreasing
	Fem_V_13d	33.14	0.2408	-3.06	0.9232	ns
	Fem_F_3d	-47.72	0.2277	-0.25	0.9946	ns
	Fem_F_13d	-63.11	0.0748	18.27	0.7011	Marginal - Linear decreasing
	Male_3d	-50.00	0.1178	-38.40	0.0801	ns
	Male_13d	-55.92	0.1373	-60.48	0.0221	Inverted U ($\cap$)

Table S2 Group-specific linear (β_1) and quadratic (β_2) effects of mean Tp on Tp variance for each temperature treatment, with inferred relationship shape (when $p < 0.05$).

Wol	E_temp	β_1	β_1 <i>p</i>-value	β_2	β_2 <i>p</i>-value	Inferred shape
W−	16	-12.59	0.5359	-85.74	0.0003	Inverted U ($\cap$)
	18–22	-58.46	0.0075	6.13	0.8018	Linear decreasing
	20	-55.50	0.0258	38.84	0.0895	Linear decreasing
	16–24	23.58	0.2019	-4.16	0.8170	ns
	24	-25.40	0.3971	5.13	0.8018	ns
W+	16	-32.20	0.1387	-65.77	0.0013	Inverted U ($\cap$)
	18–22	23.15	0.1874	51.22	0.0073	U-shape
	20	3.83	0.8132	-13.72	0.3471	ns
	16–24	23.93	0.1785	-2.02	0.9132	ns
	24	-24.26	0.2089	-2.32	0.9044	ns

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