

1 **Island species as models for small population biology**
2 **and conservation**

3 Maëva Gabrielli¹, Hernán E. Morales², Víctor Noguerales^{3,4}, Wanyi Wei⁵, Darko D. Cotoras^{6,7},
4 Nicolas Dussex⁸, Ricardo J. Pereira^{9,10}, Brent C. Emerson⁴, José Cerca^{5,11,12}

5

6 1 Centre de Recherche sur la Biodiversité et l'Environnement (CRBE), Université de Toulouse,
7 Toulouse, France

8 2 Globe Institute, University of Copenhagen, Copenhagen, Denmark

9 3 Departamento de Biología Animal, Edafología y Geología, Universidad de La Laguna, San
10 Cristóbal de La Laguna, Spain

11 4 Instituto de Productos Naturales y Agrobiología (IPNA-CSIC), San Cristóbal de La Laguna,
12 Spain

13 5 Centre for Ecological and Evolutionary Synthesis (CEES), Department of Biosciences,
14 University of Oslo, Oslo, Norway

15 6 Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Av. Libertador
16 Bernardo O'Higgins 340, Santiago 8331150, Chile

17 7 Department of Entomology, California Academy of Sciences, 55 Music Concourse Dr.,
18 Golden Gate Park, San Francisco, CA 94118, USA

19 8 Department of Population Analysis and Monitoring, Swedish Museum of Natural History,
20 Stockholm, Sweden

21 9 Department of Biodiversity Monitoring, State Museum of Natural History Stuttgart, 70191
22 Stuttgart, Germany

23 10 KomBioTa – Center for Biodiversity and Integrative Taxonomy at the University of
24 Hohenheim, Stuttgart, Germany

25 11 Department of Bioinformatics and Genetics, Swedish Museum of Natural History,
26 Stockholm, Sweden

27 12 SciLifeLab, Karolinska Institutet Science Park, Solna, Sweden

28

29

30

31

32 Corresponding author: Maëva Gabrielli (maeva.gab@hotmail.fr)

33 Abstract

34 Islands provide unparalleled natural laboratories for understanding how small, isolated
35 populations persist and evolve. Our synthesis of island population studies reveals that 50–70%
36 report effective population sizes below 100, yet many taxa have sustained such small
37 populations for millions of years. Strikingly, only 4% and 27% of studies examined genetic
38 load and genomic diversity, exposing major blind spots in our understanding of genomic
39 erosion. We argue that island systems offer critical insights into how limited populations retain
40 viability and adaptive potential. Future work should expand taxonomic and geographic
41 coverage, exploit temporal genomic data, and integrate genomic, demographic, and fitness
42 information to link evolutionary resilience on islands to the conservation of increasingly
43 fragmented populations worldwide.

44

45

46 Highlights

- 47 • Island populations offer natural experiments for understanding how long-term small
48 population sizes shape evolutionary trajectories, yet their genomic consequences
49 remain underexplored.
- 50 • Despite extensive ecological and biogeographic research on islands, studies rarely
51 quantify genetic load or temporal changes in genomic diversity, limiting insight into
52 adaptive potential.
- 53 • Many island taxa have persisted for millions of years with extremely small effective
54 population sizes, challenging assumptions about the inevitability of genomic decline.
- 55 • Integrating genomic data with life-history traits and demographic histories is essential
56 for predicting vulnerability and guiding conservation strategies for fragmented and
57 declining populations.

58 Population sizes on islands

59 The colonization of an island or archipelago typically involves a founder effect, where a small
60 number of individuals establish a new population [1–3]. The often-limited area of islands
61 constrains population sizes, and can increase the likelihood of **inbreeding** [4]. The taxon cycle,
62 in which lineages become increasingly specialized to narrower ecological niches [5], further
63 reduces available suitable habitat. Catastrophic events, such as eruptive volcanic events and
64 massive flank collapses, can further exacerbate these genetic pressures [6]. Collectively, these
65 factors result in the characteristically small population sizes observed in island ecosystems [7].

66
67 Small and bottlenecked populations experience reduced efficacy of selection and
68 increased effects of genetic drift and inbreeding, which together promote the fixation of
69 deleterious mutations as purifying selection becomes less efficient [8,9]. Over time, these
70 processes lead to **genomic erosion**, or the progressive loss of genome-wide diversity driven
71 primarily by **genetic drift** and inbreeding [10–12]. Genomic erosion generally manifests as
72 elevated **realised genetic load** (*i.e.*, the component of **genetic load** whose fitness effects are
73 expressed) and maladaptation due to mismatches between genetic variation and the prevailing
74 environmental conditions [11]. As a result, declining genetic diversity diminishes individual
75 fitness and adaptive potential, and reduced population size increases susceptibility to
76 **inbreeding depression** [8,13]. Ultimately, these combined effects heighten extinction risk and
77 may drive populations into mutational meltdown and extinction vortices [14,15]. Consistent
78 with these theoretical predictions, island populations - typically small and isolated - often
79 exhibit signatures of weaker purifying selection [7,14,16], reduced adult fitness [17–19] and
80 morphological abnormalities [20].

81
82 Although it has been suggested that island lineages face gradual extinction due to their
83 small population sizes, and the greater ecological turnover dynamics and environmental
84 catastrophes inherent to islands [21], many insular species demonstrate remarkable persistence,
85 surviving for millions of years despite long-term reduced population sizes (N_e) and significant
86 inbreeding [2,4,22,23], while exhibiting substantial genomic variation [24,25]. It may be that
87 smaller and more isolated populations typically face less intense human-driven exploitation
88 compared to larger, less isolated ones [26–28]. However, owing to the taxon-, environment-
89 and population-specific nature of inbreeding depression [29], we argue that these resilient

90 insular populations may offer crucial conservation insights into the evolution and persistence
91 of small populations.

92

93 While island systems are well studied in terms of their biogeography, evolutionary
94 history, and ecology, far less is known about their genetic diversity and genomic erosion—the
95 loss of genetic diversity, maladaptation, and accumulation of genetic load. This gap is
96 surprising given that genetic diversity determines adaptive potential and long-term population
97 viability. Here, we explore two interlinked aspects of insular biology: (i) how bottlenecks and
98 long-term small population sizes shape the evolutionary history of island lineages, and (ii) how
99 these demographic histories drive genomic erosion and influence persistence in small
100 populations. We use these dimensions to highlight insular taxa as potential models in informing
101 contemporary conservation strategies. We argue that long-term isolated lineages - sometimes
102 shaped by millions of years of evolution in geographically isolated territories - provide essential
103 insights for safeguarding biodiversity against intensifying anthropogenic threats both in insular
104 and continental contexts.

105

106 Fundamental aspects of small population sizes on islands

107 In many island vertebrates and plants (Box 1) population sizes tend to be smaller than those of
108 their mainland relatives, often as a consequence of founder events and subsequent demographic
109 fluctuations. [1,17–19]. However, this pattern cannot be mistaken as universal, as population
110 size and structure on islands depend strongly on species-specific dispersal abilities, life
111 histories, and spatial genetic structuring within and among islands and continental populations.
112 In any case, the long-term persistence of insular populations offers an opportunity to examine
113 how demographic conditions influence the **purging** of deleterious alleles. Simulations of
114 genomic data suggest that the Channel Island foxes have persisted for millennia through the
115 efficient purging of highly deleterious alleles [23]. Svalbard reindeer maintain adaptive
116 potential despite long-term genetic depletion, successfully surviving in harsh Arctic conditions
117 [30]. These examples align with theoretical predictions that purging can occur in populations
118 with effective sizes (N_e) as small as ≈ 70 [31] and with empirical evidence that suggests founder
119 effects do not necessarily constrain purifying selection [32]. Collectively, these findings
120 underscore the remarkable capacity of small insular populations to persist despite severe
121 demographic constraints (Box 2).

122 The severity of inbreeding depression varies substantially among taxa and populations,
123 according to their traits and environmental contexts [29]. Purging of deleterious alleles in
124 lineages on islands may potentially occur through two key ecological mechanisms. First,
125 reduced predator and parasite pressures on islands might lead to more constant demographic
126 dynamics, or a probability of increasing population size, and thus an increased capacity for
127 purging [23]. Indirect support for this comes from increased genetic erosion due to drift
128 following rapid bottlenecks after predator introductions [33]. Second, the dynamics of
129 population demography and connectivity of islands can enable **genetic rescue** via secondary
130 contact between isolated populations. Specifically, oceanic islands often exhibit fragmented
131 habitats resulting from climatic and geological processes, which create fragmented populations
132 and the admixture of these increase genetic diversity and might enable long-term population
133 persistence [34]. In Hawaiian kīpukas or geologically stable areas in the Canary Islands,
134 limited-dispersal organisms accumulate genetic differences across habitat patches isolated by
135 lava flows and massive flank collapses, and subsequent secondary contact could promote
136 heterosis and increased heterozygosity [35,36], potentially masking fixed deleterious mutations
137 [29,37]. In the Chatham Island black robin, historical isolation followed by secondary contact
138 and purifying selection likely purged deleterious alleles, enhancing the species' resilience [38].
139 Similarly, *Hydrobates* petrels maintain high genetic diversity through inter-colony dispersal
140 [39]. These findings suggest that managed gene flow from nearby populations could be a vital
141 tool for conserving genetically depauperate populations [21], although such interventions must
142 be carefully designed to avoid the risk of outbreeding depression.

143

144 Other than demographic and ecological factors promoted by the geographic features of
145 the archipelago, species-specific morphological and behavioural traits can play a crucial role
146 in modulating the purging of deleterious alleles. The Hawaiian hihi maintains population
147 viability and genetic diversity through extra-pair mating behaviors [40], while polyandry and
148 non-random fertilization similarly preserve genetic variation in insular Swedish adders [41].
149 Hawaiian petrels challenge conventional expectations of conservation biology by maintaining
150 stable and high genetic diversity despite small census population sizes - a phenomenon likely
151 facilitated by their extended generation times and strong dispersal ability allowing inter-colony
152 movement of individuals [42]. These cases reveal an important decoupling between census
153 population size and genetic diversity in long-lived, mobile species, offering critical insights for
154 conservation strategies targeting small populations.

155

156 Conservation, genetic load and rewilding

157 Comparatively small effective population sizes (N_e) of island endemics and native species, in
158 comparison to suitable continental species, may amplify the effect of genetic drift, increasing
159 the risk of deleterious mutation accumulation in island species [7]. This elevated genetic load
160 may erode individual fitness and threaten long-term population viability, raising concerns for
161 *in situ* and captive breeding programs [43,44]. The advent of whole-genome sequencing has
162 positioned genetic load as a critical focus in conservation biology, with whole genome data
163 now enabling a comprehensive assessment of deleterious mutations across all genomic regions
164 [45]. Nevertheless, the lack of standardized methodologies hinders comparative analyses and
165 generalization. Following the framework proposed by [46], we distinguish between **masked**
166 **load** (deleterious variants in heterozygous state) and realized load (homozygous deleterious
167 variants), assuming that most deleterious mutations are recessive. Computational simulations
168 have become essential tools for quantifying the impacts of such mutations in natural
169 populations [47]. Two major challenges remain for using genetic load in conservation studies.
170 First, accurately inferring the fitness effects of individual variants is difficult, complicating
171 efforts to assess the overall fitness consequences of elevated genetic load [48,49]. Second, more
172 empirical data are needed to quantify the relative importance of masked versus realized genetic
173 load in influencing extinction risk across different timescales [46,49]. Island species, owing to
174 their unique demographic histories, typically small effective population sizes, and limited gene
175 flow, are likely to be especially informative for studying the interplay between purging and the
176 accumulation of deleterious mutations. Well-studied systems such as the Galápagos Islands,
177 Canary Islands, Madagascar, Mauritius, New Zealand, and Hawaii offer valuable opportunities
178 to generate such empirical data, as they have long served as focal points for evolutionary,
179 ecological, and conservation genetics research.

180

181 There is growing interest in characterizing deleterious genetic variation and genetic
182 load for the conservation of small and endangered populations. Recent reviews have explored
183 how both selective purging and genetic drift shape the distribution of deleterious mutations
184 during population decline and recovery (*e.g.*, [48]). While strong bottlenecks can facilitate the
185 purging of highly deleterious alleles, weakly or mildly deleterious mutations are more likely to
186 increase in frequency, potentially reducing the overall fitness of the population [50,51]. The
187 balance between drift and selection during demographic decline and recovery thus plays a
188 critical role in shaping genetic load, with important implications for conservation management

189 (see Box 2). In small populations, genetic drift can cause previously masked deleterious alleles
190 to become homozygous and expressed, converting masked load into realized load [52].
191 Evidence from insular taxa shows elevated realized load due to enhanced drift effects, and
192 illustrates both the accumulation and purging of deleterious variation. For example, passerines
193 on islands tend to exhibit higher genetic load than their mainland counterparts [7,16], while
194 island populations of peregrine falcons show extensive runs of homozygosity and elevated drift
195 load compared to mainland populations [53]. An efficient purging of strongly deleterious
196 alleles following severe bottlenecks seems to occur on islands. For example, the recent
197 population collapse of the Reunion harrier due to anthropogenic pressures exemplifies the
198 complex interplay between genetic drift and purging: while simulations suggest that highly
199 deleterious alleles may have been purged as a result of recent bottlenecks, mildly deleterious
200 mutations persist, likely due to the reduced efficacy of selection [54]. This evidence resembles
201 that of several emblematic island species, such as the San Nicolas Island fox or the Svalbard
202 reindeer, which all show long-term persistence despite high homozygosity and apparent
203 fixation of deleterious variants, suggesting that bottlenecks may lead to purging of deleterious
204 variants and mitigating some of the negative consequences of genomic erosion [23,30,55].
205 Similarly, distinct demographic histories, including bottleneck and introgression histories, can
206 affect genetic load, as shown in the Anatolian and the Cyprian mouflons [56]. Indeed, the
207 Cyprian mouflon exhibits both lower genetic diversity and genetic load than the Anatolian
208 mouflon, likely because of founder effects, island isolation, introgression from domestic
209 lineages, or differences in their bottleneck dynamics. The San Nicolas Island fox presents a
210 particularly striking example: despite near-complete genomic monomorphism and high
211 homozygosity for putatively deleterious alleles, the population has persisted for decades
212 following a severe 20th-century bottleneck—challenging assumptions that genetic
213 depauperation inevitably leads to extinction [57]. Although few island populations have been
214 the focus of detailed load analyses, existing studies underscore the value of island systems for
215 disentangling the dynamics of deleterious variation, purging, and persistence in small
216 populations (see Box 1). More studies are needed so we are able to draw general conclusions
217 across species with diverse genomic architectures, ecological contexts, and evolutionary
218 histories.

219

220 Insular taxa can contribute to valuable insights into the poorly understood relationship
221 between genetic load and fitness. A long-term study of house sparrow metapopulations across
222 eight islands measured fitness as survival probability and lifetime reproductive success, finding

223 that inbreeding consistently reduced individual fitness when compared to mainland
224 populations. Yet, the magnitude of inbreeding depression was similar across populations
225 experiencing different levels of inbreeding [19]. In a small island population of *Glanville*
226 *fritillary* butterflies, extensive fixation of deleterious alleles contributed to reduced trait
227 performance and elevated genetic load, though inbreeding depression was not observed in
228 recent generations [58]. Recent simulation studies, applicable to both island and non-island
229 systems, highlight how the shape of the distribution of fitness effects (DFE), particularly the
230 prevalence of weakly versus strongly deleterious mutations, can influence both the
231 accumulation of load and its detectability in fitness-related traits. This may help explain why
232 high load does not always translate into observable inbreeding depression [47]. These findings
233 underscore the complex challenges of linking genetic load to immediate fitness consequences.
234 Evidence from islands shows that genomic erosion can constrain future adaptive potential. The
235 Seychelles magpie robin, which underwent historical bottlenecks followed by recent assisted
236 translocations, harbors a high realized genetic load and showcases one of the lowest genetic
237 diversity recorded in endangered birds [59]. Simulations suggest that long-term recovery may
238 require repeated translocations involving large numbers of individuals to mitigate load and
239 restore adaptive potential. The Chatham Island black robin, once reduced to a single breeding
240 pair in the 1980s, has rebounded demographically, with approximately 300 individuals now
241 present in the wild. Genomic comparisons between two extant populations and historical
242 samples predating the bottleneck suggest that some purging of deleterious alleles occurred over
243 time, potentially aided by long-term isolation across multiple islands [38]. However, evidence
244 of persistent inbreeding depression in this species underscores that even when strongly
245 deleterious alleles are purged, mildly deleterious mutations can accumulate and impair fitness
246 [18,60].

247

248 Insights from island systems can also inform genetic rescue and rewilding, particularly
249 in the context of species introductions or translocations. The extinction of the dodo on
250 Mauritius, which disrupted seed dispersal for several large-fruited tree species, some of which
251 subsequently declined, is a textbook example of how the loss of key island species can cascade
252 through ecosystems, which calls upon the importance of both rewilding and genetic rescue.
253 Islands have become key testing grounds for rewilding due to their historical loss of ecological
254 actors [61] and fewer sociopolitical barriers to reintroducing megafaunal taxa [62]. For
255 instance, giant tortoise rewilding on Indian Ocean islands demonstrates the potential to restore
256 lost ecological functions, such as seed dispersal and nutrient cycling [62]. However, evidence

257 from islands clearly shows that the genomic legacy of both reintroduced and recipient
258 populations receiving translocated individuals must be carefully considered. Reintroductions
259 from large outbred populations may unintentionally introduce recessive deleterious alleles, as
260 demonstrated in the Isle Royale wolf. In this species, an initial boost in population viability
261 followed the arrival of a single immigrant, but subsequent inbreeding among his descendants
262 led to a population collapse due to the expression of hidden genetic load [63]. While genetic
263 rescue can be a powerful tool, it also carries risks of outbreeding depression, particularly when
264 mixing highly divergent lineages or disrupting local adaptation [55]. For instance, a study on
265 the isolated population of Apennine brown bears endemic to Central Italy showed that
266 introducing individuals from Slovakia would lead to a loss of local ancestry and an increase in
267 genetic load within the Apennine population [64]. Effective rewilding or genetic rescue efforts
268 may therefore require genomic screening to identify individuals with lower realized load, fewer
269 long runs of homozygosity, and higher compatibility with recipient populations.
270

271 Conclusions

272 Earth is undergoing unprecedented environmental transformations fueled by unsustainable
273 economic paradigms that disregard planetary boundaries. This crisis is precipitating
274 biodiversity loss through habitat destruction, decreased size of populations and ultimately
275 species extinction. Within this context, island systems arise as natural and ideal models for
276 studying small population biology. Although small, isolated, insular populations are often
277 assumed to accumulate elevated genetic load, empirical evidence remains limited (only 4% of
278 our literature survey dealt with genetic load). Nonetheless, their long-term persistence,
279 particularly under benign ecological conditions (*e.g.*, reduced predation and competition),
280 highlights these systems as valuable for testing hypotheses relevant to conservation strategies.
281 A critical consideration is the temporal disparity between natural population recovery and
282 human-managed restoration efforts (see Outstanding questions). Natural populations on
283 continents likely experience a genetically sensitive post-colonization phase during bottleneck
284 recovery, a process rarely observed in contemporary island studies since most extant island
285 populations represent long-established systems. This temporal perspective highlights an
286 important gap in our understanding of early-stage recovery dynamics, which may have
287 significant implications for conservation strategies targeting recently founded or reintroduced
288 populations.

289 Future work should define thresholds at which genetic load compromises viability
290 while identifying buffering ecological contexts, translate island-derived insights into broader
291 conservation strategies for long-term viability of populations, and integrate ecological, genetic,
292 and morphological data to assess fitness consequences across population and individual levels.
293 Expanding studies beyond the current bias toward birds and mammals (Box 1) is essential.
294 Combining demographic and genomic approaches - particularly by linking effective population
295 size (N_e) estimates with direct measures of deleterious variation - will refine population health
296 assessments. Integrating genomic simulations with empirical fitness and genetic load
297 measurements can bridge island conservation genetics with general conservation theory.
298 Additionally, evaluating how human-mediated dispersal influences genetic load and adaptive
299 potential in both native and introduced island species remains crucial. Addressing these
300 priorities will elucidate how isolation, small population size, and demographic history
301 collectively shape the genomic integrity of island biotas.

302

303

304 Outstanding Questions

305

306 **1. How can we disentangle the relative contributions of demographic history, colonization** 307 **age, and ecological context to genetic load in island populations?**

308 Understanding how these factors interact is crucial for identifying when purging or drift
309 dominates and for predicting which populations are most resilient to genomic erosion.

310

311 **2. What comparative sampling designs best capture the temporal dynamics of load** 312 **accumulation and purging across islands and archipelagos?**

313 Within-archipelago comparisons—among endemic, native, and introduced taxa—offer a
314 promising avenue, but standardized frameworks are needed to separate temporal from
315 ecological effects.

316

317 **3. To what extent can genomic and demographic data from museum and ancient DNA** 318 **collections reveal historical changes in load and purging efficiency?**

319 Temporal genomic data remain underused but could uniquely test hypotheses about how small
320 populations recover or decline through time.

321

322 **4. How can forward-time simulations and variant effect prediction pipelines be integrated**
323 **to provide robust, comparable estimates of both masked and realized genetic load?**

324 Coupling empirical and simulated data will improve inference by linking specific mutational
325 burdens to demographic processes and fitness outcomes.

326

327 **5. Which measurable fitness components, such as survival, fecundity, or phenotypic**
328 **performance, most accurately reflect realized genetic load in wild island populations?**

329 Identifying reliable proxies for fitness will enable more accurate assessments of how genomic
330 erosion translates into ecological performance.

331

332 **6. How do long-term ecological stability and life-history traits modulate the relationship**
333 **between effective population size, purging, and extinction risk?**

334 Comparative analyses across species differing in longevity, reproductive rate, and habitat
335 specialization could clarify the conditions that buffer small populations from mutational
336 meltdown.

337

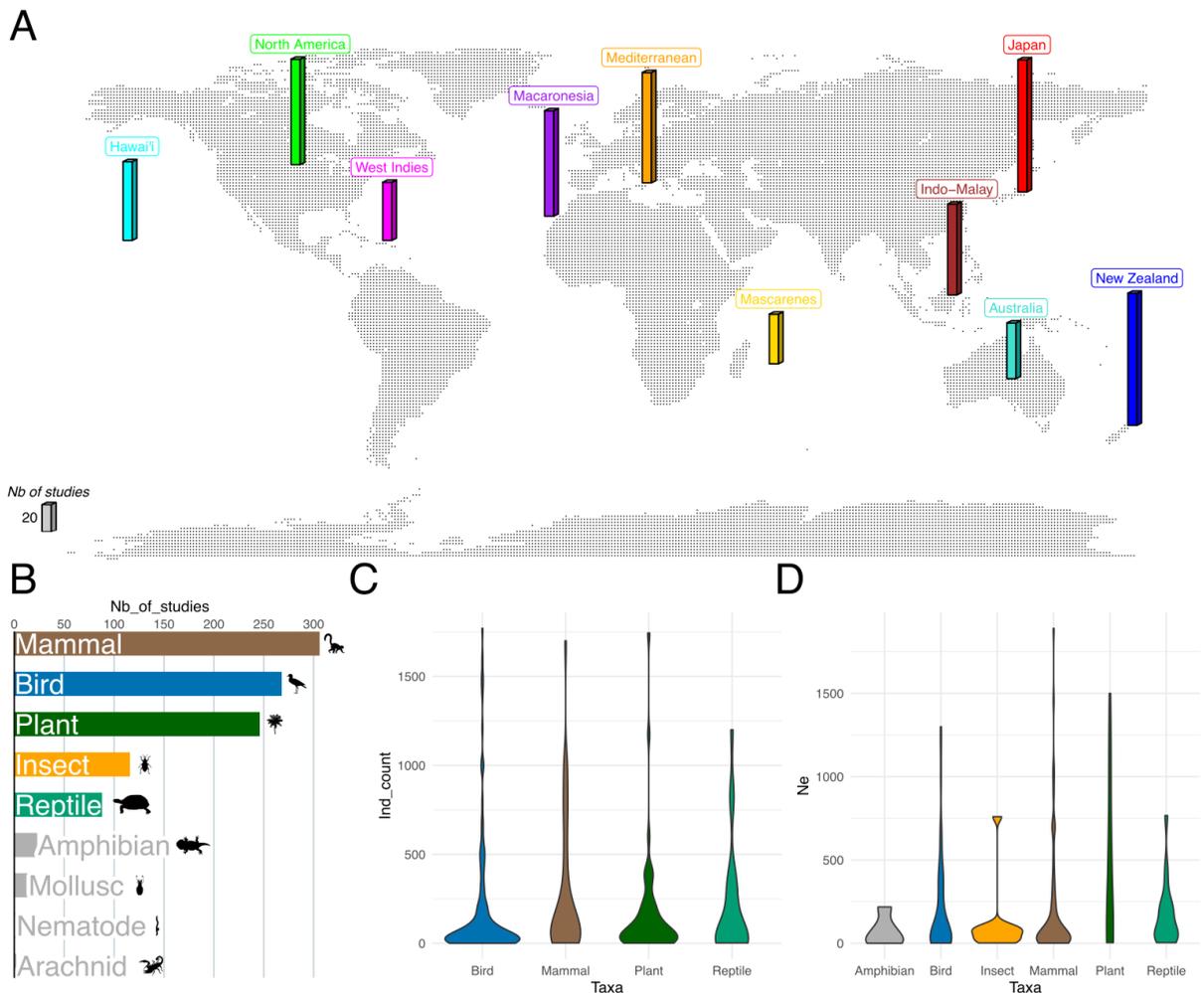
338 **7. What coordinated research frameworks can unify genomic, ecological, and**
339 **demographic perspectives to generalize patterns of load evolution in insular systems?**

340 Cross-disciplinary collaborations and shared data standards are essential to move from case
341 studies toward predictive models of genetic load in isolated populations.

342 **Box 1 - Genomic insights from islands: a global synthesis of**
343 **diversity, bottlenecks, and genetic load**

344 We conducted three literature searches using Web of Science on March 19, 2024, with the
345 following keywords as topics: (i) “genetic load” AND “island” (462 entries); (ii) “bottleneck”
346 AND “island” (1,379 entries); (iii) and “genomic diversity” AND “island” (1,934 entries)
347 (Table S1). We discarded irrelevant publications, for instance, those that focused on ‘genomic
348 islands’ of differentiation across genomes, ending up with 1,063 publications that we carefully
349 reviewed. This included 41, 733 and 289 publications for the 3 different literature searches,
350 respectively. For each of these 1,063 publications, we recorded the geographic focus (*i.e.*,
351 single islands or archipelagos), taxonomic group (*e.g.*, Amphibia, Mammalia, Plantae, Insecta),
352 the minimum reported effective population size (N_e), and the lowest reported individual counts
353 estimated by the authors (if available). For N_e estimations, we only considered cases where N_e
354 was explicitly found in the text, thus discarding the visual inference from plots reflecting N_e
355 changes through time (*e.g.*, PSMC plots). We also noted the biogeographic origin of the
356 lineages (*i.e.*, human-associated dispersal such as invasive species versus native or endemic
357 populations), and whether the study included temporal sampling (*e.g.*, ancient DNA or
358 sampling across multiple years).

359 The number of relevant publications (1,063 in total) underscores the scientific interest in
360 understanding the genomic consequences of insularity, particularly regarding genetic diversity
361 and the effects of bottlenecks. Nevertheless, only a small proportion of insular genomic studies
362 focused on genetic load (4%), suggesting that very little is known about this topic of growing
363 interest in conservation biology.



364

365 **Figure 1 (box 1)** Estimated effective population size (N_e) for different taxa across three
 366 literature searches. We conducted three separate searches in Web of Science using the
 367 following topic combinations: “genetic load” AND “island” “bottleneck” AND “island” and
 368 “genomic diversity” AND “island”. For each search, we extracted the reported N_e values for
 369 each taxon. To highlight the typically low N_e values observed in island populations, we
 370 excluded entries with $N_e > 2,000$.

371

372 The seven most frequently studied regions were: Japan and associated islands/archipelagos
 373 (109 papers), New Zealand (109), Mediterranean islands (91), North American coastal and
 374 inland islands (87), Macaronesia (86), the Indo-Malay archipelago (75), and Hawai'i (65)
 375 (Fig.1A). In terms of taxonomic focus, we identified 306 studies on mammals, 268 on birds,
 376 246 on plants, 115 on insects, 88 on reptiles, 23 on amphibians, 14 on molluscs, 2 on arachnids,
 377 and 2 on nematodes (Fig. 1B).

378 Individual count data were notably low in many cases. For birds, 23 studies reported counts
 379 ≤ 10 individuals, while 100 reported populations effective population sizes of < 100 individuals

380 (70.9% of all studies reporting individual counts). For mammals, 16 studies reported ≤ 10
381 individuals while 43 reported < 100 (51.2%). For plants, 8 studies had counts ≤ 10 and 34 had
382 < 100 individuals (66.7%). Among reptiles, 9 studies reported counts ≤ 10 , and 15 reported < 100
383 (62.5%) (Fig. 1C). The lowest reported N_e values were 3 for amphibians, 6 for birds, 2 for both
384 insects and mammals, and 4 for both plants and reptiles (Fig. 1D).

385 Overall, 268 studies focused on lineages introduced through human-associated dispersal, while
386 816 focused on native or endemic taxa. Only 166 studies incorporated any form of temporal
387 sampling.

388

389

390 **Box 2 - Genomic trade-offs in long-term small populations:** 391 **insights from island species**

392 Island species often have small long-term effective population sizes (N_e), limited gene flow,
393 and persistent isolation. These demographic features may shape their genomes in predictable
394 yet counterintuitive ways, offering critical insights into how small populations persist, and how
395 they may fail.

396 In some long-term small- N_e populations, including island endemics, inbreeding depression
397 appears weak or undetectable [23,65]. This has often been interpreted as the result of purging
398 of strongly deleterious alleles [23], although it may also result from drift eliminating
399 segregating variation, including mildly deleterious and neutral alleles, giving an illusion of
400 resilience.

401 Yet this apparent resilience is double-edged. Genetic drift in chronically small populations may
402 fix mildly deleterious mutations, thus elevating drift load and reducing fitness, while also
403 steadily eroding genome-wide diversity, compromising adaptive potential. This genomic
404 erosion is subtle, often going unnoticed in stable ecological conditions, but it may be
405 catastrophic when rapid environmental change occurs [66].

406 Thus, island species exemplify a fundamental trade-off: they may be buffered against
407 inbreeding depression due to their long-term stable but low N_e , yet this comes at the cost of
408 adaptive flexibility. These two outcomes, reduced inbreeding depression and reduced adaptive
409 potential, are two sides of the same evolutionary coin. Apparent genetic stability may conceal
410 hidden fragility, especially when populations face novel pathogens, invasive species, or
411 climatic shifts. For this reason, island species serve as powerful models for understanding how

412 long-term demography dynamics shapes both persistence and vulnerability in small
413 populations.

414

415 **Glossary**

416 **Genetic drift** Stochastic changes in allele frequencies across generations, leading to loss of
417 genetic diversity due to random sampling of alleles, particularly in small populations.

418 **Genetic load** The fraction by which the population mean fitness differs from that of a reference
419 genotype (i.e., the genotype with maximum fitness).

420 **Genetic rescue** A strategy aimed at improving the genetic diversity and fitness of a small or
421 inbred population through the introduction of individuals from another population.

422 **Genomic erosion** Cumulative genetic threats to small populations, including loss of genome-
423 wide diversity, increased genetic load, maladaptation (a mismatch between existing adaptations
424 and the environment), and genetic introgression following hybridization (introduction of non-
425 compatible or maladapted alleles).

426 **Inbreeding** Mating between related individuals, leading to increased homozygosity beyond
427 what is expected under random mating (panmixia). Inbreeding is more frequent in small or
428 fragmented populations.

429 **Inbreeding depression** Reduction in individual fitness due to increased expression of partially
430 or fully recessive deleterious alleles in homozygous form. While it can facilitate purging of
431 deleterious variants, it may also reduce overall population viability.

432 **Masked load (inbreeding load or potential load)** Component of the genetic load whose
433 fitness effects are hidden because deleterious recessive alleles are present in heterozygous
434 form.

435 **Purging** Reduction of genetic load via purifying selection against deleterious alleles exposed
436 in homozygous form, often occurring through inbreeding, population fragmentation, or positive
437 assortative mating.

438 **Realized load** Component of genetic load representing deleterious alleles that are expressed

439

440 References

- 441 1. Sendell-Price, A.T. *et al.* (2021) An island-hopping bird reveals how founder events shape
442 genome-wide divergence. *Mol. Ecol.* 30, 2495–2510
- 443 2. Wang, X. *et al.* (2023) Demographic history and genomic consequences of 10,000 generations of
444 isolation in a wild mammal. *Curr. Biol.* 33, 2051–2062.e4
- 445 3. Carson, H.L. *et al.* (1990) Extinction and recolonization of local populations on a growing shield
446 volcano. *Proc. Natl. Acad. Sci. U. S. A.* 87, 7055–7057
- 447 4. Duntsch, L. *et al.* (2023) Genomic signatures of inbreeding depression for a threatened Aotearoa
448 New Zealand passerine. *Mol. Ecol.* 32, 1893–1907
- 449 5. Wilson, E.O. (1961) The Nature of the Taxon Cycle in the Melanesian Ant Fauna. *Am. Nat.* 95,
450 169–193
- 451 6. Ramalho, R.S. *et al.* (2015) Hazard potential of volcanic flank collapses raised by new
452 megatsunami evidence. *Sci. Adv.* 1, e1500456
- 453 7. Leroy, T. *et al.* (2021) Island songbirds as windows into evolution in small populations. *Curr. Biol.*
454 31, 1303–1310.e4
- 455 8. Charlesworth, B. and Charlesworth, D. (1999) The genetic basis of inbreeding depression. *Genet.*
456 *Res.* 74, 329–340
- 457 9. Charlesworth, B. (2009) Fundamental concepts in genetics: effective population size and patterns
458 of molecular evolution and variation. *Nat. Rev. Genet.* 10, 195–205
- 459 10. Díez-Del-Molino, D. *et al.* (2018) Quantifying temporal genomic erosion in endangered species.
460 *Trends Ecol. Evol.* 33, 176–185
- 461 11. van Oosterhout, C. *et al.* (2022) Genomic erosion in the assessment of species extinction risk and
462 recovery potential *bioRxiv*, 2022.09.13.507768
- 463 12. Bosse, M. and van Loon, S. (2022) Challenges in quantifying genome erosion for conservation.
464 *Front. Genet.* 13, 960958
- 465 13. Lavanchy, E. *et al.* (2024) Too big to purge: persistence of deleterious Mutations in Island
466 populations of the European Barn Owl (*Tyto alba*). *Heredity (Edinb.)* 133, 437–449
- 467 14. Grueber, C.E. *et al.* (2013) Genetic drift outweighs natural selection at toll-like receptor (TLR)
468 immunity loci in a re-introduced population of a threatened species. *Mol. Ecol.* 22, 4470–4482
- 469 15. Gilpin, M.E. and Soulé, M.E. (1986) Minimum viable populations: processes of species extinction.
470 In *Conservation biology: The science of scarcity and diversity*, pp. 19–43, Oxford University Press
- 471 16. Kutschera, V.E. *et al.* (2020) Purifying selection in corvids is less efficient on islands. *Mol. Biol.*
472 *Evol.* 37, 469–474
- 473 17. Keller, L.F. *et al.* (2002) Environmental conditions affect the magnitude of inbreeding depression
474 in survival of Darwin’s finches. *Evolution* 56, 1229–1239
- 475 18. Kennedy, E.S. *et al.* (2014) Severe inbreeding depression and no evidence of purging in an
476 extremely inbred wild species--the Chatham Island black robin. *Evolution* 68, 987–995
- 477 19. Niskanen, A.K. *et al.* (2020) Consistent scaling of inbreeding depression in space and time in a
478 house sparrow metapopulation. *Proc. Natl. Acad. Sci. U. S. A.* 117, 14584–14592
- 479 20. Seymour, A.M. *et al.* (2001) High effective inbreeding coefficients correlate with morphological
480 abnormalities in populations of South Australian koalas (*Phascolarctos cinereus*). *Anim. Conserv.*
481 4, 211–219
- 482 21. Lawson, L.P. *et al.* (2017) Slow motion extinction: inbreeding, introgression, and loss in the
483 critically endangered mangrove finch (*Camarhynchus heliobates*). *Conserv. Genet.* 18, 159–170
- 484 22. Johnson, J.A. *et al.* (2009) Long-term survival despite low genetic diversity in the critically
485 endangered Madagascar fish-eagle. *Mol. Ecol.* 18, 54–63
- 486 23. Robinson, J.A. *et al.* (2018) Purging of strongly deleterious mutations explains long-term
487 persistence and absence of inbreeding depression in island foxes. *Curr. Biol.* 28, 3487–3494.e4
- 488 24. Losos, J.B. and Ricklefs, R.E. (2009) Adaptation and diversification on islands. *Nature* 457, 830–
489 836
- 490 25. Cerca, J. *et al.* (2023) Evolutionary genomics of oceanic island radiations. *Trends Ecol. Evol.* 38,
491 631–642
- 492 26. Voigt, M. *et al.* (2021) Emerging threats from deforestation and forest fragmentation in the

- 493 Wallacea centre of endemism. *Environ. Res. Lett.* 16, 094048
- 494 27. Struebig, M.J. *et al.* (2022) Safeguarding imperiled biodiversity and evolutionary processes in the
495 Wallacea center of endemism. *Bioscience* 72, 1118–1130
- 496 28. Aninta, S.G. *et al.* (2025) The importance of small-island populations for the long-term survival
497 of endangered large-bodied insular mammals. *Proc. Natl. Acad. Sci. U. S. A.* 122, e2422690122
- 498 29. Keller, L. (2002) Inbreeding effects in wild populations. *Trends Ecol. Evol.* 17, 230–241
- 499 30. Dussex, N. *et al.* (2023) Adaptation to the High-Arctic island environment despite long-term
500 reduced genetic variation in Svalbard reindeer. *iScience* 26, 107811
- 501 31. Caballero, A. *et al.* (2017) Inbreeding load and purging: implications for the short-term survival
502 and the conservation management of small populations. *Heredity (Edinb.)* 118, 177–185
- 503 32. Yang, H. *et al.* (2024) Consistent accumulation of transposable elements in species of the Hawaiian
504 *Tetragnatha* spiny-leg adaptive radiation across the archipelago chronosequence. *Evolutionary*
505 *Journal of the Linnean Society* 3, kzae005
- 506 33. Major, R.E. *et al.* (2021) Islands within islands: genetic structuring at small spatial scales has
507 implications for long-term persistence of a threatened species. *Anim. Conserv.* 24, 95–107
- 508 34. Reatini, B. and Vision, T.J. (2024) The two faces of secondary contact on islands: Introgressive
509 hybridization between endemics and reproductive interference between endemics and introduced
510 species. *J. Biogeogr.* 51, 483–498
- 511 35. Salces-Castellano, A. *et al.* (2020) Climate drives community-wide divergence within species over
512 a limited spatial scale: evidence from an oceanic island. *Ecol. Lett.* 23, 305–315
- 513 36. Noguerales, V. *et al.* (2024) Genetic legacies of mega-landslides: Cycles of isolation and contact
514 across flank collapses in an oceanic island. *Mol. Ecol.* 33, e17341
- 515 37. Heber, S. *et al.* (2013) The genetic rescue of two bottlenecked South Island robin populations using
516 translocations of inbred donors. *Proc. Biol. Sci.* 280, 20122228
- 517 38. von Seth, J. *et al.* (2022) Genomic trajectories of a near-extinction event in the Chatham Island
518 black robin. *BMC Genomics* 23, 747
- 519 39. Antaky, C.C. *et al.* (2020) Unexpectedly high genetic diversity in a rare and endangered seabird
520 in the Hawaiian Archipelago. *PeerJ* 8, e8463
- 521 40. Brekke, P. *et al.* (2011) High genetic diversity in the remnant island population of hihi and the
522 genetic consequences of re-introduction. *Mol. Ecol.* 20, 29–45
- 523 41. Madsen, T. *et al.* (2023) Polyandry and non-random fertilisation maintain long-term genetic
524 diversity in an isolated island population of adders (*Vipera berus*). *Heredity (Edinb.)* 130, 64–72
- 525 42. Welch, A.J. *et al.* (2012) Ancient DNA reveals genetic stability despite demographic decline:
526 3,000 years of population history in the endemic Hawaiian petrel. *Mol. Biol. Evol.* 29, 3729–3740
- 527 43. Guhlin, J. *et al.* (2023) Species-wide genomics of kākāpō provides tools to accelerate recovery.
528 *Nat. Ecol. Evol.* 7, 1693–1705
- 529 44. Jackson, H.A. *et al.* (2022) Genomic erosion in a demographically recovered bird species during
530 conservation rescue. *Conserv. Biol.* 36, e13918
- 531 45. Speak, S.A. *et al.* (2024) Genomics-informed captive breeding can reduce inbreeding depression
532 and the genetic load in zoo populations. *Mol. Ecol. Resour.* 24, e13967
- 533 46. Bertorelle, G. *et al.* (2022) Genetic load: genomic estimates and applications in non-model
534 animals. *Nat Rev Genet* 23, 492–503
- 535 47. Kyriazis, C.C. *et al.* (2023) Using computational simulations to model deleterious variation and
536 genetic load in natural populations. *Am. Nat.* 202, 737–752
- 537 48. Dussex, N. *et al.* (2023) Purging and accumulation of genetic load in conservation. *Trends Ecol.*
538 *Evol.* 38, 961–969
- 539 49. Robinson, J. *et al.* (2023) Deleterious variation in natural populations and implications for
540 conservation genetics. *Annu. Rev. Anim. Biosci.* 11, 93–114
- 541 50. Jamieson, I.G. *et al.* (2006) Inbreeding and endangered species management: is New Zealand out
542 of step with the rest of the world?: Inbreeding in New Zealand birds. *Conserv. Biol.* 20, 38–47
- 543 51. Grossen, C. *et al.* (2020) Purging of highly deleterious mutations through severe bottlenecks in
544 Alpine ibex. *Nat. Commun.* 11, 1001
- 545 52. Smeds, L. and Ellegren, H. (2023) From high masked to high realized genetic load in inbred
546 Scandinavian wolves. *Mol. Ecol.* 32, 1567–1580
- 547 53. Johnson, J.A. *et al.* (2023) Whole-genome survey reveals extensive variation in genetic diversity

- 548 and inbreeding levels among peregrine falcon subspecies. *Ecol Evol* 13, e10347
- 549 54. Bourgeois, Y. *et al.* (2024) The burden of anthropogenic changes and mutation load in a critically
550 endangered harrier from the Reunion biodiversity hotspot, *Circus maillardi*. *Mol Ecol* 33, e17300
- 551 55. Dussex, N. *et al.* (2021) Population genomics of the critically endangered kākākō. *Cell Genom.* 1,
552 100002
- 553 56. Atağ, G. *et al.* (2024) Population Genomic History of the Endangered Anatolian and Cyprian
554 Mouflons in Relation to Worldwide Wild, Feral, and Domestic Sheep Lineages. *Genome Biol Evol*
555 16
- 556 57. Robinson, J.A. *et al.* (2016) Genomic Flatlining in the Endangered Island Fox. *Curr Biol* 26, 1183–
557 1189
- 558 58. Mattila, A.L.K. *et al.* (2012) High genetic load in an old isolated butterfly population. *Proc Natl*
559 *Acad Sci U S A* 109, E2496-505
- 560 59. Cavill, E.L. *et al.* (2024) When birds of a feather flock together: Severe genomic erosion and the
561 implications for genetic rescue in an endangered island passerine. *Evol Appl* 17, e13739
- 562 60. Weiser, E.L. *et al.* (2016) Unexpected positive and negative effects of continuing inbreeding in
563 one of the world's most inbred wild animals: EFFECTS OF INBREEDING IN BLACK ROBINS.
564 *Evolution* 70, 154–166
- 565 61. Fernández-Palacios *et al.* (2021) Scientists' warning—The outstanding biodiversity of islands is in
566 peril. *Glob. Ecol. Biogeogr.* at
567 <<https://www.sciencedirect.com/science/article/pii/S2351989421003978>>
- 568 62. Falcón, W. and Hansen, D.M. (2018) Island rewilding with giant tortoises in an era of climate
569 change. *Philos Trans R Soc Lond B Biol Sci* 373
- 570 63. Hedrick, P.W. *et al.* (2014) Genetic rescue in Isle Royale wolves: genetic analysis and the collapse
571 of the population. *Conserv. Genet.* 15, 1111–1121
- 572 64. Maroso, F. *et al.* (2023) Fitness consequences and ancestry loss in the Apennine brown bear after
573 a simulated genetic rescue intervention. *Conserv. Biol.* 37, e14133
- 574 65. Uzans, A.J. *et al.* (2015) Small Ne of the isolated and unmanaged horse population on Sable Island.
575 *J. Hered.* 106, 660–665
- 576 66. Femerling, G. *et al.* (2023) Genetic load and adaptive potential of a recovered avian species that
577 narrowly avoided extinction. *Mol. Biol. Evol.* 40, msad256
- 578