

1 Consensus on future research directions in the Phylum Rotifera

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3 **Running Head:** Global priorities in rotifer research

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5 Rafael L. Macêdo^{1,2*}, Carlota Solano-Udina^{3§}, Melanie D. Borup^{4,5§}, Marco Antonio Jiménez-
6 Santos^{6,7}, Cristina Arenas-Sánchez³, Christina W.C. Branco⁸, Karen Costa⁸, Betina Kozlowsky-
7 Suzuki⁹, Claudia Bonecker¹⁰, Elizabeth J. Walsh¹¹, André R.S. Garraffoni¹², S.S.S. Sarma¹³,
8 Agnieszka Ochocka¹⁴, S. Nandini¹³, Robert Wallace¹⁵, Diego Fontaneto¹⁶, Jonathan M. Jeschke^{1,2},
9 Gissell Lacerot¹⁷

10

11 *corresponding author

12 § macedo@edu.unirio.br

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15 ¹ Leibniz Institute of Freshwater Ecology and Inland Fisheries (IGB), Müggelseedamm 310, 12587
16 Berlin, Germany17 ² Institute of Biology, Freie Universität Berlin, Königin-Luise-Str. 1–3, 14195 Berlin, Germany18 ³ Evolutionary Ecology Unit, Institut Cavanilles de Biodiversitat i Biologia Evolutiva, Universitat
19 de València, Spain20 ⁴ Institute for Marine and Antarctic Studies, University of Tasmania, Battery Point, Tasmania,
21 Australia22 ⁵ Australian Antarctic Division, Department of Climate Change, Energy, the Environment and
23 Water, Kingston, Tasmania, Australia24 ⁶ Institute of Soil Biology and Biogeochemistry, Biology Centre, Czech Academy of Sciences,
25 České Budějovice, Czech Republic26 ⁷ Laboratory of Non-Mendelian Evolution, Institute of Animal Physiology and Genetics, Academy
27 of Sciences of the Czech Republic, Liběchov, Czech Republic28 ⁸ Departamento de Zoologia, Universidade Federal do Estado do Rio de Janeiro, Rio de Janeiro,
29 Brazil30 ⁹ Departamento de Ecologia e Recursos Marinhos, Universidade Federal do Estado do Rio de
31 Janeiro, Rio de Janeiro, Brazil32 ¹⁰ Núcleo de Pesquisa em Limnologia, Ictiologia e Aquicultura (NUPÉLIA), Universidade
33 Estadual de Maringá, Paraná, Brazil34 ¹¹ Department of Biological Sciences, The University of Texas at El Paso, El Paso, TX, USA35 ¹² Laboratory of Evolutionary Meiofaunal Organisms, Department of Animal Biology, Institute of
36 Biology, Universidade de Campinas, Campinas, Brazil37 ¹³ Laboratorio de Zoología Acuática, Universidad Nacional Autónoma de México, FES Iztacala,
38 State of Mexico, Mexico39 ¹⁴ Department of Aquatic Ecosystems Research, Institute of Environmental Protection – National
40 Research Institute, Warsaw, Poland

41 ¹⁵ Department of Biology, Ripon College, Ripon, WI, USA

42 ¹⁶ National Research Council of Italy (CNR), Water Research Institute (IRSA), Verbania, Italy

43 ¹⁷ Departamento Interdisciplinario de Sistemas Costeros y Marinos, Universidad de la República,
44 Maldonado, Uruguay

45

46 **Abstract**

47

48 Rotifers play key roles in aquatic ecosystems, yet significant uncertainty remains about their
49 diversity and evolution, and basic knowledge is still lacking to address practical challenges related
50 to global change. To identify major knowledge gaps hindering progress, we carried out a Delphi
51 process both online and during the 17th International Rotifer Symposium, involving more than 40
52 experts across diverse regions and subdisciplines. A total of 133 research questions were screened
53 for relevance and clarity and reduced to 100 for online scoring. These were evaluated on a 1-to-10
54 priority scale, after which 67 questions exceeding 50% agreement were advanced to an in-person
55 workshop. Through structured discussions and round-table voting, participants identified gold,
56 silver, and bronze priority questions, while considering the feasibility of addressing them, resulting
57 in a final consensus set of high-priority questions across basic, applied, and philosophical
58 perspectives including how rotifer communities are defined, how evidence is generated and
59 integrated, and how values shape research priorities. Support for questions on knowledge transfer,
60 digital curation, and AI-assisted identification highlights the emergence of a methodological
61 subfield linking classical approaches with modern computational tools. Likewise, the emphasis on
62 improving genetic markers and connecting DNA sequences with traits shows that molecular
63 research is now viewed as a foundational component of rotiferology. This synthesis provides the
64 first community-driven roadmap for rotifer research. By articulating shared priorities and
65 clarifying persistent knowledge gaps, including unreliable phylogenies, uneven sampling

66 worldwide, and limited hypothesis-driven work, it provides a foundation for future collaborative
67 projects, funding strategies, and cross-disciplinary integration.

68

69 **Keywords:** communities, diversity, evolution, methods, populations, rotifer research agenda

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71 **1. Introduction**

72

73 Collaborative identification and prioritization of fundamental research questions have become an
74 effective way to organize knowledge in complex scientific disciplines (Sutherland et al. 2012; Dey
75 et al. 2020). A common framework for this process is the Delphi technique, which has evolved
76 since its development in the 1950s into a structured expert consultation process for many scientific
77 disciplines. This approach is useful not only to build consensus but also to identify priorities and
78 expose areas where viewpoints diverge (Hasson et al. 2000). In a typical implementation, a panel
79 of experts completes two or more rounds of questionnaires (in ecology and conservation
80 specifically, see Mukherjee et al. 2015). After each round of feedback, responses are aggregated,
81 summarized, and returned anonymously to participants, who are invited to reconsider their views
82 in light of the group's feedback. It has been used successfully in broad ecological and conservation
83 contexts (Sutherland et al. 2012; Mukherjee et al. 2015) and in more focused fields including
84 subterranean biology, invasion biology or meiofauna research, to overcome limitations of
85 bibliometric approaches and study different topics and various taxonomic groups and habitats,
86 revisit unanswered questions and foster hypothesis-driven studies (Enders et al. 2020; Mammola
87 et al. 2020; Martínez et al. 2025). By producing a shared roadmap, such exercises are particularly

88 valuable in disciplines with a long research history that are now being reshaped by new
89 technological capabilities and pressing environmental challenges.

90

91 The study of Rotifera, a phylum of microscopic metazoans, exemplifies a discipline where long-
92 standing research intersects with emerging tools and new environmental challenges. With a rich,
93 century-old history of taxonomic and ecological research (see Ehrenberg 1838; Hudson and Gosse
94 1886; Hillbricht-Ilkowska 1995), our understanding of rotifers has been profoundly shaped by
95 advancements in microscopy, molecular biology, and ecological modeling (Lampert 1997; Gómez
96 2005; Serra et al. 2019; Gansfort et al. 2020). Rotifers, inhabiting freshwater, saline, and semi-
97 terrestrial ecosystems, play pivotal roles in nutrient cycling, energy transfer, and serve as
98 bioindicators of environmental health (Arndt 1993; Fontaneto et al. 2006; Ejsmont-Karabin 2012;
99 Wallace et al. 2021). Rotifers exhibit diverse morphologies, feeding mechanisms, and life-history
100 strategies that make them a valuable system for fundamental biological inquiry. Among them,
101 bdelloid rotifers are particularly remarkable for their anhydrobiotic capabilities, obligate
102 parthenogenesis, and horizontal gene transfer (Ricci 1983; Melone and Ricci 1995; Ricci and
103 Boschetti 2003; King et al. 2005; Gladyshev and Meselson 2008; Declerck and Papakostas 2017;
104 Hespeels et al. 2023). However, important gaps in rotifer research remain. Geographic and
105 taxonomic biases continue to limit our understanding of global biogeography and biodiversity
106 patterns (Dumont 1983; Fontaneto et al. 2012). In addition, the widespread occurrence of cryptic
107 species complicates estimates of species richness, distribution, and ecological roles (May and
108 Wallace 2019; Wallace et al. 2024; Walczyńska et al. 2024).

109 Against this backdrop, we here report results of an exercise to identify the most important
110 outstanding research questions related to rotifers. Rapid environmental change, together with the

111 development of powerful new research tools (e.g., eDNA, genomics: Mohl et al. 2025; AI-assisted
112 taxonomy and ecology: Ienaga et al. 2024; Zhang et al. 2024; Diouf et al. 2026) creates new
113 opportunities to address long-standing questions in biology, ecology and conservation research.
114 This study presents the outcome of a comprehensive Delphi process designed to elicit, refine, and
115 prioritize a consensus-based list of key unanswered questions that will guide the next era of rotifer
116 research. By synthesizing the expertise of the international rotifer research community, we aim to
117 help students, researchers, and funding agencies set priorities, while also encouraging
118 collaboration across different areas of rotifer biology by highlighting shared interests and
119 persistent challenges. Articulating fundamental questions also stimulates development of new
120 hypotheses, because questions define the boundaries of inquiry, whereas hypotheses provide
121 testable explanations. Thus, this study not only outlines directions for future research but also
122 reinforces the iterative cycle of questioning and hypothesis testing that drives scientific progress.
123 Ultimately, we hope this effort will inspire a new wave of integrated and impactful research,
124 deepen our understanding of this fascinating phylum, and strengthen our ability to conserve and
125 manage freshwater ecosystems.

126

127 **2. Materials and Methods**

128

129 We followed a structured approach based on the Delphi technique to gather and prioritize expert
130 opinions on key unanswered questions in Rotifera research. The Delphi process was designed to
131 be iterative, involving multiple rounds of consultation and feedback, moving from a broad
132 collection of questions to a refined, prioritized list that reflects current challenges, emerging trends,
133 and future directions considered most important by the international rotifer research community.

134 Here, the Delphi process was carried out through both pre-symposium online forms (Phases 1 and
135 2) and in-person discussions during the workshop “Emerging Consensus and Key Questions in
136 Rotifera Research”, held at the 17th International Rotifer Symposium (IRS) in Rio de Janeiro,
137 Brazil, in August 2025 (Phase 3, see Fig. 1 for the workflow). The IRS is a triennial meeting that
138 has, since 1976, gradually become a focal point for discussions and collaborations.

139

140 During Phase 1 of the Delphi process, experts were invited to submit three to five key research
141 questions. Over 230 e-mail invitations were sent to individuals selected based on their
142 contributions to innovative, high-impact, and high-volume research on Rotifera, and their
143 participation in the International Rotifer Symposium (IRS). In addition, the invitation was shared
144 via the rotifer-family newsletter mailing list to broaden outreach. In total, 37 experts responded.
145 For each contributor, their region of work, study environment, education level, ethnicity, and
146 gender were assessed (see Fig. 2). Completion of the demographic questions was optional. In all
147 133 research questions were formulated, and these were screened for duplicates and refined for
148 clarity, consistency, and alignment with the predefined criteria of relevance and testability,
149 resulting in a list of 100 questions (see Supplementary Document S1).

150

151 Overly broad or vague topics were avoided by focusing on questions that could realistically be
152 addressed by a small research team or through a limited set of funded projects. To achieve this, we
153 adopted an established methodology based on Sutherland et al. (2011a; b), which emphasizes a
154 rigorous, democratic, and transparent approach to identify key research questions. Thus, an ideal
155 question should either directly suggest a research design or be framed in a way that allows it to be
156 translated into specific, testable research hypotheses, according to the following criteria:

- 157 (i) be answerable through a realistic research design;
- 158 (ii) address significant gaps in current knowledge;
- 159 (iii) have a factual answer that is not dependent on value judgments;
- 160 (iv) be of a spatial and temporal scale that is feasible for a research team to address;
- 161 (v) not be formulated as a broad or vague topic area;
- 162 (vi) avoid being answerable with "it depends";
- 163 (vii) not be framed as a yes/no question (e.g., avoid phrasing it like "Is Lecanidae more species-
- 164 rich than Brachionidae?");
- 165 (viii) and, if related to impacts or interventions, clearly include a subject, an intervention, and a
- 166 measurable outcome.

167

168 During Phase 2, the 100 questions were ranked through an online voting process, prior to the in-

169 person workshop. During this phase, experts rated each question on a scale from 1 to 10, where 1–

170 3 indicated low relevance, 4–6 intermediate relevance, 7–9 high relevance, and 10 very high

171 priority. The same 37 participants as in Phase 1 were instructed to base their assessments primarily

172 on their scientific knowledge and expertise rather than on personal interest, while acknowledging

173 that complete objectivity may not be attainable. Based on the online voting results, 67 questions

174 were selected according to a pre-established consensus threshold of 50% agreement on high or

175 very high priority, and these were brought forward to the experts attending the workshop at the

176 17th IRS. The prioritized research questions were subsequently categorized into six themes for in-

177 person voting. These themes broadly reflected major ecological topics or methodological

178 approaches. However, the boundaries between themes were approximate, as many questions span

179 more than one area (Sutherland et al. 2012). This was done to avoid that sorting questions into

180 themes too early might unintentionally discourage cross-cutting perspectives and novel
181 combinations of ideas.

182

183 Phase 3 consisted of an in-person workshop at the 17th IRS. All participants received the voting
184 results and were asked to reflect on the 67 most relevant questions prior to the meeting. However,
185 changes in gender balance and participant characteristics were expected during the 17th IRS due to
186 logistical and availability constraints not present in the online process. Participants were self-
187 divided into seven groups (Community and Diversity, Ecosystems and Functioning, Evolution and
188 Ecology, Human Impacts and Global Change, Methods and Populations; Fig. 1), comprising 5–10
189 experts, with two groups assigned to *Community and Diversity* because this theme contained the
190 largest number of questions. Group leaders were chosen for the in-person discussions and asked
191 to ensure that the process remained democratic, with all views respected. These individuals were
192 invited to volunteer based on their established research interests in their respective group themes.
193 Within each group, experts were instructed to rank all questions as gold, silver, or bronze, without
194 limits on each type. Gold questions generally reflected broad research across interdisciplinary
195 topics; silver questions addressed important but more focused topics; and bronze questions
196 captured research gaps that were potentially overlooked but still considered essential by the group.
197 Then, the group selected two gold, one silver, and one bronze questions. Votes during the selection
198 process of gold, silver, and bronze questions were used to understand group consensus across each
199 theme. Final decisions were made by a show-of-hands vote, requiring a consensus threshold of
200 75% for the selection of gold, silver, and bronze questions. Questions that did not reach consensus
201 were retained in the final record to acknowledge areas of expert disagreement and to reflect the
202 current diversity of perspectives within rotifer ecology. A final plenary session then determined

203 the overall top-priority questions while also accounting for the feasibility to address these
204 questions.

205

206 All analyses were performed using R Statistical Software (v 4.4.2; R Core Team 2026). Boxplots
207 were used to summarize the distribution, central tendency, and variability of question scores by
208 theme. Differences in mean question scores among themes were tested using Welch's one-way
209 ANOVA, which does not assume equal variances or balanced sample sizes. When significant
210 differences were detected, pairwise comparisons were applied. A Sankey diagram was used to
211 illustrate the flow and relative importance of questions across themes, subthemes and priority
212 levels.

213

214 **3. Results**

215

216 **3.1 Expert Panel**

217 The expert panel ($n = 37$) was geographically diverse but predominantly based in Europe, followed
218 by representation from South America and North America. Most participants specialized in
219 freshwater ecosystems and were senior researchers with over 10 years of experience, indicating
220 strong disciplinary expertise, and there was limited early-career researcher input. Ethnic
221 representation was primarily white, with lower participation from underrepresented groups, and
222 the gender balance was slightly male-skewed (56.8% male, 40.5% female, 2.7% preferred not to
223 say) (Fig. 2).

224

225 **3.2 Priority questions**

226
227 Across themes, Communities and Diversity included the largest number of high-priority questions
228 (21 out of 35; 60% within this theme), consistent with it also having the largest number of
229 submitted questions. Although it had the lowest absolute number of questions, Human Impacts
230 and Global Change showed the highest within-theme proportion of high-priority classifications (5
231 out of 6; 83.3%). Methods also contained many high-priority questions (16 out of 23; 69.6%),
232 reflecting strong consensus on the need for methodological advances. Notably, *Methods* included
233 many high-priority questions, reflecting a strong consensus that methodological advances
234 (especially molecular tools, integrative approaches, and digital identification) are urgently needed
235 to support progress across all research areas. From the full set of questions submitted by experts
236 (see demographic composition in Fig. 2), the 67 questions that reached at least 50% agreement on
237 their relevance during Phase 2 are listed below by theme (Boxes 1-6), without implying rank. Mean
238 scores on the 1–10 scale showed only small differences among themes, and confidence intervals
239 largely overlapped (Welch's ANOVA: $F_{5,26.977} = 1.17$, $p = 0.350$), indicating that all themes
240 received similar evaluations (Fig. 3).

241
242 Several subthemes clustered strongly toward high priority, including *Trait Evolution & Plasticity*,
243 *Species Interactions*, *Functional Diversity*, *Environmental Stressors*, *Molecular & Genetic Tools*,
244 and *Biodiversity Patterns & Distribution*. In contrast, *Morphology*, *Monitoring & Assessment*, and
245 *Communication* showed a higher proportion of lower-priority classifications (See Fig. 4 for the
246 full list of subthemes). The "High Priority" node dominates the flow, reflecting a strong expert
247 consensus on topic urgency. *Biodiversity Patterns & Distribution* and *Bdelloids* serve as a hub for
248 5 out of 6 themes, underscoring the multidisciplinary nature of these priorities. The subtheme

249 *Bdelloids* grouped questions that specifically addressed lineage-specific traits of bdelloid rotifers,
250 such as obligate parthenogenesis, anhydrobiosis, horizontal gene transfer, and distinctive
251 reproductive strategies, rather than broad questions applicable to Rotifera as a whole.

252

253 Consensus dynamics during Phase 3 varied across themes. Most themes (*Ecosystems and*
254 *Functioning, Evolution and Ecology, Methods, Populations*) reached full agreement on gold,
255 silver, and bronze questions. In contrast, *Communities and Diversity* showed internal
256 disagreement: one subgroup did not reach consensus on a gold question, likely reflecting
257 differences between monogonont-focused and bdelloid-focused researchers. The highest-scoring
258 question from the online voting round (Q05-What are the mechanisms behind the extreme
259 tolerance of bdelloid rotifers?) from *Ecosystems and Functioning* did not appear among the final
260 gold, silver, or bronze selections in the in-person workshop. This discrepancy illustrated how
261 expert-panel composition and in-person deliberation can shift perceived priorities. *Human Impacts*
262 *and Global Change* had only gold consensus questions, partly due to the smaller number of
263 questions generated in this theme and its broader conceptual diversity. Moreover, most gold
264 questions fell into the high-feasibility quadrant (Fig. 5). Some questions, however, were judged
265 high priority but low feasibility (e.g., Q32, Q71, Q53), indicating important topics that require
266 substantial methodological or conceptual advances. Only Q26 was rated highly feasible despite
267 being also considered of low priority, suggesting accessible research opportunities that are not
268 considered urgent.

269

270 The highlighted parts of Boxes 1–6 indicate those questions that were selected during Phase 3.
271 These questions were evaluated for both relevance and feasibility, and their classification as gold,

272 silver and bronze reached at least 75% agreement (see Fig. 5). Note that for some questions, there
273 may already be some theoretical understanding, but empirical support for the theory is still lacking
274 across taxonomic groups or contexts.

275

276 For each of the six themes ($n = 100$ questions), the share of high-priority questions was as follows:
277 *Communities and Diversity* 60% ($n = 35$); *Ecosystems and Functioning* 72.7% ($n = 11$); *Evolution*
278 *and Ecology* 58.3% ($n = 12$); *Human Impacts and Global Change* 83.3% ($n = 6$); *Methods* 69.6%
279 ($n = 23$); *Populations* 76.9% ($n = 13$).

280

281 **4. Discussion**

282

283 The thematic synthesis of the multi-stage expert consensus process (Phases 2 and 3) is presented
284 in Boxes 1–6. These Boxes highlight the prioritized research questions that achieved 'Gold' or
285 'Silver' status, representing collective agreement on the most critical challenges in rotifer ecology.
286 Together, the thematic summaries provide a structured roadmap that captures both broad
287 interdisciplinary priorities and essential technical gaps.

288

289 *Communities and Diversity*

290

291 Diversity within rotifer communities encompasses taxonomic, functional and genetic dimensions,
292 serving as indicators of environmental change and providing insights into ecological processes
293 across space and time (Wallace 2002; Obertegger et al. 2011; Kuczynska-Kippen et al. 2021;
294 Wallace et al. 2021). Rotifers inhabit a wide range of environments, from permanent aquatic

295 systems to highly ephemeral habitats lasting only a few days, exposing their communities to
296 markedly different ecological conditions across spatial and temporal scales (Wallace et al. 2006;
297 Segers 2007). This environmental heterogeneity complicates efforts to comprehensively
298 characterize the diversity and structure of rotifer communities. Integrated and interdisciplinary
299 approaches are therefore required to understand rotifer communities across the highly diverse
300 environments they inhabit. We could, for example, (1) consider the relative importance of
301 qualitative versus quantitative differences in the structure of rotifer communities, (2) analyze the
302 distribution of the physical and metabolic traits that they possess across their habitats, and (3)
303 evaluate their capacity for rapid evolution within the diversity of environments in which they are
304 found. Alongside the need for new approaches, several knowledge gaps persist. These include
305 limited spatial and temporal coverage, biases on methodological approaches, scarce information
306 on functional and genetic diversity, limited understanding of biotic interactions and responses to
307 multiple stressors. These shortcomings hinder our ability to generalize patterns, compare studies
308 across regions, and link community changes to ecosystem processes. Among the most pressing
309 topics, experts emphasize the need for intergenerational transfer of taxonomic knowledge,
310 ensuring the detection of cryptic diversity and production of comparable inventories (Q71, Box 1).
311 Moving towards trait- and interaction-based frameworks is equally essential, as these processes
312 underpin community structure and ecosystem functioning (Q97), shape how environmental
313 stressors restructure communities (Q18), and influence their roles in ecosystem processes (Q26).
314 Rotifer diversity research also has an interdisciplinary component, as patterns and processes
315 observed in rotifers can inform broader questions in other fields of research, such as ecology and
316 evolution. Similarly, the identification of reliable bioindicators and trait-based metrics (Q89) has
317 direct relevance for ecosystem monitoring and management across aquatic habitats.

Box 1. Selected questions in the Community and Diversity theme after Phase 2. Highly ranked questions resulting from Phase 3 are highlighted in orange.

Q12. How can we interpret and resolve the concept of cosmopolitanism among rotifer species?

(Gold)

Q18. What are the effects of multiple environmental stressors on rotifer development and community structure? (Gold)

Q71. What strategies can sustain intergenerational transfer of taxonomic expertise in rotifer research? (Gold)

Q97. What types of ecological interactions occur between rotifers and other organisms, and how do these interactions affect community structure and ecosystem functioning? (Gold)

Q26. What are the effects of specific environmental stressors (e.g., increased temperature, nutrient enrichment, or emerging contaminants) on rotifer community functional traits and their contribution to ecosystem processes in freshwater environments? (Silver)

Q89. Which rotifer species or trait-based groups serve as effective bioindicators in lakes and reservoirs, and which environmental gradients drive their patterns? (Silver)

Q22. Does global rotifer biodiversity reflect the biodiversity of species commonly used as indicators of environmental degradation? (Bronze)

Q91. Which quantifiable traits best represent rotifer functional diversity across diverse aquatic habitats? (Bronze)

Q08. Why has the number of rotifer species recorded in Latin America remained low, and what factors (e.g., sampling bias, taxonomic effort) explain this pattern?

- Q14.** How do taxonomic and functional diversity patterns of rotifer communities vary across environmental gradients and biogeographic regions, and what ecological or evolutionary factors explain these patterns?
- Q14.** What is the most appropriate framework to analyze beta diversity in rotifers: species contributions (SCBD) or local contributions (LCBD)?
- Q15.** What are the environmental drivers that shape beta and functional diversity of rotifers across different regions?
- Q17.** How do rotifer communities respond to rapid environmental change compared to crustaceans like copepods and *Daphnia*?
- Q23.** Which rotifer community metrics (e.g., diversity, dominance, trophic interactions) are most sensitive to early biodiversity loss caused by pond desiccation and habitat fragmentation?
- Q29.** How does cryptic diversity in monogonont rotifers vary between molecular and morphological assessments, and what does this reveal about species richness?
- Q30.** How many rotifer species likely exist globally, considering cryptic diversity and sampling gaps?
- Q31.** How does the scale of cryptic diversity differ between monogonont and bdelloid rotifers, considering their reproductive and genetic systems?
- Q39.** How does spatial connectivity influence rotifer species or genotype exchange across freshwater metacommunities, and how can mesocosm experiments simulate this?
- Q50.** How does environmental changes affect the expression of plastic traits in species complexes (e.g., *Brachionus calyciflorus*)?
- Q90.** Which rotifer taxonomic or functional groups respond most sensitively to water quality degradation in rivers and streams, and how can they serve as bioindicators?

Q94. How do anthropogenic actions alter rotifer functional diversity in streams and ponds?

319

320 *Ecosystems and Functioning*

321

322 Rotifers often dominate aquatic ecosystems in abundance playing a central role in nutrient cycling
323 and energy transfer through microbial food webs. Rotifers also connect detritus, bacteria, algae,
324 and other microorganisms to higher consumers such as crustaceans, insect larvae, and different
325 species of small fish. Due to a high reproductive capacity and short generation time, rotifers
326 populations can expand rapidly in response to fluctuations in food availability, facilitating the
327 efficient use of ephemeral or newly available resources (Walz 1987; Gilbert 2022). Such extensive
328 morphological and functional diversity (Obertegger and Flaim 2015; Balkić et al. 2017;
329 Obertegger and Wallace 2023) allows the group to inhabit a wide range of aquatic systems, from
330 temporary ponds to oligotrophic and eutrophic lakes, positioning rotifers as ideal models for
331 studying ecosystem responses to disturbance. Functional approaches have improved our
332 understanding of zooplankton ecology (Branco et al. 2023), including links between rotifer feeding
333 guilds and land use in tropical streams (Bomfim et al. 2023). However, the integration of
334 frameworks combining response and effect traits (Hébert et al. 2017) remains limited (Huỳnh et
335 al. 2024), leaving several major questions open. Relevant knowledge gaps concern how
336 environmental stressors and anthropogenic disturbances reshape rotifer functional traits,
337 community structure, and trophic roles (Q21, Q87, Box 2), how short- and long-term climate
338 variability influences community resilience (Q25), and how rotifers contribute to energy transfer
339 in tropical food webs (Q86). However, without a joint implementation across spatial and temporal

340 scales of response-and-effect trait frameworks that meaningfully connect rotifers to ecosystem
341 functioning, these key questions will remain partly unanswered.
342

Box 2. Selected questions in the Ecosystems and Functioning theme after Phase 2.

Highly ranked questions resulting from Phase 3 are highlighted in purple.

Q21. What are the effects of specific environmental stressors (e.g., temperature, nutrients, contaminants) on rotifer functional traits and their contribution to ecosystem processes? (Gold)

Q87. How do anthropogenic disturbance gradients shape the functional composition and trophic roles of rotifer communities in freshwater and transitional systems? (Gold)

Q25. To what extent does the structure of rotifer communities reflect short-term versus long-term climate-induced changes in small aquatic ecosystems? (Silver)

Q86. How do rotifers contribute to energy transfer and trophic dynamics in tropical stream food webs? (Bronze)

Q005. What are the mechanisms behind the extreme tolerance of bdelloid rotifers?

Q055. How does desiccation duration affect hatching success and development time in rotifer resting eggs with different life-history strategies?

Q56. What molecular and physiological mechanisms enable rotifer resting stages to survive desiccation, and how do these mechanisms differ among taxa from contrasting hydrological regimes?

Q57. Which morphological traits of rotifer resting eggs predict delayed hatching or reduced viability across environmental gradients (e.g. salinity, nutrients)?

346 *Evolution and Ecology*

347

348 Evolutionary aspects within a taxon of interest have been the focus of several lines of research
349 across the whole tree of life: most taxa have now reliable phylogenies that allow addressing clear
350 questions to, for example, disentangle effects of interspecific relationships, including for birds
351 (Stoddard et al. 2017), mammals (DeCasien et al. 2017), and spiders (Hopfe et al. 2024). For
352 rotifers, early phylogenetic studies did not manage to provide unambiguous relationships (Melone
353 et al. 1998; Sørensen and Giribet, 2006), and the current use of phylogenomics did not improve
354 the situation (Vasilikopoulos et al. 2024; Herlyn et al. 2025). Without a reliable phylogeny, most
355 of the relevant questions highlighted by the panel of experts cannot be addressed, given the
356 potential confounding factor of evolutionary relationships in cross-taxa comparative analyses
357 (Garamszegi 2014). Without improving conceptual and comparative phylogenetic frameworks,
358 many key evolutionary questions such as those addressing the genetic and ecological mechanisms
359 underlying bdelloid speciation and adaptation (Q32, Q36, Box 3), the evolutionary role of
360 homologous and horizontal gene transfer in asexual lineages (Q068, Q78), and the phylogenetic
361 structure and morphological innovation within the group (Q96), remain unresolved.
362 Notwithstanding such limitation, there is a broad interest in eco-evolutionary aspects of rotifers,
363 especially related to their peculiar reproductive biology, the ability to survive desiccation, and the
364 high level of horizontal gene transfer.

365

Box 3. Selected questions in the Evolution and Ecology theme after Phase 2. Highly ranked questions resulting from Phase 3 are highlighted in red.

Q32. What are the main ecological or genetic mechanisms driving speciation in bdelloid rotifers? (Gold)

Q36. Which genetic mechanisms enable rotifers to adapt to changes in salinity? (Gold)

Q78. Why are horizontally transferred genes so common and successfully integrated in bdelloid rotifers? (Silver)

Q96. What parasites and epibionts infect rotifers, how specific are these associations, and what are their impacts on host fitness? (Bronze)

Q68. In the absence of sexual recombination, how do alternative forms of homologous recombination contribute to adaptation in bdelloids?

Q79. How can phylogenetic relationships among rotifers inform our understanding of organ system evolution and morphological innovation?

Q80. What is the current phylogenetic structure of rotifers, and which clades are most closely related?

366

367 ***Human Impacts and Global Change***

368

369 Anthropogenic pressures such as habitat degradation, biological invasions, and contamination are
 370 among the most severe threats to biodiversity. Disturbance often reduces native populations,
 371 disrupts trophic links, and accelerates community-wide biodiversity loss (Molinero et al. 2006).

372 Yet much less is known about the effects of urbanization on microscopic animals, whose responses
 373 may differ across contexts and can sometimes be unpredictable (Macêdo et al. 2020; Partemi et al.

374 2024; Han et al. 2025). These impacts generate many important questions (Q38–Q42).

375 Anthropogenic pressures have created novel selective forces favoring pollution-tolerant rotifer

376 species while destabilizing population dynamics, highlighting the need for robust quantification of
377 the strength and direction of these responses. Key questions include how multiple stressors jointly
378 affect rotifer development and reproduction (Q61, Box 4), how sensitive rotifers are to
379 environmental and contaminant gradients across regions (Q04), and how such responses link to
380 functional traits. An underappreciated dimension of global change is the biological invasion of
381 microorganisms, with potentially far-reaching consequences (Macêdo et al. 2022; Oesterwind et
382 al. 2025). Current understanding remains limited, risking misinterpretation and misuse of jargon
383 in the field (Oliveira et al. 2019; Arcifa et al. 2020), with broad hypothesis testing still uncommon
384 (Branco et al. 2023). This raises further questions about how rotifer abundance and diversity shift
385 after invasions and the mechanisms underlying these changes (Q100). Meanwhile, wastewater
386 treatment systems reveal the dual roles of rotifers as bioindicators and as active agents in
387 remediation (Pajdak-Stós et al. 2023; Soto et al. 2019), prompting questions about the
388 effectiveness of rotifer-based indices in detecting subtle shifts in water quality (Q16) and their
389 contributions to nutrient recycling and microbial regulation in reuse-oriented systems (Q84).
390

Box 4. Selected questions in the Human Impacts and Global Change theme after Phase 2.

Highly ranked questions resulting from Phase 3 are highlighted in grey.

Q84. How do rotifer communities influence nutrient recycling and microbial regulation in reuse-oriented systems like wastewater or agricultural reservoirs? (Gold)

Q100. How does rotifer abundance and diversity change in response to invasive species introductions, and what mechanisms underlie these responses? (Gold)

Q61. How do multiple environmental stressors affect rotifer development, survival, and

reproduction across taxa? (Silver)

Q04. How sensitive are bdelloid rotifers to environmental stressors (salinity, temperature) and contaminants (e.g., metals, emerging contaminants), and are there locality-specific differences (e.g., Antarctic vs. Tropical)?

Q16. How effective are rotifer-based bioindicator indices in detecting subtle shifts in water quality across gradients of anthropogenic impact?

391

392 *Methods*

393

394 Accurate identification and delimitation of species are crucial elements of rotifer research. In the
395 past, most researchers received taxonomic instruction from other researchers. Unfortunately, the
396 number of rotifer taxonomists is declining, thus the opportunity for students to receive adequate
397 training is nearly non-existent (Wallace et al. 2024). To build capacity and competency among
398 new researchers, rotiferologists need to explore novel and efficient approaches (Q42, Box 5). A
399 related issue is the need of assessing many samples rapidly and accurately. Artificial intelligence
400 tools may have the capacity to do this, but we need to have the procedures developed and tested
401 and make them available at reasonable costs (Wallace et al. 2024) (Q46). Associated with accurate
402 identification of species is the use of molecular data in two areas: (1) Appropriate genetic markers
403 are needed to advance identification of species, so that changes in community structure can be
404 monitored (eDNA) (Papakostas et al. 2016; Fröbuis and Funch 2017) (Q35). (2) Genetic and
405 morphological data should be integrated to advance phylogenetic studies. Ideally, a suite of genes
406 including highly conserved, variable, and trait-specific genes for genetic studies is needed (Wilke
407 et al. 2020) (Q34).

Box 5. Selected questions in the Methods theme after Phase 2. Highly ranked questions resulting from Phase 3 are highlighted in yellow.

Q34. What steps are required to link molecular sequence data with morphological traits in rotifers? (Gold)

Q46. Which tools or AI-assisted programs can support accurate rotifer species identification, and how can their efficiency be evaluated? (Gold)

Q35. What are the appropriate genetic markers for using eDNA in rotifer detection and monitoring? (Silver)

Q42. What educational tools are most effective for training new researchers in rotifer taxonomy and identification? (Bronze)

Q27. How do species within rotifer species complexes differ genetically and ecologically, and how can integrative methods (e.g., sequencing and habitat-based experiments) clarify their species status?

Q33. How can eDNA tools be used to detect cryptic rotifer diversity?

Q37. What empirical or modeling approaches best estimate rotifer dispersal rates across freshwater habitats, and how do estimates vary by environment type?

Q40. What are the key barriers to accurate rotifer species identification in potentially high diverse regions (e.g., Australia and the Neotropics), and how can they be overcome through targeted training or integrative methods?

Q45. What strategies can address the shortage of trained rotifer taxonomists in tropical regions, and how can this gap be sustainably filled?

Q44. Which PCR primers are most effective for amplifying rotifer DNA, and how do they perform across clades?

Q60. What are the molecular and physiological mechanisms underlying transgenerational plasticity in sexual reproduction in monogonont rotifers?

Q69. How can integrative approaches combining taxonomy, molecular tools, and ecological data accelerate rotifer biodiversity research?

Q72. How can a digital platform be built to compile and update historical literature on rotifers, including rare publications?

Q76. What are the genomic consequences of DNA damage and repair during anhydrobiosis in bdelloid rotifers?

Q88. How can standardized quantification of rotifer functional traits improve detection of anthropogenic impacts in small water bodies?

Q95. What mechanisms maintain the stable coexistence of multiple rotifer species in shared aquatic environments?

409

410 *Populations*

411

412 Using rotifer population dynamics to infer abiotic and biological drivers remains a core task in
413 ecological and evolutionary research (Gilbert 1988; Lemmen et al. 2022; Réveillon and Becks
414 2024). Recent studies show that rapid evolution and phenotypic plasticity can feed back on
415 population dynamics even across short environmental gradients (Tarazona et al. 2019; Ramos-
416 Rodríguez et al. 2020). This is especially clear in traits tied to persistence (e.g., resting egg banks,
417 parthenogenesis, and dormancy timing), where small changes in hydroperiod or temperature can

418 alter cohort structure and long-term resilience. How environmental variability (e.g., hydroperiod
419 fluctuations) affects evolution of diapause traits in rotifers, and how this influences long-term
420 population persistence, remains a key question (Q53, Box 6). While correlative studies have
421 documented associations between environmental variables and community composition,
422 mechanistic understanding of such causal pathways driving population responses remains limited.
423 For instance, understanding why bdelloid rotifers give birth to live young, and what adaptive
424 advantages this confers in Antarctic environments (Q07), highlights how life-history traits may be
425 causally linked to environmental constraints. This shift toward causal frameworks is critical
426 because our ability to predict how rotifer assemblages will respond to global warming based on
427 their diversity and distribution (Q24) will determine our capacity to forecast ecosystem functioning
428 under novel conditions. Likewise, assessing whether geographically separated populations of the
429 same species exhibit niche conservatism across environmental gradients (Q09) is essential for
430 predicting their responses to environmental change. Such understanding is also important to
431 evaluate the impact of invasive species on different local communities (Haubrock et al. 2024).
432 Numerous studies on rotifers over almost a century have generated sufficient knowledge to apply
433 them in fields such as aquaculture and ecotoxicology. However, basic research on their
434 demography remains restricted to a few taxa such as *Brachionus calyciflorus* Pallas, 1776, *B.*
435 *plicatilis* Müller, 1786, *B. rubens* Ehrenberg, 1838, and *Platyonus patulus* (Müller 1786) (Lemmen
436 et al. 2022; Réveillon and Becks 2024). As a consequence, these species have become
437 indispensable as food for larval stages in aquaculture or as bioassay organisms, while the potential
438 of many other rotifer taxa, especially littoral species, has largely been neglected (e.g., in bioassays
439 of toxicants that tend to sink to the bottom of ponds, lakes, and reservoirs).
440

Box 6. Selected questions in the Populations theme after Phase 2. Highly ranked questions resulting from Phase 3 are highlighted in blue.

Q53. How does environmental variability (e.g. hydroperiod fluctuations) affect the evolution of diapause traits in rotifers, and how does this influence long-term population persistence?

(Gold)

Q09. Do geographically separated populations of the same rotifer species exhibit niche conservatism across environmental gradients? (Silver)

Q24. How well can rotifer community responses to global warming be predicted from diversity and distribution? (Silver)

Q07. Why do bdelloid rotifers give birth to live young, and what adaptive advantages does this confer in Antarctic environments? (Bronze)

Q10. What is the influence of climate change and new environmental conditions on the current distribution and abundance of rotifer species?

Q11. How have historical and evolutionary processes shaped the geographic distribution patterns of rotifers?

Q28. What is the extent and distribution of cryptic speciation across the rotifer phylum, and how can it be quantified?

Q38. How does dispersal of rotifers among connected freshwater habitats influence nutrient cycling and productivity in metacommunity frameworks?

Q52. Is population differentiation in rotifers driven more by isolation by distance or by environmental differences?

Q59. Which environmental cues or stressors trigger male production in rotifers, and how do these vary across habitats and taxa?

441

442 **4.2 Knowledge gaps and a hypothesis-driven future**

443

444 Despite substantial advances, rotifer research faces persistent gaps, including a lack of reliable

445 phylogenies and the challenge of translating high-priority questions into testable hypotheses. Our

446 findings, coupled with the uneven international engagement observed in our network (Fig. 2),

447 reflect demographic biases common in biodiversity research (Tydecks et al. 2018) and reveal

448 varying levels of consensus across thematic areas, with more divergent views in *Evolution and*

449 *Ecology* compared to *Global Change*. To address these limitations and align the field with global

450 scientific challenges, we propose a strategic framework focused on collaboration, funding, and

451 dissemination between triennial International Rotifer Symposia. Key initiatives include

452 establishing thematic working groups, seeking targeted funding for taxonomist training, and

453 fostering continuous engagement through digital platforms and researcher exchange programs.

454 These coordinated efforts aim to advance the field cohesively, ensuring that fundamental questions

455 are addressed effectively while enhancing the relevance of rotifers in ecology and conservation.

456

457 One of the most significant challenges in rotifer research pertains to fundamental questions about

458 their diversity and distribution. For instance, the persistently low number of rotifer species

459 recorded in Latin America raises questions about underlying factors such as sampling bias and

460 taxonomic effort (Fontaneto et al. 2012). This points to a broader issue of understanding global

461 rotifer biodiversity, especially when considering the pervasive nature of cryptic diversity within

462 rotifers and the potential for many species to remain undiscovered globally. The findings of this
463 study contribute to illuminating these gaps, and planning scenarios for continuous taxonomic effort
464 in multiple geographic regions, habitats and microhabitats. Implementing modern techniques will
465 be essential for achieving a more accurate assessment of the full biodiversity of Rotifera.

466

467 A core objective of this initiative was to move beyond descriptive research towards a more
468 hypothesis-driven framework. By clearly articulating fundamental questions, this study lays the
469 groundwork for generating testable hypotheses. For instance, questions about the mechanisms
470 driving speciation in bdelloid rotifers (Q32/Gold) or the genetic basis of adaptation to salinity
471 changes (Q36/Gold) directly invite the formulation and empirical testing of specific hypotheses.
472 Similarly, investigating how multiple environmental stressors affect rotifer development
473 (Q61/Silver) and community structure (Q18/Gold) requires the development of predictive
474 hypotheses that can be empirically tested. This shift is crucial for transitioning rotifer biology from
475 its foundational focus on observation and description toward a more mechanistic understanding
476 and greater predictive power. Rotifers have the potential to serve as bioindicators of ecosystem
477 health and can actively improve water quality. Recent studies (Davis et al. 2015; Pajdak-Stós et
478 al. 2020) have demonstrated that certain rotifer taxa can significantly influence the composition of
479 algal communities, including the suppression of harmful cyanobacterial blooms and toxin-
480 producing species such as *Prymnesium parvum*. Rapid reproduction, extensive grazing and
481 tolerance of changing environmental conditions make rotifers promising organisms for
482 biomanipulation and ecological restoration strategies. This shift redefines rotifer research,
483 expanding its focus beyond traditional monitoring and bioindicator roles to include the protection
484 of aquatic ecosystem functioning and resilience.

485

486 **4.3 Philosophical dimensions**

487

488 Our analysis of the 67 expert-selected questions (>50% consensus) showed that rotifer research is
489 not merely a technical pursuit but is built upon fundamental conceptual and philosophical
490 foundations. Following the framework by Heger et al. (2024), we identified several questions that
491 challenge how we define, know, and value biological data.

492

493 Ontological questions, those concerning the nature of the entities we study, are embedded in the
494 very definition of a “rotifer community” (Q26, Q97). Because rotifer studies often lack
495 standardized community boundaries (Sládeček 1983; Fontaneto et al. 2007), these questions ask
496 us to determine what actually constitutes a biological unit in space and time. Similarly, Q22 (“Does
497 global rotifer biodiversity reflect indicators of degradation?”) questions whether our chosen
498 “indicator species” are true proxies for the broader reality of biodiversity or merely convenient
499 human constructs.

500

501 Epistemological challenges, concerning the limits and methods of our knowledge, emerge when
502 we attempt to bridge different scales of evidence. For instance, Q25 (“To what extent does
503 community structure reflect short-term versus long-term climate-induced changes?”) addresses the
504 problem of temporal inference, questioning how we distinguish transient fluctuations from
505 fundamental shifts in ecological knowledge. Similarly, Q34 (“What steps are required to link
506 molecular sequence data with morphological traits in rotifers?”); linking molecular data with

507 morphology is not just a technical hurdle but a debate over methodological pluralism, asking how
508 we reconcile genetic processes with the higher-level physical traits they produce.

509
510 Furthermore, these priorities carry axiological and ethical weight. The tension between AI-assisted
511 identification emerged from Q46 (“Which tools or AI-assisted programs can support accurate
512 rotifer species identification?”) and intergenerational expertise from Q71 (“What strategies can
513 sustain intergenerational transfer of taxonomic expertise in rotifer research?”), reflecting a deep
514 shift in scientific authority. These questions ask whether taxonomic knowledge is a formalizable
515 algorithm competency that requires human mentorship to persist. At the same time, an increasing
516 reliance on AI reshapes whose expertise is valued, how authority is distributed, and what levels of
517 transparency are acceptable in ecological practice. While AI may expand data-processing capacity,
518 it also alters standards of evidence, accountability, and trust (Macêdo et al. 2023; Sugeno et al.,
519 2026; Lu et al. 2025). For instance, Sugeno et al. (2025) benchmarked Large Language Models
520 (LLMs) against the expert-labeled zooplankton dataset from Macêdo et al. (2023), finding that
521 while AI achieved 84% accuracy, human-in-the-loop oversight remains necessary for culturally
522 nuanced data. Last but not least, ethical and value-related aspects are equally evident in our
523 conservation language, as seen in Q100 (“How does rotifer abundance and diversity change in
524 response to invasive species introductions?”). Here, the choice of the term “invasive” rather than
525 “non-native” carries immediate management implications (Soto et al. 2024), demonstrating that
526 the language used to prioritize rotifer research is not value-neutral.

527

528

529 **4.4 Model organisms**

530
531 The 67 high-priority questions did not usually target individual species, but rather broader
532 taxonomic groups. Only *Brachionus* Pallas, 1776, specifically *B. calyciflorus*, appeared in two
533 questions focused on how environmental changes affect expression of plastic traits within species
534 complexes (Q50 and Q74). This reflects a critical research direction that leverages well-studied,
535 phenotypically flexible species to understand adaptation in the face of climate change and
536 environmental variability. Bdelloidea (mentioned eight times) and Monogononta (three times) also
537 highlight pervasive interest in these groups. Interest in bdelloid rotifers stems from their extreme
538 physiological tolerance, and the need to explain unique traits such as DNA repair during
539 anhydrobiosis, the high incidence of horizontal gene transfer, and their responses to environmental
540 stress and contaminants. Questions on monogonont rotifers, by contrast, often focused on resolving
541 cryptic diversity and speciation, and on the molecular and physiological bases of transgenerational
542 plasticity in sexual reproduction. Broader groups within Rotifera (see Sørensen and Giribet, 2006),
543 including acanthocephalans and seisonids, were not specifically mentioned in any of the priority
544 questions. Although bdelloid rotifers were specifically mentioned more frequently, many of the
545 questions were only relevant to monogonont rotifers and not the broader phylum (e.g. resting eggs
546 - Q55, Q56, Q57; male production - Q59; diapause - Q53, Q66).

547

548 **4.5 De-colonizing Rotifera research**

549 Reliance on expert elicitation can introduce geographical and disciplinary biases, particularly
550 when certain regions or research traditions are over-represented. Amplifying voices from
551 historically underrepresented regions will strengthen the field and foster a more just, inclusive, and
552 globally relevant vision for Rotifera research.

553
554 As in other research areas where persistent global inequities have been documented (e.g. Tydecks
555 et al. 2018; Jiang et al. 2025), our Delphi process also reflected imbalances. Experts working in
556 freshwater ecosystems and researchers based in Europe and North America constituted the
557 majority of contributors. Notably, no experts from the African continent participated, potentially
558 reinforcing pre-existing geographic blind spots. At the same time, the workshop for Phase 3 of the
559 Delphi process was held during the 17th International Rotifer Symposium (IRS), in Rio de Janeiro,
560 the first time the IRS was hosted in the southern hemisphere. This location enabled unprecedented
561 participation from researchers in South America, Central America, and the Caribbean, reducing
562 common barriers to attendance and promoting greater regional representation in the consensus
563 process. Such expanded participation is important because some research priorities—especially
564 those concerning under-studied regions—may be overlooked when shaped predominantly by
565 Global North perspectives. For instance, questions on rotifer diversity in Latin America (e.g.,
566 Q08), where sampling bias and limited taxonomic capacity strongly influence recorded species
567 richness (López et al. 2025), might not surface as priorities without meaningful regional insight.
568 By encouraging readers to examine the full list of 100 questions (Table S1), we aim to draw
569 attention to topics that may have received fewer votes not because of low relevance, but because
570 of limitations in participation, framing, or disciplinary familiarity.

571 Our study also revealed a gender imbalance among contributors in both Phases 1 and 2, with men
572 representing a higher proportion of respondents (16% difference; Fig. 2), whereas gender parity
573 was achieved during the in-person deliberation of Phase 3. The male bias in early phases likely
574 reflects the predominantly male demographics at the education and career stages represented. This
575 pattern mirrors well-documented structural inequities in academia, where women –although

576 dominating student levels— remain underrepresented in senior and decision-making positions (Ceci
577 et al. 2014; Wellenreuther and Otto 2016; Débarre et al. 2018; Salerno et al. 2019). Achieving
578 parity in Phase 3 contrasted with typical trends and may relate to the fact that the 17th International
579 Rotifer Symposium was organized by a woman, as the gender of organizers and senior authors
580 strongly predicts participant gender balance (Débarre et al. 2018; Salerno et al. 2019). Such
581 disparities are not only a matter of fairness; the composition of contributors can shape research
582 agendas and determine whose perspectives influence scientific interpretation (Débarre et al. 2018;
583 Salerno et al. 2019; Downes and Lancaster, 2020). While the potential influence of the observed
584 male bias on Rotifera research priorities was not assessed here, it may be an interesting
585 consideration for future studies. More broadly, identity-based exclusion—including gender,
586 LGBTQIA+ identity, or socio-economic constraints—can reduce both creativity and productivity
587 in science and is exacerbated by barriers such as event affordability and accessibility (Tulloch
588 2020). Although our questionnaire captured only three gender options, acknowledging these biases
589 and monitoring representation helps support more inclusive and diverse research agendas. We
590 recognize that equity is only one dimension of diversity, but addressing it is a meaningful step
591 toward broader inclusion in scientific knowledge production (Débarre et al. 2018; Tulloch 2020).

592 **4.6 Conservation efforts**

593 Small waterbodies and shallow lakes often carry high species richness (Smolak and Walsh 2022),
594 yet many of them are highly vulnerable to human impacts. Some of them are left to dry and
595 converted for agriculture, real-estate development, or other human land uses. Larger waterbodies
596 are not exempt from such pressures, and many waterbodies host endemic species and type
597 localities (Kuczyńska-Kippen et al. 2025). Their loss results in a critical loss of biodiversity – in
598 many cases biodiversity that was gone before it could have been detected and described. Such

599 cases also cause serious gaps in our understanding of natural taxonomic diversity. Poorly studied
600 systems, for example in South America, exacerbate this knowledge deficit in rotifer diversity
601 (Q08), and reliance on extraterritorial material may also lead to underestimation of the true species
602 richness within a given waterbody (Q42).

603 Conservation efforts targeting waterbodies with high species richness, high endemic taxa, or type
604 localities should involve public engagement and local governmental support (Q87). Rotifer
605 research is often hidden in limnological studies, meaning that national and international
606 conferences may carry valuable information on unique freshwater habitats that may go unnoticed.
607 Therefore, systematically compiling available data from such sources could substantially enhance
608 our understanding of rotifer diversity and support the preservation of waterbodies that harbour
609 exceptional biological value.

610

611 **Conclusion**

612

613 By leveraging a comprehensive Delphi process, our study has synthesized the collective expertise
614 of the international rotifer community to identify and prioritize fundamental questions guiding
615 rotifer research. While not intended as an exhaustive list, it represents a collective snapshot of the
616 field's current priorities and emerging directions, offering a curated roadmap for students,
617 researchers, funding agencies, and environmental policy. Our aim was to stimulate new
618 collaborations and interdisciplinary research, fostering the formulation and testing of new
619 hypotheses, and promoting integration of rotifer studies into broader ecological and evolutionary
620 contexts.

621 **Author Contributions**

622 Conceptualization: Rafael L. Macêdo, Gissell Lacerot and Jonathan M. Jeschke. Investigation,
623 methodology, validation: Rafael L. Macêdo, Gissell Lacerot and Jonathan M. Jeschke. Data
624 curation: Rafael L. Macêdo and Gissell Lacerot. Formal analysis: Rafael L. Macêdo. Writing—
625 original draft: Rafael L. Macêdo. Visualization: Rafael L. Macêdo, Gissell Lacerot, Carlota
626 Solano-Udina, Melanie D. Borup, Marco Antonio Jiménez-Santos, Diego Fontaneto. Writing—
627 review and editing: All authors wrote sections of the Results and Discussion and critically
628 reviewed and edited the entire manuscript.

629

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638

639 **Conflict of Interest**

640 None declared.

641

642 **Data Availability Statement**

643 The data that supports the findings of this study are available in the supplementary material of
644 this article.

645 **References**

646 Arcifa, M. S., B. B. de Souza, C. S. de Morais-Junior et al. 2020. “Functional Groups of Rotifers and
647 an Exotic Species in a Tropical Shallow Lake.” *Scientific Reports* 10: 14698.
648 <https://doi.org/10.1038/s41598-020-71778-1>.

649 Arndt, H. 1993. “Rotifers as Predators on Components of the Microbial Web (Bacteria, Heterotrophic
650 Flagellates, Ciliates)—A Review.” *Hydrobiologia* 255: 231–246.
651 <https://doi.org/10.1007/BF00025844>.

652 Balkić, A. G., I. Ternjej, and M. Špoljar. 2017. “Hydrology Driven Changes in the Rotifer Trophic
653 Structure and Implications for Food Web Interactions.” *Ecohydrology* 11: e1917.
654 <https://doi.org/10.1002/eco.1917>.

655 Bomfim, F. F., S. Deosti, N. Louback-Franco, R. L. M. Sousa, and T. S. Michelin. 2023. “How Are
656 Zooplankton's Functional Guilds Influenced by Land Use in Amazon Streams?” *PLoS ONE* 18:
657 e0288385. <https://doi.org/10.1371/journal.pone.0288385>.

- 658 Branco, C. W. C., E. Fintelman-Oliveira, and V. B. dos Santos Miranda. 2023. “A Review of
659 Functional Approaches for the Study of Freshwater Communities With a Focus on Zooplankton.”
660 *Hydrobiologia* 850: 4719–4744. <https://doi.org/10.1007/s10750-023-05227-1>.
- 661 Branco, C. W. C., L. C. Santos-Cabral, B. Kozlowsky-Suzuki, V. G. Lopes, A. L. Puga, and R. L.
662 Macêdo. 2024. “Persistence of the Non-Native *Kellicottia bostoniensis* (Rousselet, 1908) in a
663 Large Tropical Reservoir.” *Hydrobiologia* 851: 3039–3060. <https://doi.org/10.1007/s10750-023-05295-3>.
664
- 665 Ceci, S. J., D. K. Ginther, S. Kahn, and W. M. Williams. 2014. “Women in Academic Science: A
666 Changing Landscape.” *Psychological Science in the Public Interest* 15: 75–141.
667 <https://doi.org/10.1177/1529100614541236>.
- 668 Davis, S. L., D. L. Roelke, B. W. Brooks, V. M. Lundgren, F. Withrow, and W. C. Scott. 2015.
669 “Rotifer–*Prymnesium parvum* Interactions: Role of Lake Bloom History on Rotifer Adaptation
670 to Toxins Produced by *P. parvum*.” *Aquatic Microbial Ecology* 75: 55–68.
671 <https://doi.org/10.3354/ame01748>.
- 672 Débarre, F., N. O. Rode, and L. V. Ugelvig. 2018. “Gender Equity at Scientific Events.” *Evolution*
673 *Letters* 2: 148–158. <https://doi.org/10.1002/evl3.49>.
- 674 DeCasien, A. R., S. A. Williams, and J. P. Higham. 2017. “Primate Brain Size Is Predicted by Diet
675 but Not Sociality.” *Nature Ecology & Evolution* 1: 0112. <https://doi.org/10.1038/s41559-017-0112-0112>.
676
- 677 Declerck, S. A. J., and S. Papakostas. 2017. “Monogonont Rotifers as Model Systems for the Study
678 of Micro-Evolutionary Adaptation and Its Eco-Evolutionary Implications.” *Hydrobiologia* 796:
679 131–144. <https://doi.org/10.1007/s10750-016-2782-y>.
- 680 Dey, C. J., A. I. Rego, J. D. Midwood, and M. A. Koops. 2020. “A Review and Meta-Analysis of
681 Collaborative Research Prioritization Studies in Ecology, Biodiversity Conservation and
682 Environmental Science.” *Proceedings of the Royal Society B* 287: 20200012.
683 <https://doi.org/10.1098/rspb.2020.0012>.
- 684 Diouf, A., L. Bereziat, D. Nodier, M. Amaral, S. Haliyo, and A. Mannioui. 2026. “Rotiferometer: An
685 Automated System for Quantification of Rotifer Cultures.” *Aquacultural Engineering* 112:
686 102662. <https://doi.org/10.1016/j.aquaeng.2025.102662>.
- 687 Downes, B. J., and J. Lancaster. 2020. “Celebrating Women Conducting Research in Freshwater
688 Ecology and How the Citation Game Is Damaging Them.” *Marine and Freshwater Research* 71:
689 139–155. <https://doi.org/10.1071/MF18436>.
- 690 Dumont, H. J. 1983. “Biogeography of Rotifers.” *Hydrobiologia* 104: 19–30.
691 <https://doi.org/10.1007/BF00045948>.

- 692 Ehrenberg, C. G. 1838. Die Infusionstierchen als vollkommene Organismen: Ein Blick in das tiefere
693 organische Leben der Natur. Leipzig: L. Voss.
- 694 Ejsmont-Karabin, J. 2012. “The Usefulness of Zooplankton as Lake Ecosystem Indicators: Rotifer
695 Trophic State Index.” *Polish Journal of Ecology* 60: 339–350.
- 696 Enders, M., F. Havemann, F. Ruland, M. Bernard-Verdier, J. A. Catford, L. Gómez-Aparicio, S.
697 Haider, T. Heger, C. Kueffer, I. Kühn et al. 2020. “A Conceptual Map of Invasion Biology:
698 Integrating Hypotheses Into a Consensus Network.” *Global Ecology and Biogeography* 29: 978–
699 991. <https://doi.org/10.1111/geb.13082>.
- 700 Fontaneto, D., W. H. De Smet, and C. Ricci. 2006. “Rotifers in Saltwater Environments: Re-
701 Evaluation of an Inconspicuous Taxon.” *Journal of the Marine Biological Association of the*
702 *United Kingdom* 86: 623–656. <https://doi.org/10.1017/S0025315406013531>.
- 703 Fontaneto, D., E. A. Herniou, C. Boschetti, M. Caprioli, G. Melone, C. Ricci, and T. G. Barraclough.
704 2007. “Independently Evolving Species in Asexual Bdelloid Rotifers.” *PLoS Biology* 5: e87.
705 <https://doi.org/10.1371/journal.pbio.0050087>.
- 706 Fontaneto, D., A. M. Barbosa, H. Segers, and M. Pautasso. 2012. “The ‘Rotiferologist’ Effect and
707 Other Global Correlates of Species Richness in Monogonont Rotifers.” *Ecography* 35: 174–182.
708 <https://doi.org/10.1111/j.1600-0587.2011.06850.x>.
- 709 Fröblius, A. C., and P. Funch. 2017. “Rotiferan Hox Genes Give New Insights into the Evolution of
710 Metazoan Body Plans.” *Nature Communications* 8: 9. <https://doi.org/10.1038/s41467-017-00020-w>.
- 712 Gansfort, B., D. Fontaneto, and M. Zhai. 2020. “Meiofauna as a Model to Test Paradigms of
713 Ecological Metacommunity Theory.” *Hydrobiologia* 847: 2645–2663.
714 <https://doi.org/10.1007/s10750-020-04185-2>.
- 715 Garamszegi, L. Z. 2014. *Modern Phylogenetic Comparative Methods and Their Application in*
716 *Evolutionary Biology*. Berlin: Springer. <https://doi.org/10.1007/978-3-662-43550-2>.
- 717 Gilbert, J. J. 1988. “Suppression of Rotifer Populations by Daphnia: A Review of the Evidence, the
718 Mechanisms, and the Effects on Zooplankton Community Structure.” *Limnology and*
719 *Oceanography* 33: 1286–1303. <https://doi.org/10.4319/lo.1988.33.6.1286>.
- 720 Gilbert, J. J. 2022. “Food Niches of Planktonic Rotifers: Diversification and Implications.” *Limnology*
721 *and Oceanography* 67: 2218–2251. <https://doi.org/10.1002/lno.12199>.
- 722 Gómez, A. 2005. “Molecular Ecology of Rotifers: From Population Differentiation to Speciation.”
723 *Hydrobiologia* 546: 83–99. <https://doi.org/10.1007/s10750-005-4104-7>.
- 724 Han, S., P. J. Van den Brink, and S. A. J. Declerck. 2025. “Asymmetric Micro-Evolutionary
725 Responses in a Warming World: Heat-Driven Adaptation Enhances Metal Tolerance in a

- 726 Planktonic Rotifer, but Not Vice Versa.” *Global Change Biology* 31: e70347.
727 <https://doi.org/10.1111/gcb.70347>.
- 728 Haubrock, P. J., I. Soto, D. A. Ahmed, A. R. Ansari, A. S. Tarkan, I. Kurtul, R. L. Macêdo et al. 2024.
729 “Biological Invasions Are a Population-Level Rather than a Species-Level Phenomenon.” *Global*
730 *Change Biology* 30: e17312. <https://doi.org/10.1111/gcb.17312>.
- 731 Hébert, M.-P., B. E. Beisner, and R. Maranger. 2017. “Linking Zooplankton Communities to
732 Ecosystem Functioning: Toward an Effect-Trait Framework.” *Journal of Plankton Research* 39:
733 3–12. <https://doi.org/10.1093/plankt/fbw068>.
- 734 Heger, T., A. Elliott-Graves, M. I. Kaiser, K. H. Morrow, W. Bausman, G. P. Dietl, C. F. Dormann et
735 al. 2024. “Looking Beyond Popper: How Philosophy Can Be Relevant to Ecology.” *Oikos* 2024:
736 e10994. <https://doi.org/10.1111/oik.10994>.
- 737 Herlyn, H., A. A. Hembrom, J.-P. Tosar, K. M. Mauer, H. Schmidt, B. S. Dezfuli, T. Hankeln et al.
738 2025. “Substantial Hierarchical Reductions of Genetic and Morphological Traits in the Evolution
739 of Rotiferan Parasites.” *Genome Biology and Evolution* 17: evaf124.
740 <https://doi.org/10.1093/gbe/evaf124>.
- 741 Hillbricht-Ilkowska, A. 1995. “One Hundred Years of Polish Rotiferology—Scientists and Their
742 Work.” *Hydrobiologia* 313–314: 1–14. <https://doi.org/10.1007/BF00025925>.
- 743 Hopfe, C., B. Ospina-Jara, T. Schulze, M. Tischer, D. Morales, V. Reinhartz, R. Eshghi Esfahani, C.
744 Valderrama, J. Pérez-Rigueiro, C. Bleidorn, H. Feldhaar, J. Cabra-García, and T. Scheibel. 2024.
745 “Impact of Environmental Factors on Spider Silk Properties.” *Current Biology* 34: 56–67.e5.
746 <https://doi.org/10.1016/j.cub.2023.11.043>.
- 747 Hudson, C. T., and P. H. Gosse. 1886. *The Rotifera or Wheel-Animalcules, Both British and Foreign*.
748 London: Longmans & Green.
- 749 Huynh, T.-H., Z. Horváth, K. Pálffy, V. Kardos, B. Szabó, P. Dobosy, and C. F. Vad. 2024.
750 “Heatwave-Induced Functional Shifts in Zooplankton Communities Result in Weaker Top-Down
751 Control on Phytoplankton.” *Ecology and Evolution* 14: e70096.
752 <https://doi.org/10.1002/ece3.70096>.
- 753 Ienaga, N., T. Takashi, H. Tamamizu, and K. Terayama. 2024. “Rotifer Detection and Tracking
754 Framework Using Deep Learning for Automatic Culture Systems.” *Smart Agricultural*
755 *Technology* 9: 100577. <https://doi.org/10.1016/j.atech.2024.100577>.
- 756 Jiang, Q., Y. Sun, E. Jeppesen, J. P. Smol, D. Scavia, R. E. Hecky, T. Mehner, Y. Qin, Y. Tong, B.
757 Qin, K. D. Hambright, X. Jin, J. Li, K. Cai, Z. Wu, and Y. Liu. 2025. “Persistent Inequities in
758 Global Lake Science.” *Nature Reviews Earth & Environment* 6: 629–631.
759 <https://doi.org/10.1038/s43017-025-00722-6>.

760 Lemmen, K. D., L. Zhou, S. Papakostas and S. A. J. Declerck, 2022. “An Experimental Test of The
761 Growth Rate Hypothesis as a Predictive Framework for Microevolutionary Adaptation”. *Ecology*
762 103: e3853. <https://doi.org/10.1002/ecy.3853>.

763 Martínez, A., S. Bonaglia, M. Di Domenico, G. Fonseca, J. Ingels, K. M. Jörger, C. Laumer, F. Leasi,
764 D. Zeppilli, E. Baldrighi, H. Bik, D. Cepeda, M. Curini-Galletti, A. D. Cutter, G. dos Santos, S.
765 Fattorini, D. Frisch, S. Gollner, U. Jondelius, A. Kerbl, K. M. Kocot, N. Majdi, S. Mammola, J.
766 M. Martín-Durán, A. Menegotto, P. A. Montagna, F. J. A. Nascimento, N. Puillandre, A.
767 Rognant, N. Sánchez, I. R. Santos, A. Schmidt-Rhaesa, M. Schratzberger, F. Semprucci, M.
768 Shimabukuro, P. J. Sommerfield, T. H. Struck, M. V. Sørensen, A. Wallberg, K. Worsaae, H.
769 Yamasaki, and D. Fontaneto. 2025. “Fundamental Questions in Meiofauna Research Highlight
770 How Small but Ubiquitous Animals Can Improve Our Understanding of Nature.”
771 *Communications Biology* 8: 449. <https://doi.org/10.1038/s42003-025-07888-1>.

772 May, L., and R. L. Wallace. 2019. “An Examination of Long-Term Ecological Studies of Rotifers:
773 Comparability of Methods and Results, Insights Into Drivers of Change and Future Research
774 Challenges.” *Hydrobiologia* 844: 129–147. <https://doi.org/10.1007/s10750-019-04059-2>.

775 Melone, G., C. Ricci, H. Segers, and R. L. Wallace. 1998. “Phylogenetic Relationships of Phylum
776 Rotifera With Emphasis on the Families of Bdelloidea.” *Hydrobiologia* 387: 101–107.
777 <https://doi.org/10.1023/A:1017057619574>.

778 Mohl, J. E., P. D. Brown, A. J. Robbins, P. Lavretsky, R. Hochberg, R. L. Wallace, and E. J. Walsh.
779 2025. “Comparing Small and Large Genomes Within Monogonont Rotifers.” *Genome Biology*
780 and *Evolution* 17: evaf041. <https://doi.org/10.1093/gbe/evaf041>.

781 Molinero, J. C., O. Anneville, S. Souissi, G. Balvay, and D. Gerdeaux. 2006. “Anthropogenic and
782 Climate Forcing on the Long-Term Changes of Planktonic Rotifers in Lake Geneva, Europe.”
783 *Journal of Plankton Research* 28: 287–296. <https://doi.org/10.1093/plankt/fbi110>.

784 Mukherjee, N., J. Hugé, W. J. Sutherland, J. McNeill, M. Van Opstal, F. Dahdouh-Guebas, and N.
785 Koedam. 2015. “The Delphi Technique in Ecology and Biological Conservation: Applications
786 and Guidelines.” *Methods in Ecology and Evolution* 6: 1097–1109.
787 <https://doi.org/10.1111/2041-210X.12387>.

788 Mukhopadhyay, S. K., B. Chattopadhyay, A. R. Goswami, and A. Chatterjee. 2007. “Spatial
789 Variations in Zooplankton Diversity in Waters Contaminated With Composite Effluents.”
790 *Journal of Limnology* 66: 97–106. <https://doi.org/10.4081/jlimnol.2007.97>.

791 Obertegger, U., H. A. Smith, G. Flaim, and R. L. Wallace. 2011. “Using the Guild Ratio to
792 Characterize Pelagic Rotifer Communities.” *Hydrobiologia* 662: 157–162.

793 Obertegger, U., and G. Flaim. 2015. “Community Assembly of Rotifers Based on Morphological
794 Traits.” *Hydrobiologia* 753: 31–45. <https://doi.org/10.1007/s10750-015-2191-7>.

- 795 Obertegger, U., and R. L. Wallace. 2023. "Trait-Based Research on Rotifera: The Holy Grail or Just
796 Messy?" *Water* 15: 1459. <https://doi.org/10.3390/w15081459>.
- 797 Oesterwind, D., V. Bartolino, J. W. Behrens, M. Erlandsson, A.-B. Florin, C. Henseler, M.
798 Jakubowska-Lehrmann, C. Jaspers, M. Lehtiniemi, R. Naddafi, K. Nadolna-Ałtyn, I. Putnis, F.
799 J. Quirijns, M. Rakowski, L. Rozenfelde, D. Ustups, T. Wandzel, B. Witalis, A. Woźniczka, and
800 P. Thor. 2025. "Disentangling the Potential Effects of Four Non-Indigenous Species on
801 Commercially and Recreationally Used Fish Stocks in the Baltic Sea—A Review." *Biological*
802 *Invasions* 27: 76.
- 803 Oliveira, F. R., F. M. Lansac-Tôha, B. R. Meira, B. T. Segovia, C. Cochak, and L. F. M. Velho. 2019.
804 "Effects of the Exotic Rotifer *Kellicottia bostoniensis* (Rousselet, 1908) on the Microbial Food
805 Web Components." *Aquatic Ecology* 53: 581–594.
- 806 Pajdak-Stós, A., E. Fiałkowska, F. Hajdyła, and W. Fiałkowski. 2023. "The Potential of *Lecane*
807 Rotifers in Microplastics Removal." *Science of the Total Environment* 899: 165662.
808 <https://doi.org/10.1016/j.scitotenv.2023.165662>.
- 809 Pajdak-Stós, A., W. Fiałkowski, and E. Fiałkowska. 2020. "Rotifers Weaken the Efficiency of the
810 Cyanobacterium Defence Against Ciliate Grazers." *FEMS Microbiology Ecology* 96: fiae189.
811 <https://doi.org/10.1093/femsec/fiae189>.
- 812 Papakostas, S., E. Michaloudi, K. Proios, M. Brehm, L. Verhage, J. Rota, C. Peña, G. Stamou, V. L.
813 Pritchard, D. Fontaneto, and S. A. J. Declerck. 2016. "Integrative Taxonomy Recognizes
814 Evolutionary Units Despite Widespread Mitonuclear Discordance: Evidence From a Rotifer
815 Cryptic Species Complex." *Systematic Biology* 65: 508–524.
816 <https://doi.org/10.1093/sysbio/syw016>.
- 817 Partemi, R., N. Debortoli, A. Martínez, L. Kamburska, C. Souffreau, H. Matheve, P. Vantieghem, L.
818 De Meester, K. Van Doninck, T. Merckx, and D. Fontaneto. 2024. "Weak Effect of Urbanization
819 on Bdelloid Rotifers Living in Lichens." *Royal Society Open Science* 11: 231978.
820 <https://doi.org/10.1098/rsos.231978>.
- 821 Ramos-Rodríguez, E., E. Moreno, and J. M. Conde-Porcuna. 2020. "Intraspecific Variation in
822 Sensitivity to Food Availability and Temperature-Induced Phenotypic Plasticity in the Rotifer
823 *Keratella cochlearis*." *Journal of Experimental Biology* 223: jeb209676.
824 <https://doi.org/10.1242/jeb.209676>.
- 825 Réveillon, T., and L. Becks. 2024. "Trade-Offs Between Defense and Competitive Traits in a
826 Planktonic Predator–Prey System." *Ecology* 105: e4456. <https://doi.org/10.1002/ecy.4456>.
- 827 Ricci, C., and C. Boschetti. 2003. "Bdelloid Rotifers as a Model System to Study Developmental
828 Biology in Space." *Advances in Space Biology and Medicine* 9: 25–39.
829 [https://doi.org/10.1016/S1569-2574\(03\)09002-6](https://doi.org/10.1016/S1569-2574(03)09002-6).

- 830 Salerno, P. E., M. Páez-Vacas, J. M. Guayasamin, and J. L. Stynoski. 2019. “Male Principal
831 Investigators (Almost) Don’t Publish With Women in Ecology and Zoology.” *PLoS ONE* 14:
832 e0218598. <https://doi.org/10.1371/journal.pone.0218598>.
- 833 Segers, H. 2007. “Global Diversity of Rotifers (Rotifera) in Freshwater.” In *Freshwater Animal
834 Diversity Assessment*, edited by E. V. Balian, C. Lévêque, H. Segers, and K. Martens, 49–59.
835 *Hydrobiologia* 595. <https://doi.org/10.1007/s10750-007-9003-7>.
- 836 Serra, M., E. M. García-Roger, R. Ortells, and M. J. Carmona. 2019. “Cyclically Parthenogenetic
837 Rotifers and the Theories of Population and Evolutionary Ecology.” *Limnetica* 38: 67–93.
838 <https://doi.org/10.23818/limn.38.13>.
- 839 Sládeček, V. 1983. “Rotifers as Indicators of Water Quality.” *Hydrobiologia* 100: 169–201.
840 <https://doi.org/10.1007/BF00027429>.
- 841 Smolak, R., and E. J. Walsh. 2022. “Rotifer Species Richness in Kenyan Waterbodies: Contributions
842 of Environmental Characteristics.” *Diversity* 14: 583. <https://doi.org/10.3390/d14070583>.
- 843 Sørensen, M. V., and G. Giribet. 2006. “A Modern Approach to Rotiferan Phylogeny: Combining
844 Morphological and Molecular Data.” *Molecular Phylogenetics and Evolution* 40: 585–608.
845 <https://doi.org/10.1016/j.ympev.2006.04.001>.
- 846 Soto, F., M. A. Lopez-Ramirez, I. Jeerapan, B. E. F. de Ávila, R. K. Mishra, X. Lu, I. Chai, C. Chen,
847 D. Kupor, V. Dotson, R. Liang, B. Waters, and J. Wang. 2019. “Rotibot: Use of Rotifers as Self-
848 Propelling Biohybrid Microcleaners.” *Advanced Functional Materials* 29: 1900658.
849 <https://doi.org/10.1002/adfm.201900658>.
- 850 Stoddard, M. C., E. H. Yong, D. Akkaynak, C. Sheard, J. A. Tobias, and L. Mahadevan. 2017. “Avian
851 Egg Shape: Form, Function, and Evolution.” *Science* 356: 1249–1254.
852 <https://doi.org/10.1126/science.aaj1945>.
- 853 Sugeno, H., K. Inoshita, K. Nojiri, and T. Taga. 2026. “Introducing Large Language Models to
854 Human-Based Etymological Classification in Zooplankton.” *Journal of Plankton Research*
855 bioRxiv. <https://doi.org/10.1101/2025.05.08.652882>.
- 856 Sutherland, W. J., E. Fleishman, M. B. Mascia, J. Pretty, and M. A. Rudd. 2011. “Methods for
857 Collaboratively Identifying Research Priorities and Emerging Issues in Science and Policy.”
858 *Methods in Ecology and Evolution* 2: 238–247. [https://doi.org/10.1111/j.2041-
859 210X.2010.00083.x](https://doi.org/10.1111/j.2041-210X.2010.00083.x).
- 860 Sutherland, W. J., R. P. Freckleton, H. C. J. Godfray, S. R. Beissinger, T. Benton, D. D. Cameron, Y.
861 Carmel, D. A. Coomes, T. Coulson, M. C. Emmerson, R. S. Hails, G. C. Hays, D. J. Hodgson,
862 M. J. Hutchings, D. Johnson, J. P. G. Jones, M. J. Keeling, H. Kokko, W. E. Kunin, X. Lambin,
863 O. T. Lewis, Y. Malhi, N. Mieszkowska, E. J. Milner-Gulland, K. Norris, A. B. Phillimore, D.
864 W. Purves, J. M. Reid, D. C. Reuman, K. Thompson, J. M. J. Travis, L. A. Turnbull, D. A.

- 865 Wardle, and T. Wiegand. 2013. "Identification of 100 Fundamental Ecological Questions."
866 *Journal of Ecology* 101: 58–67. <https://doi.org/10.1111/1365-2745.12025>.
- 867 Tarazona, E., C. Hahn, L. Franch-Gras, E. M. García-Roger, M. J. Carmona, and A. Gómez. 2019.
868 "Ecological Genomics of Adaptation to Unpredictability in Experimental Rotifer Populations."
869 *Scientific Reports* 9: 19646. <https://doi.org/10.1038/s41598-019-56100-y>.
- 870 Tulloch, A. I. T. 2020. "Improving Sex and Gender Identity Equity and Inclusion at Conservation and
871 Ecology Conferences." *Nature Ecology & Evolution* 4: 1311–1320.
872 <https://doi.org/10.1038/s41559-020-1255-x>.
- 873 Tydecks, L., J. M. Jeschke, M. Wolf, G. Singer, and K. Tockner. 2018. "Spatial and Topical
874 Imbalances in Biodiversity Research." *PLoS ONE* 13: e0199327.
875 <https://doi.org/10.1371/journal.pone.0199327>.
- 876 Vasilikopoulos, A., H. Herlyn, D. Fontaneto, C. G. Wilson, R. W. Nowell, J.-F. Flot, T. G.
877 Barraclough, and K. Van Doninck. 2024. "Whole-Genome Analyses Converge to Support the
878 Hemirotifera Hypothesis Within Syndermata (Gnathifera)." *Hydrobiologia* 851: 2795–2826.
879 <https://doi.org/10.1007/s10750-023-05451-9>.
- 880 Walczyńska, A., D. Fontaneto, A. Kordbacheh, S. Hamil, M. A. Jimenez-Santos, S. Paraskevopoulou,
881 A. Pociecha, and W. Zhang. 2024. "Niche Differentiation in Rotifer Cryptic Species Complexes:
882 A Review of Environmental Effects." *Hydrobiologia* 851: 2909–2926.
883 <https://doi.org/10.1007/s10750-023-05291-7>.
- 884 Wallace, R. L. 2002. "Rotifers: Exquisite Metazoans." *Integrative and Comparative Biology* 42: 660–
885 667. <https://doi.org/10.1093/icb/42.3.660>.
- 886 Wallace, R. L., T. W. Snell, C. Ricci, and T. Nogrady. 2012. "Rotifera: Volume 1—Biology, Ecology
887 and Systematics (2nd edn)." *Journal of Paleolimnology* 47: 171–172.
888 <https://doi.org/10.1007/s10933-011-9539-4>.
- 889 Wallace, R. L., E. J. Walsh, S. Nandini, and S. S. S. Sarma. 2021. "A Meta-Analysis of Benthic Rotifer
890 Community Structure as a Function of Lake Trophic State." *Aquatic Ecology* 55: 1297–1304.
891 <https://doi.org/10.1007/s10452-020-09825-2>.
- 892 Wallace, R. L., R. Hochberg, and E. J. Walsh. 2024. "The Undiscovered Country: Ten Grand
893 Challenges in Rotifer Biology." *Hydrobiologia* 851: 3225–3248. <https://doi.org/10.1007/s10750-023-05247-x>.
- 894
- 895 Walz, N. 1987. "Comparative Population Dynamics of the Rotifers *Brachionus angularis* and
896 *Keratella cochlearis*." *Hydrobiologia* 147: 209–213. <https://doi.org/10.1007/BF00025744>.
- 897 Wellenreuther, M., and S. Otto. 2016. "Women in Evolution—Highlighting the Changing Face of
898 Evolutionary Biology." *Evolutionary Applications* 9: 3–16. <https://doi.org/10.1111/eva.12343>.

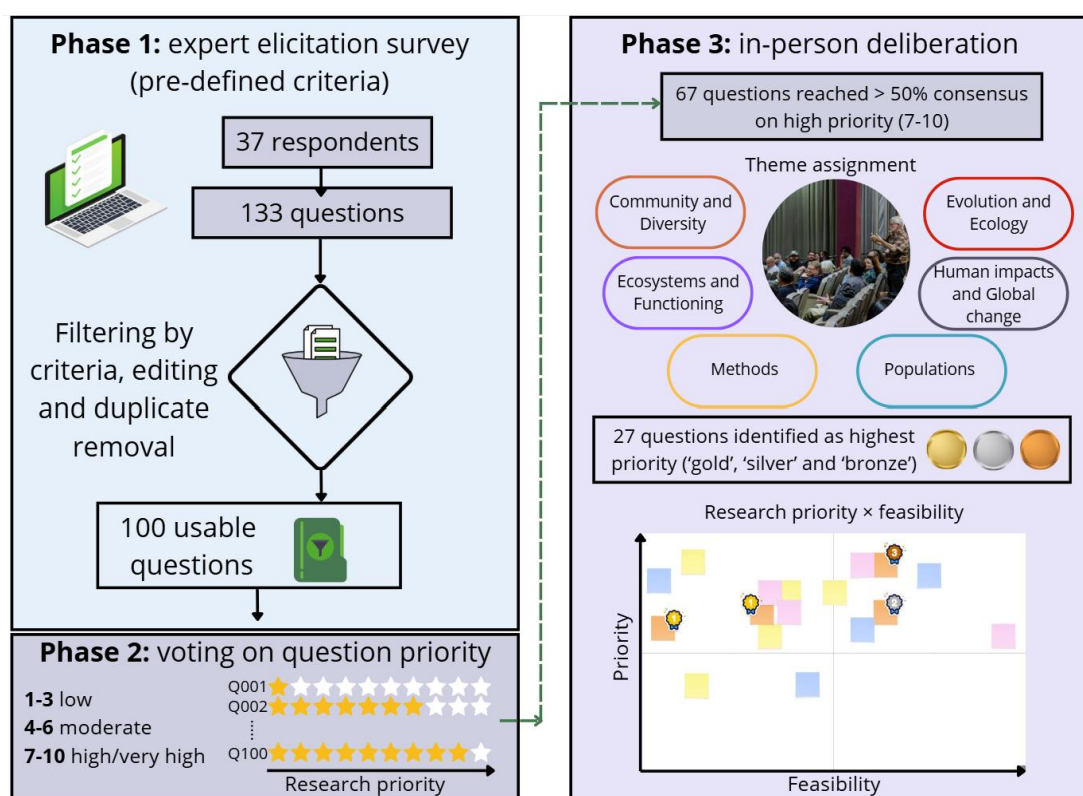
899 Wilke, T., W. H. Ahlrichs, and O. R. P. Bininda-Emonds. 2020. “The Evolution of Synchaetidae
 900 (Rotifera: Monogononta) With a Focus on Synchaeta: An Integrative Approach Combining
 901 Molecular and Morphological Data.” *Journal of Zoological Systematics and Evolutionary*
 902 *Research* 58: 823–857. <https://doi.org/10.1111/jzs.12378>.

903 Zhang, W., H. Bi, D. Wang, X. Cheng, Z. Cai, and K. Ying. 2024. “Automated Zooplankton Size
 904 Measurement Using Deep Learning: Overcoming the Limitations of Traditional Methods.”
 905 *Frontiers in Marine Science* 11: 1341191. <https://doi.org/10.3389/fmars.2024.1341191>.

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907 FIGURES

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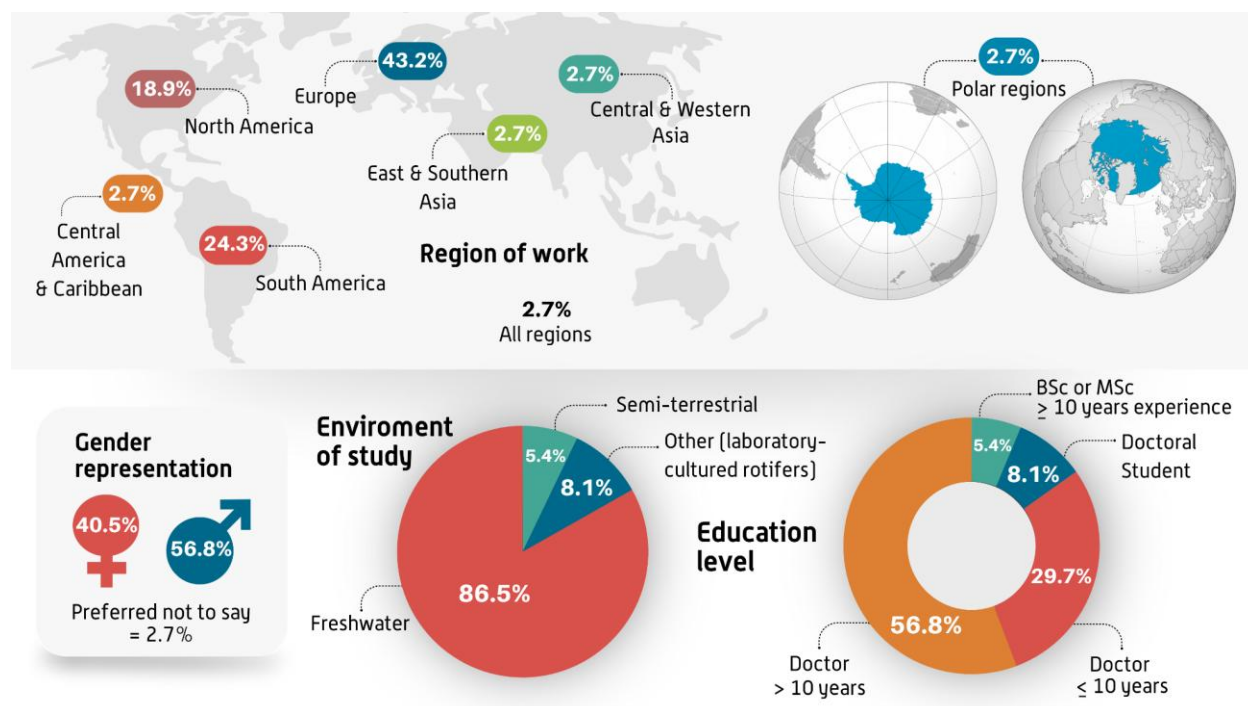
910 **Figure 1.** Workflow of our approach to identify research priorities in Rotifera research. Phase 1

911 involved expert elicitation and filtering of 133 proposed questions into 100 usable ones (see

912 Supplementary Document S1). Phase 2 focused on expert scoring to prioritize the questions, and

913 Phase 3 involved in-person deliberation, theme assignment, and identification of the top 27 high-

914 priority questions.



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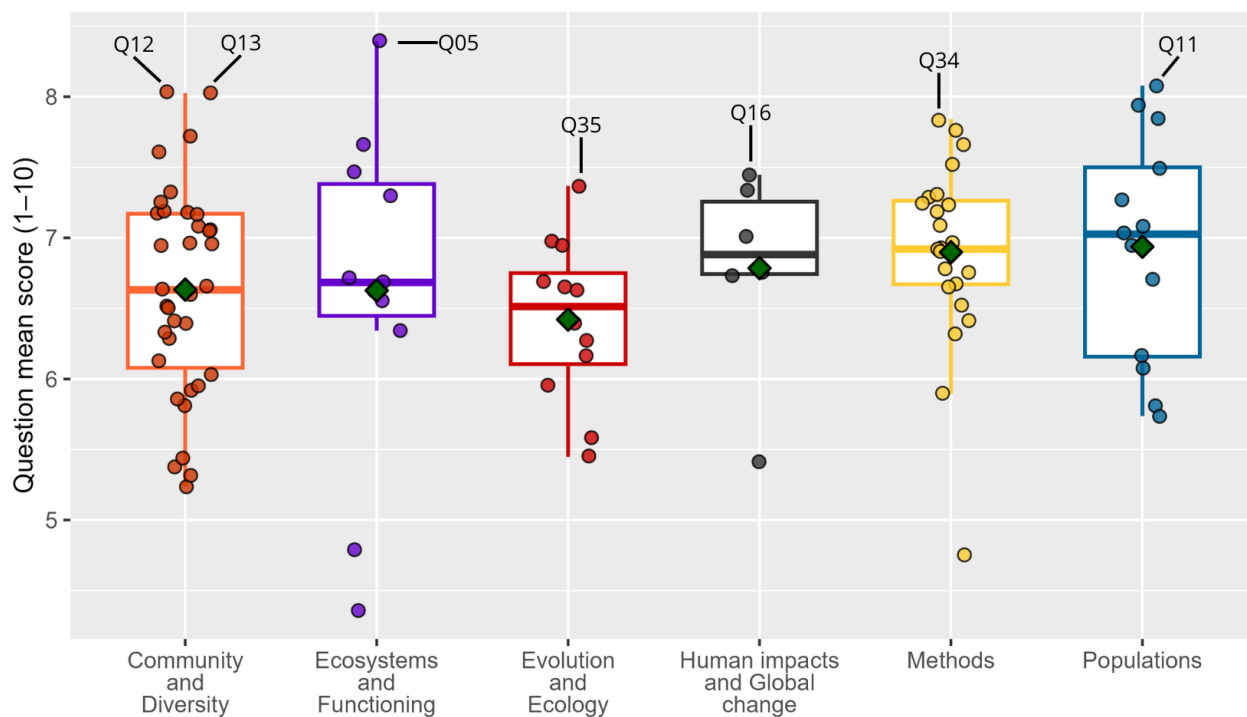
916 **Figure 2.** Demographic composition of the expert panel ($n = 37$) during Phase 1. The map shows
 917 the geographic distribution of experts by region of work, expressed as percentages of total
 918 participants. Accompanying charts present the academic backgrounds, education levels, and
 919 gender representation. Note that “all regions” represents experts whose research focus
 920 encompasses multiple or all geographic regions rather than a single specific territory.

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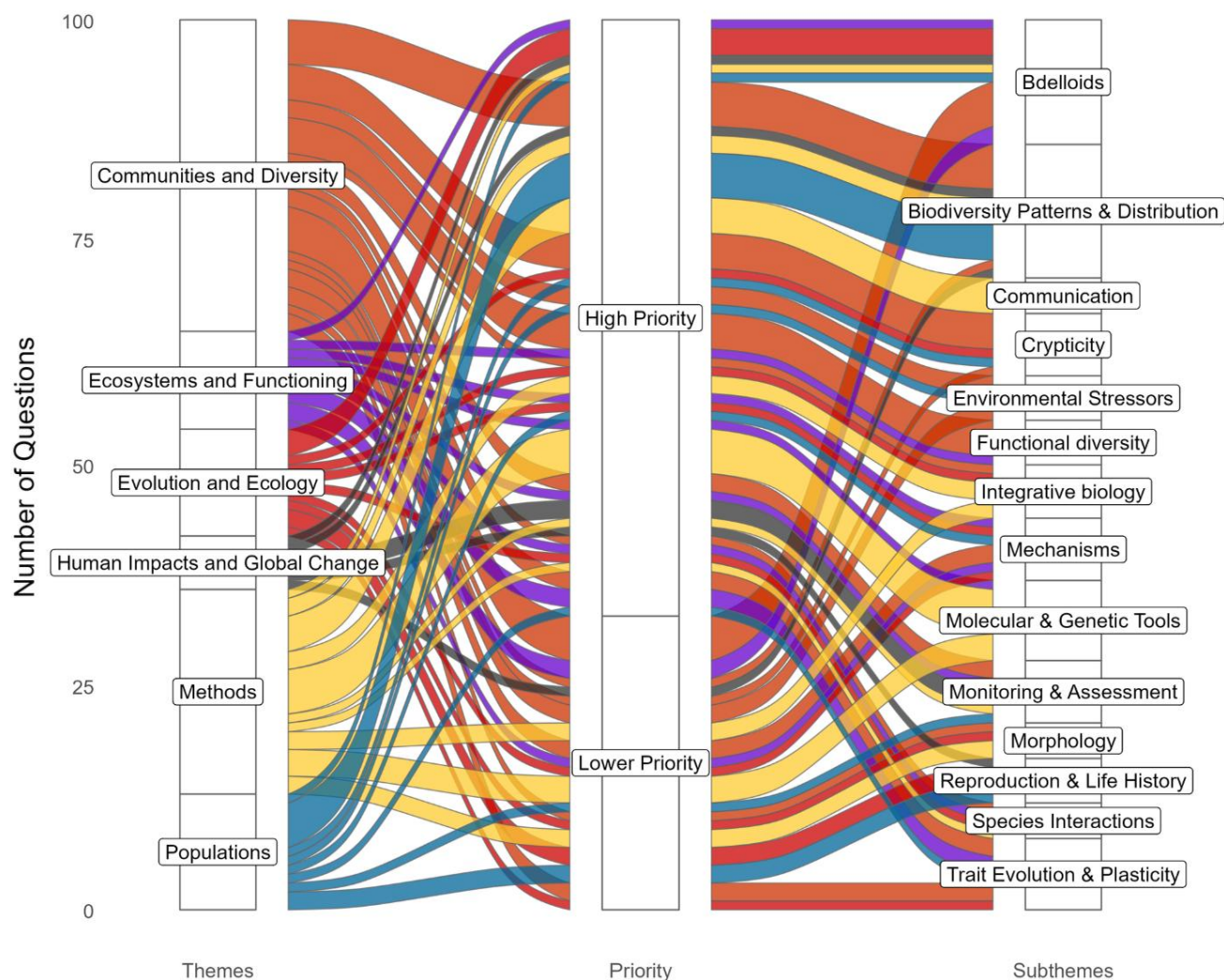
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926 **Figure 3.** Distribution of mean scores for the 100 research questions (Q1–Q100) across research
 927 themes. Each point represents the mean expert score for one question during Phase 2, boxplots
 928 indicate the interquartile range, whiskers represent the maximum values within 1.5 interquartile
 929 ranges, horizontal lines within each box show the median, and diamonds mark the mean values.
 930 This visualization highlights variation in perceived priority across themes. Scores did not differ
 931 significantly among themes (Welch’s ANOVA, $p = 0.35$). The highest-scoring questions within

932 each thematic group are labeled by ID; multiple labels are shown in the event of tied scores.



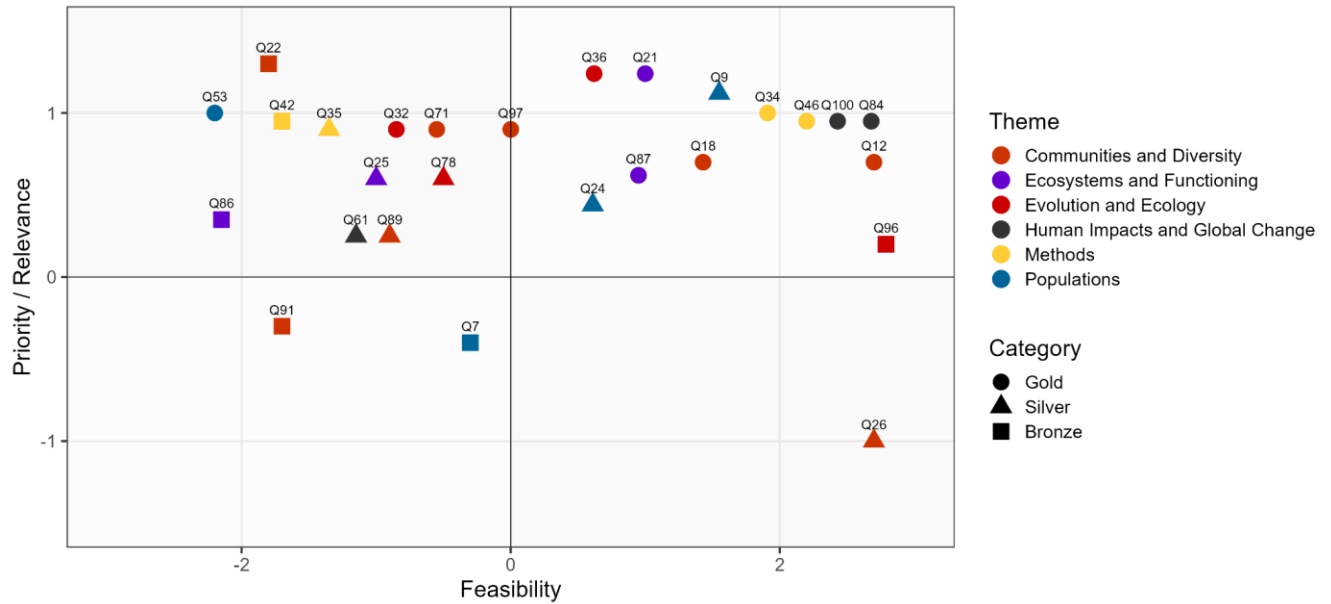
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934 **Figure 4.** Sankey diagrams showing flows from Themes → Priority classification → Subthemes

935 from Phase 1 to 3. Widths reflect the number of questions considered during Phase 3,

936 highlighting absolute research focus. Subthemes were grouped based on shared conceptual

937 keywords.



938

939 **Figure 5.** Phase 3 results: highest-priority questions placed on a two-axis grid (priority ×
 940 feasibility), enabling experts to visually compare and negotiate the relative importance and
 941 practicality of candidate research questions. The original panel resulting from the in-person
 942 exercise during the workshop is available at [Genially.com](https://genially.com) for close inspection.

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944