#### Perspective – Conservation Physiology 1

2 **Evaluating the vulnerability of critical early life stages in plants** 3 during heat extremes 4 5

6

Pieter A. Arnold 1,\*, Tara J. Walker 1, Ella V. Wishart 1, Lydia K. Guja 2,3,4

7

- <sup>1</sup> Division of Ecology and Evolution, Research School of Biology, The Australian 8
- National University, Canberra, ACT Australia 9
- <sup>2</sup> Centre for Australian National Biodiversity Research (a joint venture between 10
- 11 CSIRO and Parks Australia), CSIRO, Canberra, ACT, Australia
- <sup>3</sup> National Seed Bank, Australian National Botanic Gardens, Parks Australia, 12
- 13 Canberra, ACT, Australia
- <sup>4</sup> Umwelt Environmental and Social Consulting, Canberra, ACT Australia 14

15

16 \* Author for correspondence: Pieter Arnold (pieter.arnold@anu.edu.au)

- Lay summary: Early and reproductive life stages of plants are exposed to very 18
- 19 different conditions than established vegetation. Physiological differences among
- 20 these stages mean that they tolerate and respond to heat exposure differently. We
- 21 argue the need to consider physiology and ecology of these vulnerable life stages for
- 22 enhancing conservation and restoration.

# **Abstract**

23

24 Plants, their seeds, and their gametes show remarkable resilience and 25 responsiveness to environmental conditions. However, worsening climate change with more severe and frequent extreme climatic events, like heatwaves and hot 26 27 droughts, will likely push beyond physiological limits of many species. If such events occur during important points of development and reproduction – rather than mature 28 29 vegetative growth – the direct impact on individual fitness can be high, with potential to bottleneck recruitment in populations. Here we take an ecophysiological 30 31 perspective to discuss what is known of the effects of extreme heat on four critical early life stage transitions in wild plant development that affect fitness and 32 33 recruitment. These life stages are pollen development, pollen germination to seed set, dormant to imbibed seeds, and seed-to-seedling transition. We use the recently 34 developed thermal load sensitivity framework to showcase how these critical points 35 of ontogeny could be exposed to vastly different microclimate conditions and have 36 37 different physiological heat tolerance. Assessing sensitivity of these life stage transitions to increasing thermal load with the additional stressors of limited soil 38 39 moisture and drying atmosphere could be an effective approach to identify at-risk populations or species. We argue that vulnerable developmental stages and narrow 40 reproductive windows that affect recruitment must considered for effective 41 conservation and restoration of plant populations under climate change. 42

43

44

### Keywords

Heat, drought, heatwave, reproduction, extreme climatic event, thermal death time,

46 thermal load sensitivity

47

48

### **Abbreviations**

49	ECE	Extreme Climatic Event
50	ROS	Reactive Oxygen Species
51	TDT	Thermal Death Time
52	TLS	Thermal Load Sensitivity
53	VPD	Vapour Pressure Deficit

## Introduction

Extreme climatic events (sometimes called ECEs) are now occurring more frequently across Earth with far-reaching impacts for life (Perkins-Kirkpatrick et al., 2024, Perkins-Kirkpatrick et al., 2020, Thakur et al., 2022). Against a long-term background of steadily increasing average temperatures through climate warming, extreme heat events including individual hot days can act as 'pulses' that punctuate the impact of the rising 'press' of anthropogenic climate change (Harris et al., 2018). The severity of extreme heat events increases hyper-allometrically with average warming, meaning that expected extremes in future are greater than a simple increase above background temperatures (Seneviratne et al., 2021). 

The effects of extreme heat events like hot droughts and heatwaves are most often studied on adult populations of standing vegetation without considering flow on impacts to reproduction and recruitment of future populations. Yet, while extreme heat events are inherently rare occurrences (Perkins *et al.*, 2013), they are powerful forces for selection at life stages that have limited capacity to buffer against intense or repeated thermal stress (Buckley *et al.*, 2016). Drought also often co-occurs with extreme heat due to precipitation deficits and changes in evapotranspiration that lead to lack of sufficient soil moisture. As temperature increases, the vapour pressure deficit (VPD) climbs exponentially, leading to greater atmospheric evaporative demands and amplification of drought stress (Grossiord *et al.*, 2020, Novick *et al.*, 2024). All these stressors represent significant challenges for physiological functions. Mortality of mature plants obviously limits potential recruitment into populations. Yet the impacts of drought, heat, and/or high VPD on the production of viable seed and seedling establishment are more limiting in wild plants, with profound implications for population persistence.

### The impact of heat extremes and their timing on recruitment

- There is high potential for recruitment failure at early life stages facing extreme heat.
- 82 Reproductive and newly developing tissues are typically more susceptible to heat
- failure than vegetative tissues in plants (Ladinig et al., 2015, Lohani et al., 2020,
- Tushabe et al., 2025b). In many ecosystems there are times of year that are typically
- optimal for plant growth and reproduction, referred to here as growing season (which
- may not be contiguous). This period often coincides with gradually increasing

daylength, adequate soil moisture from rainfall or snowmelt, and release from cold limitation by warmer conditions facilitating growth. Germinating seeds and seedlings can be pushed beyond physiological tipping points from extremes occurring early in the growing season (Cinto Mejía *et al.*, 2023, Fernández-Pascual *et al.*, 2019, Niu *et al.*, 2014). Phenology – the timing of biological events – can be highly responsive to average warming, where earlier germination and earlier flowering are typical responses among plants (Anderson *et al.*, 2012, Parmesan *et al.*, 2015, Wadgymar *et al.*, 2018). However, there is substantial risk that extremes that occur 'unexpectedly' (e.g., early in the growing season) will disproportionately impact reproductive fitness and more vulnerable early life stages (Cinto Mejía *et al.*, 2023). Plants with predictable growing seasons may be more at risk from out-of-season (a-seasonal) extremes due to the phenology of physiological stress tolerance conferral matching seasonal change (Grossman, 2023).

Here we contend that research on the effects of heat at critical early life stages needs attention in wild plants. We briefly discuss developmental stages and physiological transitions that are vulnerable to heat and drought stress during a typical plant's life cycle: 1) pollen development, 2) pollen germination to seed set, 3) dormant to imbibed seeds, and 4) seed-to-seedling transition. We discuss how physiology can build understanding of the sensitivity of these important stages to extremes, focusing on heat (and not the potentially damaging effects of cold temperatures). We then use the thermal load sensitivity modelling framework to showcase potential impacts of heat on these life stages. Effective conservation requires that we understand not only the effects of heat on adult vegetation, but across other life stages. Conservation and restoration efforts can likely be enhanced by considering heat sensitivity from a whole-life-cycle perspective to develop predictive and protective approaches during establishment and reproduction periods.

# Key life stages for fitness are less resilient to heat stress

# Pollen development

Pollen develops through a complex, multi-stage process that requires coordinated function of multiple tissues (Hafidh *et al.*, 2016). The complex nature of pollen development means it is easily disrupted by abiotic stress, ultimately causing fewer viable pollen grains to be released (Chaturvedi *et al.*, 2021, De Storme *et al.*, 2014).

Heat or drought stress, either in isolation or combination, during flowering can result in abnormal anther morphology and function (Rang *et al.*, 2011, Tushabe *et al.*, 2025a); altered carbohydrate metabolism and inadequate nutrient reserve accumulation in pollen (Hu *et al.*, 2020, Pressman *et al.*, 2002); and oxidative damage to reproductive tissues (Djanaguiraman *et al.*, 2018). When prolonged, even relatively mild increases in growth temperature can drastically reduce pollen viability and the number of grains released in tomato (Sato *et al.*, 2006). Short periods of extreme stress can cause defects in mature pollen by disrupting specific processes. For example, chromosome segregation and cytokinesis during meiosis (De Storme *et al.*, 2020, Pécrix *et al.*, 2011) and the timing of tapetal cell degeneration, which is crucial for normal pollen development (Saini *et al.*, 1984). Stress exposure during early pollen development often results in greater reductions in pollen production and viability compared to later stages (Begcy *et al.*, 2019, Chaturvedi *et al.*, 2021).

By limiting the number of viable pollen grains available, stress during pollen development drastically diminishes the chances of successful fertilisation, with subsequent impacts on the success of sexual reproduction (Fig. 1). For fertile seed to be produced, viable pollen must be transferred to a compatible stigma and then germinate to deliver sperm cells to the ovule – all within a crucial window in which female tissue remains receptive (Hedhly et al., 2009). Very few of the pollen grains produced ever adhere to a compatible stigma in outcrossing species (Harder et al., 2008), and subsequent pollen germination is both competitive and highly sensitive to abiotic conditions (Hedhly et al., 2009, Stokes et al., 2025). Reduced availability of viable pollen due to stress-affected development constrains the number of possible fertilisation events. As such, heat stress contributes to and exacerbates pollen limitation – whereby seed production is limited by the quantity and quality of pollen grains reaching the stigma (Ashman et al., 2004, Rosenberger et al., 2024). Low seed set stemming from inviable pollen production contributes to the rarity of several threatened plant species (Burne et al., 2003, Kimpton et al., 2002), and has implications for population persistence, gene flow, and species distributions (Fernández-Pascual et al., 2019, Harder et al., 2010).

### Pollen development



- · Fewer viable pollen grains released
- Diminished pool of viable pollen
- Altered anther morphology and function
- Indehiscence (grains not released)
- Carbohydrate metabolism and nutrient accumulation altered
- Reduced starch required to germinate
- Oxidative stress and damage

# Pollen germination to seed set



- \* Fewer successful fertilisation events
- · Inhibited pollen tube growth
- Turgor loss alters pollen tube morphology
- Pistil unable to support pollen germination
- · Earlier loss of female receptivity

- Pollen limitation
- Seed set constrained
- \* Reproductive failure
- Reduced recruitment opportunities
- Decreased gene flow and dispersal capacity
- · Loss of genetic diversity
- Declines in abundance
- Localised extinctions

### Moderating factors:

- Long flowering duration
- Flexible flowering time
- Long lifespan
- Self-fertilisation or mixedmating strategy

### Dormant to imbibed seeds



- Fewer successful germinating seeds
- · Disruption of metabolic processes
- · Oxidative stress and damage
- Imbibition reduces protection from small heat shock proteins
- Re-drying of imbibed seeds beyond critical fraction leads to mortality
- Embryonic cell death and halted germination

- · Recruitment failure
- Seedbank depletion
- Loss of genetic diversity
- · Declines in abundance
- Localised extinctions

### Moderating factors:

- Dormancy type
- Maternal effects
- Sensitivity to and triggers of germination

# Seed-to-seedling transition



- Fewer established seedlings
- \* "False starts"
- Prioritise biosynthesis for water acquisition over photosynthesis
- Disruption of metabolic processes
- · Oxidative stress and damage
- · Reduced photosynthetic biomass
- Reduced cooling capacity via transpiration

- Recruitment failure
- Impaired growth and reproduction later in life
- · Loss of genetic diversity
- Declines in abundance
- Localised extinctions

### Moderating factors:

- Maternal effects
- Seed and soil microbiome

150 151

152

153

154

155

156

**Figure 1.** Four critical life stage transitions during a typical plant's development. Some of the major (non-exhaustive) effects of extreme heat (with associated drying and high VPD) on physiology changes at each stage, and the fitness impacts for individuals and populations. Some moderating factors that could affect the impacts of heat on fitness are inset in grey boxes. Note that pollen development and pollen germination to seed set are grouped together to simplify fitness impacts and the coloured horizontal dividing lines match the colours of life stages used in Figures 2 and 3.

### Pollen germination and seed set

Upon adhering to a compatible and receptive stigma, viable pollen rehydrates and germinates. It then forms a pollen tube that grows rapidly through the style and ruptures once reaching the embryo sac of an ovule to deposit sperm cells for fertilisation (Firon et al., 2012, Johnson et al., 2019). Abiotic stress can reduce fertilisation success by inhibiting pollen tube growth and limiting the ability of the pistil to support pollen germination (Hedhly et al., 2009, Snider et al., 2011b). Pollen germination and the rate of pollen tube growth increase with temperature up to an optimum, then rapidly decline beyond a thermal maximum where tube growth fails (Kakani et al., 2002, Kakani et al., 2005, Steinacher et al., 2012). This thermal maximum is less than 45°C in many plant species and can be as low as 30°C (Stokes et al., 2025, Tushabe et al., 2025b). High temperatures are associated with increased reactive oxygen species (ROS) production in pollen tubes, leading to growth arrest or premature rupture (Muhlemann et al., 2018); altered respiration rates (Karapanos et al., 2009); and altered cell wall synthesis due to disturbed cytoskeletal function and anomalous enzyme distributions (Parrotta et al., 2016). Heat stress can also shorten the receptivity period of the stigma (Hedhly et al., 2005), and reduce the availability of soluble carbohydrates in the pistil to sustain pollen tube growth (Snider et al., 2011b). Heat coupled with drought can cause osmotic stress from dehydration of female tissues that alters the volume and morphology of pollen tubes (Qian et al., 2025, Zonia et al., 2004).

Declines in fertilisation success caused by stress-impaired pollen germination can constrain the production of fertile seed (Snider *et al.*, 2011a). This limits opportunities for recruitment, with cascading ecological and evolutionary consequences for populations (Rosbakh *et al.*, 2018) (**Fig. 1**). Short-lived species are also likely to be more affected by pollen-driven declines in seed set, since adults have fewer opportunities to reproduce over their lifetime (Iler *et al.*, 2021). Self-compatible species may be less vulnerable to pollen limitation and subsequent reproductive failure than self-incompatible species (Busch *et al.*, 2008). Though self-incompatibility can be beneficial for adaptation and avoiding inbreeding, the inability to self-fertilise ultimately means self-incompatible plants have fewer sources of compatible pollen and a greater reliance on pollinator activity; both of which can be detrimentally impacted by abiotic stress (Burd, 1994, Busch *et al.*, 2008). Seed

limitation can exert a strong influence on the growth and dynamics of wild plant populations (Clark *et al.*, 2007, Ehrlén *et al.*, 2006, Price *et al.*, 2008). Lack of viable seed set due to impaired pollen tube growth and failed fertilisation can then set population boundaries, lead to localised extinctions, and alter community composition (Pigott *et al.*, 1981, Rosbakh *et al.*, 2018). As pollen is a vessel for gene transfer, fertilisation can shape the genetic structure and adaptation in subsequent generations (Ravikumar *et al.*, 2003, Rounds *et al.*, 2011). Impaired pollen availability and performance due to abiotic stress may also impact the capacity for plants to respond to future abiotic regimes (Hedhly *et al.*, 2009).

#### Dormant to imbibed seeds

In seeds with physical dormancy, a water-impermeable seed coat keeps the seeds in an anhydrobiotic state until dormancy break occurs (Baskin et al., 2014). Anhydrobiosis allows orthodox seeds to dry and survive in a dehydrated, dormant state where metabolic activity has declined to near-zero, and long periods of drought and extreme heat can be endured. Seeds with morphological and/or physiological dormancy have water-permeable seed coats and thus can readily imbibe water while still in a dormant state (Baskin et al., 2014). Recalcitrant seeds do not undergo anhydrobiosis and thus retain a higher moisture content while in a quiescent state. For simplicity, we do not discuss recalcitrant seeds further here, but it is worth noting that their retained moisture content likely confers greater thermal vulnerability from dispersal until germination than orthodox seeds. The lead up to imbibition therefore differs among seed types. Recalcitrant seeds maintain a high seed moisture content relative to orthodox seeds. Orthodox seeds undergo anhydrobiosis and can either: i) persist in a state of quiescence prior to imbibition (non-dormant seeds), ii) remain in a dry state until dormancy is alleviated, transferring them to a guiescent state prior to imbibition (all physically dormant seeds), or iii) readily imbibe water while still in a dormant state.

Imbibition is the process that reverses anhydrobiosis, rehydrating the seed and facilitating repair of damage incurred during prolonged anhydrobiosis to allow germination to progress. The germination process is often described in three phases (Bewley *et al.*, 2013, Nonogaki *et al.*, 2010). Briefly, Phase I: imbibition is the first step in the germination process and involves cell rehydration and expansion. Phase II: an uptick in cellular respiration and the resumption of various metabolic processes

(e.g., restoration of cellular integrity, and mitochondrial and DNA repair) that had been suspended or reduced prior to imbibition. Phase III: germination is complete once the radicle or other embryonic tissue has emerged from the seed coat.

Dry seeds can tolerate extreme temperatures before being detrimentally impacted (Bell et al., 1998, Hanley et al., 2000, Ooi et al., 2014). However, if desiccation from drought or heat occurs during imbibition, this tolerance can be altered. Partial re-drying of imbibed seeds during the early stages of imbibition before reaching the critical fraction – beyond which the seed is committed to germination – can be favourable to harden seeds (Hanson, 1973). However, once beyond the critical fraction, orthodox seeds lose their desiccation tolerance and can no longer survive re-drying (Daws et al., 2007). Once seeds are imbibed, they become more vulnerable to abiotic factors including high temperature (Ruprecht et al., 2016, Tangney et al., 2019). This may be a result of available free water becoming heated to the extent that cellular machinery is damaged (Tangney et al., 2019). Additionally, small heat shock proteins may provide protection of key cellular machinery while seeds are in a dry state, but these reduce in abundance as seeds hydrate, thereby conferring less protection (Leprince et al., 2017). ROS play a crucial role in signalling throughout seed germination, however heat stress can greatly increase ROS production and lead to substantial oxidative stress and cell death (Gomes et al., 2013, Kranner et al., 2010).

While most dry seeds have a high thermal tolerance, the increase of extreme heat events (Perkins-Kirkpatrick *et al.*, 2020) and hazard-reduction burns (Agee *et al.*, 2005) occurring out-of-season when soil (and seed) moisture content is higher (Tangney *et al.*, 2022), suggests that seeds will increasingly be exposed to warm temperatures while in a vulnerable state. This could lead to widespread mortality that reduces the soil seedbank from which populations regenerate (**Fig. 1**). Seed and germination traits can adapt to the changing climate to shift towards trait values conducive to successful establishment, as long as the rate at which climatic changes occur does not outpace species' adaptive capacity (Everingham *et al.*, 2021). Water-impermeable seed coats may enhance persistence and thus shift community composition towards species that possess physical dormancy as a bet-hedging strategy (Pausas *et al.*, 2022). Maternal resource provisioning and stress priming can be conferred to seeds to enhance seedling establishment (Brunel-Muguet *et al.*, 2025). However, dormancy loss due to exposure to increased soil temperatures may

diminish the extent to which physical dormancy can moderate the impact of extreme heat on seed bank persistence (Cochrane, 2017, Ooi *et al.*, 2009). Finally, species that rely solely on regeneration from the soil seedbank may be more vulnerable to subsequent extreme events than species with resprouting ability. If a mortality event depleting the soil seedbank is followed by a disturbance event that diminishes the standing vegetation before adequate contributions are made to restock the seedbank, there is an enhanced threat of both immaturity risk and localised extinctions (Keeley *et al.*, 1999, Nolan *et al.*, 2021, Zedler *et al.*, 1983).

### Seed-to-seedling transition

The seedling stage – referring to the period during which seed reserves are still partially utilised, ending when first true foliage is mature (Winkler et al., 2024) - is one of the most vulnerable stages of a plant's development (Leck et al., 2008, Saatkamp et al., 2019). The seed-to-seedling transition marks a fundamental shift from heterotrophic to autotrophic metabolism (Henninger et al., 2022). Biosynthesis of chlorophyll to develop autotrophic competence is initiated by light and highly responsive to environmental cues (Ha et al., 2017a). Both chlorophyll biosynthesis and response to abiotic stressors produce ROS, which can accumulate to cause oxidative damage and cell death (Ha et al., 2017a, 2017b, Mohanty et al., 2006), but are also essential for stress signalling (Mittler et al., 2011). The elongation of seedling shoot apical meristems and cotyledons occurs with exposure to warm temperatures (Lee et al., 2014), which can facilitate swift emergence through the soil and enable leaf thermoregulation by increasing distance from soil that radiates heat (Crawford et al., 2012). Developing seedlings need to rapidly expand to establish autotrophic growth, where biosynthesis within the cotyledons establishes photosynthetic machinery and begins carbon fixation while seed reserves are still being mobilised (Kandar et al., 2024).

Plants have evolved remarkable developmental plasticity in response to their environment (Sultan, 1995, 2000) – germinating seeds can slow metabolic processes to conserve energy, activate stress responses, and delay the shift to autotrophic metabolism until conditions improve (Rosental *et al.*, 2014). Nonetheless, "false starts" occur due to sensitive germination response during temporarily favourable conditions that are followed by periods of stress. Abiotic stress, especially heat and drought can quickly prove lethal to emerging seedlings or impair seedling

growth with lasting fitness consequences (Dekkers *et al.*, 2015, Smolikova *et al.*, 2021, Winkler *et al.*, 2024). As less predictable extremes occur more frequently and a-seasonally due to climate change progression, poorly timed germination could cause failure of entire cohorts of seedlings (Fernández-Pascual *et al.*, 2019, Orsenigo *et al.*, 2014). Early *vs* late extreme heat can have differential impacts on fecundity and survival of plants (Cope *et al.*, 2023, Dreesen *et al.*, 2015). Even arid dryland grasses that are highly adapted to taking advantage of narrow germination windows are threatened by both heat and water stress, such that extreme heat during water limitation can cause >95% recruitment failure (Lewandrowski *et al.*, 2021). Dry soil exposed to high solar input greatly exceeds air temperatures (Qiu *et al.*, 1998), resulting in inescapable extreme heat conditions for small plants, which accumulate to exceed physiological tolerance thresholds (Hankin *et al.*, 2025, Kolb *et al.*, 1996). Prolonged dry conditions with concurrently high temperatures presents a lethal combination for seedlings well after initial establishment (Hankin *et al.*, 2025, Moran *et al.*, 2019).

Stressful conditions during development can diminish individual- and population-level fitness (**Fig. 1**). To endure extreme heat and drought, seedlings must slow their metabolism and prioritise biosynthesis for water acquisition over photosynthetic tissues. Consequently, these stressors early in plant development can lead to impaired growth and poor reproductive outcomes that reduce individual fitness (Hou *et al.*, 2014, Shevtsova *et al.*, 2009, Winkler *et al.*, 2019, Zhang *et al.*, 2008). Recruitment of seedlings is already a substantial bottleneck for populations due to their limited resources, vulnerability to biotic and abiotic factors, and exposure to high competition (Eriksson *et al.*, 2008). Further, the microbiome of the seed and the soil it germinates in plays a potentially significant role in facilitating seedling establishment (Nelson, 2018). Ultimately, if cohorts of seedlings are unable to establish, or have reduced fitness later in life, then population persistence is likely to be jeopardised.

Seedlings that do manage to establish then enter an extended phase of vegetative growth, where they photosynthesise to accumulate biomass to support later reproductive phases. The vegetative stage is generally much more robust than the life stages discussed here due to established root systems and autotrophy. Further, leaves of vegetative plants are often numerous, and are replaceable modules rather than whole individuals (i.e., seeds and seedlings). While the thermal

tolerance of mature vegetative tissue is outside the focus of this perspective, there is a vast literature for readers to explore further (e.g., Geange *et al.*, 2021, Lancaster *et al.*, 2020, Perez *et al.*, 2025).

# Modelling thermal load sensitivity across vulnerable life stages

There are now sophisticated biophysical models to estimate plant temperatures through time based on high-resolution microclimate data sources (Kearney *et al.*, 2024, Kearney *et al.*, 2020, Klinges *et al.*, 2022, Meyer *et al.*, 2023). Here we provide a simplified demonstration of potential ways to model probability of mortality or 'failure' due to heat at early life stages using biophysical models and physiological parameters within the Thermal Load Sensitivity (TLS) framework (Arnold *et al.*, 2025a). Here, we use a location near Richmond, New South Wales, Australia (150.7379 °E, 33.6187 °S) for these illustrative examples. This location is classified as a humid subtropical climate but can have extreme heat, with record maximum temperatures ≥ 40°C between October and March, with a highest recorded 47.4°C in January 2020.

To model relevant environment conditions for the different life stages, we used micro silo: an implementation of the microclimate model in NicheMapR that uses a high-resolution weather database for Australia (Kearney et al., 2020). We simulated a generic small plant's microclimate: for pollen at 50 cm, for imbibed seeds at the soil surface (0 cm), and seedlings at 1 cm aboveground (relevant for newly emerged seedling height). For all simulations, we set a minimum of 0% shade and maximum of 60% shade to simulate a relatively thin canopy that could be expected in many ecosystems during periods of drought and heat. Solar radiation, rainfall effects on the soil moisture, relative humidity, and wind speed, among others, are all modelled as part of the *micro silo* implementation (Kearney et al., 2017). We fitted 'ectotherm' models in 'leaf mode', following the procedure outlined in Kearney et al. (2024). The same principles can be applied across life stages if different aspects of the plant's microclimate are captured in the fitted parameters. For simplification purposes, we fitted the model for pollen as a single stage, along with imbibed seed ('seed') and seedling stages. All R code with annotation of modelling procedures, outputs, and to replicate the simulations is openly available

Here we simulate individual- and population-level heat failure risk using reasonable initial estimates from our own preliminary data. Although ecophysiological data for seedlings is very limited, the heat tolerance limits of photosystem II in cotyledons and first leaves appear similar to that of adult plants (Alvarez et al., 2025). The key difference is that these tissues are essentially the entirety of productive biomass for seedlings, while they are a replaceable module of vegetative, mature plants. We fitted TLS parameters (akin to thermal death time or TDT) to the three life stages based to show how the highest temperature each can tolerate (heat tolerance) differ but clearly decreases when exposed to that temperature for longer durations (Fig. 2a). For simulations of mortality through time, we calculate net damage based on the difference between a damage accumulation model (Ørsted et al., 2024) and an Arrhenius-type repair function (Arnold et al., 2025a). The net damage rate can then be separated into two parts. The first is a range of 'permissive' temperatures in which repair outweighs damage, resulting in net repair. Then, beyond the permissive range, temperatures are 'stressful' in which damage outweighs repair and damage accumulates exponentially (Ørsted et al., 2022) (Fig. 2b). Heat failure rate in the stressful range escalates extremely rapidly (> 100% per 1°C) due to exceeding physiological tipping points (Jørgensen et al., 2022). Including repair allows heat failure probability to decline as damage is repaired during periods when temperatures are permissive.

359

360

361

362

363

364

365

366

367

368

369

370

371

372

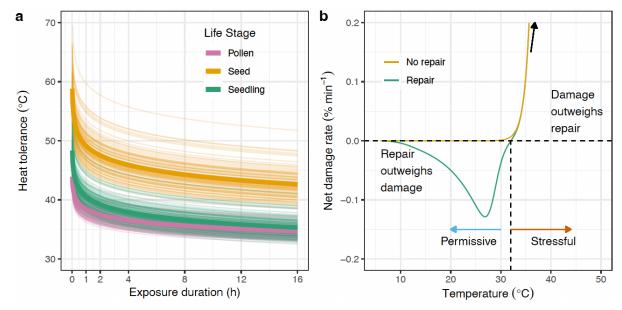
373

374

375

376

377



**Figure 2.** Thermal load sensitivity modelling inputs. **a)** Thermal death time (TDT) curves showing the relationship between heat tolerance and exposure duration across the three life stages that differ in heat tolerance and sensitivity. As exposure duration increases, heat tolerance declines but to different extents across the life stages. Thick lines show the mean population-level (*n*=100) responses of the thin lines that are simulated individuals. **b)** Example relationship between net damage and temperature, showing that for a given minute, repair outweighs damage below ~31°C (the 'permissive' range), while damage outweighs repair (the 'stressful' range) above it and damage accumulates exponentially, well beyond the maximum repair rate (black arrow indicates exponential increase).

By simulating the microclimates at relevant vertical stratifications for the different life stages, we illustrate how radically different microclimate temperatures emerge from biophysical principles, especially on hot days (Fig. 3a). All life stages reach temperatures well above the standard 1.2 m air temperature, especially seeds and seedlings that are closest to the soil surface under high solar load (Fig. 3a). This approach allows us to model heat failure probability across the time series of microclimate temperatures by determining the hazard of heat accumulation over time (Fig. 3b). The notable spikes in heat failure probability align with the hottest daytime temperatures reached but clearly vary among the population (Fig. 3b) even with relatively minor differences in their sensitivity to heat exposure (i.e., Fig. 2a). Despite the much higher temperatures in the seed microclimate, their higher tolerance reduces heat failure probability to well below pollen and akin to seedlings at the end of the simulation (Fig. 3b). Here, the individuals that have heat failure probability < 1 can repair damage during the cooler period with rainfall. However, those individuals that had relatively more damage remain more likely to succumb to accumulated damage, such that the recurrent heat event requires less damage to reach a physiological tipping point, for damage to accumulate, and for heat failure probability to reach 1 (Fig. 3b).

389

390

391

392

393

394

395

396

397

398

399

400

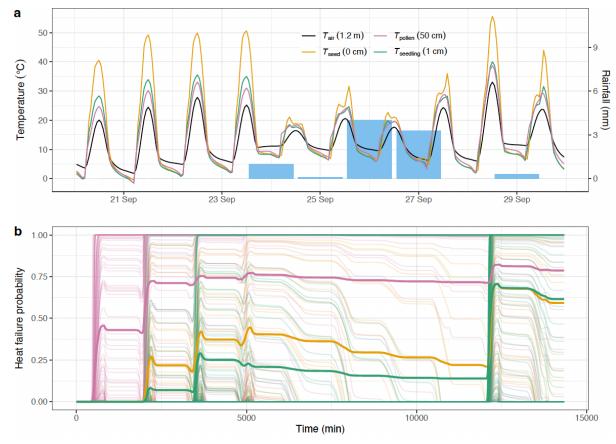
401

402

403

404

405



**Figure 3.** Simulated microclimate temperatures and heat failure probability across life stages. **a)** Temperature at different heights above the ground drastically differ, especially on hot days with high solar radiation input and without rainfall.  $T_{air}$  at 1.2 m is equivalent to standard air temperature weather forecasts. Taking the first day as an example, an imbibed seed right at the soil surface could reach temperatures 20°C above  $T_{air}$ , while a seedling at 1 cm above it could be much cooler but still 8°C above  $T_{air}$ . **b)** By modelling the damage accumulation over time and allowing for repair at permissive temperatures, we can estimate the heat failure probability at the individual- (thin lines) and mean population-level (thick lines) among life stages across the time series. Due to the differences in thermal load sensitivity each individual and life stage has a different heat failure probability under identical (within-stage) environmental conditions.

We note that our illustrative example models are a starting point for generating hypotheses and stimulating ideas for further modelling and empirical science. They are generalisations of the biophysical forces acting on simplified structures with arbitrarily set properties. The discrete heights assumes that seeds are germinating directly at the soil surface and simplified effects of sparse surrounding vegetation to simulate shade. Nonetheless, microclimate models are a vast improvement over assuming nearby air temperature is equivalent to plant temperature. The extreme thermal deviations and vertical stratification at biologically relevant fine scales cannot be overlooked anymore (Klinges et al., 2025, Körner et al., 2018). Leaf heat tolerance is generally weakly predicted by species-level traits, phylogenetics, and origin macroclimates has been limited (Bison et al., 2024, Briceño et al., 2025, Perez et al., 2021). However, we now know that the thermal conditions to which plant tissues are directly exposed four days prior to tolerance assays can predict heat tolerance thresholds with high accuracy (Pottinger et al., 2025). As these life stages occur sequentially, at the individual- or population-level the cumulative hazard of heat failure may be additive across these developmental stages, exacerbating the impact of extremes on fitness.

Empirical validation of how damage and repair processes manifest to affect heat failure probability in early and reproductive life stages of plants need to be investigated from biochemical and energetic perspectives (Buerger *et al.*, 2023, Sokolova, 2021). The different scales and modularity of plants also need to be considered further. For example, a plant may produce hundreds to thousands of flowers with millions of individual pollen grains, so damage could be a proportion of grains rendered inviable, or number of flowers aborted per individual, while a seed or seedling is a discrete unit. Nonetheless, the procedural damage-repair approaches certainly offer an improvement over single point critical thresholds and thermal safety margin approaches that do not include exposure duration (Arnold *et al.*, 2025a, Cook *et al.*, 2024, Lutterschmidt *et al.*, 1997, Ørsted *et al.*, 2022). Integrating life stage-specific sensitivity among species using a systems modelling approach seems promising for developing understanding of potential community responses in complex ecological systems (Noble *et al.*, 2025).

# **Emerging themes for empirical inquiry**

Clearly there needs to be empirical effort to explore damage and repair, and to ground-truth both thermal exposure through microclimates and thermal load sensitivity for key life stages in populations of interest, not just mature vegetation. There is now a comprehensive handbook for standardised protocols to measure regenerative plant functional traits that ecologists should utilise for empirical data collection (Poschlod *et al.*, 2025). We also need to evaluate the abiotic and biotic modifiers for TLS across life stages. For example, warming or drought (soil or atmospheric) conditions prior to an extreme heat event could enhance TLS via cross-tolerance, stress memory, or priming that improves heat tolerance from the upregulation of other protective mechanisms (Bryant *et al.*, 2024, Harris *et al.*, 2024, Hossain *et al.*, 2018, Rodgers *et al.*, 2025).

Plants can display astonishing phenotypic plasticity in response to growth conditions (Arnold *et al.*, 2019, 2024, Nicotra *et al.*, 2010). Some species have capacity to rapidly upregulate heat tolerance in photosynthetic tissues in response to heat in hours to days (Andrew *et al.*, 2023, 2024, Havaux, 1993, Zhu *et al.*, 2018, 2024), even seedlings (Alvarez *et al.*, 2025). However, inducible plasticity and ecoevolutionary responses to abiotic stressors will have costs and limits that are not well characterised for mature wild plants, let alone for early and reproductive life stages thereof (DeWitt *et al.*, 1998, Hendry, 2016). Alternative strategies that limit heat transfer through passive structural traits and active thermoregulation rather than altering physiological heat tolerance could be effective for persisting through heat events, and the diversity of responses needs further exploration (Arnold *et al.*, 2025b, Guo *et al.*, 2025, van Zanten *et al.*, 2021). Generating empirical data to validate parameters and simulations such as our example could facilitate forecasting heat failure probability, which can be integrated across life stages and systems based on both realised exposure and physiological limits (Noble *et al.*, 2025).

# Conservation and restoration need physiologically resilient early and reproductive life stages

It is increasingly evident that meeting conservation goals will require consideration of how new plants can establish and build resilient, self-sustaining populations (Miller *et al.*, 2017). The early and reproductive life stages discussed here are essential for the

success of both retaining ecological values in currently conserved sites, and in reestablishing those values in degraded areas requiring active management and restoration (Tomlinson *et al.*, 2022). Identifying the conditions that limit the establishment, growth, and survival of plants at early life stages are fundamental. Understanding the ecophysiology of these systems will assist, even if the resultant actions of managers and practitioners will differ depending on conservation or restoration context. Heat exposure is modified by the conditions of the plant's surrounding micro-environment, physical, chemical, biotic, and hydrological aspects of the soil substrate and the surrounding biota (Sharma *et al.*, 2023). We need to consider how natural or artificial structures (to add shade or protection) or treatments (addition of water, soil microbiota, or pre-stress conditioning) can be applied to ameliorate heat exposure during key life stages at appropriate scales (Miller *et al.*, 2017, Tomlinson *et al.*, 2022).

The range of possible environments that support seed germination through to complete seedling establishment (i.e., regeneration niche breadth) needs greater attention. Experimental approaches like determining the thermal or hydrothermal germination niche of seeds for conservation or restoration can evaluate the window of opportunity for seed germination (Hirst *et al.*, 2025, Rajapakshe *et al.*, 2024). By integrating physiological windows such as these with microclimate windows from mechanistic niche mapping tools, we can generate heat failure probabilities for different life stages across time and space. Building spatiotemporal maps of heat failure probabilities based on the TLS framework has potential to integrate with spatial prioritisation tools (e.g., Unnithan Kumar *et al.*, 2025) to explore the effectiveness or feasibility of intervention treatments.

Restoration and conservation efforts requires careful consideration of species and provenances of seeds and seedlings. For sustainable restoration and longer-term conservation, these need to be tolerant to the most extreme microclimatic conditions to which they are (and will be) exposed at key points of ontogeny, not just the macroclimatic averages (Gross *et al.*, 2017, Miller *et al.*, 2017). This is especially important in restoration of degraded landscapes that are inherently more open and less protected from extremes (Valliere *et al.*, 2022). Ensuring that the chosen plants have reasonable likelihood to reproduce successfully also needs to consider the timing window of reproductive events and their sensitivity to extremes. Achieving sustainable conservation and restoration targets requires ecophysiology of early and

reproductive life stages and extremes to be explicitly considered to increase long-term resilience at the plant, population, and ecosystem level (Tomlinson *et al.*, 2022, Valliere *et al.*, 2022).

### Conclusion

Plants face a more extreme future that threatens individual- and population-level fitness. Here, we have highlighted the need for focus on early and reproductive life stages as key transitions throughout development. We have showcased how the evaluation of thermal exposure and thermal load sensitivity can lead to strong differential impacts of extreme heat across life stages. Physiology is central to understanding how biological entities and ecological systems will cope with the prolonged, intense heat and other extreme environmental perturbations in future. Conservation and restoration efforts need to ensure that the vulnerability of early and reproductive life stages to climate extremes, more than averages, is considered and supported so that populations have high likelihood to establish and reproduce in the short-term to be self-sustaining in the long-term.

## **Acknowledgements**

We acknowledge that this work was conducted on Ngunnawal Country and we pay our respects to the traditional custodians of these lands and elders past, present, and emerging. We thank the many colleagues who have been influential in shaping these ideas over years of study on plant ecophysiology and heat responses, notably Adrienne Nicotra, Andy Leigh, Loeske Kruuk, Verónica Briceño Rodríguez, Alicia Cook, Rosalie Harris, Phillipa Alvarez, Renée Marchin Prokopavicius, Kristine Crous, Samuel Andrew, Jerónimo Vásquez-Ramírez, Nicole Bison, Joanne Bennett, Daniel Falster, Sean Michaletz, Enrico Rezende, Michael Kearney, and Daniel Noble.

### **Author Contributions**

P.A.A. conceived the ideas, designed the modelling simulations, and led the writing of the manuscript. T.J.W., E.V.W., and L.K.G. all contributed sections and critical revisions to the draft and gave final approval for publication.

551	
552	Conflicts of Interest
553	None declared.
554	
555	Funding
556	This work was supported by an Australian Research Council Discovery Project Grant
557	(DP240100177) funded by the Australian Government.
558	
559	Data Availability
560	No specific data is associated with this article, however R code for simulations is
561	openly available on GitHub at <a href="https://pieterarnold.github.io/Life_stage_TLS/">https://pieterarnold.github.io/Life_stage_TLS/</a> . This will
562	also be published as a release via Zenodo.
563	
564	
565	References
566 567	Agee JK, Skinner CN (2005) Basic principles of forest fuel reduction treatments.
568	Forest Ecology and Management 211: 83-96. 10.1016/j.foreco.2005.01.034.
569	Alvarez PR, Harris RJ, Cook AM, Briceño VF, Nicotra AB, Leigh A (2025) Native
570	Australian seedlings exhibit novel strategies to acclimate to repeated heatwave
571	events. Oecologia 207: 84. 10.1007/s00442-025-05704-5.
572	Anderson JT, Inouye DW, McKinney AM, Colautti RI, Mitchell-Olds T (2012)
573	Phenotypic plasticity and adaptive evolution contribute to advancing flowering
574	phenology in response to climate change. Proceedings of the Royal Society B:
575	Biological Sciences 279: 3843-3852. 10.1098/rspb.2012.1051.
576	Andrew SC, Arnold PA, Simonsen AK, Briceño VF (2023) Consistently high heat
577	tolerance acclimation in response to a simulated heatwave across species from the
578	broadly distributed Acacia genus. Functional Plant Biology 50: 71-83.
579	10.1071/FP22173.
580	Andrew SC, Simonsen AK, Coppin CW, Arnold PA, Briceño VF, McLay TGB,
581	Jackson C.I. Gallagher RV. Mokany K. (2024) Expression-environment

- associations in transcriptomic heat stress responses for a global plant lineage.
- 583 *Molecular Ecology* 33: e17473. 10.1111/mec.17473.
- Arnold PA, Kruuk LEB, Nicotra AB (2019) How to analyse plant phenotypic
- plasticity in response to a changing climate. *New Phytologist* 222: 1235-1241.
- 586 10.1111/nph.15656.
- 587 Arnold PA, Noble DWA, Nicotra AB, Kearney MR, Rezende EL, Andrew SC,
- 588 Briceño VF, Buckley LB, Christian KA, Clusella-Trullas S et al. (2025a) A
- framework for modelling thermal load sensitivity across life. Global Change Biology
- 590 31: e70315. 10.1111/gcb.70315.
- 591 Arnold PA, Wang S, Notarnicola RF, Nicotra AB, Kruuk LEB (2024) Testing the
- evolutionary potential of an alpine plant: phenotypic plasticity in response to growth
- temperature outweighs parental environmental effects and other genetic causes of
- variation. *Journal of Experimental Botany* 75: 5971-5988. 10.1093/jxb/erae290.
- 595 Arnold PA, White MJ, Cook AM, Leigh A, Briceño VF, Nicotra AB (2025b) Plants
- originating from more extreme biomes have improved leaf thermoregulation. *Annals*
- 597 of Botany 136: 199-213. 10.1093/aob/mcaf080.
- 598 Ashman T-L, Knight TM, Steets JA, Amarasekare P, Burd M, Campbell DR,
- 599 Dudash MR, Johnston MO, Mazer SJ, Mitchell RJ et al. (2004) Pollen limitation of
- 600 plant reproduction: Ecological and evolutionary causes and consequences. *Ecology*
- 601 85: 2408-2421. 10.1890/03-8024.
- Baskin CC, Baskin JM (2014) Seeds: Ecology, Biogeography, and Evolution of
- 603 Dormancy and Germination. Academic Press, San Diego, CA, USA.
- 604 Begcy K, Nosenko T, Zhou L-Z, Fragner L, Weckwerth W, Dresselhaus T (2019)
- Male sterility in maize after transient heat stress during the tetrad stage of pollen
- development. *Plant Physiology* 181: 683-700. 10.1104/pp.19.00707.
- 607 **Bell DT, Williams DS** (1998) Tolerance of thermal shock in seeds. *Australian Journal*
- 608 of Botany 46: 221-233. 10.1071/BT97010.

- 609 **Bewley D, Bradford KJ, Hilhorst HWM, Nonogaki H** (2013) Seeds: Physiology of
- development, germination and dormancy. Springer, New York, NY, USA.
- Bison NN, Michaletz ST (2024) Variation in leaf carbon economics, energy balance,
- and heat tolerance traits highlights differing timescales of adaptation and
- 613 acclimation. *New Phytologist* 242: 1919-1931. 10.1111/nph.19702.
- 614 Briceño VF, Arnold PA, Cook AM, Courtney Jones SK, Gallagher RV, French K,
- 615 Bravo LA, Nicotra AB, Leigh A (2025) Drivers of thermal tolerance breadth of
- plants across contrasting biomes. Journal of Ecology 113: 3812-3829. 10.1111/1365-
- 617 2745.70198.
- 618 Brunel-Muguet S, Baránek M, Fragkostefanakis S, Sauvage C, Lieberman-
- 619 Lazarovich M, Maury S, Kaiserli E, Segal Na, Testillano PS, Verdier J (2025)
- Maternal environmental effects and climate-smart seeds: unlocking epigenetic
- inheritance for crop innovation in the seed industry. *The Plant Journal* 123: e70407.
- 622 10.1111/tpj.70407.
- 623 Bryant C, Harris RJ, Brothers N, Bone C, Walsh N, Nicotra AB, Ball MC (2024)
- 624 Cross-tolerance: Salinity gradients and dehydration increase photosynthetic heat
- tolerance in mangrove leaves. Functional Ecology 38: 897-909. 10.1111/1365-
- 626 2435.14508.
- 627 **Buckley LB, Huey RB** (2016) How extreme temperatures impact organisms and the
- evolution of their thermal tolerance. *Integrative and Comparative Biology* 56: 98-109.
- 629 10.1093/icb/icw004.
- 630 Buerger P, Buler M, Yeap HL, Edwards OR, van Oppen MJ, Oakeshott JG, Court
- 631 **L** (2023) Flow cytometry-based biomarker assay for *in vitro* identification of
- 632 microalgal symbionts conferring heat tolerance on corals. Frontiers in Marine
- 633 *Science* 10: 1094792. 10.3389/fmars.2023.1094792.
- 634 **Burd M** (1994) Bateman's principle and plant reproduction: The role of pollen
- limitation in fruit and seed set. *The Botanical Review* 60: 83-139.
- 636 10.1007/BF02856594.

637 Burne HM, Yates CJ, Ladd PG (2003) Comparative population structure and reproductive biology of the critically endangered shrub Grevillea althoferorum and 638 639 two closely related more common congeners. Biological Conservation 114: 53-65. 10.1016/S0006-3207(02)00420-2. 640 641 Busch JW, Schoen DJ (2008) The evolution of self-incompatibility when mates are 642 limiting. Trends in Plant Science 13: 128-136. 10.1016/j.tplants.2008.01.002. 643 Chaturvedi P, Wiese AJ, Ghatak A, Záveská Drábková L, Weckwerth W, Honys 644 **D** (2021) Heat stress response mechanisms in pollen development. New Phytologist 231: 571-585. 10.1111/nph.17380. 645 Cinto Mejía E, Wetzel WC (2023) The ecological consequences of the timing of 646 extreme climate events. Ecology and Evolution 13: e9661. 10.1002/ece3.9661. 647 648 Clark CJ, Poulsen JR, Levey DJ, Osenberg CW (2007) Are plant populations seed 649 limited? A critique and meta-analysis of seed addition experiments. *The American* 650 Naturalist 170: 128-142. 10.1086/518565. 651 **Cochrane A** (2017) Are we underestimating the impact of rising summer 652 temperatures on dormancy loss in hard-seeded species? Australian Journal of 653 Botany 65: 248-256. 10.1071/BT16244. Cook AM, Rezende EL, Petrou K, Leigh A (2024) Beyond a single temperature 654 655 threshold: Applying a cumulative thermal stress framework to plant heat tolerance. Ecology Letters 27: e14416. 10.1111/ele.14416. 656 657 Cope OL, Zehr LN, Agrawal AA, Wetzel WC (2023) The timing of heat waves has 658 multiyear effects on milkweed and its insect community. *Ecology* 104: e3988. 659 10.1002/ecy.3988. 660 Crawford AJ, McLachlan DH, Hetherington AM, Franklin KA (2012) High temperature exposure increases plant cooling capacity. Current Biology 22: R396-661 662 R397. 10.1016/j.cub.2012.03.044. Daws MI, Bolton S, Burslem DFRP, Garwood NC, Mullins CE (2007) Loss of 663 664 desiccation tolerance during germination in neo-tropical pioneer seeds: implications

665 for seed mortality and germination characteristics. Seed Science Research 17: 273-281. 10.1017/S0960258507837755. 666 667 **De Storme N, Geelen D** (2014) The impact of environmental stress on male reproductive development in plants: biological processes and molecular 668 669 mechanisms. *Plant. Cell & Environment* 37: 1-18. 10.1111/pce.12142. 670 De Storme N, Geelen D (2020) High temperatures alter cross-over distribution and 671 induce male meiotic restitution in Arabidopsis thaliana. Communications Biology 3: 672 187. 10.1038/s42003-020-0897-1. Dekkers BJW, Costa MCD, Maia J, Bentsink L, Ligterink W, Hilhorst HWM 673 (2015) Acquisition and loss of desiccation tolerance in seeds: from experimental 674 model to biological relevance. Planta 241: 563-577. 10.1007/s00425-014-2240-x. 675 676 **DeWitt TJ, Sih A, Wilson DS** (1998) Costs and limits of phenotypic plasticity. *Trends* in Ecology & Evolution 13: 77-81. 10.1016/S0169-5347(97)01274-3. 677 678 Djanaguiraman M, Perumal R, Jagadish SVK, Ciampitti IA, Welti R, Prasad PVV (2018) Sensitivity of sorghum pollen and pistil to high-temperature stress. *Plant, Cell* 679 680 & Environment 41: 1065-1082. 10.1111/pce.13089. Dreesen FE, De Boeck HJ, Horemans JA, Janssens IA, Nijs I (2015) Recovery 681 dynamics and invasibility of herbaceous plant communities after exposure to 682 experimental climate extremes. Basic and Applied Ecology 16: 583-591. 683 10.1016/j.baae.2015.05.002. 684 Ehrlén J, Munzbergova Z, Diekmann M, Eriksson OVE (2006) Long-term 685 assessment of seed limitation in plants: results from an 11-year experiment. Journal 686

of Ecology 94: 1224-1232. 10.1111/j.1365-2745.2006.01169.x.

Cambridge University Press, pp 239-254

Eriksson O, Ehrlén J (2008) Seedling recruitment and population ecology. In Leck

MA, Parker VT, Simpson RL eds, Seedling Ecology and Evolution, Cambridge:

687

688

689

690

- 691 Everingham SE, Offord CA, Sabot MEB, Moles AT (2021) Time-traveling seeds
- reveal that plant regeneration and growth traits are responding to climate change.
- 693 *Ecology* 102: e03272. 10.1002/ecy.3272.
- 694 **Fernández-Pascual E, Mattana E, Pritchard HW** (2019) Seeds of future past:
- 695 climate change and the thermal memory of plant reproductive traits. Biological
- 696 Reviews 94: 439-456. 10.1111/brv.12461.
- 697 **Firon N, Nepi M, Pacini E** (2012) Water status and associated processes mark
- 698 critical stages in pollen development and functioning. Annals of Botany 109: 1201-
- 699 1214. 10.1093/aob/mcs070.
- 700 Geange SR, Arnold PA, Catling AA, Coast O, Cook AM, Gowland KM, Leigh A,
- Notarnicola RF, Posch BC, Venn SE et al. (2021) The thermal tolerance of
- 702 photosynthetic tissues: a global systematic review and agenda for future research.
- 703 New Phytologist 229: 2497-2513. 10.1111/nph.17052.
- 704 **Gomes MP, Garcia QS** (2013) Reactive oxygen species and seed germination.
- 705 *Biologia* 68: 351-357. 10.2478/s11756-013-0161-y.
- 706 **Gross CL, Fatemi M, Simpson IH** (2017) Seed provenance for changing climates:
- early growth traits of nonlocal seed are better adapted to future climatic scenarios,
- but not to current field conditions. *Restoration Ecology* 25: 577-586.
- 709 10.1111/rec.12474.
- 710 Grossiord C, Buckley TN, Cernusak LA, Novick KA, Poulter B, Siegwolf RTW,
- 711 Sperry JS, McDowell NG (2020) Plant responses to rising vapor pressure deficit.
- 712 New Phytologist 226: 1550-1566. 10.1111/nph.16485.
- 713 **Grossman JJ** (2023) Phenological physiology: seasonal patterns of plant stress
- tolerance in a changing climate. New Phytologist 237: 1508-1524.
- 715 10.1111/nph.18617.
- 716 **Guo J-J, Gong X-W, Hao G-Y** (2025) Leaf transpirational cooling and thermal
- 717 tolerance vary along the spectrum of iso-anisohydric stomatal regulation in sand-
- 718 fixing shrubs. *Plant, Cell & Environment* 48: 2053-2066. 10.1111/pce.15279.

- 719 Ha J-H, Han S-H, Lee H-J, Park C-M (2017a) Environmental adaptation of the
- 720 heterotrophic-to-autotrophic transition: The developmental plasticity of seedling
- establishment. Critical Reviews in Plant Sciences 36: 128-137.
- 722 10.1080/07352689.2017.1355661.
- 723 Ha J-H, Lee H-J, Jung J-H, Park C-M (2017b) Thermo-induced maintenance of
- 724 photo-oxidoreductases underlies plant autotrophic development. Developmental Cell
- 725 41: 170-179.e174. 10.1016/j.devcel.2017.03.005.
- Hafidh S, Fíla J, Honys D (2016) Male gametophyte development and function in
- angiosperms: a general concept. Plant Reproduction 29: 31-51. 10.1007/s00497-
- 728 015-0272-4.
- Hankin LE, Barrios-Masias FH, Urza AK, Bisbing SM (2025) Lethal combination
- for seedlings: extreme heat drives mortality of drought-exposed high-elevation pine
- 731 seedlings. *Annals of Botany* 135: 293-304. 10.1093/aob/mcae064.
- 732 Hanley ME, Lamont BB (2000) Heat pre-treatment and the germination of soil- and
- 733 canopy-stored seeds of south-western Australian species. Acta Oecologica 21: 315-
- 734 321. 10.1016/S1146-609X(00)01087-0.
- 735 **Hanson AD** (1973) The effects of imbibition drying treatments on wheat seeds. *New*
- 736 *Phytologist* 72: 1063-1073. 10.1111/j.1469-8137.1973.tb02083.x.
- 737 **Harder LD, Aizen MA** (2010) Floral adaptation and diversification under pollen
- 738 limitation. Philosophical Transactions of the Royal Society B: Biological Sciences
- 739 365: 529-543. 10.1098/rstb.2009.0226.
- 740 Harder LD, Johnson SD (2008) Function and evolution of aggregated pollen in
- angiosperms. International Journal of Plant Sciences 169: 59-78. 10.1086/523364.
- Harris RJ, Alvarez PR, Bryant C, Briceño VF, Cook AM, Leigh A, Nicotra AB
- 743 (2024) Acclimation of thermal tolerance in juvenile plants from three biomes is
- suppressed when extremes co-occur. Conservation Physiology 12: coae027.
- 745 10.1093/conphys/coae027.

- 746 Harris RMB, Beaumont LJ, Vance TR, Tozer CR, Remenyi TA, Perkins-
- 747 Kirkpatrick SE, Mitchell PJ, Nicotra AB, McGregor S, Andrew NR et al. (2018)
- 748 Biological responses to the press and pulse of climate trends and extreme events.
- 749 Nature Climate Change 8: 579-587. 10.1038/s41558-018-0187-9.
- 750 **Havaux M** (1993) Rapid photosynthetic adaptation to heat stress triggered in potato
- leaves by moderately elevated temperatures. *Plant, Cell & Environment* 16: 461-467.
- 752 10.1111/j.1365-3040.1993.tb00893.x.
- 753 **Hedhly A, Hormaza JI, Herrero M** (2005) The effect of temperature on pollen
- germination, pollen tube growth, and stigmatic receptivity in peach. *Plant Biol (Stuttg)*
- 755 7: 476-483. 10.1055/s-2005-865850.
- 756 **Hedhly A, Hormaza JI, Herrero M** (2009) Global warming and sexual plant
- 757 reproduction. *Trends in Plant Science* 14: 30-36. 10.1016/j.tplants.2008.11.001.
- 758 **Hendry AP** (2016) Key questions on the role of phenotypic plasticity in eco-
- evolutionary dynamics. *Journal of Heredity* 107: 25-41. 10.1093/jhered/esv060.
- Henninger M, Pedrotti L, Krischke M, Draken J, Wildenhain T, Fekete A, Rolland
- 761 **F, Müller MJ, Fröschel C, Weiste C et al.** (2022) The evolutionarily conserved
- kinase SnRK1 orchestrates resource mobilization during Arabidopsis seedling
- 763 establishment. *The Plant Cell* 34: 616-632. 10.1093/plcell/koab270.
- Hirst MJ, Vázquez-Ramírez J, Rendall AR, Venn SE (2025) Thermal germination
- 765 niche: implications for seed-based restoration in climate-sensitive alpine
- 766 environments. Restoration Ecology 33: e70031. 10.1111/rec.70031.
- 767 Hossain MA, Li Z-G, Hoque TS, Burritt DJ, Fujita M, Munné-Bosch S (2018) Heat
- or cold priming-induced cross-tolerance to abiotic stresses in plants: key regulators
- and possible mechanisms. *Protoplasma* 255: 399-412. 10.1007/s00709-017-1150-8.
- Hou Q-Q, Chen B-M, Peng S-L, Chen L-Y (2014) Effects of extreme temperature
- on seedling establishment of nonnative invasive plants. *Biological Invasions* 16:
- 772 2049-2061. 10.1007/s10530-014-0647-8.

- 773 Hu W, Huang Y, Loka DA, Bai H, Liu Y, Wang S, Zhou Z (2020) Drought-induced
- disturbance of carbohydrate metabolism in anthers and male abortion of two
- 775 Gossypium hirsutum cultivars differing in drought tolerance. Plant Cell Reports 39:
- 776 195-206. 10.1007/s00299-019-02483-1.
- 777 Iler AM, CaraDonna PJ, Forrest JRK, Post E (2021) Demographic consequences
- of phenological shifts in response to climate change. *Annual Review of Ecology*,
- 779 Evolution, and Systematics 52: 221-245. 10.1146/annurev-ecolsys-011921-032939.
- Johnson MA, Harper JF, Palanivelu R (2019) A fruitful journey: Pollen tube
- 781 navigation from germination to fertilization. Annual Review of Plant Biology 70: 809-
- 782 837. 10.1146/annurev-arplant-050718-100133.
- Jørgensen LB, Ørsted M, Malte H, Wang T, Overgaard J (2022) Extreme
- escalation of heat failure rates in ectotherms with global warming. *Nature* 611: 93-98.
- 785 10.1038/s41586-022-05334-4.
- 786 Kakani VG, Prasad PVV, Craufurd PQ, Wheeler TR (2002) Response of in vitro
- pollen germination and pollen tube growth of groundnut (*Arachis hypogaea* L.)
- 788 genotypes to temperature. Plant, Cell & Environment 25: 1651-1661. 10.1046/j.1365-
- 789 3040.2002.00943.x.
- 790 Kakani VG, Reddy KR, Koti S, Wallace TP, Prasad PVV, Reddy VR, Zhao D
- 791 (2005) Differences in *in vitro* pollen germination and pollen tube growth of cotton
- 792 cultivars in response to high temperature. *Annals of Botany* 96: 59-67.
- 793 10.1093/aob/mci149.
- 794 **Kandar CC, Pal D** (2024) Relation between seed life cycle and cell proliferation.
- Metabolic changes in seed germination. In Pal D ed, Seeds: Anti-proliferative
- 796 Storehouse for Bioactive Secondary Metabolites, Singapore: Springer Nature
- 797 Singapore, pp 49-79
- 798 Karapanos IC, Akoumianakis KA, Olympios CM, Passam HC (2009) The effect of
- substrate, ADP and uncoupler on the respiration of tomato pollen during incubation
- in vitro at moderately high temperature. Sexual Plant Reproduction 22: 133-140.
- 801 10.1007/s00497-009-0098-z.

- 802 **Kearney MR, Leigh A** (2024) Fast, accurate and accessible calculations of leaf
- temperature and its physiological consequences with NicheMapR. *Methods in*
- 804 *Ecology and Evolution* 15: 1516-1531. 10.1111/2041-210X.14373.
- 805 **Kearney MR, Porter WP** (2017) NicheMapR an R package for biophysical
- modelling: the microclimate model. *Ecography* 40: 664-674. 10.1111/ecog.02360.
- 807 **Kearney MR, Porter WP** (2020) NicheMapR an R package for biophysical
- modelling: the ectotherm and Dynamic Energy Budget models. *Ecography* 43: 85-96.
- 809 10.1111/ecog.04680.
- Keeley JE, Ne'eman G, Fotheringham C (1999) Immaturity risk in a fire-dependent
- pine. Journal of Mediterranean Ecology 1: 41-48.
- 812 Kimpton SK, James EA, Drinnan AN (2002) Reproductive biology and genetic
- 813 marker diversity in *Grevillea infecunda* (Proteaceae), a rare plant with no known
- seed production. Australian Systematic Botany 15: 485-492. 10.1071/SB01029.
- 815 Klinges DH, Duffy JP, Kearney MR, Maclean IMD (2022) mcera5: Driving
- microclimate models with ERA5 global gridded climate data. *Methods in Ecology and*
- 817 Evolution 13: 1402-1411. 10.1111/2041-210X.13877.
- 818 Klinges DH, Muñoz MM, Domínguez-Guerrero SF, Maclean IMD, Kearney MR,
- 819 **Skelly DK** (2025) Matching climate to biological scales. *Trends in Ecology &*
- 820 *Evolution*: 10.1016/j.tree.2025.11.015.
- 821 Kolb PF, Robberecht R (1996) High temperature and drought stress effects on
- survival of *Pinus ponderosa* seedlings. *Tree Physiology* 16: 665-672.
- 823 10.1093/treephys/16.8.665.
- Körner C, Hiltbrunner E (2018) The 90 ways to describe plant temperature.
- 825 Perspectives in Plant Ecology, Evolution and Systematics 30: 16-21.
- 826 10.1016/j.ppees.2017.04.004.
- Kranner I, Minibayeva FV, Beckett RP, Seal CE (2010) What is stress? Concepts,
- definitions and applications in seed science. New Phytologist 188: 655-673.
- 829 10.1111/j.1469-8137.2010.03461.x.

- 830 Ladinig U, Pramsohler M, Bauer I, Zimmermann S, Neuner G, Wagner J (2015)
- ls sexual reproduction of high-mountain plants endangered by heat? *Oecologia* 177:
- 832 1195-1210. 10.1007/s00442-015-3247-0.
- 833 Lancaster LT, Humphreys AM (2020) Global variation in the thermal tolerances of
- plants. Proceedings of the National Academy of Sciences 117: 13580-13587.
- 835 10.1073/pnas.1918162117.
- Leck MA, Simpson RL, Parker VT (2008) Why seedlings? In Leck MA, Parker VT,
- 837 Simpson RL eds, Seedling Ecology and Evolution, Cambridge: Cambridge University
- 838 Press, pp 3-14
- 839 Lee H-J, Jung J-H, Cortés Llorca L, Kim S-G, Lee S, Baldwin IT, Park C-M (2014)
- FCA mediates thermal adaptation of stem growth by attenuating auxin action in
- Arabidopsis. Nature Communications 5: 5473. 10.1038/ncomms6473.
- Leprince O, Pellizzaro A, Berriri S, Buitink J (2017) Late seed maturation: drying
- without dying. *Journal of Experimental Botany* 68: 827-841. 10.1093/jxb/erw363.
- 844 Lewandrowski W, Stevens JC, Webber BL, L. Dalziell E, Trudgen MS, Bateman
- 845 **AM, Erickson TE** (2021) Global change impacts on arid zone ecosystems: Seedling
- establishment processes are threatened by temperature and water stress. *Ecology*
- 847 and Evolution 11: 8071-8084. 10.1002/ece3.7638.
- 848 Lohani N, Singh MB, Bhalla PL (2020) High temperature susceptibility of sexual
- reproduction in crop plants. *Journal of Experimental Botany* 71: 555-568.
- 850 10.1093/jxb/erz426.
- Lutterschmidt WI, Hutchison VH (1997) The critical thermal maximum: history and
- 852 critique. Canadian Journal of Zoology 75: 1561-1574. 10.1139/z97-783.
- Meyer AV, Sakairi Y, Kearney MR, Buckley LB (2023) A guide and tools for
- selecting and accessing microclimate data for mechanistic niche modeling.
- 855 *Ecosphere* 14: e4506. 10.1002/ecs2.4506.
- 856 Miller BP, Sinclair EA, Menz MHM, Elliott CP, Bunn E, Commander LE, Dalziell
- 857 **E, David E, Davis B, Erickson TE et al.** (2017) A framework for the practical

- science necessary to restore sustainable, resilient, and biodiverse ecosystems.
- 859 Restoration Ecology 25: 605-617. 10.1111/rec.12475.
- Mittler R, Vanderauwera S, Suzuki N, Miller G, Tognetti VB, Vandepoele K,
- **Gollery M, Shulaev V, Van Breusegem F** (2011) ROS signaling: the new wave?
- 862 Trends in Plant Science 16: 300-309. 10.1016/j.tplants.2011.03.007.
- 863 **Mohanty S, Grimm B, Tripathy BC** (2006) Light and dark modulation of chlorophyll
- biosynthetic genes in response to temperature. *Planta* 224: 692-699.
- 865 10.1007/s00425-006-0248-6.
- Moran EV, Das AJ, Keeley JE, Stephenson NL (2019) Negative impacts of
- summer heat on Sierra Nevada tree seedlings. *Ecosphere* 10: e02776.
- 868 10.1002/ecs2.2776.
- 869 Muhlemann JK, Younts TLB, Muday GK (2018) Flavonols control pollen tube
- growth and integrity by regulating ROS homeostasis during high-temperature stress.
- 871 Proceedings of the National Academy of Sciences 115: E11188-E11197.
- 872 10.1073/pnas.1811492115.
- Nelson EB (2018) The seed microbiome: Origins, interactions, and impacts. *Plant*
- and Soil 422: 7-34. 10.1007/s11104-017-3289-7.
- Nicotra AB, Atkin OK, Bonser SP, Davidson AM, Finnegan EJ, Mathesius U,
- Poot P, Purugganan MD, Richards CL, Valladares F et al. (2010) Plant phenotypic
- plasticity in a changing climate. *Trends in Plant Science* 15: 684-692.
- 878 10.1016/j.tplants.2010.09.008.
- Niu S, Luo Y, Li D, Cao S, Xia J, Li J, Smith MD (2014) Plant growth and mortality
- under climatic extremes: An overview. *Environmental and Experimental Botany* 98:
- 881 13-19. 10.1016/j.envexpbot.2013.10.004.
- Noble DWA, Mayfield MM, Hoffmann AA, Chen Z-H, Lade S, Bai X, Way D,
- 883 **Medlyn BE, Atkin O, Nicotra A et al.** (2025) A systems-modelling approach to
- predict biological responses to extreme heat. *EcoEvoRxiv*: 10.32942/X2BP8N.

- Nolan RH, Collins L, Leigh A, Ooi MKJ, Curran TJ, Fairman TA, Resco de Dios
- **V, Bradstock R** (2021) Limits to post-fire vegetation recovery under climate change.
- 887 Plant, Cell & Environment 44: 3471-3489. 10.1111/pce.14176.
- Nonogaki H, Bassel GW, Bewley JD (2010) Germination—Still a mystery. *Plant*
- 889 *Science* 179: 574-581. 10.1016/j.plantsci.2010.02.010.
- 890 Novick KA, Ficklin DL, Grossiord C, Konings AG, Martínez-Vilalta J, Sadok W,
- Trugman AT, Williams AP, Wright AJ, Abatzoglou JT et al. (2024) The impacts of
- 892 rising vapour pressure deficit in natural and managed ecosystems. Plant, Cell &
- 893 Environment 47: 3561-3589. 10.1111/pce.14846.
- 894 **Ooi MKJ, Auld TD, Denham AJ** (2009) Climate change and bet-hedging:
- interactions between increased soil temperatures and seed bank persistence. Global
- 896 Change Biology 15: 2375-2386. 10.1111/j.1365-2486.2009.01887.x.
- 897 Ooi MKJ, Denham AJ, Santana VM, Auld TD (2014) Temperature thresholds of
- 898 physically dormant seeds and plant functional response to fire: variation among
- species and relative impact of climate change. *Ecology and Evolution* 4: 656-671.
- 900 10.1002/ece3.973.
- 901 Orsenigo S, Mondoni A, Rossi G, Abeli T (2014) Some like it hot and some like it
- 902 cold, but not too much: plant responses to climate extremes. Plant Ecology 215: 677-
- 903 688. 10.1007/s11258-014-0363-6.
- 904 Ørsted M, Jørgensen LB, Overgaard J (2022) Finding the right thermal limit: a
- 905 framework to reconcile ecological, physiological and methodological aspects of
- 906 CT<sub>max</sub> in ectotherms. *Journal of Experimental Biology* 225: jeb244514.
- 907 10.1242/jeb.244514.
- 908 Ørsted M, Willot Q, Olsen AK, Kongsgaard V, Overgaard J (2024) Thermal limits
- of survival and reproduction depend on stress duration: A case study of *Drosophila*
- 910 *suzukii. Ecology Letters* 27: e14421. 10.1111/ele.14421.
- 911 Parmesan C, Hanley ME (2015) Plants and climate change: complexities and
- 912 surprises. *Annals of Botany* 116: 849-864. 10.1093/aob/mcv169.

- Parrotta L, Faleri C, Cresti M, Cai G (2016) Heat stress affects the cytoskeleton
- and the delivery of sucrose synthase in tobacco pollen tubes. *Planta* 243: 43-63.
- 915 10.1007/s00425-015-2394-1.
- Pausas JG, Lamont BB, Keeley JE, Bond WJ (2022) Bet-hedging and best-bet
- 917 strategies shape seed dormancy. *New Phytologist* 236: 1232-1236.
- 918 10.1111/nph.18436.
- 919 **Pécrix Y, Rallo G, Folzer H, Cigna M, Gudin S, Le Bris M** (2011) Polyploidization
- 920 mechanisms: temperature environment can induce diploid gamete formation in Rosa
- 921 sp. Journal of Experimental Botany 62: 3587-3597. 10.1093/jxb/err052.
- 922 **Perez TM**, **Feeley KJ** (2021) Weak phylogenetic and climatic signals in plant heat
- 923 tolerance. *Journal of Biogeography* 48: 91-100. 10.1111/jbi.13984.
- 924 Perez TM, Kullberg A, Rehm E, Feeley K (2025) A database of plant heat
- tolerances and methodological matters. *bioRxiv*: 10.1101/2025.09.14.676187.
- 926 **Perkins SE, Alexander LV** (2013) On the measurement of heat waves. *Journal of*
- 927 Climate 26: 4500-4517. 10.1175/JCLI-D-12-00383.1.
- 928 Perkins-Kirkpatrick SE, Barriopedro D, Jha R, Wang L, Mondal A, Libonati R,
- 929 Kornhuber K (2024) Extreme terrestrial heat in 2023. Nature Reviews Earth &
- 930 Environment 5: 244-246. 10.1038/s43017-024-00536-y.
- 931 **Perkins-Kirkpatrick SE, Lewis SC** (2020) Increasing trends in regional heatwaves.
- 932 Nature Communications 11: 3357. 10.1038/s41467-020-16970-7.
- 933 **Pigott CD, Huntley JP** (1981) Factors controlling the distribution of *Tilia cordata* at
- the nothern limits of its geographical range III. Nature and causes of seed sterility.
- 935 New Phytologist 87: 817-839. 10.1111/j.1469-8137.1981.tb01716.x.
- 936 Poschlod P, Mašková T, Chen SC, Phartyal S, Rosbakh S, Silveira FA,
- 937 Saatkamp A, Dalling JW, Dalziell E, Dickie J et al. (2025) A handbook for
- 938 standardised measurements of regenerative plant functional traits. *EcoEvoRxiv*:
- 939 10.32942/X27W7D.

- 940 Pottinger C, Arnold PA, Bird M, Danzey LM, Geange SR, Herdean A, Nicotra AB,
- Leigh A (2025) Machine learning reveals the effect of leaf temperature extremes on
- shifts in plant photosystem heat tolerance thresholds. *EcoEvoRxiv*:
- 943 10.32942/X2N36T.
- Pressman E, Peet MM, Pharr DM (2002) The effect of heat stress on tomato pollen
- characteristics is associated with changes in carbohydrate concentration in the
- 946 developing anthers. *Annals of Botany* 90: 631-636. 10.1093/aob/mcf240.
- 947 Price MV, Campbell DR, Waser NM, Brody AK (2008) Bridging the generation gap
- 948 in plants: Pollination, parental fecundity, and offspring demography. *Ecology* 89:
- 949 1596-1604. 10.1890/07-0614.1.
- 950 Qian D, Wang M, Niu Y, Yang Y, Xiang Y (2025) Sexual reproduction in plants
- under high temperature and drought stress. *Cell Reports* 44: 115390.
- 952 10.1016/j.celrep.2025.115390.
- 953 **Qiu GY, Yano T, Momii K** (1998) An improved methodology to measure evaporation
- 954 from bare soil based on comparison of surface temperature with a dry soil surface.
- 955 *Journal of Hydrology* 210: 93-105. 10.1016/S0022-1694(98)00174-7.
- 956 Rajapakshe RPVGSW, Tomlinson S, Tudor EP, Turner SR, Elliott CP,
- 957 **Lewandrowski W** (2024) Same, same, but different: dissimilarities in the
- 958 hydrothermal germination performance of range-restricted endemics emerge despite
- 959 microclimatic similarities. *Conservation Physiology* 12: coae009.
- 960 10.1093/conphys/coae009.
- 961 Rang ZW, Jagadish SVK, Zhou QM, Craufurd PQ, Heuer S (2011) Effect of high
- temperature and water stress on pollen germination and spikelet fertility in rice.
- 963 Environmental and Experimental Botany 70: 58-65.
- 964 10.1016/j.envexpbot.2010.08.009.
- 965 Ravikumar RL, Patil BS, Salimath PM (2003) Drought tolerance in sorghum by
- 966 pollen selection using osmotic stress. *Euphytica* 133: 371-376.
- 967 10.1023/A:1025702709095.

968 Rodgers EM, Baldanzi S, Collins M, Dowd WW, Feugere L, Mottola G, 969 Vermandele F, Gomez Isaza DF (2025) Protective multi-stressor interactions in the 970 Anthropocene: Key considerations for investigating cross-tolerance in a conservation 971 context. Conservation Physiology 13: coaf052. 10.1093/conphys/coaf052. 972 Rosbakh S, Pacini E, Nepi M, Poschlod P (2018) An unexplored side of 973 regeneration niche: seed quantity and quality are determined by the effect of 974 temperature on pollen performance. Frontiers in Plant Science 9: 1036. 10.3389/fpls.2018.01036. 975 976 Rosenberger NM, Hemberger JA, Williams NM (2024) Heatwaves exacerbate 977 pollen limitation through reductions in pollen production and pollen vigour. AoB Plants 16: plae045. 10.1093/aobpla/plae045. 978 979 Rosental L, Nonogaki H, Fait A (2014) Activation and regulation of primary 980 metabolism during seed germination. Seed Science Research 24: 1-15. 981 10.1017/S0960258513000391. Rounds CM, Winship LJ, Hepler PK (2011) Pollen tube energetics: respiration, 982 fermentation and the race to the ovule. AoB Plants 2011: plr019. 983 10.1093/aobpla/plr019. 984 985 Ruprecht E, Lukács K, Domokos P, Kuhn T, Fenesi A (2016) Hydration status 986 influences seed fire tolerance in temperate European herbaceous species. Plant 987 Biology 18: 295-300. 10.1111/plb.12394. Saatkamp A, Cochrane A, Commander L, Guja LK, Jimenez-Alfaro B, Larson J, 988 989 Nicotra AB, Poschlod P, Silveira FAO, Cross AT et al. (2019) A research agenda 990 for seed-trait functional ecology. New Phytologist 221: 1764-1775. 991 10.1111/nph.15502. 992 Saini HS, Sedgley M, Aspinall D (1984) Development anatomy in wheat of male 993 sterility induced by heat stress, water deficit or abscisic acid. Australian Journal of 994 Plant Physiology 11: 243-253. 10.1071/PP9840243. 995 Sato S, Kamiyama M, Iwata T, Makita N, Furukawa H, Ikeda H (2006) Moderate 996 increase of mean daily temperature adversely affects fruit set of Lycopersicon

997 esculentum by disrupting specific physiological processes in male reproductive development. Annals of Botany 97: 731-738. 10.1093/aob/mcl037. 998 999 Seneviratne SI, Zhang X, Adnan M, Badi W, Dereczynski C, Di Luca A, Ghosh S, Iskandar I, Kossin J, Lewis S et al. (2021). Weather and Climate Extreme Events 1000 in a Changing Climate. In Masson-Delmotte V, Zhai P, Pirani A, Connors SL, Péan C, 1001 Berger S, Caud N, Chen Y, Goldfarb L, Gomis MI et al. eds, Climate Change 2021: 1002 1003 The Physical Science Basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change, Cambridge 1004 1005 University Press, Cambridge, UK and New York, NY, USA, pp 1513–1766 1006 Sharma PK, Kumar S (2023) Soil Physical Environment and Plant Growth, 1007 Berlin/Heidelberg, Germany: Springer, pp 175-196 1008 Shevtsova A, Graae BJ, Jochum T, Milbau ANN, Kockelbergh F, Beyens L, Nijs I 1009 (2009) Critical periods for impact of climate warming on early seedling establishment 1010 in subarctic tundra. Global Change Biology 15: 2662-2680. 10.1111/j.1365-1011 2486.2009.01947.x. Smolikova G, Leonova T, Vashurina N, Frolov A, Medvedev S (2021) Desiccation 1012 tolerance as the basis of long-term seed viability. International Journal of Molecular 1013 Sciences 22: 101. 10.3390/ijms22010101. 1014 1015 Snider JL, Oosterhuis DM (2011a) How does timing, duration, and severity of heat 1016 stress influence pollen-pistil interactions in angiosperms? Plant Signaling & Behavior 1017 6: 930-933. 10.4161/psb.6.7.15315. Snider JL, Oosterhuis DM, Loka DA, Kawakami EM (2011b) High temperature 1018 1019 limits in vivo pollen tube growth rates by altering diurnal carbohydrate balance in field-grown Gossypium hirsutum pistils. Journal of Plant Physiology 168: 1168-1175. 1020 1021 10.1016/j.jplph.2010.12.011. 1022 Sokolova I (2021) Bioenergetics in environmental adaptation and stress tolerance of 1023 aquatic ectotherms: linking physiology and ecology in a multi-stressor landscape. 1024 Journal of Experimental Biology 224: jeb236802. 10.1242/jeb.236802.

1025 Steinacher G, Wagner J (2012) Effect of temperature on the progamic phase in 1026 high-mountain plants. *Plant Biology* 14: 295-305. 10.1111/j.1438-8677.2011.00498.x. 1027 **Stokes M, Geitmann A** (2025) Screening methods for thermotolerance in pollen. Annals of Botany 135: 71-88. 10.1093/aob/mcae067. 1028 1029 Sultan SE (1995) Phenotypic plasticity and plant adaptation. Acta Botanica 1030 Neerlandica 44: 363-383. 10.1111/j.1438-8677.1995.tb00793.x. 1031 Sultan SE (2000) Phenotypic plasticity for plant development, function and life 1032 history. Trends in Plant Science 5: 537-542. 10.1016/S1360-1385(00)01797-0. 1033 Tangney R, Merritt DJ, Fontaine JB, Miller BP (2019) Seed moisture content as a 1034 primary trait regulating the lethal temperature thresholds of seeds. Journal of 1035 Ecology 107: 1093-1105. 10.1111/1365-2745.13095. Tangney R, Merritt DJ, Miller BP (2022) Environmental factors driving seed 1036 1037 hydration status of soil seed banks and the implications for post-fire recruitment. 1038 Frontiers in Plant Science Volume 12 - 2021: 10.3389/fpls.2021.795003. 1039 Thakur MP, Risch AC, van der Putten WH (2022) Biotic responses to climate extremes in terrestrial ecosystems. iScience 25: 104559. 10.1016/j.isci.2022.104559. 1040 1041 Tomlinson S, Tudor EP, Turner SR, Cross S, Riviera F, Stevens J, Valliere J, Lewandrowski W (2022) Leveraging the value of conservation physiology for 1042 1043 ecological restoration. Restoration Ecology 30: e13616. 10.1111/rec.13616. Tushabe D, Altmann F, Koehler E, Woods S, Kahl S, Rosbakh S (2025a) 1044 1045 Adaptation and acclimation of gametophytic traits to heat stress in a widely distributed wild plant along a steep climatic gradient. Ecology and Evolution 15: 1046 1047 e71199. 10.1002/ece3.71199. 1048 Tushabe D, Rosbakh S (2025b) Patterns and drivers of pollen temperature tolerance. Plant, Cell & Environment 48: 1366-1379. 10.1111/pce.15207. 1049 1050 Unnithan Kumar S, Baker DJ, Maclean IMD, Gaston KJ (2025) Spatial 1051 prioritisation for nature recovery with multiple options for habitat creation. Journal of

Applied Ecology 62: 2688-2700. 10.1111/1365-2664.70144.

- 1053 Valliere JM, Ruscalleda Alvarez J, Cross AT, Lewandrowski W, Riviera F, 1054 Stevens JC, Tomlinson S, Tudor EP, Wong WS, Yong JWH et al. (2022) 1055 Restoration ecophysiology: an ecophysiological approach to improve restoration strategies and outcomes in severely disturbed landscapes. Restoration Ecology 30: 1056 1057 e13571. 10.1111/rec.13571. 1058 van Zanten M, Ai H, Quint M (2021) Plant thermotropism: an underexplored thermal 1059 engagement and avoidance strategy. Journal of Experimental Botany 72: 7414-7420. 10.1093/jxb/erab209. 1060 Wadgymar SM, Ogilvie JE, Inouye DW, Weis AE, Anderson JT (2018) 1061 1062 Phenological responses to multiple environmental drivers under climate change: insights from a long-term observational study and a manipulative field experiment. 1063 1064 New Phytologist 218: 517-529. 10.1111/nph.15029. Winkler DE, Garbowski M, Kožić K, Ladouceur E, Larson J, Martin S, Rosche C, 1065 Roscher C, Slate ML, Korell L (2024) Facilitating comparable research in seedling 1066 1067 functional ecology. Methods in Ecology and Evolution 15: 464-476. 10.1111/2041-1068 210X.14288. 1069 Winkler DE, Lin MY-C, Delgadillo J, Chapin KJ, Huxman TE (2019) Early life history responses and phenotypic shifts in a rare endemic plant responding to 1070 1071 climate change. Conservation Physiology 7: coz076. 10.1093/conphys/coz076. 1072 Zedler PH, Gautier CR, McMaster GS (1983) Vegetation change in response to 1073 extreme events: The effect of a short interval between fires in California chaparral 1074 and coastal scrub. *Ecology* 64: 809-818. 10.2307/1937204. Zhang H, Zhou D, Matthew C, Wang P, Zheng W (2008) Photosynthetic 1075
- Zhu L, Bloomfield KJ, Hocart CH, Egerton JJG, O'Sullivan OS, Penillard A,
  Weerasinghe LK, Atkin OK (2018) Plasticity of photosynthetic heat tolerance in

contribution of cotyledons to early seedling development in *Cynoglossum* 

divaricatum and Amaranthus retroflexus. New Zealand Journal of Botany 46: 39-48.

1076

1077

1078

10.1080/00288250809509752.

1081	plants adapted to thermally contrasting biomes. Plant, Cell & Environment 41: 1251-
1082	1262. 10.1111/pce.13133.
1083	Zhu L, Scafaro AP, Vierling E, Ball MC, Posch BC, Stock F, Atkin OK (2024) Heat
1084	tolerance of a tropical-subtropical rainforest tree species Polyscias elegans: time-
1085	dependent dynamic responses of physiological thermostability and biochemistry.
1086	New Phytologist 241: 715-731. 10.1111/nph.19356.
1087	Zonia L, Munnik T (2004) Osmotically induced cell swelling versus cell shrinking
1088	elicits specific changes in phospholipid signals in tobacco pollen tubes. Plant
1089	Physiology 134: 813-823. 10.1104/pp.103.029454.
1090	
1091	
1092	

# Figure Legends

Figure 1. Four critical life stage transitions during a typical plant's development. Some of the major (non-exhaustive) effects of extreme heat (with associated drying and high VPD) on physiology changes at each stage, and the fitness impacts for individuals and populations. Some moderating factors that could affect the impacts of heat on fitness are inset in grey boxes. Note that pollen development and pollen germination to seed set are grouped together to simplify fitness impacts and the coloured horizontal dividing lines match the colours of life stages used in Figures 2 and 3.

**Figure 2.** Thermal load sensitivity modelling inputs. **a)** Thermal death time (TDT) curves showing the relationship between heat tolerance and exposure duration across the three life stages that differ in heat tolerance and sensitivity. As exposure duration increases, heat tolerance declines but to different extents across the life stages. Thick lines show the mean population-level (*n*=100) responses of the thin lines that are simulated individuals. **b)** Example relationship between net damage and temperature, showing that for a given minute, repair outweighs damage below ~31°C (the 'permissive' range), while damage outweighs repair (the 'stressful' range) above it and damage accumulates exponentially, well beyond the maximum repair rate (black arrow indicates exponential increase).

**Figure 3.** Simulated microclimate temperatures and heat failure probability across life stages. **a)** Temperature at different heights above the ground drastically differ, especially on hot days with high solar radiation input and without rainfall.  $T_{air}$  at 1.2 m is equivalent to standard air temperature weather forecasts. Taking the first day as an example, an imbibed seed right at the soil surface could reach temperatures 20°C above  $T_{air}$ , while a seedling at 1 cm above it could be much cooler but still 8°C above  $T_{air}$ . **b)** By modelling the damage accumulation over time and allowing for repair at permissive temperatures, we can estimate the heat failure probability at the individual- (thin lines) and mean population-level (thick lines) among life stages across the time series. Due to the differences in thermal load sensitivity each individual and life stage has a different heat failure probability under identical (within-stage) environmental conditions.