

No effect of ocean acidification on individual-level variation in behaviour and susceptibility to predation in a Great Barrier Reef damselfish

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Abstract

- 1) Ocean acidification, caused by rising carbon dioxide (CO₂) in the atmosphere, has been reported to negatively impact a wide variety of behaviours in fishes, including activity, exploration, and predator avoidance.
- 2) These effects have been documented at the population level, but many animal species naturally show large and repeatable individual-level differences in behaviour. How environmental stressors, such as ocean acidification, affect behavioural variation at the individual level remains largely unknown but is critically important to understand adaptation given natural selection operates on variation at the individual rather than population level.
- 3) Using a statistical approach allowing variation in means and variation in variance to be modeled within a single framework, we quantified individual-level differences across five behaviours in the coral reef damselfish *Pomacentrus amboinensis* (emergence time, activity level, time spent sheltering, thigmotaxis, novel object inspection). We measured behaviour in a novel environment assay, twice before (CO₂ ~450 µatm) and twice following acclimation to predicted end-of-century ocean acidification conditions (~1,100 µatm).
- 4) Following behavioural assays, we tested individual survival in a live predation experiment. We used predatory rock cod, *Cephalopholis microprion*, acclimated to the same CO₂ treatments as Ambon damsel and examined predictors of survival probability.
- 5) All behaviours in damselfish were moderately and significantly repeatable, with no marked differences in repeatability estimates between the ambient CO₂ and elevated CO₂ treatment groups. Exposure to end-of-century ocean acidification conditions had no effect on any of

the five behaviours measured, both in terms of group means and residual (within-individual) variance.

- 6) The probability of survival in the predation trials was similar for damselfish in the elevated and ambient CO₂ treatment groups. Smaller damselfish as well as those that spent a greater amount of time inspecting a novel object (i.e., bolder individuals) had a lower probability of survival regardless of their CO₂ treatment.
- 7) Our results challenge assumptions about the impacts of ocean acidification on coral reef fish behaviour and susceptibility to predation, both at the population and individual level. They also provide support for a trade-off between boldness and predation risk in fish.

Introduction

The partial pressure of carbon dioxide (CO₂) in the oceans has increased by ~45% over the past century, driving a process known as ocean acidification (OA) (Lüthi *et al.* 2008; Hönlisch *et al.* 2012). This environmental change shows no signs of stopping: forecasts predict that oceanic CO₂ levels at the beginning of the next century will reach those not seen in the last 30 million years (Lüthi *et al.* 2008). Among the range of detrimental impacts reported for animals under ocean acidification, altered species interactions — including between predators and their prey — have received considerable attention (Draper *et al.* 2019). For example, lab-based experiments have suggested that end-of-century levels of CO₂ cause coral reef fishes to become highly attracted to the chemical cues of their predators (Dixon *et al.* 2010; Munday *et al.* 2010), implying increased predation risk under these scenarios (but see Clark *et al.* 2020, which questions the repeatability of this finding). Indeed, small demersal fishes exposed to end-of-century CO₂ and then returned to their natural coral reefs were reported to disappear faster than control individuals (Munday *et al.*

2010; Munday *et al.* 2012). The exact fate of these fish was not recorded, but suggestions for their disappearance included predator attraction, increased activity/boldness, and poorer homing/sheltering ability, all leading to increased predation (Munday *et al.* 2010; Briffa *et al.* 2012; Devine *et al.* 2012; Devine *et al.* 2013). However, a limited number of studies have directly tested the hypothesis that ocean acidification leads to decreased prey survival during a predator interaction. Interestingly, the few laboratory studies on predator-prey interactions between coral reef fishes following exposure to end-of-century levels of CO₂ suggest that the performance of both predators and prey may be altered (e.g. Allan *et al.* 2013; Allan *et al.* 2017). Conversely, and contrary to early findings (e.g. Dixon *et al.* 2010; Munday *et al.* 2010), there is an emerging consensus that ocean acidification may have only negligible effects on coral reef fish behaviour (Clark *et al.* 2020; Clements *et al.* 2022). This uncertainty makes it difficult to predict the impact of future ocean acidification on predator-prey relationships on coral reefs.

Many experiments exploring fish behaviour following exposure to ocean acidification report extremely low inter-individual variability in behavioural responses (e.g. Dixon *et al.* 2010; Munday *et al.* 2010; Clements *et al.* 2022). This is particularly alarming given that the resilience of a species to environmental perturbations depends on variation in individual responses (Tuomainen *et al.* 2011). Phenotypic variation is an inherent biological trait that forms the basis for natural selection and enables an adaptive response to stressors across generations.

Individual behaviours that are repeatable across time and contexts are often referred to as animal “personality” (Wolf *et al.* 2012; Carter *et al.* 2013; Roche *et al.* 2016), and interest in understanding the implications of these individual-level differences on trait evolution has grown over the last 20 years. Behavioural differences influence both the ways in which individuals interact with their environment as well as the outcome of biotic interactions such as predation,

competition, parasitism, cooperation and mate-choice (Wolf *et al.* 2012; Roche *et al.* 2016; Dubois *et al.* 2022; Gradito *et al.* 2024). For example, individuals that are consistently ‘bold’ may expend more energy on activity, increasing their susceptibility to predation but also providing greater foraging opportunities compared with ‘shy’ individuals that remain in hiding for longer durations. Thus, in order to understand how a species might respond to environmental stressors, including ocean acidification, variability in individual responses should be explicitly considered rather than overlooked.

Focusing on individual-level variation as opposed to population mean responses to environmental perturbations has major conservation implications by allowing us to understand the traits responsible for conferring resilience to stressful conditions (Browman 2016; Killen *et al.* 2016). As such, behavioural ecologists are increasingly adopting a reaction norm approach to the study of behavioural variation in order to examine how behaviour and phenotypic plasticity are correlated and/or under selection in a given environment (Dingemanse *et al.* 2010; Dingemanse *et al.* 2013; Roche *et al.* 2016; Fig. 1). Hypothetically, a population-level increase in a behaviour (e.g., activity level) due to elevated CO₂ exposure can occur from all individuals increasing their activity score (Fig. 1 A1), or only some individuals exhibiting a marked increase in activity (Fig. 1 A2, A3). In this example, individuals that do not show a change in activity after elevated CO₂ exposure may be considered tolerant. Identifying tolerant individuals and their associated behaviours is a critical step in predicting how species will respond and adapt to future ocean acidification conditions.

To address several of the knowledge gaps highlighted above, we studied individual-level behavioural differences and reaction norms in the Ambon damselfish, *Pomacentrus amboinensis*, exposed to current day, ambient CO₂ conditions and elevated CO₂ conditions approximating end-

of-century levels of ocean acidification. We also performed a live predation experiment with the same individuals to explicitly test the effect of ocean acidification on fitness while considering the influence of individual behavioural differences on the outcome of predator-prey interactions. Ours is the first study with the explicit aim of understanding how individual-level behavioural variation affects resilience and susceptibility to ocean acidification while incorporating a direct test of the fitness consequences of any CO₂-induced behavioural changes. The approaches used here enable us to address key outstanding questions in this field of research: (1) Are individual-level behavioural differences in Ambon damselfish repeatable?; (2) How does ocean acidification affect population- and individual-level differences in behaviour?; (3) Do effects of ocean acidification on behaviour lead to higher predation rates?; and (4) Do behavioural traits predict susceptibility to predation? We hypothesized that the repeatability of the behaviours we examined would depend on CO₂ treatment and that exposure to elevated CO₂ would induce behavioural differences compared with individuals in the ambient CO₂ treatment at both the population and individual levels. We also hypothesized that behavioural traits and CO₂ exposure would impact survival in a predation trial with bolder fish acclimated to elevated CO₂ being the most likely to be eaten.

Materials and methods

Fish collection and husbandry

Animals were collected and cared for under Marine Parks Permit no. G13/35909.1 issued by the Australian Government's Great Barrier Reef Marine Park Authority. All experiments were approved by the Animal Experimentation Ethics Committee at James Cook University (permit no.: A1924) according to the Australian Code of Practice for the Care and Use of Animals for Scientific Purposes, 7th edition 2007.

All experiments were conducted between January 12th and February 1st, 2016. Post-settlement juvenile *P. amboinensis* were collected by SCUBA divers on January 12-13 from lagoon reefs at Lizard Island on the northern Great Barrier Reef, Australia (14°40' S; 145° 28' E) using hand nets and a barrier net (10 mm stretch monofilament). Fish were transported within 90 minutes of capture to the aquarium facilities at the Lizard Island Research Station (LIRS) in 20 L buckets containing seawater aerated with battery-operated pumps. At LIRS, the fish were placed in 30 L holding tanks receiving seawater at $\sim 750 \text{ ml min}^{-1}$ from one of two flow-through 32 L header tanks (receiving seawater pumped directly from the reef) diffused with ambient air. At 24-48 h post-capture, fish were weighed (Mettler Toledo PL602-S, $d = 0.01 \text{ g}$) and measured for standard length in a water-filled plastic bag to the nearest 0.1 mm with digital callipers. Seventy-four *P. amboinensis* ($33.1 \pm 2.5 \text{ mm}$; $1.43 \pm 0.35 \text{ g}$; mean \pm SD) were then randomly placed individually in 1 L flow-through plastic aquaria ($17 \times 12 \times 7 \text{ cm}$, L \times W \times H) using the randomisation tool in Excel. These aquaria were supplied with seawater pumped directly from the reef. Each aquarium contained a 4 cm long white PVC pipe for shelter. The outer sides of each plastic aquarium were covered in black tape to prevent visual interaction and disturbance. Fish were fed daily with 0.5 ml of a commercial fish flake-saltwater slurry (TetraMin Tropical Flakes, Tetra, Blacksburg, VA). All animals fed readily by day three post-capture. Fish were habituated in their aquaria for three days prior to the start of the experiment. Seawater temperature in the aquaria was $29.5 \pm 1^\circ\text{C}$. Behavioural assessments for Experiment 1 (below) were performed on each individual twice under ambient CO₂ conditions, then half the fish were acclimated to elevated CO₂ for 4-6 days and half were maintained at ambient CO₂ conditions for the same length of time before behavioural assessments were performed another two times on all individuals. Predation

experiments for Experiment 2 (below) were conducted 1-4 days following the last (4th) behavioural trial.

Experiment 1: behavioural trials before and after CO₂ exposure

We examined behavioural differences among *P. amboinensis* using a novel environment assay. This assay is a modified version of the open-field test in which an individual is introduced into an unfamiliar environment, where the environment also included a novel object (Carter *et al.* 2013; Roche *et al.* 2016). Each assay lasted 25 min and was repeated twice (two days between trials) on all fish at ambient CO₂ (458 ± 17.9 μ atm; mean \pm standard deviation) and then two more times (two days between trials) after half the fish were acclimated to elevated CO₂ or maintained at ambient CO₂ conditions (see CO₂ exposure below and Fig. S1). All trials were conducted between 9:00-16:00, and the time of day was randomized across replicates to control for diurnal patterns in activity. Each trial was filmed (top view) with a digital camera (GoPro Hero 3+ black; GoPro, San Mateo USA). The stock lens of the camera was replaced with a commercially available lens to avoid image distortion (4.14 mm f/3.0 86° HFOV 5MP GP41430; Peau Productions Inc., San Diego, USA). We presented a note with the fish ID and trial number at the start of each video (Clark *et al.* 2016). Fish were transferred from their individual aquaria to the experimental arena using their water-filled shelter (the experimenter blocked both ends of the PVC shelter with the palm of their hands). Fish either entered their shelter when the experimenter approached their aquaria or were carefully placed into the shelter by the experimenter. This procedure minimized air exposure and reduced handling stress prior to introduction into the experimental arena. In instances where a fish was accidentally air exposed, we noted the handling time to account for differences in handling stress prior to the start of the experiment.

The experimental arena ($38 \times 28 \times 30$ cm, $L \times W \times H$) had white, opaque sides and was devoid of structure other than a novel object affixed in the centre. Four different novel objects were used (one object was used per arena at any given trial), each measuring approximately 2 cm in diameter and 8 cm in height. The object was changed when the assay was repeated, and the order of objects was randomized between fish and trials such that individual fish were presented with each object only once. The experimenter positioned the shelter and the fish at a fixed, pre-determined location in the arena and exited the room for the duration of the trial to minimize disturbance. The fish was then free to exit its shelter and explore its surroundings over the 25-minute duration of the trial. Water height (seawater pumped directly from the reef) was maintained at 7 cm and the arena was emptied and rinsed with water between each trial. At the end of each trial, the fish was returned to its individual aquarium with its shelter.

To avoid observer bias, we extracted behavioural data from the videos using the automated tracking software ViewPoint (ZebraLab, Lyon, France). For each trial, we recorded five behavioural measurements: 1) emergence time (the time at which the fish's entire body first exited the shelter in seconds); 2) activity level (the distance covered during the trial [25 min] after emergence in mm per minute), 3) sheltering (the time spent in the vicinity of the shelter [within 3 cm; approximately 1 BL] in seconds); 4) thigmotaxis (the time spent within 3 cm [approximately 1 BL] of the arena walls in seconds); and 5) novel object inspection (time spent within 5 cm i.e. <2 body lengths, of the novel object) in seconds.

CO₂ exposure

Once all individuals had repeated the behavioural assay twice under ambient CO₂, half of the fish ($n = 37$) were maintained in ambient water at present-day $p\text{CO}_2$ (ambient; 458 ± 17.9 μatm ; mean

\pm SD) and the other half ($n = 37$) were exposed to end-of-century $p\text{CO}_2$ levels (elevated; $1104 \pm 175 \mu\text{atm}$; mean \pm SD) for 4-6 days. This exposure time was chosen to be consistent with studies reporting altered behaviour in coral reef fishes and reports that longer exposure durations (10 or 25 days) do not alter the response any further (e.g. Munday *et al.* 2010; Allan *et al.* 2013; Munday *et al.* 2013). Control aquaria received seawater from one of two flow-through 32 L header tanks diffused with ambient air and filled at $\sim 750 \text{ ml min}^{-1}$ from seawater pumped from the reef. CO_2 -treatment aquaria received water from one of two additional aerated header tanks (32 L, flow-through filled at $\sim 750 \text{ ml min}^{-1}$) in which the $p\text{CO}_2$ level was gradually increased over 24 h using pH stat computers (Aqua Medic GmbH, Bissendorf, Germany) connected to solenoid valves regulating administration of 100% CO_2 . We monitored $p\text{CO}_2$ in the header tanks and holding tanks multiple times daily using a handheld CO_2 meter (Vaisala GMT 222, Finland) connected to an aspiration pump (Vaisala GM 70, Finland) and a submerged gas-permeable PFTE probe (Qubit Systems, Kingston, Canada) following (Hari *et al.* 2008; Jutfelt *et al.* 2013). The Vaisala CO_2 meter was factory calibrated prior to experiments. The experimental design and CO_2 -dosing system thus followed best practices for ocean acidification research (Reibesell *et al.* 2011; Cornwall *et al.* 2015).

Experiment 2: predation trials

We collected 22 dot-head rock cod (*Cephalopholis microprius*; hereafter rock cod) between January 12th and 17th 2016 from lagoon reefs nearby LIRS using hook-and-line. Rock cod are common predators of small fishes on the Great Barrier Reef (Vail *et al.* 2011). They were transported to the aquarium facility in 20 L aerated buckets within two hours of capture and housed in large ($\sim 300 \text{ l}$) flow-through aquaria. We selected 14 individuals that began feeding within 12h

of capture and moved them to individual 32 L white opaque plastic aquaria ($38 \times 28 \times 30$ cm, L \times W \times H). Each aquarium contained a 12.5 cm long pvc pipe (55 mm diameter) providing shelter for the rock cod at one end of the tank. At the other end, we positioned six pieces of half pvc pipe (55 mm diameter) glued together to create a three-dimensional refuge for *P. amboinensis*. This allowed us to standardize the refuge size and location across all aquaria. The predator shelter and prey refuge were affixed to the bottom of the tank with silicone. To habituate rock cod in these aquaria, we fed them pieces of cut, defrosted pilchard during four days prior to the experiments. During this time, seven rock cod were kept in present-day control water (508 ± 48.4 μ atm; mean \pm SD) and seven were exposed to end-of-century $p\text{CO}_2$ levels (1229 ± 252 μ atm; mean \pm SD) with seawater temperature at approximately $29.5 \pm 1^\circ\text{C}$. CO_2 dosing and measurements followed the procedures as detailed in the *CO₂ Exposure* section above. Two header tanks were used for the control fish and four for the CO_2 fish.

Predator trials commenced when experiment 1 had been completed (January 28 – February 1). Twenty minutes before the onset of a predation trial, we introduced an opaque partition in the middle an aquarium containing a rock cod to restrict *C. microprion* to the area of the tank containing the shelter. One *P. amboinensis* from experiment 1 was introduced to the half of the tank, using its PVC shelter as in Experiment 1, and allowed a 20-minute exploration period. We then lifted the partition, and allowed the predator and prey to interact for 60 min. We filmed interactions with the same cameras as in Experiment 1 a digital camera mounted above the aquarium (GoPro Hero 3+ black fitted with a commercial lens). Prey that were not consumed within 60 min were returned to their holding aquarium. Control and CO_2 -treated *P. amboinensis* were paired with a predator receiving the same experimental treatment using stratified randomization so that each predator received approximately the same number of prey, which

spanned a range of behavioural scores. Each predator was tested between 5 and 6 times with different prey individuals. Each predator was tested only once in a given day and all predators were fed one piece of pilchard daily, in the evening.

Statistical analysis

Experiment 1: behavioural trials before and after CO₂ exposure

We checked the distributions of each of the five behavioural measurements (Fig. S2) and determined appropriate transformations to comply with the normality assumption of model residuals. We used the R package *lme4* (Bates *et al.* 2015) to fit five general Linear Mixed-effects Models (LMM), one for each behavioural measurement. We specified the following fixed effects in each model: treatment (binary variable: ambient vs. elevated CO₂); trial number (ordinal variable from 1 to 4 with trials 3 and 4 corresponding to elevated CO₂ exposure for the treatment group); the interaction between treatment and trial number; and the standard length of fish (continuous variable). The random effects for these models included trial number as random slopes and fish identity as random intercepts. Activity level and thigmotaxis (time spent close to the wall) were square-root-transformed; latency to approach a novel object was log-transformed after adding 0.5; time to emergence from the shelter was log-transformed; time spent in shelter was not transformed (Fig. S2).

We used the *brms* package (Bürkner 2017) to fit double-hierarchical general linear mixed-effects model (DHGLM), one for each of the five behavioural measurements. A DHGLMs comprises two parts, allowing variation in means and variation in variances to be modelled within a single framework (Cleasby *et al.* 2015; O'Dea *et al.* 2022). The mean component of the DHGLMs was identical to the LMMs above, and the variation component included the fixed effects

treatment, trial, and their interaction, as well as the random effect fish identity. The DGHLM allowed us to investigate the effect of CO₂ treatment on both mean and residual variance (or within-individual variance), and to compute an estimate of population-wide variation in predictability (CV_P, the coefficient of variation in predictability) (Cleasby *et al.* 2015). Predictability refers to the level of variability in an individual's behaviour in a given environment (i.e., within-individual variation in behaviour), in contrast to repeatability, which is a measure of the consistency in how individuals differ in their average phenotypes (i.e., between-individual variation in behaviour; see below). CV_P represents standardized variation in within-individual variance among individuals and can serve as a standardized effect size for comparisons across traits, groups, or studies.

We used the *rptR* package (Stoffel *et al.* 2017) with 10,000 bootstrapping iterations to calculate three estimates of repeatability (*R*) for each of the five behavioural measurements: one for all fish, irrespective of treatment group (N=73); one for fish in the ambient CO₂ treatment group (N=37); and one for fish in the elevated CO₂ treatment group (N=36). In total, we computed 15 *R* estimates. *R* ranges between 0 and 1 and is the proportion of the total phenotypic variance attributable to differences between individuals; it provides a standardized measure of the consistency of phenotypes across time or contexts (Nakagawa *et al.* 2010; Roche *et al.* 2016). We note that *R* estimates can be obtained from DGHLMs, but these models could not provide separate estimates for the ambient and elevated CO₂ treatment groups in our study (see Cleasby *et al.* 2015). The *rptR* package uses mixed models from the *lme4* package to obtain the repeatability of behavioural measurements; we obtained ‘adjusted’ *R* estimates by fitting trial number as a fixed effect to control for time-related changes in behaviour (Mitchell *et al.* 2019). We tested whether *R* differed between the ambient and elevated CO₂ treatment groups for each behavioural measurement by calculating contrasts between the two groups (five contrasts in total). When the

95% confidence interval (CI) for a given contrast excluded zero, we considered the two R estimates to be statistically different.

Experiment 2: predation trials

We examined differences in the survival of elevated CO₂-exposed (N=34) and ambient CO₂ (N=34) Ambon damselfish in predation trials with a Cox Proportional Hazards model, using the *survival* package (Therneau 2020). Five Ambon damselfish used in *Experiment 1* were excluded from this analysis due to issues during the predation trials (i.e., camera malfunction, manipulation error). Seven explanatory variables were included in the model: CO₂ treatment, prey body size, and the mean of each of the five behavioural measurements recorded four times per Ambon damselfish in *Experiment 1*. Predator identity was specified as a cluster (i.e., random) variable to account for repeated measurements. We examined the assumption of proportional hazards using `cox.zph()` in the package *survival* and `ggcoxdiagnostics()` in the package *survminer* (Therneau 2020).

We examined differences in the number of predatory attacks resulting in capture for fish in the elevated CO₂ (N=30) and ambient CO₂ (N=28) groups with a Generalized Linear Mixed-effects Model (GLMM), specifying a Poisson error distribution. Fixed and random effects in the model were the same as in the survival analysis. Marginal ($R^2_{\text{GLMM(m)}}$) and conditional ($R^2_{\text{GLMM(c)}}$) R-squared values were computed with `r.squaredGLMM()` in the package *MuMIn* (Barton 2016). We examined model assumptions with a qqplot of residuals and other diagnostic plots of residuals using `simulateResiduals()` in the package *DHARMa* (Hartig 2018).

The assumptions of both models were met although there was an indication of overdispersion and slight quantile deviations for the GLMM (Dispersion test $P=0.040$). All analyses were done

in R 4.2.2 (R Core Team 2023). We used the packages *ggplot2* (Wickham 2016), *tidybayes* (Kay 2024), *emmeans* (Lenth 2025), and *survminer* (Therneau 2020) for data visualization.

Results

Experiment 1: behavioural trials before and after CO₂ exposure

Exposure to projected end-of-century CO₂-induced aquatic acidification had no effect on any of the five behaviours measured in *P. amboinensis*, both in terms of group means (i.e., population-level) (Fig. 2, Table 1) and residual (i.e., within-individual) variance (Fig. 3, Table 1). Thigmotaxis decreased, and time spent near the novel object increased with trial number for fish in both the ambient CO₂ and elevated CO₂ groups, indicating habituation to the experimental setup and/or procedure in both treatment groups (Fig. 2, Table 1). No such population-level trends were observed for emergence time, activity level, or time spent sheltering (Fig. 2, Table 1). Intra-individual variability decreased with trial number for all five behaviours for fish in both the ambient CO₂ and elevated CO₂ groups (Fig. 3), indicating some degree of habituation to the setup/manipulation.

All behaviours were moderately and significantly repeatable, with no marked differences in repeatability estimates between CO₂ groups (Table 2). There was substantial variability in predictability (i.e., within-individual variance) for all five behavioural measurements for both groups (CV_p; see Table 1).

Experiment 2: predation trials

The probability of survival in a predation trial was similar for Ambon damselfish in the elevated CO₂ and ambient groups (Fig. 4), with fish in both treatments having a similar hazard of death (ambient

CO₂ treatment hazard ratio relative to elevated CO₂ treatment group = 0.88, 95% CI = 0.43–1.78, $P=0.719$; Table 3). Body size, emergence time, activity level, time sheltering and thigmotaxis were unrelated to survival probability (Table 3). Ambon damselfish that spent a greater amount of time inspecting a novel object had a lower probability of survival (hazard ratio = 2.66, 95% CI = 1.36–5.20, $P=0.004$; Table 3).

The number of predatory attacks leading to capture was similar for fish in the elevated CO₂ (mean \pm SD: 5.96 ± 3.18) and ambient CO₂ groups (mean \pm SD: 5.39 ± 3.86) (GLMM estimate \pm SE: -0.040 ± 0.182 , $P=0.827$; Table S1, Fig. S3). Predators required more attacks to capture large Ambon damselfish (estimate \pm SE: 0.147 ± 0.067 , $P=0.027$) as well as Ambon damselfish that spent more time near a shelter (estimate \pm SE: 0.490 ± 0.243 , $P=0.044$) and in the vicinity of the novel object during the repeated behavioural trials (estimate \pm SE: 0.386 ± 0.137 , $P=0.005$) (Table S1). The proportion of the total variance explained by the model's fixed effects ($R^2_{\text{GLMM(m)}}$) was 0.19 and the proportion of the variance explained by fixed and random effects ($R^2_{\text{GLMM(m)}}$) was 0.35.

Discussion

Studies on the impacts of end-of-century ocean acidification on coral reef fish behaviour are largely based on comparisons between treatment means rather than individual-level effects. Although it has been argued that some individuals are potentially more tolerant to elevated CO₂ exposure than others (Tresguerres *et al.* 2017; Esbaugh 2018), the extent to which individual behaviour affects responses to ocean acidification has never been tested.

We investigated the repeatability, individual- and population-level differences in five commonly measured behaviours as well as survival outcomes during a predation trial in the context of ocean acidification. All individuals habituated to the experimental set-up as evidenced by

decreased intra-individual variability over the course of the four behavioural trials (Fig. 3). Habituation to experimental arenas is common when repeatedly measuring behaviour, especially in laboratory settings when the timing between assays is short (Martin *et al.* 2008; Blumstein 2016). Nevertheless, we found that all five behaviours measured were significantly repeatable in both treatments, indicating that our assays measured consistent behavioural differences in Ambon damsels. Our repeatability estimates (0.278 – 0.497; Table 2) are relatively high considering that when measured in the laboratory and on ectotherms, estimates tend to be lower than when measured in the field and/or on endotherms (Bell *et al.* 2009). Contrary to widespread assumptions, this behavioural consistency was not affected by CO₂ treatment: both control and elevated CO₂ groups had similar estimates of repeatability in all behaviours, and population reaction-norms followed similar trends in both groups even after CO₂ exposure (scenario B1 in Fig. 1). This suggests that individual behavioural traits are not differentially impacted by CO₂ exposure, providing evidence refuting the suggestion that some individual phenotypes are more tolerant or susceptible to elevated CO₂ exposure than others.

When population-level differences in behaviours were considered, exposure to projected end-of-century CO₂ had no effect on any of the five behaviours measured in Ambon damselfish. Our results are in line with Clark *et al.* (2020) suggesting that CO₂ exposure does not meaningfully alter coral reef fish behaviour. This contrasts with earlier literature showing large effects of CO₂ exposure on many aspects of fish behaviour (reviewed in Briffa *et al.* 2012; Clements *et al.* 2015; Clements *et al.* 2022; Clark *et al.* 2024). A lack of a CO₂ treatment effect is perhaps not surprising since fishes have a well-developed acid-base regulatory system, which allows them to maintain tissue pH under acidified conditions (Ishimatsu *et al.* 2005). In fact, more than a century of research has shown that fish physiology is generally tolerant to elevated CO₂ levels including those that

exceed climate change projections (Clark *et al.* 2024). That behavioural responses are similarly resilient is consistent with this idea, and there is growing empirical evidence of a decline effect in results testing for behavioural differences between control individuals and individuals exposed to ocean acidification (Clements *et al.* 2022; Clements *et al.* 2023). The results from our carefully designed study provide robust evidence that fish behaviour is not altered following exposure to elevated CO₂ whether individual or population-level effects are considered. Although ocean acidification may indeed have negative impacts on some marine life, especially calcifying organisms (Kroeker *et al.* 2010; Clark *et al.* 2024), our results highlight the growing agreement around the behavioural resilience of fishes to ocean acidification, as well as the value of transparent and reproducible research practices in achieving consensus around controversial research topics (O'Dea *et al.* 2021; Roche *et al.* 2022).

CO₂ treatment had no effect on the probability of prey fish surviving in a predation trial; Ambon damselfish from both treatment groups had a similar hazard of death (Fig 4). The number of predatory attacks leading to capture was also similar between treatment groups. These results are consistent with several other studies suggesting that CO₂ exposure has limited effects on predator-prey interactions in coral reef fishes (Allan *et al.* 2017; Ferrari *et al.* 2017; McCormick *et al.* 2018), refuting early assertions, based on indirect evidence, that ocean acidification dramatically increases prey susceptibility to predation (e.g. Munday *et al.* 2010; Devine *et al.* 2013). Indeed, Allan *et al.* (2013) found that when both the predator and prey had been exposed to elevated CO₂ (as in our study), the capture rate was similar to that of the control group (where both predators and prey were exposed to present-day CO₂). Similarly, a study exploring the effect of elevated CO₂ on predation in freshwater fishes found no effect of CO₂ treatment on predator success or time to survival in prey (Midway *et al.* 2017). Our results further highlight the importance of

behavioural tests on interacting animals when assessing the impact of anthropogenic stressors such as ocean acidification on complex ecological interactions such as predation.

Although elevated CO₂ had no impact on survival outcomes during predation trials in our study, we did observe small differences in the survival of Ambon damsel based on individual traits. We found that predators required more attacks to capture large Ambon damsel, which may be due to minor differences in mouth gape requirements of the predators and/or higher absolute burst swimming speeds in larger versus smaller Ambon damsel conspecifics (Taylor *et al.* 1985). While emergence time, activity level, time spent sheltering, and thigmotaxis behaviours were not related to individual survival, Ambon damsels that spent a greater amount of time inspecting the novel object had a lower probability of survival. This trend was independent of CO₂ treatment. Our results are consistent with the notion that bolder individuals face higher predation risk than shy individuals (Hulthén *et al.* 2017). However, as four out of five behaviours tested had no relationship with survival, the role of individual behavioural traits in predicting predation risk should be interpreted with caution. Furthermore, although we predicted higher mortality in bolder fish from the elevated CO₂ treatment group compared to the control group, the effect of behaviour on survival outcome was independent of CO₂ exposure. This result runs counter to those of several studies that have linked elevated CO₂ to increased boldness in fishes (e.g. Ferrari *et al.* 2011; Munday *et al.* 2014; Ferrari *et al.* 2017; Mitchell *et al.* 2022). Again, our results align with more recent studies demonstrating negligible effects of ocean acidification on fish behaviour (Clark *et al.* 2020; Clements *et al.* 2022; Clements *et al.* 2023; Clark *et al.* 2024) and do not support the notion of differences in individual tolerance or susceptibility to ocean acidification within a fish population.

Conclusion

Ocean acidification presents a growing threat to marine ecosystems globally as a result of anthropogenic activities. Concerns for how this change in water chemistry will impact aquatic organisms is warranted and an important area of continued research. Yet, there are limited resources to understand the complexities of ocean acidification impacts on marine life and thus researchers should focus their time and effort on species, lifestages, interactions and physiological processes that are most likely to suffer negative consequences as a result of ocean acidification in order to inform decision making and conservation strategies (Buxton *et al.* 2021). Based on our results, as well as the growing body of evidence from different species and study systems, we suggest that fish behaviour is not likely to be meaningfully impacted by near-future ocean acidification levels. Thus, future research should prioritize studying how other aspects of elevated atmospheric CO₂ impact marine fishes, including the rapid rise in the temperature of the global oceans.

Author contributions

DGR, BSR, JS, TDC and SAB conceived the study and designed methodology; DGR, BSR and SAB collected the data; TDC processed the data; DGR and SN analysed the data; DGR and SAB led the writing of the manuscript with substantial input from JS and TDC. All authors contributed critically to the drafts and gave final approval for publication. Our study brings together authors from a number of different countries, including two scientists based in Australia where the study was carried out.

Data and code availability

The data and analytical code to reproduce the results of the study are publicly available on the Open Science Framework and were shared with the editor and reviewers on manuscript submission: <https://doi.org/10.17605/OSF.IO/CZVSF>.

Conflicts of Interest

None to declare.

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References

- Allan, B.J.M., Domenici, P., McCormick, M.I., Watson, S.-A. & Munday, P.L. (2013) Elevated CO₂ affects predator-prey interactions through altered performance. *PLoS ONE*, **8**, e58520.
- Allan, B.J.M., Domenici, P., Watson, S.A., Munday, P.L. & McCormick, M.I. (2017) Warming has a greater effect than elevated CO₂ on predator-prey interactions in coral reef fish. *Proceedings of the Royal Society B: Biological Sciences*, **284**, 20170784.
- Barton, K. (2016) Package "MuMIn": Multi-modal inference. *R package version 1.15.6*, <https://cran.r-project.org/web/packages/MuMIn/MuMIn.pdf>.
- Bates, D., Mächler, M., Bolker, B. & Walker, S. (2015) Fitting linear mixed-effects models using lme4. *Journal of Statistical Software*, **67**, 1 - 48.
- Bell, A.M., Hankison, S.J. & Laskowski, K.L. (2009) The repeatability of behaviour: a meta-analysis. *Animal Behaviour*, **77**, 771-783.
- Blumstein, D.T. (2016) Habituation and sensitization: new thoughts about old ideas. *Animal Behaviour*, **120**, 255-262.
- Briffa, M., de la Haye, K. & Munday, P.L. (2012) High CO₂ and marine animal behaviour: potential mechanisms and ecological consequences. *Marine Pollution Bulletin*, **64**, 1519-1528.
- Browman, H.I. (2016) Applying organized scepticism to ocean acidification research. *ICES Journal of Marine Science*, **73**, 529-536.
- Bürkner, P.-C. (2017) brms: An R package for Bayesian multilevel models using Stan. *Journal of Statistical Software*, **80**, 1 - 28.

- Buxton, R.T., Nyboer, E.A., Pigeon, K.E., Raby, G.D., Rytwinski, T., Gallagher, A.J., Schuster, R., Lin, H.-Y., Fahrig, L., Bennett, J.R., Cooke, S.J. & Roche, D.G. (2021) Avoiding wasted research resources in conservation science. *Conservation Science and Practice*, **3**, e329.
- Carter, A.J., Feeney, W.E., Marshall, H.H., Cowlshaw, G. & Heinsohn, R. (2013) Animal personality: what are behavioural ecologists measuring? *Biological Reviews*, **88**, 465-475.
- Clark, T.D., Binning, S.A., Raby, G.D., Speers-Roesch, B., Sundin, J., Jutfelt, F. & Roche, D.G. (2016) Scientific Misconduct: the elephant in the lab. A Response to Parker *et al.* *Trends in Ecology & Evolution*, **31**, 899-900.
- Clark, T.D., Clements, J.C., Jutfelt, F. & Sundin, J. (2024) Ocean acidification. *Behavioural Responses to a Changing World* (eds B.B.M. Wong & U. Candolin), pp. 72. Oxford University Press.
- Clark, T.D., Raby, G.D., Roche, D.G., Binning, S.A., Speers-Roesch, B., Jutfelt, F. & Sundin, J. (2020) Ocean acidification does not impair the behaviour of coral reef fishes. *Nature*, **577**, 370-375.
- Cleasby, I.R., Nakagawa, S. & Schielzeth, H. (2015) Quantifying the predictability of behaviour: statistical approaches for the study of between-individual variation in the within-individual variance. *Methods in Ecology and Evolution*, **6**, 27-37.
- Clements, J.C. & Hunt, H.L. (2015) Marine animal behaviour in a high CO₂ ocean. *Marine Ecology Progress Series*, **536**, 259-279.
- Clements, J.C., Sundin, J., Clark, T.D. & Jutfelt, F. (2022) Meta-analysis reveals an extreme “decline effect” in the impacts of ocean acidification on fish behavior. *Plos Biology*, **20**, e3001511.

- Clements, J.C., Sundin, J., Clark, T.D. & Jutfelt, F. (2023) Extreme original data yield extreme decline effects. *Plos Biology*, **21**, e3001996.
- Cornwall, C.E. & Hurd, C.L. (2015) Experimental design in ocean acidification research: problems and solutions. *ICES Journal of Marine Science*, **73**, 572-581.
- Devine, B.M. & Munday, P.L. (2013) Habitat preferences of coral-associated fishes are altered by short-term exposure to elevated CO₂. *Marine Biology*, **160**, 1955-1962.
- Devine, B.M., Munday, P.L. & Jones, G.P. (2012) Homing ability of adult cardinalfish is affected by elevated carbon dioxide. *Oecologia*, **168**, 269-276.
- Dingemanse, N.J. & Dochtermann, N.A. (2013) Quantifying individual variation in behaviour: mixed-effect modelling approaches. *Journal of Animal Ecology*, **82**, 39-54.
- Dingemanse, N.J., Kazem, A.J., Réale, D. & Wright, J. (2010) Behavioural reaction norms: animal personality meets individual plasticity. *Trends in Ecology & Evolution*, **25**, 81-89.
- Dixon, D.L., Munday, P.L. & Jones, G.P. (2010) Ocean acidification disrupts the innate ability of fish to detect predator olfactory cues. *Ecology Letters*, **13**, 68-75.
- Draper, A.M. & Weissburg, M.J. (2019) Impacts of global warming and elevated CO₂ on sensory behavior in predator-prey interactions: a review and synthesis. *Frontiers in Ecology and Evolution*, **7**.
- Dubois, F. & Binning, S.A. (2022) Predation and parasitism as determinants of animal personalities. *Journal of Animal Ecology*, **91**, 1918-1928.
- Esbaugh, A.J. (2018) Physiological implications of ocean acidification for marine fish: emerging patterns and new insights. *Journal of Comparative Physiology B*, **188**, 1-13.
- Ferrari, M.C.O., Dixon, D.L., Munday, P.L., McCormick, M.I., Meekan, M.G., Sih, A. & Chivers, D.P. (2011) Intrageneric variation in antipredator responses of coral reef fishes

- affected by ocean acidification: implications for climate change projections on marine communities. *Global Change Biology*, **17**, 2980-2986.
- Ferrari, M.C.O., McCormick, M.I., Watson, S.-A., Meekan, M.G., Munday, P.L. & Chivers, D.P. (2017) Predation in high CO₂ waters: prey fish from high-risk environments are less susceptible to ocean acidification. *Integrative and Comparative Biology*, **57**, 55-62.
- Gradito, M., Dubois, F., Noble, D.W.A. & Binning, S.A. (2024) Double trouble: host behaviour influences and is influenced by co-infection with parasites. *Animal Behaviour*, **215**, 31-44.
- Hari, P., Pumpanen, J., Huotari, J., Kolari, P., Grace, J., Vesala, T. & Ojala, A. (2008) High-frequency measurements of productivity of planktonic algae using rugged nondispersive infrared carbon dioxide probes. *Limnology and Oceanography: Methods*, **6**, 347-354.
- Hartig, F. (2018) DHARMa: residual diagnostics for hierarchical (multi-level/mixed) regression models. *R Packag version 020*, <https://cran.r-project.org/package=DHARM>.
- Hönisch, B., Ridgwell, A., Schmidt, D.N., Thomas, E., Gibbs, S.J., Sluijs, A., Zeebe, R., Kump, L., Martindale, R.C., Greene, S.E., Kiessling, W., Ries, J., Zachos, J.C., Royer, D.L., Barker, S., Marchitto, T.M., Moyer, R., Pelejero, C., Ziveri, P., Foster, G.L. & Williams, B. (2012) The geological record of ocean acidification. *Science*, **335**, 1058-1063.
- Hulthén, K., Chapman, B.B., Nilsson, P.A., Hansson, L.-A., Skov, C., Brodersen, J., Vinterstare, J. & Brönmark, C. (2017) A predation cost to bold fish in the wild. *Scientific Reports*, **7**, 1239.
- Ishimatsu, A., Hayashi, M., Lee, K.-S., Kikkawa, T. & Kita, J. (2005) Physiological effects on fishes in a high-CO₂ world. *Journal of Geophysical Research: Oceans*, **110**.

- Jutfelt, F. & Hedgärde, M. (2013) Atlantic cod actively avoid CO₂ and predator odour, even after long-term CO₂ exposure. *Frontiers in Zoology*, **10**, 81.
- Kay, M. (2024) tidybayes: Tidy Data and Geoms for Bayesian Models. *R package version 3.0.7*, <http://mjskay.github.io/tidybayes/>.
- Killen, S.S., Adriaenssens, B., Marras, S., Claireaux, G. & Cooke, S.J. (2016) Context dependency of trait repeatability and its relevance for management and conservation of fish populations. *Conservation Physiology*, **4**, cow007.
- Kroeker, K.J., Kordas, R.L., Crim, R.N. & Singh, G.G. (2010) Meta-analysis reveals negative yet variable effects of ocean acidification on marine organisms. *Ecology Letters*, **13**, 1419-1434.
- Lenth, R.V. (2025) emmeans: Estimated Marginal Means, aka Least-Squares Means. *R package version 1.10.7*, <https://rvlenth.github.io/emmeans/>.
- Lüthi, D., Le Floch, M., Bereiter, B., Blunier, T., Barnola, J.-M., Siegenthaler, U., Raynaud, D., Jouzel, J., Fischer, H., Kawamura, K. & Stocker, T.F. (2008) High-resolution carbon dioxide concentration record 650,000–800,000 years before present. *Nature*, **453**, 379.
- Martin, J.G.A. & Réale, D. (2008) Temperament, risk assessment and habituation to novelty in eastern chipmunks, *Tamias striatus*. *Animal Behaviour*, **75**, 309-318.
- McCormick, M.I., Watson, S.-A., Simpson, S.D. & Allan, B.J.M. (2018) Effect of elevated CO₂ and small boat noise on the kinematics of predator-prey interactions. *Proceedings of the Royal Society B: Biological Sciences*, **285**, 20172650.
- Midway, S.R., Hasler, C.T., Wagner, T. & Suski, C.D. (2017) Predation of freshwater fish in environments with elevated carbon dioxide. *Marine and Freshwater Research*, **68**, 1585-1592.

- Mitchell, A., Booth, D.J. & Nagelkerken, I. (2022) Ocean warming and acidification degrade shoaling performance and lateralization of novel tropical–temperate fish shoals. *Global Change Biology*, **28**, 1388-1401.
- Mitchell, D.J., Dujon, A.M., Beckmann, C. & Biro, P.A. (2019) Temporal autocorrelation: a neglected factor in the study of behavioral repeatability and plasticity. *Behavioral Ecology*, **31**, 222-231.
- Munday, P.L., Cheal, A.J., Dixon, D.L., Rummer, J.L. & Fabricius, K.E. (2014) Behavioural impairment in reef fishes caused by ocean acidification at CO₂ seeps. *Nature Climate Change*, **4**, 487-492.
- Munday, P.L., Dixon, D.L., McCormick, M.I., Meekan, M., Ferrari, M.C.O. & Chivers, D.P. (2010) Replenishment of fish populations is threatened by ocean acidification. *Proceedings of the National Academy of Sciences*, **107**, 12930-12934.
- Munday, P.L., McCormick, M.I., Meekan, M., Dixon, D.L., Watson, S.-A., Chivers, D.P. & Ferrari, M.C. (2012) Selective mortality associated with variation in CO₂ tolerance in a marine fish. *Ocean Acidification*, **1**, 1-5.
- Munday, P.L., Pratchett, M.S., Dixon, D.L., Donelson, J.M., Endo, G.G., Reynolds, A.D. & Knuckey, R. (2013) Elevated CO₂ affects the behavior of an ecologically and economically important coral reef fish. *Marine Biology*, **160**, 2137-2144.
- Nakagawa, S. & Schielzeth, H. (2010) Repeatability for Gaussian and non-Gaussian data: a practical guide for biologists. *Biological Reviews*, **85**, 935-956.
- O'Dea, R.E., Noble, D.W.A. & Nakagawa, S. (2022) Unifying individual differences in personality, predictability and plasticity: A practical guide. *Methods in Ecology and Evolution*, **13**, 278-293.

- O'Dea, R.E., Parker, T.H., Chee, Y.E., Culina, A., Drobniak, S.M., Duncan, D.H., Fidler, F., Gould, E., Ihle, M., Kelly, C.D., Lagisz, M., Roche, D.G., Sánchez-Tójar, A., Wilkinson, D.P., Wintle, B.C. & Nakagawa, S. (2021) Towards open, reliable, and transparent ecology and evolutionary biology. *BMC Biology*, **19**, 68.
- Reibesell, U., Fabry, V.J., Hansson, L. & Gattuso, J.-P. (2011) Guide to best practices for ocean acidification research and data reporting. pp. 258. Luxembourg.
- Roche, D.G., Careau, V. & Binning, S.A. (2016) Demystifying animal ‘personality’ (or not): why individual variation matters to experimental biologists. *Journal of Experimental Biology*, **219**, 3832-3843.
- Roche, D.G., Raby, G.D., Norin, T., Ern, R., Scheuffele, H., Skeeles, M., Morgan, R., Andreassen, A.H., Clements, J.C., Louissaint, S., Jutfelt, F., Clark, T.D. & Binning, S.A. (2022) Paths towards greater consensus building in experimental biology. *Journal of Experimental Biology*, **225**, jeb243559.
- Stoffel, M.A., Nakagawa, S. & Schielzeth, H. (2017) rptR: repeatability estimation and variance decomposition by generalized linear mixed-effects models. *Methods in Ecology and Evolution*, **8**, 1639-1644.
- Taylor, E.B. & McPhail, J.D. (1985) Burst swimming and size-related predation of newly emerged coho salmon *Oncorhynchus kisutch*. *Transactions of the American Fisheries Society*, **114**, 546-551.
- R Core Team (2023) R: A Language and Environment for Statistical Computing. *R Foundation for Statistical Computing, Vienna, Austria*.
- Therneau, T. (2020) A Package for Survival Analysis in R. . <https://CRAN.R-project.org/package=survival>.

Tresguerres, M. & Hamilton, T.J. (2017) Acid–base physiology, neurobiology and behaviour in relation to CO₂-induced ocean acidification. *Journal of Experimental Biology*, **220**, 2136-2148.

Tuomainen, U. & Candolin, U. (2011) Behavioural responses to human-induced environmental change. *Biological Reviews*, **86**, 640-657.

Vail, A.L. & McCormick, M.I. (2011) Metamorphosing reef fishes avoid predator scent when choosing a home. *Biology Letters*, **7**, 921-924.

Wickham, H. (2016) *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York.

Wolf, M. & Weissing, F.J. (2012) Animal personalities: consequences for ecology and evolution. *Trends in Ecology & Evolution*, **27**, 452-461.

1 TABLES

2 **Table 1.** Estimates and 95% confidence intervals (CI) for predictors included in five double hierarchical linear mixed-effects models
3 (DHGLMs) assessing how end-of-century CO₂-induced aquatic acidification affects five behavioural measurements in the Ambon
4 damselfish (*Pomacentrus amboinensis*): activity level (Activity), emergence time (Emergence), time spent inspecting a novel object
5 (Novel), time sheltering (Sheltering), time close to the arena walls (Thigmotaxis). Predictors 1-4 relate to the mean component of the
6 DHGLMs and predictors 5-7 relate to the dispersion component (i.e., residual variances for individuals). Estimates in bold have a 95%
7 credible interval non-overlapping zero.

8

| Predictor | Activity (95% CI) | Emergence (95% CI) | Novel (95% CI) | Sheltering (95% CI) | Thigmotaxis (95% CI) |
|-----------------|-------------------------|-----------------------|-------------------------|-------------------------|-------------------------|
| Intercept | 57.74 (53.40, 61.90) | 2.45 (1.74, 3.18) | 0.70 (0.39, 1.02) | 32.17 (27.69, 36.69) | 3.55 (3.17, 3.92) |
| sigma_Intercept | 2.37 (2.10, 2.63) | 0.73 (0.50, 0.95) | -0.44 (-0.66, -0.22) | 2.41 (2.19, 2.62) | 0.23 (0.01, 0.43) |

| | | | | | | |
|--------------------------------|--------------------------------|--------------------------------|-----------------------------|-------------------------|--------------------------------|---|
| 1. groupcontrol | 1.05 (-4.65, 7.00) | 0.09 (-0.92, 1.08) | 0.27 (-0.17, 0.73) | -1.37 (-7.76, 4.90) | -0.48 (-1.01, 0.04) | 9 |
| 2. trial | -0.96 (-6.60, 4.83) | -0.13 (-0.80, 0.56) | 0.51 (0.18, 0.83) | 2.53 (-3.08, 8.17) | -0.77 (-1.31, -0.21) | |
| 3. SL | 0.24 (-5.60, 6.13) | 0.6 (-0.40, 1.61) | 0.19 (-0.24, 0.63) | -5.09 (-11.48, 1.46) | 0.16 (-0.36, 0.66) | |
| 4. groupcontrol:trial | 3.42 (-4.51, 11.30) | -1.64 (-2.54, -0.70) | 0.24 (-0.21, 0.69) | -4.5 (-12.29, 3.09) | 0.30 (-0.48, 1.07) | |
| 5. sigma_groupcontrol | 0.01 (-0.34, 0.37) | -0.13 (-0.43, 0.19) | 0.06 (-0.24, 0.36) | -0.06 (-0.35, 0.22) | -0.12 (-0.39, 0.15) | |
| 6. sigma_trial | -0.56 (-1.01, -0.11) | -0.2 (-0.58, 0.15) | -0.24 (-0.69, 0.23) | -0.22 (-0.66, 0.26) | -0.24 (-0.64, 0.16) | |
| 7. sigma_groupcontrol:trial | 0.15 (-0.48, 0.80) | 0.27 (-0.25, 0.82) | 0.13 (-0.5, 0.78) | -0.34 (-1.01, 0.28) | -0.23 (-0.78, 0.31) | |

Table 2. Estimates of repeatability (R) and variation in predictability (CVp) for five behavioural traits measured experimentally four times in the Ambon damselfish, *Pomacentrus amboinensis*. Behaviour was assessed in all fish under ambient CO₂ conditions twice, and thereafter twice more after having exposed half of the fish to elevated CO₂. The difference in R between the ambient CO₂ group (control) and elevated CO₂ treatment group for each behaviour is presented as a contrast (Contrast). 95% confidence intervals are indicated in parentheses. The behaviours measured were activity level (Activity), emergence time (Emergence), time spent inspecting a novel object (Novel), time sheltering (Sheltering), and time close to the arena walls (Thigmotaxis).

| Behaviour | Overall (95% CI) | R Control (95% CI) | R Treatment (95% CI) | Contrast (95% CI) | CVp (95% CI) |
|--------------|----------------------------|----------------------------|----------------------------|------------------------------|----------------------------|
| Activity | 0.278 (0.149, 0.402) | 0.255 (0.068, 0.427) | 0.303 (0.113, 0.472) | 0.046 (-0.211, 0.302) | 0.805 (0.625, 0.997) |
| Emergence | 0.298 (0.169, 0.424) | 0.317 (0.124, 0.488) | 0.303 (0.118, 0.478) | -0.015 (-0.269, 0.247) | 0.754 (0.533, 0.967) |
| Novel object | 0.497 (0.363, 0.605) | 0.568 (0.390, 0.703) | 0.405 (0.210, 0.571) | -0.162 (-0.403, 0.082) | 0.636 (0.324, 0.856) |

| | | | | | |
|-------------|---------|---------|---------|----------|---------|
| Shelter | 0.381 | 0.427 | 0.340 | -0.085 | 0.545 |
| | (0.247, | (0.230, | (0.147, | (-0.335, | (0.202, |
| | 0.502) | 0.588) | 0.509) | 0.174) | 0.775) |
| Thigmotaxis | 0.245 | 0.243 | 0.225 | -0.020 | 0.568 |
| | (0.118, | (0.066, | (0.043, | (-0.271, | (0.175, |
| | 0.368) | 0.413) | 0.398) | 0.229) | 0.805) |

Table 3. Hazard ratios (HR) for fixed-effects predictors in a Cox Proportional Hazards model examining differences in survival probability for CO₂-exposed (N=34) and control (N=34) Ambon damselfish (*Pomacentrus amboinensis*) (prey). A hazard ratio lower than 1 indicates reduced hazard of death whereas a hazard ratio greater than 1 indicates an increased hazard of death. 95% confidence intervals, Z scores, and p-values are indicated. The predator (*Cephalopholis microprion*) was exposed to the same conditions of water chemistry as the prey.

| Predictor | HR | 95% CI | Z | P |
|----------------------|------|------------|--------|-------|
| Prey standard length | 1.11 | 0.87, 1.42 | 0.833 | 0.405 |
| Treatment_control | 0.88 | 0.43, 1.78 | -0.360 | 0.719 |
| Emergence time | 1.11 | 0.84, 1.47 | 0.746 | 0.456 |
| Activity level | 1.36 | 0.87, 2.14 | 1.349 | 0.177 |
| Time sheltering | 2.99 | 0.90, 9.96 | 1.785 | 0.074 |
| Thigmotaxis | 1.60 | 0.69, 3.71 | 1.084 | 0.278 |
| Novel object | 2.66 | 1.36, 5.20 | 2.871 | 0.004 |

FIGURES

Figure 1. Examples of hypothetical behavioural reaction norms from exposure to high CO₂. On average, individuals might exhibit either (A) an increase or (B) no change in a given behavioural trait (e.g. activity, anxiety). Different combinations of individual-level responses can underpin population-level patterns, where individuals might or might not respond similarly to high CO₂ exposure. A population-level increase in behaviour can result from (A1) all individuals increasing their behaviour or (A2-A3) some individuals exhibiting a sharp increase in behaviour and others, no change in behaviour. A lack of population-level change in behaviour can result from: (B1) the behaviour of all individuals remaining unchanged; (B2) differences in the behaviour of individuals accentuating; or (B3) no consistent changes in behaviour across individuals.

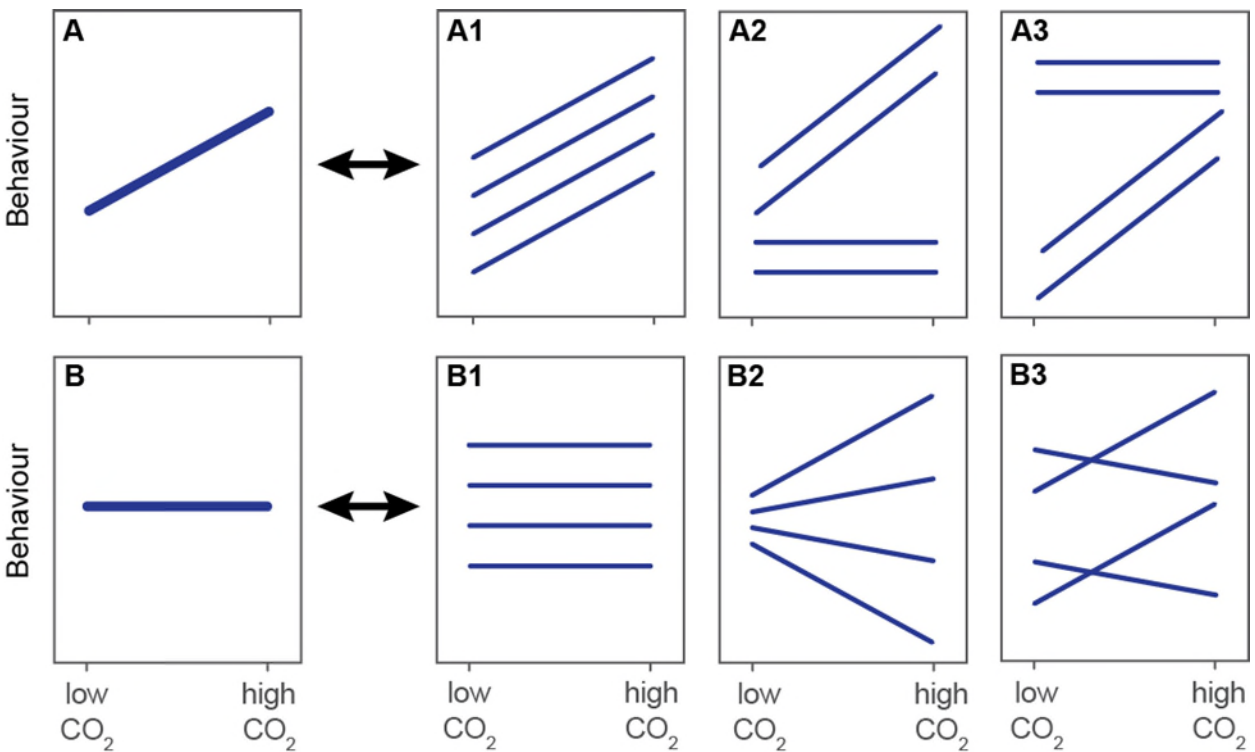
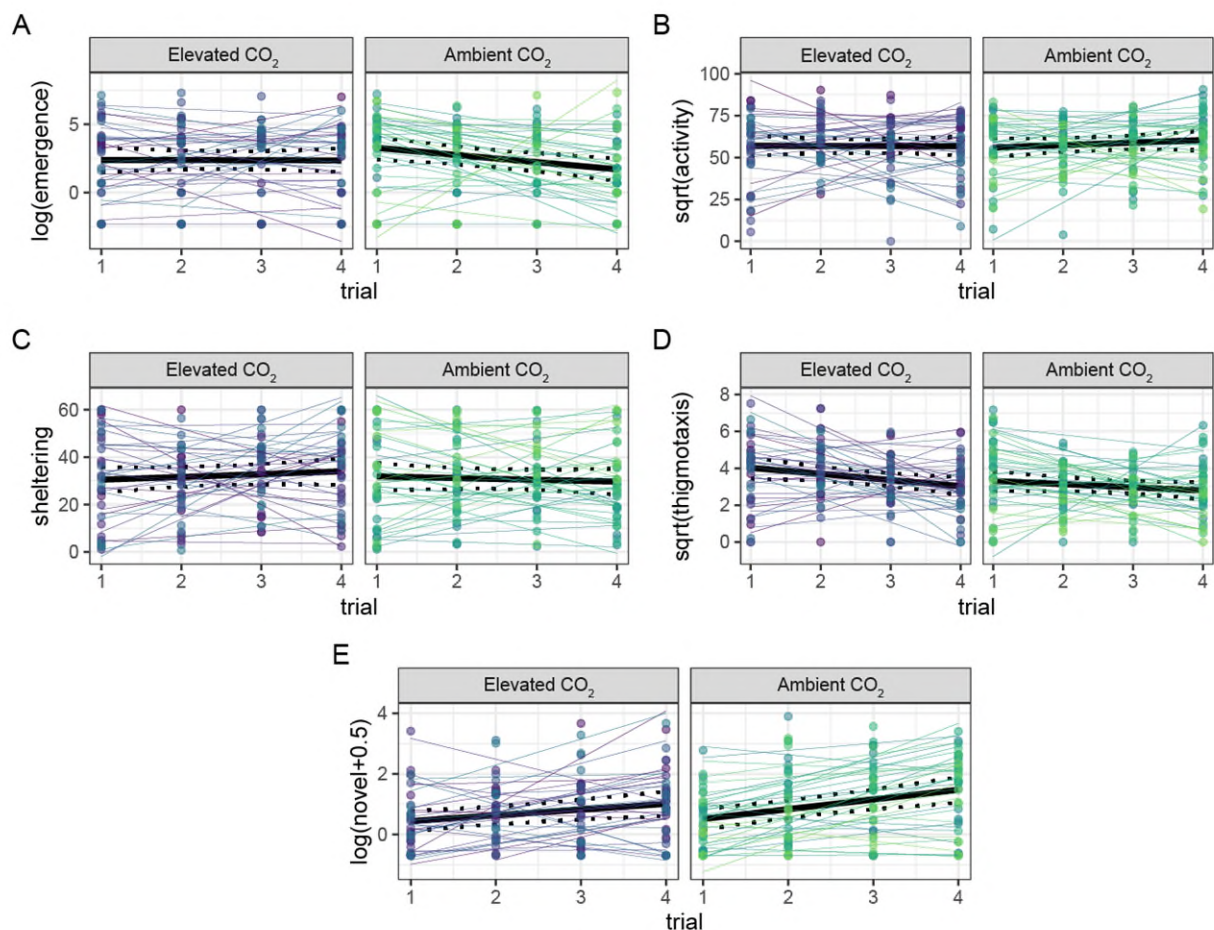
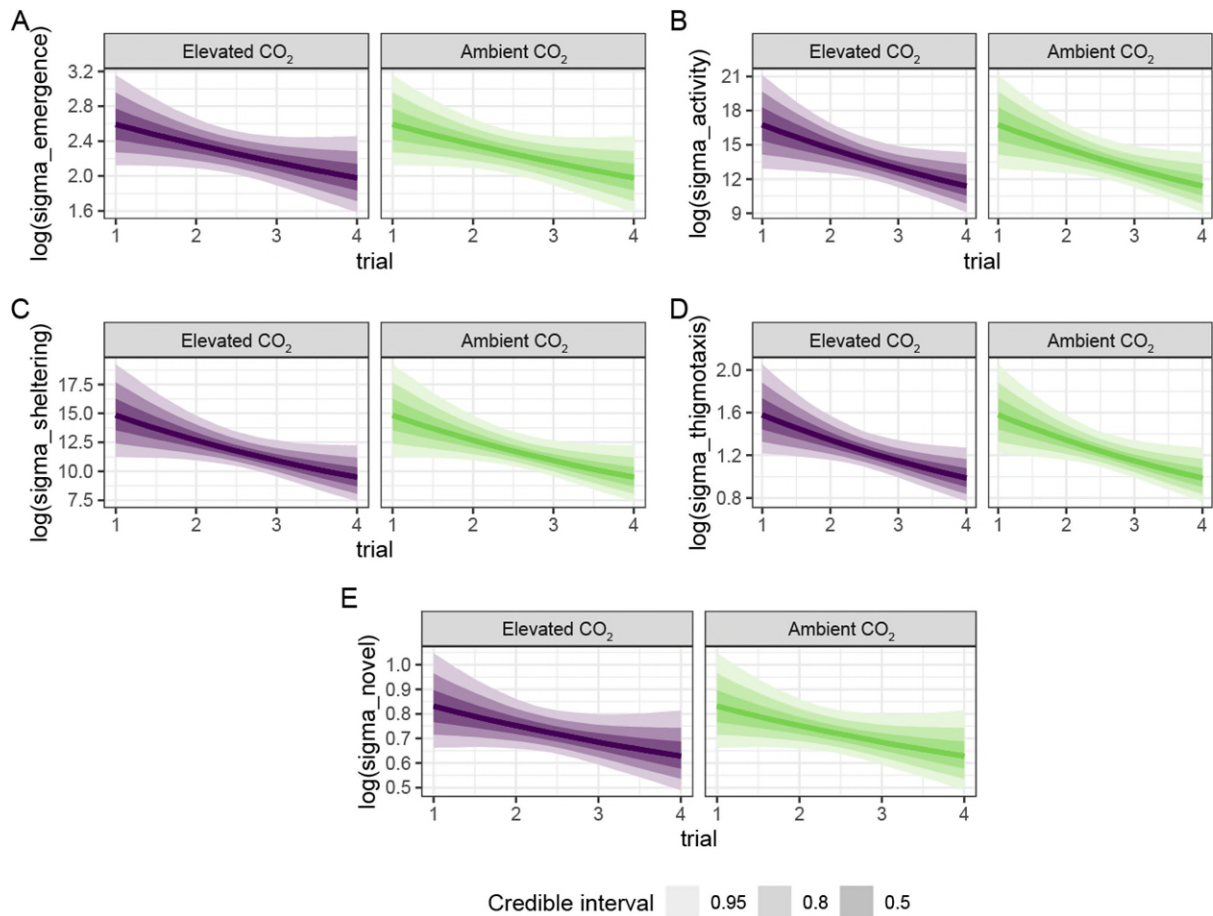


Figure 2. Reaction norm plots of 5 behavioural measurements recorded across 4 trials for Ambon damselfish (*P. amboinensis*) exposed to present-day levels of dissolved CO₂ (Ambient CO₂; n=36) and projected end-of-century CO₂-induced aquatic acidification (Elevated CO₂, n=37): (A) emergence, (B) activity, (C) time sheltering, (D) thigmotaxis and (E) inspection of a novel object. Fish in the control group were exposed to present-day CO₂ levels in all 4 trials, whereas fish in the CO₂ group were exposed to present-day CO₂ levels in trials 1-2 and projected end-of-century CO₂ levels in trials 3-4. Data points and coloured lines represent individual fish; the black line is the population-level (i.e., mean) response with the 95% CI credible interval indicated by dotted lines.

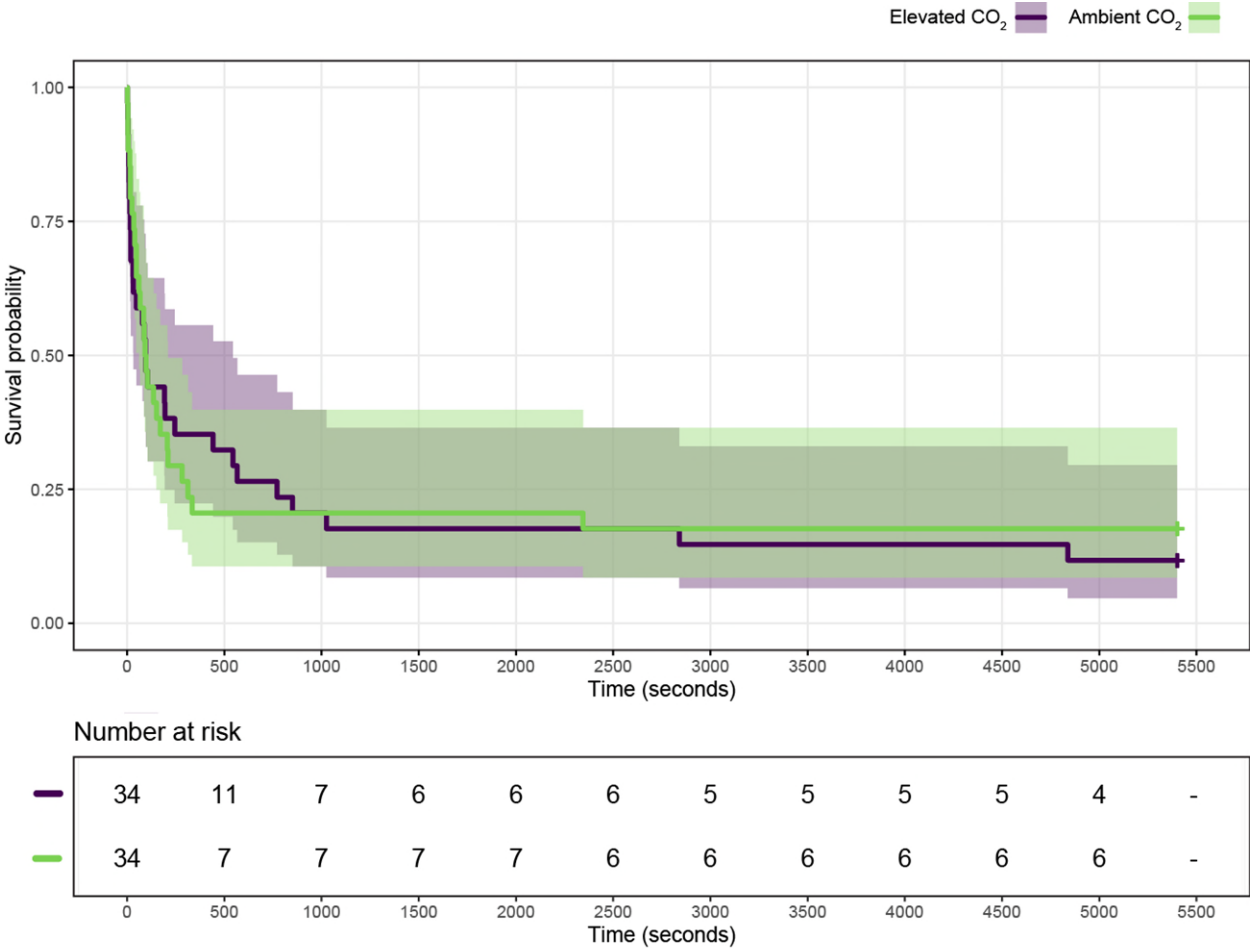


49 **Figure 3.** Intra-individual variation (residual variance for individual fish) in 5 behavioural
 50 measurements across 4 trials in Ambon damselfish (*P. amboinensis*) exposed to present-day levels
 51 of dissolved CO₂ (Ambient CO₂, n=36) and projected end-of-century CO₂-induced aquatic
 52 acidification (Elevated CO₂, n=37): (A) activity, (B) time sheltering, (C) thigmotaxis, (D)
 53 inspection of a novel object, and (E) time to emergence from the shelter. Fish in the control group
 54 were exposed to present-day CO₂ levels in all 4 trials, whereas fish in the CO₂ group were exposed
 55 to present-day CO₂ levels in trials 1 and 2, and projected end-of-century CO₂ levels in trials 3 and
 56 4.



57

Figure 4. Kaplan Meier survival curves for Ambon damselfish (*P. amboinensis*) exposed to present-day levels of dissolved CO₂ (Ambient CO₂, n=34) and projected end-of-century CO₂-induced aquatic acidification (Elevated CO₂, n=34). Fish were subjected to a 90 min predation trial involving a predator, the rock cod *Cephalopholis microprion*, exposed to the same conditions of water chemistry.

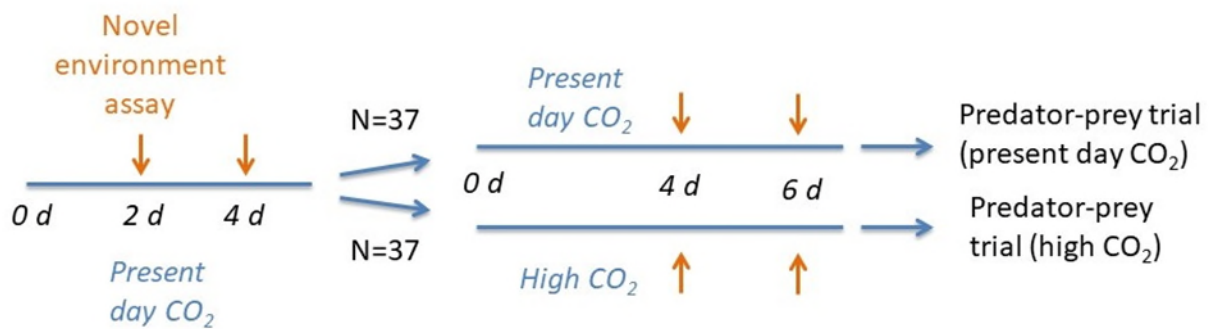


SUPPLEMENTARY MATERIAL

Table S1. Summary table with estimates, standard errors (SE), *Z* and *P* values for fixed effects included in a Poisson generalized linear mixed-effects model examining differences in number of predatory attacks that resulted in the capture of Ambon damsel (*Pomacentrus amboinensis*) in ambient CO₂ (N=28) and elevated CO₂ conditions (N=30). The sample size excludes Ambon damsel that were not captured. Predators (*Cephalopholis microprion*) were exposed to the same conditions of water chemistry as their prey. Predator identity was included as a random effect.

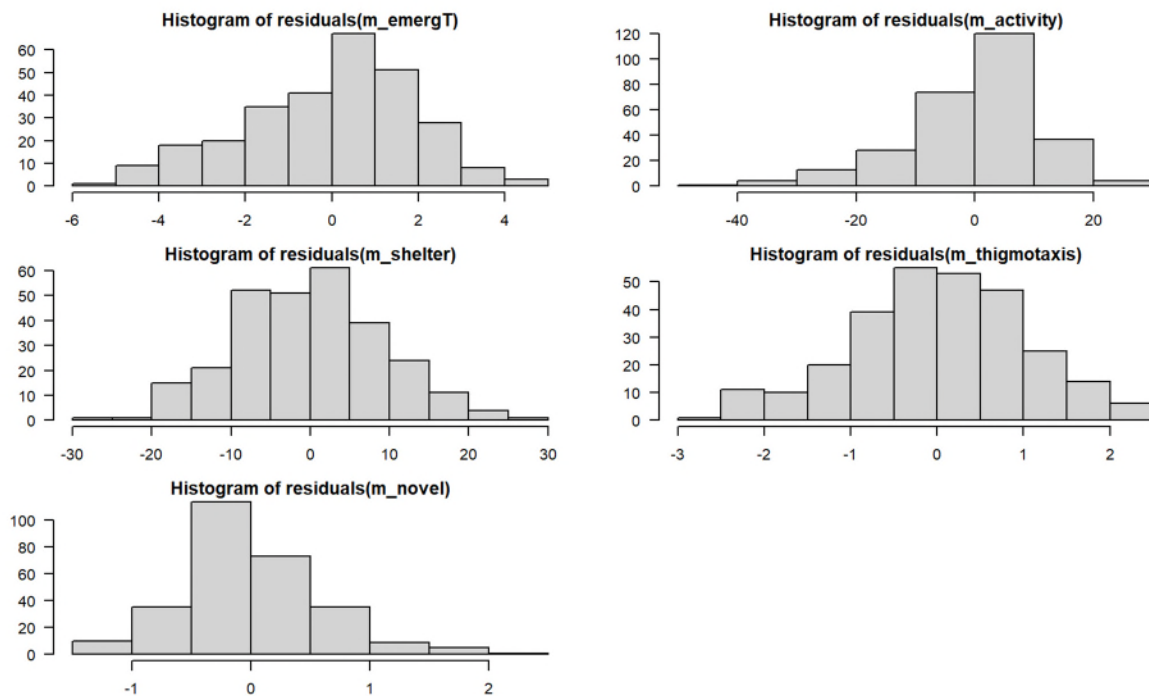
| Explanatory variable | Estimate | SE | <i>Z</i> | <i>P</i> |
|----------------------------------|----------|-------|----------|----------|
| Prey standard length | 0.147 | 0.067 | 2.210 | 0.027 |
| Mean emergence time | 0.017 | 0.075 | 0.223 | 0.823 |
| Mean time active | 0.168 | 0.108 | 1.555 | 0.120 |
| Mean time in/near shelter | 0.490 | 0.243 | 2.012 | 0.044 |
| Mean time displaying thigmotaxis | 0.260 | 0.146 | 1.785 | 0.074 |
| Mean time near novel object | 0.386 | 0.137 | 2.815 | 0.005 |
| Treatment_control | -0.040 | 0.182 | -0.219 | 0.827 |

Figure S1. Graphical representation of the experimental timeline used to examine the effect of exposure to end-of-century CO₂ levels on personality and susceptibility to predation in *Pomacentrus amboinensis*. Fish personality was assessed by repeatedly measuring behaviour in the same fish in a novel environment assay on specific days (orange arrows) during 4 days exposure to present day, ambient, CO₂ (control, n=74) followed the next day by treatment with either present day CO₂ or elevated CO₂ for 6 days (n=37 each). At the end of the CO₂ treatments, each fish was exposed to a predator-prey trial (using *Cephalopholis microprion* as predator) under the same CO₂ treatment.



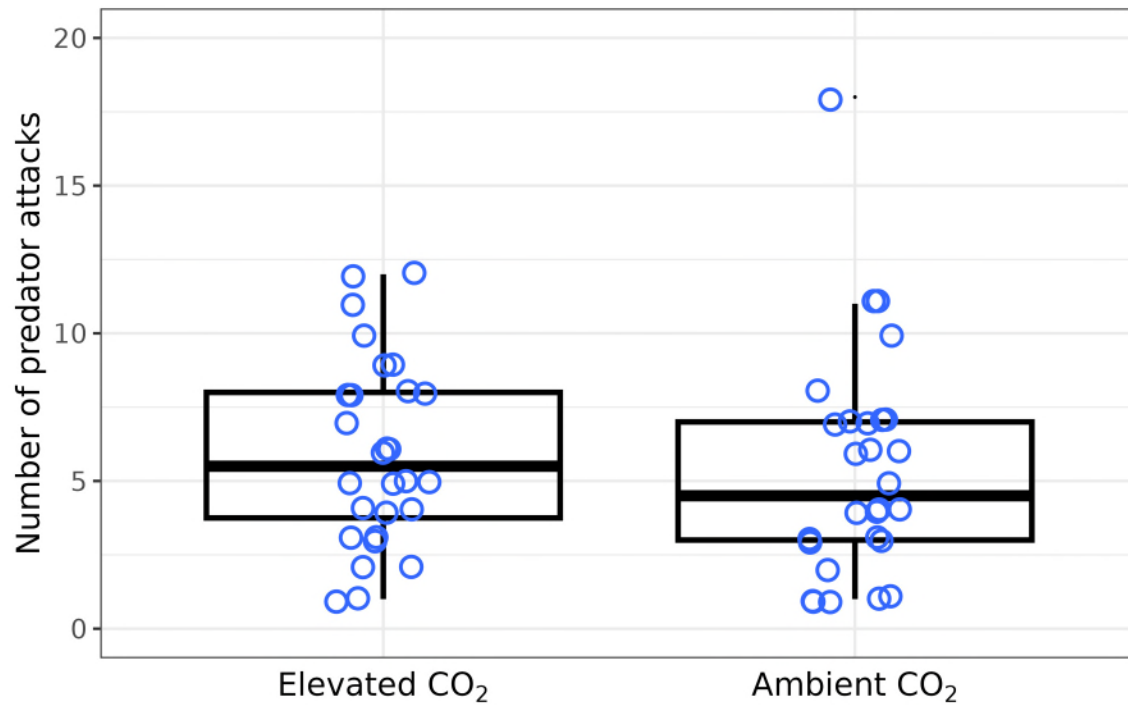
84 **Figure S2.** The distribution of model residuals (m_) after the following transformations: time to
85 emergence from the shelter (emergT) was log-transformed; activity levels (activity) and
86 thigmotaxis (thigmotaxis) were square-root-transformed; time spent sheltering (shelter) was not
87 transformed; and time near a novel (novel) object was log-transformed after adding 0.5.

88



89

90 **Figure S3.** The number of predatory attacks by dot-head rock cod (*Cephalopholis microprion*)
91 resulting in the capture of Ambon damsel (*Pomacentrus amboinensis*) in ambient CO₂ (N=28) and
92 elevated CO₂ (N=30) treatment groups. Ambon damsel that were not captured are not shown.



93